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PROCEEDINGS OF THE THIRTY-FIRST ANNUAL CON- VENTION OF THE ASSOCIATION OF OFFICIAL AGRICULTURAL CHEMISTS, 1914.

THIRD DAY.

WEDNESDAY—MORNING SESSION.

No report was made by the referee on the general subject of separation of nitrogenous substances.

REPORT ON SEPARATION OF NITROGENOUS SUBSTANCES (MILK AND CHEESE).

BY LEROY S. PALMER (Agricultural Experiment Station, Columbia, Mo.),
*Associate Referee.*¹

The referee on separation of nitrogenous substances (milk and cheese) was appointed in March to fill the vacancy due to the resignation of O. B. Winter, and there was not time for definite coöperative work before the close of this fiscal year.

No recommendations have been made since 1911 and, as far as could be learned by the associate referee, no work has been done along the line of the separation of the amino acids in cheese recommended at that time. It was decided, therefore, with the consent of the referee on the separation of nitrogenous substances, to lay plans for taking up work of a more general nature, where it was felt there was some need.

The associate referee has been impressed for some time with the need of a careful study of the determination of the so-called albumin of milk, leading if possible to the adoption of an accurate, reliable method, and, as associate referee, at once saw the opportunity of obtaining the coöperation of the association in this subject as no official method has ever been adopted. L. L. Van Slyke, who devised the present provisional method, as referee in 1908, recommended for study during the following year the selection of an official method for the determination of milk albumin. The recommendation either was not approved or did not come up in time for approval. Further evidence of need along this line is shown by the frequently recurring articles in the chemical journals, of improved methods

¹ Presented by P. F. Trowbridge.

for milk albumin. One which appeared during the past year¹ pointed out the varying results obtained by the provisional method of this association. To obtain further evidence a letter was sent to the chemists who had indicated their willingness to coöperate in the milk and cheese protein work containing the following inquiries:

1. Do you use the provisional method of the Association of Official Agricultural Chemists for milk albumin?

2. If so, do you find it entirely satisfactory?

3. If not, what modification do you use and what is its basis?

Three of the four coöperators replied as follows:

R. E. Stallings: We use the provisional method in this laboratory.

C. O. Swanson: We do not find the provisional method for milk albumin satisfactory because it is almost impossible to get constant results. The amount of albumin obtained depends to such a large extent upon conditions of temperature, time of heating, and the personal equation that it is very difficult to get comparative results. We have not worked out any modification of the method.

A. W. Bosworth: We use the provisional method for the determination of albumin in milk. The method originated in this laboratory and, although it is not entirely satisfactory, it seems to be the best one available.

It is now purposed to make some recommendations to the association in regard to milk albumin, but before doing so attention is called to a point in the nomenclature of the milk proteins, especially in regard to the so-called albumin. To any one familiar with the composition of milk it is obviously not strictly correct to call the heat-coagulable protein of milk, albumin, when there is recognized in milk the presence of a heat-coagulable globulin which is precipitated with the albumin in the present provisional method. It has always seemed that the more correct nomenclature of the milk proteins, as determined by the present methods, should contain the terms casein and heat-coagulable proteins (albumin and globulin) in place of the present terms casein and albumin.

If the association approves the recommendation to make studies leading to the selection of an official method for milk albumin, the question arises as to the definition of the term albumin, whether strictly as lact-albumin, or in the sense in which it is used at present, that is, heat-coagulable protein. The associate referee is inclined to think that a separate determination of albumin and globulin would not be wise, but that better success lies in the improvement of the present provisional method for the so-called albumin. Should the association agree, it is suggested that the matter of changing the nomenclature be taken up with the view of indicating the correct character of the protein obtained on coagulation with heat.

¹ *J. Ind. Eng. Chem.*, 1914, 6: 573.

Another matter concerning the milk proteins that is believed would be profitable for the association to consider is the selection of official methods for the determination of the protein substances in milk other than casein and the heat-coagulable albumin and globulin. Reference has not been made to all the association reports, but in 1901 Mr. Le Clere, referee on dairy products, called attention to the fact that the protein substances in milk, not precipitated by acetic acid or heat, amount in many cases to from 10 to 12 per cent of the total protein. Mr. Le Clere urged at that time that some attention be given to the determination of these bodies. The determination of these substances which we recognize as caseoses, albumoses, globuloses, peptones and other protein-like substances and nitrogenous extractives, is important not only because of their presence in fresh milk, but especially because of their increase in milk after it is drawn, due to the action of proteolytic enzymes. A study¹ of their relative proportion in milk of various states of freshness and under other conditions leads to important results from the standpoint of the nutritive value of the milk. While it would probably not be feasible for the food chemist to go into the separation of the individual members of this group of substances, it would be a very simple matter to separate them into two groups: namely, (1) the proteoses and peptones; and (2) the remaining nitrogenous substances, which might be called residual proteins. Such a separation is readily made by tannic acid. A number of analyses made in this manner have shown that tannic acid divides the non-casein non-heat-coagulable proteins into approximately equal parts, both of which are usually greatly increased in old milk.

In conclusion attention is called to the fact that studies in the preservation of milk for chemical analysis, which have long been of interest to this association, are often useless inasmuch as the methods for the analysis of the proteins remain unsatisfactory. It is urged, therefore, that the association take up these matters for study. The following recommendations are accordingly offered for approval.

RECOMMENDATIONS.

It is recommended—

(1) That the nomenclature of the so-called albumin of milk be changed to heat-coagulable proteins (albumin and globulin) in order that the true character of the protein thus obtained be shown.

(2) That studies be made leading to the adoption of an official method for the determination of the heat-coagulable proteins (albumin and globulin) of milk.

(3) That studies be made leading to the adoption of methods for the determination of the proteoses, peptones, and residual nitrogenous substances in milk.

¹ *J. Biol. Chem.*, 1913, 16: 331.

THE SO-CALLED NEUTRALIZATION PRECIPITATE OF COWS' MILK.

BY LEROY S. PALMER (Agricultural Experiment Station, Columbia, Mo.).

Van Slyke and Hart,¹ in 1903, called the attention of this association to the fact that the filtrate from the acetic acid precipitation of the casein of cows' milk (by the official association method) gives more or less of a precipitate on neutralization. They stated that this precipitate usually amounted to 0.1 per cent of casein. The explanation offered for this phenomenon was as follows: When cows' milk is treated with 1.5 per cent acetic acid, as in the official method, the precipitate formed is wholly casein-diacetate. A small portion of this is dissolved by the excess acetic acid used and reappears when its solution, the filtrate from the casein precipitation, is neutralized. Whether this neutralization precipitate is in the form of casein or its acetic acid compound was not stated.

Later work by Van Slyke and Van Slyke² on the action of dilute acids upon casein, would indicate that this explanation requires some modification. These investigators definitely showed that casein does form acetic acid, as well as other acid compounds, which are somewhat soluble in an excess of the acid used. That these acid compounds are of a definite character, however, was not borne out by the investigation. They appeared to be adsorption compounds rather than chemical combinations. It was also shown that the acid-casein compounds, when in solution, give up their acid on neutralization. It would appear from this that the neutralization precipitate, if it arises from an acid-soluble casein-acetate, should be free from acetic acid.

Interest in the so-called neutralization precipitate grew out of its invariable appearance, in greater or less amount, in some studies of new methods for the determination of the heat-coagulable proteins of cows' milk, in all of which the casein was first removed by acetic acid. It seemed possible that the protein character of the neutralization precipitate, as indicated by the previously mentioned work, might have a bearing on the determination of the casein or the heat-coagulable proteins.

Van Slyke and Van Slyke showed that the precipitate, obtained on careful neutralization of solutions of casein-acid compounds, had all the properties of casein, being readily soluble in an excess of alkali, and reappearing again on neutralization. They also found that the precipitation of the casein on addition of alkali was practically complete before the neutral point was reached, but could be prevented entirely if the alkali was added rapidly.

¹ Bur. Chem. Bull. 81, p. 91.

² *Am. Chem. J.*, 1907, 38: 383.

A brief study of the properties of the neutralization precipitate obtained on addition of sodium hydroxid to the casein filtrate showed that it also possessed the property of being precipitated before neutrality was reached, and of dissolving again on addition of acid. It was not found possible, however, to bring about a solution of the precipitate in sodium hydroxid. This insolubility in sodium hydroxid did not appear to be affected in any way either (a) by the manner of adding the alkali, whether rapidly or drop by drop, or (b) by the amount of sodium hydroxid added, or (c) by the temperature of the solution. In fact practically no solution occurred even on long boiling in sodium hydroxid solution. This treatment, however, resulted in rendering some of the precipitate insoluble in acetic acid.

The question at once arose, is the so-called neutralization precipitate casein?

As a preliminary investigation, casein was prepared from milk by the addition of acetic acid, according to the method used by Van Slyke and Van Slyke. Solutions of this casein and also of Merck's c. p. casein (according to Hammarsten) were made in sodium hydroxid and in glacial acetic acid. These solutions were then neutralized with acetic acid and sodium hydroxid, respectively, and the casein precipitated from the neutral solutions by acetic acid under conditions as nearly identical as possible with those which apparently cause the formation of the neutralization precipitate in the case of milk. The filtrates were then examined for neutralization precipitate. None was found in any case.

This result also throws great doubt upon the supposition that the neutralization precipitate is casein. On the other hand, the properties of the neutralization precipitate already described were so suggestive of tricalcium phosphate that a calcium and phosphorus analysis seemed warranted. For this analysis the neutralization precipitate was isolated from one liter of skim milk as follows:

One liter of fresh skim milk was diluted with one-half liter of water, and 150 cc. of 10 per cent acetic acid added, the diluted milk being at a temperature of 40°C. After cooling, the casein was filtered off. Several filtrations were required to get a clear filtrate. Sodium hydroxid solution (10 to 20 per cent) was now added to the filtrate to a faint alkaline reaction (to phenolphthalein). A heavy flocculent precipitate appeared. It was filtered off, washed somewhat with water, and then suspended in water to which 40 cc. of 10 per cent acetic acid had been added. All but traces of the precipitate dissolved readily in this solution. The acid solution was now placed on the steam-bath for an hour and then boiled for several minutes. The coagulated proteins thus formed were removed by filtration. The filtrate was treated with 10 to 20 per cent sodium hydroxid solution to faint alkalinity. A heavy flocculent precipitate came down, most of it before the neutral point was reached. The neutralization precipitate was filtered off, washed thoroughly on the paper, and then suspended in water and dissolved by the addition of dilute

acetic acid. Sodium hydroxid was added to this solution, bringing about reprecipitation. This precipitate was washed many times by decantation. It was finally dried on a porous plate in the electric oven at 105°C., yielding a fine, pure white powder, which was dried to constant weight at 105°C. before analysis. About 1.8 grams of dry material were obtained, no allowance being made for losses in isolation.

The results of the analysis of the neutralization precipitate are here given. On ignition at red heat it gave off water and blackened. After burning off the carbonaceous matter the ash was analyzed for calcium and phosphorus.

	<i>Per cent</i>
Total loss on ignition.....	16.92
Water lost on ignition.....	6.95
Nitrogen in dry substance.....	0.46
Calcium in ash.....	36.16
Phosphorus in ash.....	21.12

The results were found rather difficult to interpret because of the large loss on ignition. A calculation based on the results obtained, however, shows that a mixture of 72.5 per cent of $\text{Ca}_3(\text{PO}_4)_2$, 24.5 per cent of CaHPO_4 , 2 per cent of H_2O and 2.8 per cent of protein (casein or albumin) would give results conforming very closely to those obtained. The relation between the theoretical analysis of such a mixture and the results found is indicated in the following table:

	<i>Theoretical mixture per cent</i>	<i>Found for neutralization precipitate per cent</i>
Calcium.....	33.55	32.60
Phosphorus.....	19.55	19.10
Water lost on ignition.....	6.57	6.57
Nitrogen.....	0.46	0.46

If the protein in the precipitate be considered as casein, the relation between the theoretical results obtained from the ash of such a mixture and the results obtained for the neutralization precipitate would be those given in the following statement:

ASH ANALYSIS.

	<i>Theoretical mixture per cent</i>	<i>Neutralization precipitate per cent</i>
Calcium.....	36.18	38.45
Phosphorus.....	21.13	22.58

The greater difference in this case is due to the unexplained loss on ignition.

These results make untenable the previously advanced explanation of the origin and character of the so-called neutralization precipitate of cows' milk. Instead of being a portion of the casein which has been dissolved by the excess of acid used, it appears to be almost entirely a mixture of di- and tricalcium phosphates. Whether the protein obtained in the analyses is a contamination or an essential part of the precipitate is not apparent.

It may be stated, however, that the method of isolation would seem to preclude a protein contamination, which is also borne out by the ready solubility of the isolated precipitate in acetic acid.

The source of the neutralization precipitate is a question of considerable interest, especially since some evidence can be presented in regard to it. Lindet¹ has recently shown that casein, which has been precipitated from cows' milk by rennet, will yield CaO and P_2O_5 on treatment with dilute acetic acid, equivalent to 3.45 per cent $\text{Ca}_3(\text{PO}_4)_2$ and an additional 1.84 per cent CaO . Since this result is obtained merely by washing with dilute acid, it would seem that the explanation of the origin of the calcium phosphate of the neutralization precipitate is very simple. It is possible, of course, that some of the calcium phosphate, especially the dicalcium phosphate, may result from preëxisting phosphates in the milk serum. In view of Lindet's results and the fact that it was not found possible to bring about a further liberation of calcium phosphate (neutralization precipitate) from casein which had been thrown down by acetic acid, it would seem to be a very easy matter to demonstrate whether the source of the neutralization precipitate is the caseinogen of the milk or preëxisting phosphates of the serum, or whether it arises from both of these sources. According to Lindet's results, casein precipitated by rennet should yield a neutralization precipitate after treatment with acetic acid; and if this is the only source of the neutralization precipitate, the filtrate from the rennet coagulation should yield no neutralization precipitate. In the acetic acid coagulation of casein, the filtrate from the coagulation of the albumin and globulin by heat will usually show the neutralization precipitate as readily as the filtrate from the casein coagulation.

No difficulty was experienced in showing the absence of neutralization precipitate in the filtrate from the rennet coagulation of casein, either before or after coagulation of the albumin and globulin with heat. On the other hand, neither was it found possible to show that rennet-coagulated casein will yield a neutralization precipitate after treatment with acetic acid. In this study practically the same procedure was followed as in the previous study of the acid-coagulated casein, with the exception that ammonium hydroxid² was used for the alkaline solvent in place of sodium hydroxid. Calcium phosphate being somewhat soluble in ammonium acetate solution, it was necessary in this case to remove the ammonia by distillation in the presence of fixed alkali, before testing for the neutralization precipitate.

The explanation of this result is not apparent. Possibly the conditions were not right, or the calcium phosphates obtained in the usual

¹ Proc. 8th Intern. Cong. Appl. Chem., 1912, 19: 199.

² Lindet states that rennet-coagulated casein dissolves in ammonium hydroxid without liberation of calcium phosphate.

neutralization precipitate are the result of a chemical reaction and not of a mere washing with dilute acetic acid. Even then, it is not apparent why a repetition of Lindet's work did not yield a precipitate of calcium phosphate on neutralization.

CONCLUSIONS.

(1) The so-called neutralization precipitate obtained from the filtrate of the acetic acid coagulation of cows' milk appears to be largely a mixture of di- and tricalcium phosphates. The precipitate isolated in this study was mixed with a small proportion of protein, probably casein or albumin, either as a contamination or as an essential part of the precipitate.

(2) The origin of the calcium phosphates of the neutralization precipitate would presumably be largely the caseinogen of the milk, but attempts to show this fact, using casein coagulated by rennet, have so far been unsuccessful.

(3) The very small amount of protein contained in the neutralization precipitate precludes its being an important factor in the determination of casein or the heat-coagulable proteins.

REPORT ON DAIRY PRODUCTS.

BY LEWIS I. NURENBERG (State Board of Health, Boston, Mass.), *Referee*.¹

The following recommendations were made for collaborative work:

(1) That the copper sulphate method of preparing milk serum be studied with a view of its adoption as an optional provisional method.

(2) That the Harding-Parkin method for fat determination be studied in comparison with the present official and provisional methods.

(3) That in U. S. Bur. Chem. Bull. 107, Rev., p. 122, after the paragraph on Cream, the following paragraph be inserted under the heading "Condensed Milk (Unsweetened)": "Dilute 40 grams of the homogeneous material with 60 grams of distilled water, and proceed as directed under 'Milk', and correct the results for dilution"; and that the word "Sweetened" be inserted before the word "Condensed" in the subsequent heading.

(4) That subjects now under consideration be given further study.

No work on the copper sulphate method has been reported to the association since 1909. In the study of the copper sulphate method it was the referee's idea to collect and compile figures from all available sources. Twenty-two letters were sent out to the various food and dairy departments, asking for figures on known purity milk by the copper refraction

¹ Presented by H. C. Lythgoe.

method, together with figures on solids, fat, etc., on the same milk, and if the milk was from individual cows, the name of breed and amount of milk given at the time of milking; if from a herd, the number and breed of cows and the amount of milk produced at the time of milking. Figures were also requested on the comparison of the copper and acetic methods for the preparation of milk serum, comparison of the copper method on night's and morning's milk, and on the different stages of lactation. Eleven replies were received, two of which contained some of the desired figures. One reply was from H. Louis Jackson of the Idaho Board of Health, and the other was from E. M. Bailey of the Connecticut Agricultural Experiment Station. The referee has obtained figures on 834 samples of known purity milk, examined during the past eight years in the laboratory of food and drug inspection of the Massachusetts Board of Health, and, unless otherwise stated, all analyses are from that laboratory.

In 698 cases where the samples came from individual cows the breed was known and is distributed as follows: Jersey 47; Guernsey 38; Grade Jersey 45; Grade Guernsey 26; Grade Durham 27; Grade Ayrshire 54; Swiss 12; Grade Holstein 198; other grades 13; Ayrshire 48; Dutch Belt 51; and Holstein 139. A little more than one-half of the total number, therefore, represent low grade milk. The samples were collected during different seasons of the year and during varying periods of lactation, so that it is proper to consider them as fairly representative of all influencing conditions.

PREPARATION OF THE COPPER SERUM.¹

Dissolve 72.5 grams of copper sulphate in water and make up to 1 liter. Adjust this solution to read 36 at 20°C. on the scale of the Zeiss immersion refractometer. Add 4 volumes of milk to 1 of the copper sulphate solution, shake well and filter. Determine the index of refraction of the filtrate at 20°C. A reading below 36 is indicative of added water.

Table 1 presents the refractions of the copper sulphate serums on 834 samples of known purity milk. The variation in readings extends from 36–40.4. The largest number of samples, 16.7 per cent, refracts between 37.8 and 38.0, only 3.1 per cent from 36–36.2 and practically none above 40. The range of readings is, therefore, small, much less than that of the acetic serum which extends from 39–46.

¹ Mass. State Board of Health Rept., 1908, p. 594; U. S. Bur. Chem. Bull. 132, p. 122.

TABLE 1.

Refractions of copper sulphate serum on 834¹ samples of milk of known purity.

READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES
36.0	14	37.5	27	39.0	28
36.1	9	37.6	36	39.1	4
36.2	3	37.7	38	39.2	8
36.3	8	37.8	28	39.3	8
36.4	10	37.9	39	39.4	5
36.5	15	38.0	72	39.5	5
36.6	19	38.1	32	39.6	1
36.7	10	38.2	41	39.7	7
36.8	22	38.3	40	39.8	1
36.9	13	38.4	33	39.9	..
37.0	24	38.5	33	40.0	1
37.1	18	38.6	23	40.1	2
37.2	28	38.7	24	40.2	..
37.3	28	38.8	22	40.3	1
37.4	33	38.9	20	40.4	1

¹ This figure includes 53 samples from H. Louis Jackson, State board of health, Boise, Idaho, 53 from W. G. Tice, State board of health, Trenton, N. J. (U. S. Bur. Chem. Bull., 132, p. 130) and 14 from E. M. Bailey, agricultural experiment station, New Haven, Conn.

TABLE 2.

Composition of milk of known purity.

(H. Louis Jackson, Idaho.)

SPECIFIC GRAVITY AT 15.5°C. (WESTPHAL)	FAT BY BABCOCK METHOD (TESTED BOTTLES)	TOTAL SOLIDS ¹	SOLIDS NOT FAT	CORRECTED COPPER SERUM READING AT 20°C.
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	
1.0307	4.1	12.61	8.51	37.2
1.0313	4.5	13.24	8.74	38.1
1.0314	4.2	12.90	8.70	38.0
1.0314	6.5	15.66	9.16	37.9
1.0296	5.0	13.41	8.41	38.5
1.0327	7.9	17.67	9.77	38.8
1.0293	4.5	12.74	8.24	37.4
1.0318	5.5	14.57	9.07	38.7
1.0324	5.5	14.72	9.22	38.4
1.0297	3.6	11.76	8.16	37.2
1.0316	4.5	13.31	8.81	37.9
1.0312	5.1	13.94	8.84	37.6
1.0324	4.1	13.03	8.93	38.2

¹ Leach. Food Inspection and Analysis. 1913, 3rd ed., p. 151.

TABLE 3.
Composition of milk of known purity.
(H. Louis Jackson, Idaho.)

FAT BY BABCOCK METHOD (TESTED BOTTLES)	TOTAL SOLIDS (5 GRAMS FOR 3 HOURS ON THE WATER-BATH)	* SOLIDS NOT FAT	CORRECTED COPPER SERUM READING AT 20°C.
<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	
3.80	12.57	8.77	38.9
3.55	11.39	7.84	38.7
3.35	12.41	9.06	38.5
3.25	11.77	8.52	38.4
3.40	11.36	7.96	36.3
3.30	11.73	8.43	38.5
3.60	11.58	7.98
4.60	13.52	8.92	38.0
4.40	13.31	8.91	38.0
3.40	11.52	8.12
3.80	12.76	8.96	38.0
3.95	12.60	8.65	38.2
3.70	12.80	9.10	38.3
6.40	15.25	8.85	39.5
3.50	12.29	8.79	38.3
3.50	12.23	8.73	38.0
3.30	11.87	8.57	37.5
3.80	12.61	8.81	37.8
0.88 ¹	9.34	8.46	36.5
3.60 ²	12.71	9.11	38.5
10.00 ³	18.53	8.53	38.3
3.85 ⁴	12.38	8.53	37.8
3.35 ⁵	11.60	8.25	37.7
3.95 ⁶	12.90	8.95	38.0
4.00 ⁷	12.86	8.86	37.7

¹ Fore milk. Just after milking began.² Drawn at the middle of the milking.³ Strippings.⁴ From first 10 gallon can filled.⁵ From next 5 gallon can filled.⁶ From next 5 gallon can filled.⁷ From next 5 gallon can three-fourths filled.

TABLE 4.
Composition of milk of practically known purity.
(H. Louis Jackson, Idaho.)

SPECIFIC GRAVITY AT 15° C. 15° (WESTPHAL)	FAT BY BABCOCK METHOD (TESTED BOTTLES)	TOTAL SOLIDS BY CALCULATION	SOLIDS NOT FAT	CORRECTED COPPER SERUM READING AT 20°C.	REMARKS
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>		
1.0335	3.9	12.06	8.16	38.4	
.....	4.6	38.6	Night's milk.
.....	5.6	38.75	Morning's milk.
.....	4.28	38.2	Morning's milk.
.....	6.4	38.9	Morning's milk.
.....	3.3	38.96	Morning's milk.
.....	4.6	38.0	Morning's milk.
.....	5.1	40.3	Morning's milk.
.....	4.8	37.0	Morning's milk.
.....	6.0	40.1	
.....	4.25	38.6	Composite of 4 cows. ¹

¹ No. 1, $\frac{1}{2}$ Black Jersey $\frac{1}{2}$ Holstein, 1 gallon of milk; No. 2, full blood Black Jersey, 2 gallons of milk; No. 3, $\frac{1}{2}$ Short Horn $\frac{1}{2}$ Durham, $\frac{1}{2}$ gallon of milk; No. 4, $\frac{1}{2}$ Black Jersey $\frac{1}{2}$ Short Horn, 3 gallons of milk.

Milk from pure Guernsey from registered stock (age 10-11 years).

PART OF UDDER	FAT BY BAB- COCK METHOD (TESTED BOTTLES)	TOTAL SOLIDS (5 GRAMS FOR 3 HOURS (N WATER-BATH)	SOLIDS NOT FAT	CORRECTED COPPER SERUM READING AT 20°C.
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	
Left hind teat.....	6.13	14.88	8.75	37.62
Left front teat.....	6.3	14.96	8.66	37.79
Right front teat.....	4.15	13.45	9.30	38.36
Right hind teat.....	8.23	17.24	9.01	37.95

In the preceding tables it will be noticed that the lowest copper refraction from the milk of individual cows is 36.3, the highest 40.1, while the herd milk refracts at practically 38.0.

E. M. Bailey of New Haven sent in copper refraction figures on the milk of 14 herds of Holstein and Grade Guernsey cows. The range of the copper readings extended from 36.8-39.4.

The copper serum method besides having a narrow range of readings has several other advantages; it is simple, the serum is very quickly prepared without the use of heat, no loss by evaporation and the determination may be made on a small quantity of milk, sufficient serum being obtained with 17.6 cc. of milk and 4.4 cc. of copper sulphate solution.

PREPARATION OF ACETIC SERUM.¹

The refractions of the acetic serums on 554 samples of known purity milk are shown in Table 5. From this table it will be noted that the greatest percentage of samples refracts between 43 and 43.5, practically none below 40, and from 4-5 per cent about 45.

During the past five years in the laboratory of the Massachusetts Board of Health the refractive index has been determined on the sour serum of milk.

PREPARATION OF SOUR SERUM.²

Allow the milk to sour spontaneously, filter and determine the index of refraction of the clear serum at 20°C. by means of the Zeiss immersion refractometer. Refractive indices have been made on 515 samples of known purity milk and are given in Table 6. About 50 per cent of these samples refract between 41.0 and 43.0.

The number of refractions between 40.0 and 41.0 and between 43.0 and 44.0 is nearly the same, that is, 15 per cent. Five per cent of the samples are between 38.0 and 39.0 and 1 per cent is above 45.0. This method has been found especially useful in cases where a sample of milk presents the least sign of souring.

¹ Provisional method. U. S. Bur. Chem. Bull. 107, Rev., p. 120.

² *Z. offent. Chem.*, 1903, 9: 173.

TABLE 5.

Refraction of acetic acid serum on 557¹ samples of milk of known purity.

READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES
39.0	1	41.0	13	43.0	42	45.0	4
39.1	..	41.1	4	43.1	14	45.1	2
39.2	1	41.2	4	43.2	13	45.2	4
39.3	2	41.3	9	43.	12	45.3	1
39.4	1	41.4	6	43.4	14	45.4	4
39.5	..	41.5	11	43.5	12	45.5	2
39.6	2	41.6	13	43.6	13	45.6	1
39.7	1	41.7	14	43.7	10	45.7	1
39.8	1	41.8	17	43.8	10	45.8	3
39.9	2	41.9	2	43.9	9	45.9	5
40.0	5	42.0	17	44.0	11		
40.1	..	42.1	13	44.1	13		
40.2	4	42.2	15	44.2	9		
40.3	3	42.3	20	44.3	7		
40.4	7	42.4	11	44.4	7		
40.5	5	42.5	16	44.5	11		
40.6	4	42.6	14	44.6	9		
40.7	5	42.7	9	44.7	4		
40.8	3	42.8	21	44.8	5		
40.9	5	42.9	19	44.9	5		

¹ This figure includes 49 samples from W. G. Tice, State board of health, Trenton, N. J. (U. S. Bur. Chem. Bull., 132, p. 130).

TABLE 6.

Refractions of sour serum on 504 samples of milk of known purity.

READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES	READING	NUMBER OF SAMPLES
38.3	4	40.3	11	42.3	9	44.3	1
38.4	2	40.4	9	42.4	19	44.4	3
38.5	2	40.5	6	42.5	6	44.5	3
38.6	3	40.6	8	42.6	7	44.6	1
38.7	2	40.7	9	42.7	10	44.7	2
38.8	2	40.8	10	42.8	10	44.8	1
38.9	1	40.9	6	42.9	5	44.9	2
39.0	9	41.0	18	43.0	14	45.0	2
39.1	5	41.1	10	43.1	8	45.1	1
39.2	6	41.2	11	43.2	8	45.2	1
39.3	2	41.3	15	43.3	5	45.3	..
39.4	9	41.4	10	43.4	7	45.4	..
39.5	7	41.5	17	43.5	10	45.5	1
39.6	2	41.6	10	43.6	5	45.6	..
39.7	4	41.7	13	43.7	11	45.7	1
39.8	5	41.8	13	43.8	4	45.8	..
39.9	8	41.9	11	43.9	3	45.9	..
40.0	11	42.0	15	44.0	4		
40.1	3	42.1	18	44.1	2		
40.2	8	42.2	19	44.2	4		

TABLE 7.
Relation between refractions of copper sulphate and acetic acid serum on 325 samples of milk of known purity.

REFRACTION OF ACETIC SERUM	REFRACTION OF COPPER SERUM													PER CENT OF SAMPLES	
	Number of samples														
	39.0-39.2	39.3-39.5	39.6-39.8	39.9-37.1	37.2-37.4	37.5-37.7	37.8-33.0	38.1-38.3	38.4-38.6	38.7-38.9	39.0-39.2	39.3-39.5	39.6-39.8		Totals
Reading at 20°C.	1 ¹	..	1	2	0.6
39.0-39.5	3	..	1	4	1.2
39.6-40.1	1	6	3	3	2	15	4.6
40.2-40.7	..	5	5	10	6	1	27	8.3
40.8-41.3	3	4	9	9	5	..	1	1	34	10.5
41.4-41.9	3	1	6	10	22	14	2	51	15.7
42.0-42.5	3	5	10	25	17	8	2	2	69	21.3
42.6-43.1	..	2	1	6	10	11	4	8	42	12.9
43.2-43.7	1	4	5	9	9	6	4	1	..	39	12.0
43.8-44.3	1	1	..	2	4	7	6	5	1	..	27	8.3
44.4-44.9	1	..	1	5	3	2	1	13	4.0
45.0-45.5	1	..	1	5	2	0.6
45.6-46.1	1	2	0.6
Totals.....	5	13	16	22	31	35	66	46	38	23	23	4	3	325	
Per cent of samples.....	1.5	4.0	4.9	6.8	9.5	10.8	20.3	14.2	11.7	7.1	7.1	1.2	0.9	100	

¹ Lowest sample: copper serum 36.1; acetic serum 39.0.

Tables 7, 8 and 9, present the relations existing between (a) the copper and acetic serum, (b) the copper and sour serum, and (c) the acetic and sour serum. In Table 7 the range of the acetic refractions is twice as large as that of the copper refraction, so that the ratio of 3 copper refractions to the copper limit is the same as 6 acetic refractions to the acetic limits. The table has, therefore, been arranged in that manner. In Table 8 the ratio is 4 copper refractions to 9 sour serum refractions, while in Table 9, 6 sour serum refractions equal 7 acetic refractions.

In Table 7, 20 per cent of the copper samples refracted between 37.8 and 38.0 and 21 per cent of the acetic between 42.6 and 43.1. Twenty-five samples, or 7.7 per cent of the total samples, had both copper and acetic refractions within the above-stated limits. From this point the relationship between the 2 serums is fairly uniform.

TABLE 8.

Relation between refractions of copper and sour serum on 364 samples of milk of known purity.

REFRACTION OF SOUR SERUM	REFRACTION OF COPPER SERUM										PER CENT OF SAMPLES	
	36.0- 36.3	36.4- 36.7	36.8- 37.1	37.2- 37.5	37.6- 37.9	38.0- 38.3	38.4- 38.7	38.8- 39.1	39.2- 39.5	39.6- 39.9		Totals
Reading at 20°C.												
	Number of samples											
37.9-38.7	4 ¹	4	1	8	2.2
38.8-39.6	5	13	6	3	1	28	7.7
39.7-40.5	6	8	20	6	4	2	46	12.6
40.6-41.4	4	2	7	23	18	11	4	2	71	19.5
41.5-42.3	..	2	4	11	21	32	19	7	96	26.4
42.4-43.2	1	3	9	20	17	13	4	..	67	18.4
43.3-44.1	1	..	1	7	11	5	4	2	31	8.5
44.2-45.0	..	1	1	2	..	6	1	2	13	3.6
45.1-45.9	1	1	2	4	1.1
Totals.....	20	30	40	46	54	75	51	33	9	6	364	
Per cent of samples.....	5.5	8.2	11.0	12.7	14.8	20.5	14.0	9.1	2.5	1.6		100

¹ Lowest sample: sour serum 38.3; copper serum 36.0.

In Table 8, 20 per cent of the copper refractions were between 38.0 and 38.3, and 26 per cent of the sour refractions between 41.5 and 42.3; 32 samples, or 8.7 per cent of the total, gave copper and sour serum readings between the above-stated limits. The relationship here is not so uniform as in the case of the copper and acetic comparison, but this is due, undoubtedly, to the unusual forms of fermentation occurring during the souring of the milk.

In Table 9, 19.8 per cent of the acetic refractions were between 43.0 and 43.5; 22.1 per cent of the sour refractions between 41.5 and 42.1; 10 samples, or 6 per cent of the total, gave sour and acetic readings between

TABLE 9.

Relation between refractions of acetic and sour serums on 167 samples of milk of known purity.

REFRACTION OF sour serum	REFRACTION OF ACETIC SERUM										TOTALS	PER CENT OF SAMPLES
Reading at 20°C.	40.0- 40.5	40.6- 41.1	41.2- 41.7	41.8- 42.3	42.4- 42.9	43.0- 43.5	43.6- 44.1	44.2- 44.7	44.8- 45.3	45.4- 45.9		
	Number of samples											
38.0-38.6	1 ¹	1	2	1.2
38.7-39.3	1	...	1	2	1	5	3.0
39.4-40.0	1	1	3	...	1	6	3.6
40.1-40.7	5	1	6	6	1	2	1	22	13.2
40.8-41.4	...	3	4	4	6	4	1	2	24	14.4
41.5-42.1	...	1	4	13	7	7	3	...	2	...	37	22.1
42.2-42.8	6	5	10	4	4	29	17.3
42.9-43.5	4	3	7	4	2	2	22	13.2
43.6-44.2	1	2	1	3	3	...	2	1	13	7.8
44.3-44.9	1	...	1	1	1	1	2	7	4.2
Totals.....	8	6	19	32	25	33	20	11	7	6	167
Per cent of samples.....	4.8	3.6	11.3	19.1	15.0	19.8	12.0	6.6	4.2	3.6	...	100

¹ Lowest sample: sour serum 38.3; acetic serum 40.1.

the above-stated limits. In general, the sour serum refracts lower than the acetic. The largest number of samples (13 or 7.8 per cent) with acetic refractions of 41.8 and 42.3 and sour serum refractions of 41.5 to 42.1 would, without doubt, not occur at this point if a larger number of samples had been examined.

During the past three years determinations have been made of the ash of the sour serum.

Allow the milk to sour spontaneously, pipette 25 cc. of the clear serum into a flat-bottomed platinum dish and evaporate to dryness over the water-bath. Heat cautiously the contents of the dish over a small flame (to avoid sputtering) until charred. Perform the final heating in the electric muffle (with a pyrometer connected) at a heat not greater than 500°C. or 900°F. A white ash is almost invariably obtained after once leaching.

In the original article as described by A. Burr and F. M. Berberich,¹ the authors have outlined the following method:

Measure 50 cc. of the serum into a platinum dish, evaporate to dryness and carbonize over a low flame. Extract the char with hot water, burn the insoluble residue, add the solution to this ash, evaporate to dryness, ignite at a low temperature and weigh.

Experience has shown, however, that with the exercise of care and an accurate pyrometer the former stated method is as accurate as the latter and involves less manipulation.

¹ *Chem. Ztg.*, 1908, **32**: 617.

Table 10 gives the ash of the sour serum, expressed in grams per 100 cc., of 371 samples of known purity milk.

TABLE 10.
Ash of the sour serum on 371 samples of milk of known purity.
(Expressed in grams per 100 cc.)

ASH	NUM- BER OF SAMP- LES	ASH	NUM- BER OF SAMP- LES	ASH	NUM- BER OF SAMP- LES	ASH	NUM- BER OF SAMP- LES
0.730-0.734	22	0.790-0.794	20	0.850-0.854	2	0.910-0.914	..
0.735-0.739	8	0.795-0.799	16	0.855-0.859	4	0.915-0.919	..
0.740-0.744	26	0.800-0.804	22	0.860-0.864	6	0.920-0.924	..
0.745-0.749	12	0.805-0.809	13	0.865-0.869	6	0.925-0.929	..
0.750-0.754	14	0.810-0.814	19	0.870-0.874	3	0.930-0.934	1
0.755-0.759	10	0.815-0.819	8	0.875-0.879	1	0.935-0.939	..
0.760-0.764	26	0.820-0.824	17	0.880-0.884	2	0.940-0.944	2
0.765-0.769	11	0.825-0.829	4	0.885-0.889	3		
0.770-0.774	16	0.830-0.834	4	0.890-0.894	1		
0.775-0.779	9	0.835-0.839	2	0.895-0.899	1		
0.780-0.784	28	0.840-0.844	5	0.900-0.904	2		
0.785-0.789	17	0.845-0.849	7	0.905-0.909	1		

As will be noticed, practically the same percentage of samples, 18 per cent, was found with ashes between the following limits: 0.730 and 0.749; 0.750 and 0.769; 0.770 and 0.789; 0.790 and 0.810; or, in 72 per cent of the cases the limits of the ash extended from 0.730-0.810. Thirteen per cent of the samples had ashes from 0.810-0.830. Five per cent of the ashes were from 0.830-0.850 and 5 per cent from 0.850-0.870, making 10 per cent between 0.830 and 0.870. The remaining 5 per cent gave figures above 0.830 which were distributed as extensively as the first 72 per cent.

TABLE 11.
Composition of 67 samples of herd milk.
(Arranged in order of Total Solids.)

TOTAL SOLIDS	FAT	SOLIDS NOT FAT	COPPER REFRACTION	ACETIC REFRACTION	SOUR SERUM REFRACTION	SOUR SERUM ASH
<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>reading</i>	<i>reading</i>	<i>reading</i>	<i>grams per 100 cc.</i>
15.32	5.30	10.02	38.2	41.4
14.80	5.90	8.90	38.5	43.3	0.964
14.57	5.40	9.17	38.5	44.1
14.40	5.00	9.40	38.6	44.6
14.19	5.00	9.19	38.3	42.7	0.792
13.84	4.50	9.34	38.0	41.9	0.808
13.73	4.60	9.13	37.7	42.4	41.8	0.806
13.68	5.10	8.58	38.8	42.7	0.960
13.68	4.80	8.88	37.8	41.6	42.1	0.796
13.60	4.45	9.15	38.7	45.4

TABLE 11.—Continued.

TOTAL SOLIDS	FAT	SOLIDS NOT FAT	COPPER REFRACTION	ACETIC REFRACTION	SOUR SERUM REFRACTION	SOUR SERUM ASH
<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>reading</i>	<i>reading</i>	<i>reading</i>	<i>grams per 100 cc.</i>
13.48	4.40	9.08	38.0	42.3	41.9	0.788
13.48	4.10	9.38	37.8	42.3	42.4	0.810
13.46	4.50	8.96	38.0	42.4	42.0	0.809
13.45	4.35	9.10	43.6	42.9
13.44	4.10	9.34	38.2	43.4
13.40	4.20	9.20	38.5	42.2	0.780
13.35	4.00	9.35	44.3	42.8
13.34	4.30	9.04	38.3	42.2	0.800
13.32	5.00	8.32	38.0	43.5	0.800
13.28	4.50	8.78	38.2	43.1
13.27	4.40	8.87	37.7
13.08	4.75	8.33	37.7	42.2
13.08	4.30	8.78	38.2	42.1	0.740
13.08	4.10	8.98	37.7	41.3	0.788
13.06	4.05	9.01	38.1	42.3	42.6	0.812
12.97	4.40	8.57	38.0	42.3	0.760
12.96	4.00	8.96	42.1	41.0
12.95	4.00	8.95	37.7	42.0
12.92	4.20	8.72	38.0	42.0	41.6	0.796
12.88	4.00	8.88	37.5	41.4	0.771
12.80	3.80	9.00	37.9	42.3	0.776
12.76	4.40	8.36	42.5	41.5
12.76	3.80	8.96	37.8
12.73	3.80	8.93	38.0	41.4	0.800
12.72	3.90	8.82	38.8	43.0	0.792
12.67	3.90	8.77	43.5
12.64	3.60	9.04	38.2	43.0	0.800
12.62	3.85	8.77	37.2
12.59	3.70	8.89	38.3	41.4	0.768
12.58	3.60	8.98	38.0	42.0	0.812
12.57	4.00	8.57	38.1	43.1
12.57	4.00	8.57	37.2	41.5	0.766
12.56	4.20	8.36	37.9	43.1
12.55	3.65	8.90	43.4
12.53	3.70	8.83	37.7	41.5	0.808
12.52	4.00	8.52	37.2	41.0
12.49	3.65	8.84	43.0	42.3
12.45	3.60	8.85	38.3
12.44	3.70	8.74	38.0	42.0	0.750
12.36	3.90	8.46	38.0	40.2	40.3	0.794
12.26	3.70	8.56	37.5	40.7	0.820
12.25	3.70	8.55	42.9
12.24	3.70	8.54	41.3
12.22	3.50	8.72	43.9	42.6
12.18	3.80	8.38	37.9	42.0
12.07	3.40	8.67	37.1
12.03	3.60	8.43	38.5	42.6
12.02	3.40	8.62	37.7	41.8	41.9	0.764
12.00	3.90	8.10	36.7	40.0	0.788
11.96	3.35	8.61	37.2	42.2

TABLE 11.—Continued.

TOTAL SOLIDS	FAT	SOLIDS NOT FAT	COPPER REFRACTION	ACETIC REFRACTION	SOUR SERUM REFRACTION	SOUR SERUM ASH
<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>reading</i>	<i>reading</i>	<i>reading</i>	<i>grams per 100 cc.</i>
11.84	3.50	8.34	37.7
11.80	3.55	8.25	42.5	41.0
11.74	3.40	8.34	37.0	39.6	0.780
11.74	3.30	8.44	37.4	40.3	0.786
11.28	3.20	8.08	37.1	39.5	0.752
11.25	3.60	7.65	36.6
11.06	3.30	7.76	36.6
Total sam- ples 67	67	67	56	34	42	34
Maximum 15.32	5.90	10.02	38.8	45.4	43.5	0.964
Minimum 11.06	3.20	7.65	36.6	40.2	39.5	0.740
Average 12.83	4.06	8.77	37.9	42.7	41.7	0.798

The above 67 herds are representative of all the breeds mentioned on page 9 of this report.

Here, as would be expected in herd milk, the copper, acetic and sour serum refractions and the sour serum ash figures are higher than the minimum obtained from individual cows. The lowest figure approaching nearest to the minimum copper limit (36.0) is 36.6; to the minimum acetic limit (39.0) is 40.2; and to the sour serum limit (38.3) is 39.5; and the nearest approach to the sour serum ash limit (0.730) is 0.740. The sour serum ash figures are high, not only in the greater majority of cases, but particularly wherever there is a tendency for the copper, acetic or sour serums to refract lower than the average figures given above.

COMMENTS OF ANALYSTS ON THE COPPER METHOD.

A. G. Woodman: My work on milk has been almost entirely on isolated samples for educational purposes, and practically no samples of undoubted authenticity, so I can give you no figures that would be of use to you. Personally, I much prefer the copper method to the acetic acid as we get a clear serum with much less trouble, and I hope that the copper method will be adopted by the association.

Wm. Frear: In Pennsylvania we are using the Woodman acetic acid method, not because of any particular preference, except that the records from its use give a wider range for comparison than are available from any other copper sulphate method.

W. C. Woodward: Owing to our extremely limited laboratory facilities and the large number of analyses made, we have accomplished little research work of any kind, and therefore have no data available on the subject. The laboratory has, however, tested this method to some extent, and although comparing favorably

TABLE 12.
Composition of a sample of milk systematically watered.¹

PER CENT TOTAL SOLIDS ADDED WATER	PER CENT FAT	PER CENT ASH	ACETIC SERUM				COPPER SER. M				SOUR SERUM					
			Re- fraction at 20°C.	Sp.gr. at 20°C. 4°	Percent Solids $\frac{n^2-1}{n^2+2} \cdot \frac{1}{d}$	$\frac{n-n^1}{C}$	Re- fraction at 20°C.	Sp.gr. at 20°C. 4°	Per cent Sol- ids	$\frac{n^2-1}{n^2+2} \cdot \frac{1}{d}$	$\frac{n-n^1}{C}$	Refracti- on at 20°C.	Sp. gr. at 20°C. 4°	Percent Ash $\frac{n^2-1}{n^2+2} \cdot \frac{1}{d}$		
0	12.19	3.60	0.71	1.0269	6.49	0.20589	0.00159	37.3	1.0261	5.61	0.20523	0.00153	39.9	1.0254	0.740	0.20588
10	10.97	3.24	0.61	1.0241	5.89	0.20589	0.00157	35.4	1.0243	5.15	0.20518	0.00156	37.5	1.0224	0.666	0.20601
20	9.75	2.88	0.57	1.0208	5.16	0.20592	0.00157	33.5	1.0223	4.62	0.20518	0.00158	34.7	1.0197	0.572	0.20597
30	8.53	2.50	0.50	1.0181	4.45	0.20592	0.00160	31.5	1.0204	4.14	0.20516	0.00158	32.0	1.0169	0.520	0.20597
40	7.31	2.16	0.43	1.0151	3.85	0.20594	0.00156	29.5	1.0118	3.65	0.20507	0.00158	29.4	1.0142	0.428	0.20597

n = Refractive index (not scale reading) of serum; n^1 = Refractive index of water; n^2 = square of refractive index of serum; d = Specific gravity; c = Concentration as percent.

¹ Proc. 8th Intern. Cong. Appl. Chem., 1912, 1: 311.

with the acetic acid method as to efficiency we prefer the latter as the serum is uncolored, the line of refraction more clearly defined, and therefore much more easily and accurately read, especially in sera which are not absolutely clear.

H. H. Hanson: Although we are using the copper sulphate method in our laboratory, we have no determinations upon milk of known purity.

H. Louis Jackson: I abandoned the acetic acid method of preparing the serum several years ago and much prefer the copper method. I feel that it should most certainly be made at least an optional provisional method.

E. M. Bailey: I made the suggestion in regard to the copper serum at the meeting last year because so much data had already been collected and because so many analysts seemed to be using it in preference to the acetic acid serum. Mr. Conn of our State board of health has used it for some years and his experience coincides with mine as regards range of readings. I have never found a milk which I thought was straight which read below 36.

Lythgoe¹ has criticized the copper method, and subsequently Ackermann,² on account of the dilution necessary in the preparation of the serum, but this is offset by less variation in the serum from different samples. Another disadvantage is that it is not possible to use the values of the ash of the copper serum in detecting added water because the amount of copper in the serum is higher in watered milk than in unwatered milk, and consequently the ash in the serum of watered samples is but little less than in the serum of the original milk before watering.

RECOMMENDATIONS.

It is recommended—

(1) That the following be adopted as auxiliary provisional methods:

DETECTION OF ADDED WATER.

(a) *Copper serum.*³—To 1 volume of copper sulphate solution (72½ grams of copper sulphate per liter, adjusted if necessary to read 36.0 at 20°C. on the scale of the Zeiss immersion refractometer, or, to a specific gravity of 1.0443 at $\frac{20^\circ\text{C.}}{4^\circ}$) add 4 volumes of milk. Shake well and filter. Determine the refractive index of the clear serum by means of the Zeiss immersion refractometer. A reading below 36.0 indicates added water.

(b) *Sour serum.*⁴—Allow the milk to sour spontaneously, filter and determine the index of refraction of the clear serum by means of the Zeiss immersion refractometer. A reading below 38.0 indicates added water.

(c) *Ash of sour serum.*⁵—Allow the milk to sour spontaneously and filter. Transfer 25 cc. of this serum to a flat-bottomed platinum dish and evaporate to dryness over the water-bath. Heat the contents of the dish over a small flame (to avoid sputtering) until charred. Place the dish in an electrically heated muffle (with pyrometer connected) and ash at a temperature not greater than 500°C. or 900°F. Cool and weigh. Express results as grams per 100 cc. An ash below 0.730 indicates added water.

(d) *Ash of acetic serum.*—Transfer 25 cc. of the serum to a flat-bottomed platinum

¹ Proc. 8th Intern. Cong. of Appl. Chem., 1912, 1: 308.

² Z. Nahr. Genussm., 1912, 24: 612.

³ Mass. State Board of Health Rep., 1908, p. 594.

⁴ Z. offent. Chem., 1903, 9: 173.

⁵ Chem. Ztg., 1908, 32: 617.

dish and proceed as directed under (c). An ash figure below 0.715 grams per 100 cc. indicates added water.

It is recommended in conjunction with the copper, acetic or sour serum refraction method that the determination of the ash of the sour serum or of the acetic serum be made in all cases where the indices of refraction fall below the minimum limit so as to eliminate all possibility of the sample being abnormal milk from a sick cow. The acetic serum ash multiplied by the factor 1.021 equals the sour serum ash (dilution of the acetic serum being 2 per cent).

(2) That further study be made on the Harding-Parkin¹ method for fat determination in comparison with the present official and provisional methods.

(3) That in U. S. Bur. Chem. Bull. 107, Rev., p. 122, after the paragraph on Cream, the following paragraph be inserted under the heading "Condensed Milk (Unsweetened)": "Dilute 40 grams of the homogeneous material with 60 grams of distilled water, proceed as directed under 'Milk' and correct the results for dilution"; and the word "Sweetened" be inserted before the word "Condensed" in the subsequent heading.

(4) That further study be given to enzyme reactions of milk.

REPORT ON SEPARATION OF NITROGENOUS SUBSTANCES.

By A. D. EMMETT (Agricultural Experiment Station, Urbana, Ill.),
*Associate Referee.*²

During the past year samples with detailed outline of directions were sent out to five laboratories. Owing to pressure of other duties, only two reports were received, and but one of these was complete. As a result, the referee feels that the data given in the following tables are inadequate for definite deductions, and suggests that the work be continued along these lines another year.

It is suggested that the Folin method for creatin and creatinin determinations, as outlined for meats and meat extracts, be made official but that the standard used be the usual potassium dichromate, until more work has been done on obtaining perfectly satisfactory salts of creatinin.

It would appear from the ammonia determinations (Table 3) that the use of sodium hydroxid has a definite and distinct effect in comparison with potassium carbonate. Whether this additional liberation of ammonia is due solely to the breaking up of the ammonium magnesium phosphate is not as yet clear from these data. This point should be studied more thoroughly.

¹ *J. Ind. Eng. Chem.*, 1913, 5: 131.

² Read by P. F. Trowbridge.

TABLE 1.

Percentage of creatin and creatinin in meat extracts and meats (Folin method).

SAMPLE	CREATIN			CREATININ		
	F. C. Cook	C. R. Moulton	A. D. Emmett	F. C. Cook	C. R. Moulton	A. D. Emmett
MEAT EXTRACT:						
1	3.31	2.95	2.82	4.11	4.41	4.66
2	3.07	2.96	2.99	4.13	4.47	4.46
3	3.69	3.69 ¹	4.19	4.33	4.59
Average.....	3.16	2.95	2.90	4.14	4.41	4.57
DRIED MEAT:						
1	1.30	1.84	1.23
2	1.31	1.87	1.25
3	1.40	1.83
Average.....	1.33	1.85	1.24

¹ Omitted from average.

TABLE 2.

Percentage of amino nitrogen in meat extracts and meats.

SAMPLE	VAN SLYKE METHOD		KOBER METHOD
	F. C. Cook	A. D. Emmett	F. C. Cook
MEAT EXTRACT:			
1	1.16	0.70	0.41
2	1.22	0.50	0.47
DRIED MEAT:			
1	0.22	0.59	0.33
2	0.25	0.70	0.30

TABLE 3.

Percentage of ammonia nitrogen in meat extracts and meats.

SAMPLE	F. C. COOK AND J. B. WILSON			A. D. EMMETT		
	Aeration method		Magnesium oxid method	Aeration method		Magnesium oxid method
	Folin (K ₂ CO ₃)	Steel-Gies (NaOH)		Folin (K ₂ CO ₃)	Steel-Gies (NaOH)	
MEAT EXTRACT:						
1	0.402	0.498	0.468	0.644	0.712	0.479
2	0.393	0.458	0.485	0.633	0.702	0.481
3	0.332	0.361	0.507	0.612	0.713
Average.....	0.376	0.439	0.487	0.633	0.709	0.480
DRIED MEAT:						
1	0.055	0.197	0.225	0.086	0.257	0.111
2	0.055	0.203	0.225	0.091	0.272	0.147
3	0.053
Average.....	0.054	0.200	0.225	0.088	0.264	0.129

TABLE 4.
Percentage rate of liberation of ammonia by aeration methods.
 (By A. D. Emmett.)

SAMPLE	FOLIN METHOD					STEEL-GIES METHOD				
	1 hour	20 min- utes	20 min- utes	1 hour	Total 2½ hours	1 hour	20 min- utes	20 min- utes	1 hour	Total 2½ hours
BEEF EXTRACT:										
1	0.491	0.069	0.042	0.042	0.644	0.513	0.081	0.052	0.066	0.712
2	0.466	0.095	0.036	0.036	0.633	0.511	0.077	0.049	0.065	0.702
3	0.482	0.057	0.040	0.033	0.612	0.525	0.082	0.045	0.061	0.713
Average.....	0.483	0.074	0.039	0.037	0.633	0.516	0.080	0.049	0.064	0.709
Percentage....	76.3	11.7	6.2	5.8	100.0	72.8	11.3	6.9	9.0	10.00
Percentage in- crease due to Sugiura- Gies method	6.4	0.7	20.4	41.2	10.7

PLAN FOR COÖPERATIVE WORK.

It is suggested that the work be undertaken along the following lines: (a) Further study of the determination of creatin and creatinin nitrogen; (b) determination of amino nitrogen (amino acids); and (c) ammonia nitrogen.

The determinations should be made in triplicate if possible, that is, on three portions of the original sample.

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SAMPLES.

The samples to be studied consist of desiccated lean beef and beef extract.

The meat was first thoroughly dried by a special procedure at room temperature, then in vacuo. Care should be taken to make all weighings by difference and to keep the bottle covered when not in use. If possible, keep the samples in a refrigerator until ready to analyze.

Before making any weighings mix carefully each sample. In the case of the meat, rotate the bottle in various directions and in the case of the beef extract, warm the jar on the edge of the steam-bath for 5-10 minutes, then stir and transfer a portion to a weighing bottle and allow it to cool for weighing.

PREPARATION OF BEEF EXTRACT SOLUTIONS.

Weigh off (by difference) about 5 grams of the sample into 150 cc. beakers. Dissolve in cold (20°-25°C.) ammonia-free distilled water. Transfer the solution carefully to a 250 cc. measuring flask. Dilute to the mark and mix thoroughly.

Take in duplicate the following quantities, mixing each time before measuring: (a) creatinin, 5 cc.; (b) creatin, 20 cc.; (c) amino acids, 20-40 cc.; (d) ammonia (aeration method), 25 cc.; (magnesium oxid method), 50 cc.

I. CREATIN AND CREATININ.¹

Solutions and reagents.—Ten per cent solution of sodium hydroxid, carbonate-free; picric acid, saturated solution, 1.2 per cent; potassium dichromate, and standard creatin solution.

Apparatus.—Two hundred cc. volumetric flasks; 100 cc. volumetric flasks; autoclave; colorimeter, Duboscq if possible.

(a) CREATIN.

Determination.

(a) *Meats.*—Transfer 5 grams of the fresh meat, or 2 grams of the dried meat (weighed by difference), to a 200 cc. Erlenmeyer flask. Add 100 cc. of N/2 sulphuric acid. Cover the flask with tinfoil or a crucible lid and heat in the autoclave at 130°-135°C. for 30 minutes. Allow the temperature of the autoclave to fall and remove the flask. Cool it under running water and transfer the solution to a 200 cc. volumetric flask, using cold distilled water to aid in the transfer. Shake, to break up the flocculent skeletons of the tissue, dilute to the mark, and mix thoroughly.

Pour the solution on a quantitative filter, collecting filtrate in a dry flask or beaker. Titrate 10 cc. with 10 per cent sodium hydrate, using phenolphthalein as indicator. Measure 10 cc. into a 100 cc. measuring flask, add 20 cc. of the saturated picric acid and "enough 10 per cent sodium hydrate solution to give 1.5 cc. over and above that required for the neutralization of the sulphuric acid"; let stand 10 minutes, dilute to the mark, mix, and read in the colorimeter.

Use the standard creatinin-zinc chlorid solution sent out with the sample (1.389 grams per liter or 1 mg. of creatin per cc.). Set the standard at 10 mm.

(b) *Beef extract.*—Transfer 20 cc. of the beef extract to a 200 cc. volumetric flask

¹ Reprints by Folin accompanied instructions to collaborators.

add 100 cc. N/2 sulphuric acid and proceed exactly as directed under (a) for creatin in meat. Set the standard at 15 mm.

(b) CREATININ.

Determination.

Beef extract.—Transfer 5 cc. of the beef extract solution to a 100 cc. volumetric flask, add 20 cc. of the saturated picric acid and then 1.5 cc. of the 10 per cent sodium hydrate solution. Mix and let stand for 10 minutes, after which dilute to the mark and mix and read in colorimeter. Set the standard at 20 mm. If the readings are too low adjust the standard at 15 mm. The creatin determination in the beef extract includes the creatinin and proper corrections should be made for obtaining the actual creatin, namely, subtract the preformed creatinin from the total creatinin and multiply the difference by 1.16.

NOTE.—In all of the work on creatin and creatinin if the readings are less than two-thirds or more than one and one-half those of the standard, the determinations should be repeated with more or less of the solution.

II. AMMONIA.

(a) AERATION METHOD.¹

Solutions and reagents.—Saturated solution of potassium oxalate (325 grams per liter), potassium carbonate (817 grams per liter), sodium hydrate (506 grams per liter), and kerosene to retard foaming. Sulphuric acid (1 to 4) for wash bottle. Seventieth-normal acid and seventieth-normal alkali. Alizarin red as indicator.

Apparatus.—Test tubes, 10 x 1 inch, or lipless cylinder 8 x 1½ inches; 150 cc. Florence flasks; rubber disk to prevent splashing into receiving flask; suction pump. (See *J. Biol. Chem.*, 1912, 11: 499).

Determination.

Transfer about 5 grams of dried meat or 10 grams of fresh meat (weighed by difference) or 25 cc. of the beef extract solution to a test tube or cylinder. Add 3 to 5 grams of thoroughly washed sand, 10 cc. of distilled ammonia-free water and mix with a glass rod, then add 10 cc. more of water and stir. Remove the rod, washing it down with 5 cc. of distilled water and 10 cc. of the oxalate solution. Add 3 to 4 drops of kerosene and then 5 cc. either of the potassium carbonate or sodium hydroxid solution. At once connect up the apparatus, and start the suction or pressure, gently increasing it during the first minute or two. Allow the air current to run at a high rate for 45 minutes. Then replace the receiving flask with a second one which contains 5 cc. of the standard acid. Continue the aeration for 20 minutes longer. Replace the second receiving flask with another containing 5 cc. of acid and let the operation continue for 20 minutes. Replace the third flask and continue the aeration for an hour. Titrate each distillate with seventieth-normal sodium hydroxid, using 5 drops of alizarin red.

NOTE.—Make a comparison of the effect of using potassium carbonate and sodium hydrate.

(b) MAGNESIUM OXID METHOD.

(See U. S. Bur. Chem. Bull. 107, Rev., p. 9.) Use 50 cc. of the beef extract solution.

¹ Reprints by Folin and Farmer and Folin and Macallum, and Shulansky and Gies accompanied instructions to collaborators.

III. AMINO ACID NITROGEN.

(a) VAN SLYKE METHOD.

Solutions and reagents.—Alkaline permanganate, 50 grams of potassium permanganate and 25 grams of potassium hydroxid per liter; sodium nitrite, 300 grams per liter; glacial acetic acid; caprylic alcohol; sulphuric acid, 1 per cent solution; 25 per cent (316 grams per liter) sodium hydrate solution; 50 per cent acetic acid solution; and 95 and 80 per cent ethyl alcohol.

Apparatus.—A special apparatus is required for this method. This can be procured from Emil Greiner, 45 Cliff Street, New York City. The micro-apparatus is preferable. Mr. Van Slyke suggests that in ordering, it be specified that a 2 cc. cylinder be attached to either apparatus for the caprylic alcohol. It is sometimes necessary in working with tissue to add the alcohol quickly to stop foaming.

Five hundred and 1000 cc. double-necked distilling flasks; and 25 cc. volumetric flasks.

Determination.

(a) *Meats.*—Take in duplicate 10 grams (dried meat) or 40 grams (fresh meat), weighed by difference. Transfer it to 200 cc. Erlenmeyer flasks. Add 100 cc. of boiling water to which 1 cc. of 50 per cent acetic acid per liter is added. Stir and digest for 15 minutes at 100°C. Decant the solution through a glass wool filter, taking care to prevent the particles of meat from passing over. Add to the residue 75 cc. of the hot acid water and heat for 5 minutes. Decant the solution on the same filter. Repeat the extraction 4 more times. Transfer the extract to a liter double-necked distilling flask and concentrate under diminished pressure until the volume of the solution is about 20 cc. Allow the distillation to continue as rapidly as possible without loss of solution by foaming. Transfer the solution to a 250 cc. volumetric flask, using 2 portions of about 5 cc. each of water, then 3 portions of 10 cc. each of 95 per cent alcohol, dilute to the mark with 95 per cent alcohol, mix thoroughly and measure off 100 cc. for the Kober-Sugiura method, and let the remainder stand overnight. Filter the alcoholic solution through a folded filter into a liter distilling flask, wash with 80 per cent alcohol. Add, drop by drop, to the filtrate enough 25 per cent sodium hydrate to make the filtrate alkaline to phenolphthalein. Again concentrate under diminished pressure to remove both ammonia and alcohol. When the volume of the concentrating solution has reached 10 or 20 cc. the distillation is interrupted and enough 50 per cent acetic acid added to acidify the solution. About 50 cc. of water are also added, in order to insure that the last traces of alcohol are driven off, and the concentration continued. Evaporate to 10-15 cc. and transfer cautiously with water to a 25 cc. volumetric flask. Dilute to the mark and mix.

Take 10 cc. portions of this solution for the large apparatus and 2 cc. portions for the micro-apparatus. Continue the procedure exactly as described by Van Slyke (*J. Biol. Chem.* 1911, 9: 189; 1912, 12: 279; and 1914, 16: 121.) The time for the reaction should be maintained carefully; for temperatures 15°-20°C., 4-5 minutes; for 20°-25°C., 3 minutes; and for 25°-30°C., 2½ minutes. In calculating the results consult the table given in the *J. Biol. Chem.*, 1912, 12: 234.

(b) *Beef extract.*—Take 25-40 cc. of the original solution and transfer it to a 250 cc. volumetric flask, dilute to the mark with 95 per cent alcohol. Measure out 100 cc. for the Kober-Sugiura method, and let the remainder stand overnight. Filter on a folded filter and proceed exactly as directed under (a) for this stage of the work.

(b) KOBER AND SUGIURA METHOD.¹

Solutions and reagents.—Cupric hydroxid; anhydrous cupric chlorid solution, 5 per cent; barium hydroxid, saturated solution; sodium hydroxid, normal solution; alkaline sodium boric acid solution; hydrochloric acid, tenth-normal solution; glacial acetic acid; phenolphthalein, 0.10 per cent solution; four-thousandths-normal sodium thiosulphate solution; starch solution; potassium iodid crystals; alcoholic solution zinc chlorid, saturated.

In the preparation of the cupric hydroxid and "buffer" solution the details should be followed exactly as given by the authors.

Determination.

(a) *Meats.*—Take the 100 cc. of the alcoholic solution prepared by the Van Slyke method as directed under (a), transfer to a 100 cc. volumetric flask and add a few drops of the alcoholic zinc chlorid to assist in removing proteins and coloring matter (Folin, *J. Biol. Chem.* 1912, 11: 529). Dilute to the mark with 95 per cent ethyl alcohol and mix. Let stand for a couple of hours, until the precipitate separates out. Filter through dry, folded filter. Take 75 cc. of the filtrate, transfer to a double-necked distillation flask and remove ammonia according to Van Slyke process; that is, add drop by drop to the filtrate enough 25 per cent sodium hydrate to make the filtrate alkaline to phenolphthalein. Again concentrate under diminished pressure to remove both ammonia and alcohol until the volume is 10-20 cc. Then add to the flask 50 per cent acetic acid until the solution is acid. Also add about 50 cc. of water and continue the concentration. Evaporate to 10-15 cc., and very cautiously transfer with water to a 50 cc. volumetric flask. Dilute to the mark and mix.

Take 25 cc. of the solution. Transfer to a 50 cc. volumetric flask. Make neutral or slightly alkaline to phenolphthalein. Dissolve any insoluble substances with the aid of tenth-normal sodium hydroxid, using not more than 5-6 cc. Keep the flask stoppered to avoid the presence of carbon dioxid. Add 20 cc. of the "buffer" solution and a little of the freshly made cupric hydroxid. Shake the mixture vigorously for about a minute. If there is an excess of the cupric hydrate, after bringing it to room temperature, dilute to the mark, and shake for 2-3 minutes. Filter through dry filters (S. and S. No. 590 11 cm.). The filtrate is supposed to contain all the soluble complexes and the insoluble residue on the filter the insoluble complexes and the excess of copper hydroxid.

An aliquot portion (25 cc.) of the filtrate is then taken and after acidification with 1-3 cc. of glacial acetic acid, 5 cc. of the potassium-iodid-starch solution are added and the solution titrated with four-thousandths-normal thiosulphate solution. Every cubic centimeter of thiosulphate solution is equivalent to 0.0003184 gram of cupric oxid or 0.0001120 gram of amino acid nitrogen or 1 cc. of one-thousandth-normal thiosulphate solution is equivalent to 0.000280 gram of amino acid nitrogen.

(b) *Beef extracts.*—Take the 100 cc. of the alcoholic solution prepared by the Van Slyke method, transfer it to a 110 cc. volumetric flask and add a few drops of the alcoholic zinc chlorid solution. Dilute to the mark with 95 per cent ethyl alcohol and mix. Continue exactly as outlined for meats by this method.

¹ Reprints by Kober and Sugiura accompanied instructions to collaborators.

REPORT ON FEEDS AND FEEDING STUFFS.

By G. L. BIDWELL (Bureau of Chemistry, Washington, D. C.), *Referee*.

The subject of the nitrogen factor has been before the association for several years. The factor 6.25 has been used for nearly 70 years and, when recent additions to the knowledge of the proteins are considered, is it not proper to ask if that value is not antiquated, and if it is not time to change to one which will give results nearer the truth? When substances containing a few well-known proteins are considered, such as dairy products or wheat flours, there can be no argument but that the change should be made from 6.25 to a factor that is known to be nearer correct. But in the case of feeding stuffs, especially from a control or inspection standpoint, there is a different set of conditions. In the first place, there are about 100 different products that enter into the composition of stock feeds. Ten or fifteen different products are derived from corn alone. From wheat we get bran, middlings, red dog, ship stuff and wheat screenings. Because wheat has a factor of 5.7, it does not follow that bran, middlings, and red dog, have the same factor nor is it likely that bran has the same factor as red dog. As for screenings, how is the factor to be determined? If it should be determined, would there ever be another sample of screenings which would have the same factor?

Now, consider the labor involved in determining the factor to apply to any one substance. This is not a problem to be solved by some assistant in a few weeks. It would be necessary to determine the amount of each protein in that substance, then to prepare it in a pure condition so that the per cent of nitrogen might be determined therein. This is a problem for the specialist who has available almost unlimited time and money. But if there were factors of all the ingredients of feeds could they be used? About 45 per cent of the feeds examined in the Bureau of Chemistry for the past two years were mixed feeds, and even when coarsely ground, the trained microscopist could not tell, except within very wide limits, the amounts of the various ingredients present. If different factors were used for the various feed stuffs, it would still be necessary to have a general factor for mixtures. If the factor for corn meal should be 6 and wheat by-products 5.7 and a mixture of 97 per cent middlings and 3 per cent corn meal were being analyzed, the general factor would still have to be used, whatever that might be. Although 5.7 would be the factor that would give nearest correct results, it would be impossible to tell the relative amounts of middlings and corn meal and therefore the only course left would be to use the general factor. Now, keeping in mind that we are considering this from the control and inspection standpoint, is it ever desired to know the actual amount of protein in a feed? It is known that the different proteins vary a great deal in composition,

some having less than 15 per cent nitrogen and others having over 17 per cent. But is there anything to show that a pound of one of these is equal to a pound of the other in feeding value? It seems reasonable to suppose that the amount of nitrogen may be a better index of the value of the proteins of a feed than the actual weight of the protein itself. For nearly 70 years the per cent of nitrogen has been multiplied by 6.25 and the result called protein, but all this time it is the nitrogen percentages that have been compared. If another factor were adopted the same ratios would still be used when comparing the protein content of feeds; that is, the percentages of nitrogen would still be used even though they were multiplied by a factor. There are several hundred thousands of analyses of feeds on record and nearly all of them employ the factor 6.25. If the factor were changed the results of analyses would no longer be comparable with former analyses. It is too much to hope that all chemists would adopt the new factor. Experience has also shown that it would be impossible to persuade all chemists to indicate the factor they employed. It seems, therefore, that a change would cause endless confusion and give no compensating advantages. Of course it should be understood that these arguments do not apply to investigational work or work that is restricted to single well-known substances such as those mentioned in the first part of this report.

RECOMMENDATIONS.

It is recommended—

(1) That 6.25 be retained as the factor to change nitrogen to protein in all feeding stuffs control and inspection work.

(2) That the subject of improvements in the determination of fiber be further studied.

REPORT ON CRUDE FIBER.

By CHARLES K. FRANCIS (Agricultural Experiment Station, Stillwater, Okla.), *Associate Referee*.

The work on crude fiber has been confined to determinations on one sample of cottonseed meal. The sample was sent to the collaborators without special instructions, other than to determine the crude fiber and report the details of the method used.

The well-known official method for determining crude fiber has been subjected to criticism by many chemists. The chief trouble seems to be with the filtering materials, linen, asbestos, or glass wool. These substances vary in their physical characteristics and necessarily in filtering efficiency. The determinations reported show a sufficient range of materials, and certainly the results suggest a need for further study. Except where otherwise indicated, the official method was used by the collaborating chemists.

RESULTS OF COÖPERATIVE WORK.

Crude fiber in cottonseed meal.

COLLABORATOR	CRUDE FIBER	REMARKS	COLLABORATOR	CRUDE FIBER	REMARKS
	<i>per cent</i>			<i>per cent</i>	
J. M. Pickel, Raleigh, N. C.....	6.94 6.72 6.05 6.69 6.21 6.62	Filtration, for acid digestion, upward suction through linen stretched over a funnel.	A. S. Wells, Portland, Ore.....	10.96 11.42 11.20	Silk filter.
Average.....	6.54		Average.....	11.19	
C. G. Remsburg, College Park, Md.....	7.99 7.88 8.07 8.15	Muslin filter.	C. Kennedy, St. Paul, Minn.....	10.77 10.35	Sweeney method, Kennedy modification. Linen filter.
Average.....	8.02		Average.....	10.56	
C. Cutler, Lafayette, Ind.....	7.85 8.09 7.76 8.15 7.91	Dried at 100°C. Linen filter.	A. A. Jones, Stillwater, Okla.....	9.50 9.54	Sweeney method, modified. Asbestos filter.
Average.....	7.95		Average.....	9.52	
J. H. Roop, Lafayette, Ind.....	8.33 8.15 8.30	Dried at 100°C. Linen filter.	O. C. Smith, Stillwater, Okla.	10.55 10.83 10.25	Sweeney method, modified. Asbestos filter.
Average.....	8.26		Average.....	10.54	
C. E. Mangels, Columbia, Mo...	8.54 8.86 8.56 8.51	Muslin filter.	G. Merry, Stillwater, Okla.....	9.76 9.14 9.55	Sweeney method, modified. Asbestos filter.
Average.....	8.62		Average.....	9.48	
C. E. Mangels, Columbia, Mo...	8.59 8.53	Sweeney method.	J. W. Chewning, College Station, Texas.....	8.76	Asbestos filter.
Average.....	8.56		G. W. Roark, College Station, Texas.....	8.99 8.67	Asbestos filter.
			Average.....	8.83	
			General average.....	9.07	

RECOMMENDATIONS.

It is recommended—

- (1) That the Sweeney method, and some of its modifications, be studied.
- (2) That asbestos be specified as the filtering material.
- (3) That the official method be compared, with linen and asbestos filters specified.

THE DETERMINATION OF CRUDE FIBER.

BY G. L. BIDWELL AND G. P. WALTON (Bureau of Chemistry,
Washington, D. C.).

DISCUSSION.

The official method for the determination of crude fiber is considered far from satisfactory by many feed analysts. This is apparently due to difficulties in manipulation and to the time necessary to complete the determination. Several modifications of the method have been proposed with the view of eliminating various objectionable features or of shortening the time required. An investigation of the official method and one of the more promising of the proposed modifications has been undertaken.

For this purpose 4 representative feeding stuffs, having a fairly wide range in crude fiber, were selected.

Description of samples used in this investigation.

SAMPLE	CRUDE FIBER ¹
	<i>per cent</i>
Oats (containing oat hulls).....	11.30
Corn (Argentine).....	1.80
Cottonseed meal.....	12.33
Roughage (a composite of several forage plants).....	26.57

¹ Average of 8 separate determinations by the official method.

A large sample of each was carefully prepared by grinding, sifting and thoroughly mixing.

Each sample was extracted with ether before the crude fiber determination was made.

Kennedy has proposed a modification of the Sweeney method, endeavoring to eliminate the acid filtration which is considered one of the principal drawbacks to the official method. Her method is as follows:

Boil fat-free material for 30 minutes under a reflux condenser with 200 cc. of a 1.25 per cent solution of sulphuric acid, add 200 cc. of boiling 3.52 per cent solution of sodium hydroxid; boil 30 minutes longer, filter through linen, wash first with a little hot water, then thoroughly with a boiling 1.25 per cent solution of sulphuric

acid, and then with hot water, transfer to a Gooch crucible, finish washing with hot water, then with 95 per cent alcohol and finally with ether. Dry to constant weight at 110° C., weigh, incinerate, weigh, and the difference is crude fiber.

The following table gives the results obtained by this method:

Per cent of crude fiber.

SAMPLE	METHOD		
	Kennedy modification	Official	Excess of Kennedy over official
Oats.....	12.30	11.30	1.00
Corn.....	2.29	1.80	0.49
Cottonseed meal.....	13.56	12.33	1.23
Roughage.....	26.70	26.57	0.13

From the writers' experience the official method takes less time than the Kennedy modification. The slow final filtration, the time consumed by washing with the dilute sulphuric acid and transferring the residue from the linen to the crucible more than make up for the time consumed in the acid filtration in the official method. To throw light on the high result by the Kennedy method another set in duplicate was run varying from the official method only by the addition of a sufficient amount of sodium sulphate to both solutions to give the same concentration of this salt as that resulting from the neutralization of the sulphuric acid in the Kennedy method. Although this solution boiled at 102°C. its dissolving or disintegrating action was less than that of the regular reagent. The average percentage figures obtained were: oats 11.87; corn 2.18; cottonseed meal 13.51; and roughage 27.72.

A charge of cottonseed meal was boiled with the acid as in the official method. After filtration the acid filtrate was treated with 200 cc. of 3.52 per cent of sodium hydroxid solution. The precipitate formed was filtered and washed with boiling 1.25 per cent solution of sulphuric acid; 0.85 per cent, calculated on the original charge of cottonseed meal, remained undissolved.

Since crude fiber is not the name of a definite substance but only a convenient and somewhat descriptive term used to denote the result obtained by treating a feed by the official method, several determinations were made to show the effect of variations of those factors which define that method. When using 400 cc. for each of the reagents instead of 200 cc., the results were only a trifle less than by the official method. Changing the concentration of the reagent affected the results appreciably, as is shown in the following table:

Per cent of crude fiber obtained with solutions of varying percentage composition.

STRENGTH SOLUTION	OATS	CORN	COTTONSEED MEAL	ROUGHAGE
0.78	12.23	1.97	13.44	28.38
1.25	11.30	1.80	12.33	26.57
2.00	10.52	1.77	11.01	24.73

The percentage of crude fiber obtained is inversely proportional to the concentration of the reagents.

To show the effect of variation in the time of boiling, sets were run with the regular acid and alkali solutions, for the following periods of time: 20 minutes; 45 minutes; 60 minutes; and 90 minutes. The results showed a continuous decrease in the percentage of crude fiber with no indication that a point might be reached beyond which further boiling would cease to cause a decrease in crude fiber.

The rôle performed by each of the reagents was next investigated. Sets were run in duplicate with acid alone for 20, 30 and 45 minutes.

Per cent of residue obtained.

TIME OF BOILING	OATS	CORN	COTTONSEED MEAL	ROUGHAGE
<i>minute:</i>				
20	23.00	9.97	26.74	48.94
30	20.22	9.00	24.15	46.56
45	18.50	8.26	22.63	44.64

Considering the total material dissolved in the official method as 100 per cent, the amount dissolved by acid in the oats is 89.94 per cent; in the corn 92.67 per cent; in the cottonseed meal 86.85 per cent; and in the roughage 72.78 per cent.

SUMMARY.

Inasmuch as crude fiber designates simply a numerical value obtained by following the official method which is strictly an empirical one, any deviation therefrom will give incorrect results. This is the strongest and all important objection to any material change in a method for official endorsement. The writers have found that by observing certain precautions, the results by the official method will check very closely and the difficulties and tediousness will be eliminated. In fact, they have been unable to make the determination by any of the proposed modifications in any less time than by the official method.

PRECAUTIONS.

- (1) The solutions should be of proper strength and mixed thoroughly.
- (2) The charge should be practically free from fat and about 1-2 grams of ignited asbestos added to render the filtration quicker.

(3) The boiling should be under a reflux condenser to prevent concentration of the reagent.

(4) The time of boiling and filtration should be uniform, as the lengthening of either tends to lower results. The linen should be selected carefully. It is possible to filter and wash a set of 10 determinations obtaining a clear filtrate in about 15 minutes. This is true of cottonseed meals as well as the more readily filtered materials.

(5) The washings should be made with very hot water.

(6) The wash water after the acid filtration should be allowed to drain so as to avoid diluting the alkali solutions.

If these precautions are observed in conjunction with the directions found in U. S. Bur. Chem. Bull. 107, Rev., it is believed that little fault will be found with the official method.

GOSSYPOL, A TOXIC SUBSTANCE IN THE COTTONSEED.

A PRELIMINARY NOTE BY W. A. WITHERS AND FRANK E. CARRUTH
(Agricultural Experiment Station, Raleigh, N. C.).

Gossypol occurs in the peculiar resin glands of the cottonseed. Oil was removed from ground up cottonseed kernels by petroleum ether, leaving gossypol undissolved. This was subsequently extracted by ethyl ether. The gossypol extract was dissolved in purified cottonseed oil and fed to rabbits by catheter and also by intraperitoneal injection. It was fatal in every case in a few hours. The extract was mixed with corn meal and molasses and fed to rabbits. It was uniformly fatal in 9 to 16 days.

A product precipitated from the "gossypol extract" by petroleum ether was also poisonous.

A crystalline product corresponding in properties to the phenolic substance gossypol, isolated by Marchlewski¹ from crude cottonseed oil was dissolved in purified cottonseed oil and administered to 2 rabbits intraperitoneally. It caused death within 6 hours.

It is difficult to extract gossypol completely as this material is held dye-like by the seed tissue and some of the resin glands are perhaps not penetrated by the solvent. Thoroughly extracted kernels were fed over long periods without fatal results.

The properties of gossypol may be used to explain the fact that boiling alcoholic alkali removes the toxicity of cottonseed meal,² and iron salts act as *antidotes* for cottonseed meal.³ In alkaline solutions the substance *oxidizes* with a purple coloration which subsequently disappears. Ferrous sulphate forms an insoluble lake with gossypol.

The occurrence and properties of gossypol are being further studied and will form the basis of a later report.

¹ *J. prakt. Chem.*, 1899, **60**: 80.

² *Science*, 1912, **36**: 31.

³ *J. Biol. Chem.*, 1913, **15**: 161.

REPORT ON FEED ADULTERATION.

By CARLETON CUTLER (Experiment Station, Lafayette, Ind.), *Associate Referee*.

The study of feed adulteration was added by the association at its last meeting to the subjects for coöperative investigation. The growing importance of this subject is due to the many new sources of feeding stuffs resulting from increased demand and higher prices received for concentrated feeds. The utilization of numerous and varied by-products affords many opportunities for adulteration.

Because of the small amount of other data available, this report is confined to feeding stuffs inspected in the laboratory of the associate referee. Methods of adulteration have been outlined and a brief résumé prepared of some of the adulterations with methods for their detection.

For the examination of feeding stuffs for adulteration the following apparatus is necessary: a set of sieves from 20 to 70 mesh inclusive, a hand lens, a compound microscope with low and high power lenses, and a set of instruments, including dissecting needles, scalpels, etc. Certain reagents are necessary, such as clearing reagents and stains for cellulose, woody fiber, fats, oils, starch, proteid matter, etc. It is also desirable to have apparatus for making permanent mounts of substances of known purity for comparisons.

TYPES OF ADULTERATION AND METHODS OF DETECTION.

(1) *Total substitution of one material for another.*—Corn feed meal is sometimes substituted for hominy feed. Upon sieving the sample, a preponderance of fine starchy material and an absence of corn germ indicates adulteration. A quantitative determination of crude fat and protein is necessary for confirmation. Thirty-two samples of corn feed meal analyzed in the laboratory of the associate referee carried 3.9 per cent fat and 8.6 per cent protein as against 7.7 per cent fat and 11.2 per cent protein as the average of 72 samples of hominy feed.

(2) *Use of a substance of an inferior quality.*—Linseed meal containing ground flaxseed screenings is sometimes found. Upon sieving and examination a large amount of finely ground weed seed coatings is found. The characteristic odor of this adulterant is generally sufficient for detection. A protein determination generally gives confirmation.

(3) *Intentional addition.*—Cocoa shells may be added to a compounded feed. The cocoa shells can be separated from the coarser portion of the sample after sieving and can be identified by the practiced eye with certainty under the hand lens. The shell, chocolate brown on the outer surface and a silver brown on the inner surface, is thin and brittle to the touch of the dissecting needle. For confirmation one should clear.

mount, and examine with the compound microscope. Some of the most prominent characteristics are the longitudinally elongated polygonal cells underlying the cross cells of the endocarp, the spiral cells of the spongy parenchyma and the thickened stone cells. The elements must be familiar from the examination of the pure substance.

Peat dried at a high temperature may be added to mixtures containing tankage, molasses and the like. No satisfactory method has been discovered for detecting this adulterant.

(4) *Accidental addition*.—Often in milling the flour stuff is not thoroughly removed from the bran coating. The microscopist who is familiar with wheat by-products can generally detect this with the hand lens after sieving. For confirmation, a quantitative determination shows a low content in both fat and protein.

(5) *Omission*.—Often in a proprietary feed one of the most expensive ingredients is omitted, the manufacturer, no doubt, trusting that because of the complexity of the mixture it will not be missed. Feeds of this class may be rubbed gently in the palm of the hand before sieving. This does not destroy the character of the individual ingredients, but renders them more recognizable for the examination under the hand lens. Frequently in finely ground feeds it is necessary to make repeated mounts for examination with the compound microscope before the absence or presence of the ingredient can be confirmed.

(6) *Intentional abstraction and addition*.—A linseed meal may contain 15 per cent of pressed flaxseed screenings and 10 per cent of medium cottonseed meal and be sold for linseed meal, as the fat and protein content will approximate closely with that of pure linseed meal. The odor will indicate flaxseed screenings. If some of the adulterated sample is placed on the moistened finger and rubbed, the characteristic yellow of the cottonseed meal is noticed. After rubbing gently in the palm of the hand and sieving, one can easily detect weed seed coatings, fragments of cottonseed hulls and fine particles of meal by use of the hand lens, and by the use of a dissecting needle can pick out a considerable percentage of cottonseed product.

Other adulterations observed are as follows:

Finely ground corn cobs added to hominy feed; white corn meal to wheat middlings; ground and whole wheat screenings to wheat bran and middlings; corn screenings, containing considerable cob, to chop feeds; clover seed and wheat screenings to poultry feeds; oat clipping by-product to proprietary feeds; and cottonseed hulls to cottonseed meal.

By the use of sieves, the hand lens and a compound microscope, the microscopist is able to detect these adulterations with considerable accuracy. A quantitative determination of protein, fat and fiber acts as a guide in detecting adulteration. The chances of adulteration are so

numerous that the microscopist should be continually on the alert for new adulterants, and methods for their detection. It is important that he should be thoroughly familiar with feeding stuffs, their process of manufacture and the by-products used in the compounding of feeds.

The economic importance of adulteration is worthy of consideration, both from the viewpoint of the manufacturer and the consumer.

RECOMMENDATIONS.

It is recommended—

(1) That samples be sent out for the quantitative determination of adulterants in amounts varying from 5-25 grams to determine the smallest amount necessary to give concordant results.

(2) That samples be sent out containing unknown adulterants for qualitative detection of the same.

(3) That methods for the detection of peat dried at high temperature in feeding stuffs be investigated.

(4) That the maximum percentage of foreign materials permissible in mill by-products be investigated.

No report was made by the referee on sugar.

REPORT ON TESTING CHEMICAL REAGENTS.

BY J. B. RATHER (College Station, Texas), *Referee*.¹

The work for the past year dealt with the following chemicals:

1. Crude caustic soda.
 - a. Determination of nitrogen.
 - b. Methods for the determination of sodium hydroxid and sodium carbonate.
2. Molybdic acid.
 - a. Determination of molybdenum trioxid.
 - b. Determination of phosphoric acid.
3. Ethyl ether.
 - a. Determination of solids non-volatile at 100°C.
4. Citric acid.
 - a. Determination of impurities.

CRUDE CAUSTIC SODA.

Determination of Nitrogen.

Method (A).—This method is similar to the method of Krauch-Merck for the determination of nitrogen in caustic soda, using, however, a 40

¹ Presented by G. S. Fraps.

gram sample, which is the approximate amount of solid caustic soda used in the ordinary nitrogen determination.

Method (B).—In this method the above described procedure was modified by distilling the ammonia into *distilled water containing no acid*, then redistilling the distillate into fifth-normal hydrochloric acid in the usual manner.

Method (C).—In this method the procedure, outlined in (B), was further modified by the use of a blank titration of fifth-normal hydrochloric acid, which was intended as a correction for error of standard solution, titration, etc.

The results of the single distillation process (A) were quite variable, ranging from 0.03 to 1.75 cc. of fifth-normal acid. This was probably due to traces of alkali carried over in the distillate. The results by the redistillation process (B), which was devised to correct for this defect of method (A), vary from 0.03–0.75 cc. of fifth-normal acid.

Determination of Sodium Hydroxid and Sodium Carbonate.

The direct titration method with phenolphthalein and methyl orange for the determination of sodium hydroxid and sodium carbonate in caustic soda was compared with the method of Tillmans and Heublein (*Z. Angew. Chem.*, 1911, **24**: 874), and it was found that the latter method gave better results.

MOLYBDIC ACID.

Determination of Molybdenum Trioxid.

The method used was the lead molybdate method, recommended by Krauch-Merck, as modified by the referee as follows: After solution of the molybdenum trioxid in ammoniacal water, it was filtered to remove iron and alumina; and further, the precipitate was washed by decantation a number of times before filtering through a Gooch. The results by different analysts agreed well except in a few instances.

Effect of varying some conditions of the Krauch-Merck method.—The possible effect of a number of factors in the determination of molybdenum trioxid in molybdic acid by the lead method were studied in order to find a possible reason for the variations sometimes occurring in the analyses. The results of the investigation showed: (1) that the quantity of lead acetate used does not greatly affect the results; (2) that the purity of the lead acetate does not affect the determination (The same results were obtained using a sample of "technical" lead acetate as when using a so-called "chemically pure" crystallized sample.); (3) that the addition of enough sulphates to cause an increase of 0.25 per cent in the supposed molybdenum trioxid content of the sample, caused an increase of 0.20 per

cent in the results (Larger amounts of sulphates caused larger increases, but not in proportion to the amount of sulphates added. Further study will be required to determine the effect of the presence of sulphates.); (4) that the acidity of the solution may be varied within wide limits without affecting the accuracy of the method.

Determination of Phosphoric Acid.

The method for the determination of phosphoric acid in molybdic acid as outlined in last year's report was further tested.

The results showed that small amounts of phosphoric acid (1 mg. or less) can not be determined always when the solution is diluted to 75 cc. or more. At a volume of 50 cc., all of the added phosphoric acid was recovered.

ETHYL ETHER.

Determination of Solids Non-Volatile at 100°C.

The method consisted in evaporating to dryness 100 cc. in a tared dish on the steam-bath and drying to constant weight at 100°C. The solids obtained varied from 0.0006–0.0034 gram. This would affect the results of a fat determination, calculated on the basis of a 2 gram sample, from 0.02–0.17 per cent. The results indicate that if ether be used for fat determinations, it should be tested for non-volatile solids regardless of its supposed purity.

CITRIC ACID.

Determination of Impurities.

This work was devised to study the accuracy of the methods used last year for the determination of sulphuric acid and oxalic acid in commercial citric acid.

Sulphuric acid.—This was determined in the ordinary way by precipitation with barium chlorid.

Oxalic acid.—This was determined by neutralizing with ammonia and precipitating with calcium chlorid in a strongly acidified acetic acid solution. After standing a few hours the precipitate was filtered, washed and ignited in the usual manner. If the solution is not strongly acid, calcium citrate precipitates when the solution is heated or stands overnight. Any moderate amount of the citrate will be removed in the washing of the precipitate of calcium oxalate.

The results show that the method for the determination of sulphates is adequate for amounts varying between 0.01 and 3.00 per cent. The results in the oxalate determination were not quite satisfactory.

RECOMMENDATIONS.

It is recommended—

(1) That the double distillation method (*B* including *C*) for the determination of nitrogen in crude caustic soda, the modified lead method for the determination of MoO_3 in molybdic acid, and the method for the determination of solids in ethyl ether non-volatile at 100°C ., as given in this report, be studied further with the view of incorporating them in the official methods for nitrogen, phosphoric acid, and crude fat or ether extract, respectively.

(2) That the direct titration method and the Tillmans and Heublein method for the determination of NaOH and Na_2CO_3 in crude caustic soda and the method for the determination of P_2O_5 in molybdic acid, as given in this report, be studied coöperatively next year, and that the effect of the presence of sulphates on the determination of MoO_3 in molybdic acid, together with the causes of the variations in the determination of MoO_3 in molybdic acid by the modified lead method, be studied further.

(3) That the study of the nature and amount of the impurities in commercial citric acid be discontinued for the present.

A paper "On the Colorimetric Determination of Small Quantities of Hydrocyanic Acid" by Arno Viehoveer and C. O. Johns of the Bureau of Chemistry, was read by Mr. Johns. This information has since been published (*J. Am. Chem. Soc.*, 1915, **37**: 601).

C. H. Jones presented the following recommendations of the Executive Committee, which were approved:

(1) That the proceedings of this association be published in the form of a quarterly.

(2) That a one-half day session be given to section meetings to be arranged by the Executive Committee.

(3) That the matter of the revision of the Constitution and By-Laws relating to annual dues be left until the 1915 meeting.

REPORT OF COMMITTEE A ON RECOMMENDATIONS OF REFEREES.

By B. B. Ross, (Agricultural Experiment Station), *Chairman*.

(Phosphoric acid, nitrogen, potash, soils, inorganic plant constituents, insecticides and water.)

PHOSPHORIC ACID.

It is recommended—

(1) That further work be done on the methods for basic slag as reported this year.

Approved.

(2) That special attention be given to standardizing the alkali solution used in the volumetric method.

Approved.

(3) That the methods given in this year's report be tried out on as many samples as possible.

Approved.

(4) That the associate referee give attention to the question of whether neutral ammonium citrate, sodium citrate, or citric acid solution should be employed as a solvent in the determination of reverted phosphoric acid in fertilizers.

Approved.

NITROGEN.

It is recommended—

(1) That the zinc-ferrous-sulphate-soda method for nitrates be further studied during the coming year with a view of its final adoption as official in 1916 and that it be now adopted as provisional.

Adopted, final action as provisional; approved for final action as official in 1916.

(2) That the Jones and Street methods for the determination of organic nitrogen activity be further studied during the coming year with the special purpose in view of improving or modifying the manipulations in the conduct of each process, so as to increase the accuracy of the water-insoluble organic nitrogen determinations, and, in the case of the Jones method, to overcome the difficulties experienced by most analysts in the distillation with alkaline permanganate; and that they be now adopted as official.

Adopted, final action.

POTASH.

It is recommended—

(1) That further study of the perchlorate method be made with special reference to the method for washing the potassium perchlorate precipitate.

Approved.

(2) That coöperation be secured to test the necessity for the addition of hydrochloric acid to the potash extract, with special reference to the reason why the hydrochloric acid is added.

Approved.

(3) That denatured alcohol made up according to formula 1 (U. S. Internal Rev. Reg. No. 30, Rev., Aug. 22, 1911, p. 45. To 100 parts by volume of ethyl alcohol, not less than 180° proof, there shall be added 10 parts by volume of approved wood alcohol and one-half of one part by volume of approved benzine) with sufficient water added to make 80 per

cent alcohol by volume, be further tested for washing potassium chloroplatinate, with a view of its adoption officially in 1915.

Approved for final action as official in 1915.

SOILS.

It is recommended—

(1) That a further test of methods for the determination of soil carbonates be made, comparing the Marr procedure with methods which involve the use of dilute hydrochloric acid (1 to 15) and constant aspiration of air with and without heating.

Approved.

(2) That the wet combustion method with the mixture of chromic and sulphuric acids for the estimation of organic carbon be further compared with combustion of the soil in furnace.

Approved.

(3) That, at the suggestion of O. M. Shedd of Kentucky, a study be made by the association of a method for lime requirement of soils by H. B. Hutchinson and K. MacLennan (*Chem. News*, Aug. 7, 1914, p. 61).

Approved.

(4) That the methods for nitrates, nitrites, and ammonia, as adopted for waters, in conformity with the recommendations of Committee A in 1913, be now adopted as official methods for the determinations of these constituents in aqueous soil extracts.

Approved for final action as official in 1915.

(5) That the official process for the determination of carbon dioxide in soils be stricken from the methods of the association.

Approved.

(6) That the official method for alkali waters be provisionally adopted for alkali soils.

Approved for final action in 1915.

INORGANIC PLANT CONSTITUENTS.

No report or recommendations.

INSECTICIDES.

It is recommended—

(1) That Method I for total arsenious oxid in Paris green, as described in U. S. Bur. Chem. Bull. 107, Rev., pp. 25-26, be changed as described in this year's report, and that the method thus modified be studied further with a view of its adoption as an official method in 1915.

Approved.

(2) That Methods II and III for total arsenious oxid in Paris green (U. S. Bur. Chem. Bull. 107, Rev., pp. 26-27) be discarded.

Approved.

(3) That Methods II and III for total arsenic in Paris green, described in this year's report, be studied further with a view of their adoption as official in 1915.

Approved.

(4) That the official method for the determination of moisture in Paris green be extended to include determinations of moisture in Bordeaux mixture, Bordeaux and Paris green mixtures, and Bordeaux and lead arsenate mixtures, when these materials are in the form of a powder, and that Method (b) for the determination of moisture in Bordeaux mixture, Bordeaux and Paris green mixtures, and Bordeaux and lead arsenate mixtures, when in the form of pastes, as described in this year's report, be studied further with a view of its adoption at the proper time as an official method.

Approved.

(5) That the method for the determination of carbon dioxide in Bordeaux mixture, Bordeaux and Paris green mixtures, and Bordeaux and lead arsenate mixtures, as described in this year's report, be studied further with a view of its adoption as an official method in 1915.

Approved.

(6) That the electrolytic method for the determination of copper in Bordeaux mixture, as described in this year's report, be studied further with a view of its adoption as an official method in 1915.

Approved.

(7) That the thiosulphate titration method for the determination of copper in Bordeaux mixture, as described in this year's report, be studied further with a view of its adoption as an official method in 1915.

Approved.

(8) That the method for water-soluble arsenious oxid in Bordeaux and Paris green mixtures, as described in this year's report, be studied further with a view of its future adoption as an official method.

Approved.

(9) That the distillation method for the determination of total arsenic in Bordeaux and Paris green mixtures, as described in this year's report, be studied further with a view of its future adoption as an official method.

Approved.

(10) That the methods of C. M. Smith and C. C. Hedges for the determination of total arsenious oxid in Paris green, as described in this year's report, be studied further with a view of their future adoption as official methods for the determination of arsenious oxid in Bordeaux and Paris green mixtures.

Approved.

(11) That the electrolytic method for the determination of copper in Bordeaux and lead arsenate mixtures, as described in this year's report,

be studied further with reference to its applicability to the determination of copper in both Bordeaux and Paris green mixtures and Bordeaux and lead arsenate mixtures.

Approved.

(12) That the thiosulphate titration method for the determination of copper in Bordeaux and Paris green mixtures, as described in this year's report, be studied further.

Approved.

(13) That the method for water-soluble arsenic oxid in Bordeaux and lead arsenate mixtures, as described in this year's report, be studied further with a view of its future adoption as an official method.

Approved.

(14) That the method for the determination of lead oxid in Bordeaux and lead arsenate mixtures, as described in this year's report, be studied further.

Approved.

(15) That Procedure (b) of the silicotungstate method for the determination of nicotin, as described in this year's report, be studied further with a view of its adoption as official in 1915.

Approved.

(16) That further work on the Lloyd method for the determination of nicotin be discontinued.

Adopted.

(17) That the coöperative work on insecticides for the coming year comprise a study of the following:

(a) Methods for the determination of As^{iii} and As^v in the presence of each other and in the presence of one or more of the following: lead, copper, zinc, and calcium.

(b) A method other than an electrolytic one for the determination of copper, lead, and arsenic in a Bordeaux and lead arsenate mixture.

(c) Methods for the determination of the principal ingredients in zinc-arsenic compounds, alone and in combination with Bordeaux mixture.

Approved.

WATER.

It is recommended—

(1) That methods for the determination of strontium be studied further during the coming year.

Approved.

REPORT OF COMMITTEE B ON RECOMMENDATIONS OF
REFEREES.

By P. F. TROWERIDGE (Agricultural Experiment Station, Columbia,
Mo.), *Acting Chairman*.

(Dairy products, feeds and feeding stuffs, sugar, water in foods and feeding stuffs, organic and inorganic phosphorus in foods, feeding stuffs, and drugs, separation of nitrogenous substances, testing chemical reagents, tannin, medicinal plants and drugs.)

DAIRY PRODUCTS.

It is recommended—

(1) That the copper serum method, as given in this year's report, be adopted as an optional provisional method for the detection of added water.

Adopted, final action.

(2) That the sour serum method, as given in this year's report, be adopted as an optional provisional method for the detection of added water.

Approved for final action as provisional in 1915.

(3) That the ash of the sour serum method, as given in this year's report, be adopted as an optional provisional method for the detection of added water.

Approved for final action as provisional in 1915.

(4) That the ash of the acetic serum method, as given in this year's report, be adopted as an optional provisional method for the detection of added water.

Approved for final action as provisional in 1915.

(5) That in conjunction with the copper acetic or sour serum refraction method, the determination of the ash of the sour serum or of the acetic serum be made in all cases where the indices of refraction fall below the minimum limit so as to eliminate all possibility of abnormal milk from a sick cow.

Approved.

(6) That further study be made of the Harding-Parkin method for fat determination (*J. Ind. Eng. Chem.*, 1913, **5**: 131) in comparison with the present official and provisional methods.

Approved.

(7) That in U. S. Bur. Chem. Bull. 107, Rev., p. 122, after the paragraph on Cream, the following paragraph be inserted under the heading "Condensed Milk (Unsweetened)"; "Dilute 40 grams of the homogeneous material with 60 grams of distilled water, proceed as directed

under 'Milk', and correct the results for dilution"; and the word "Sweetened" be inserted before the word "Condensed" in the subsequent heading.

Adopted.

(8) That further study be given to enzyme reactions of milk.

Approved.

FEEDS AND FEEDING STUFFS.

It is recommended—

(1) That no change be made in the nitrogen factor for protein.

Approved.

(2) That there be a thorough investigation of the methods for crude fiber determination.

Approved.

(3) That samples be sent out for the quantitative determination of adulterants in amounts varying from 5-25 grams to determine the smallest amount necessary to give concordant results.

Approved.

(4) That samples be sent out containing unknown adulterants for qualitative detection of the same.

Approved.

(5) That methods for the detection of peat dried at high temperature in feeding stuffs be investigated.

Approved.

(6) That the maximum percentage of foreign materials permissible in mill by-products be investigated.

Approved.

SUGAR.

No report or recommendations.

WATER IN FOODS AND FEEDING STUFFS.

It is recommended—

(1) That the investigation of desiccating agents, with and without increase in temperature, be continued.

Approved.

(2) That moisture determination by the vacuum method over sulphuric acid (U. S. Bur. Chem. Bull. 122, p. 219) be made an optional official method.

Recommended in 1913 and unconsidered in 1914.

ORGANIC AND INORGANIC PHOSPHORUS IN FOODS, FEEDING STUFFS, AND DRUGS.

It is recommended—

(1) That the magnesia mixture method of E. B. Forbes and his associates for the determination of water-soluble inorganic phosphorus in animal substances, as described in this year's report, be adopted as an official method.

Approved for final action as official in 1915.

(2) That further effort be made on methods of separation of organic and inorganic phosphorus in vegetable substances.

Approved.

SEPARATION OF NITROGENOUS SUBSTANCES (MEAT PROTEINS).

It is recommended—

(1) That work on ammonia and amino nitrogen be continued.

Approved.

(2) That the Folin method for estimating creatin and creatinin in meat and beef extract, as given in the 1913 report, be made official.

Adopted, final action.

(3) That the proposed method for determining the coagulable protein in meats (U. S. Bur. Chem. Bull. 107, Rev., p. 108, 7 (d)), be made optional.

Adopted, final action.

SEPARATION OF NITROGENOUS SUBSTANCES (VEGETABLE PROTEINS).

No report or recommendations.

SEPARATION OF NITROGENOUS SUBSTANCES (MILK AND CHEESE).

It is recommended—

(1) That studies be made leading to the adoption of methods for the determination of the noncasein proteins and the products of protein decomposition in milk.

Approved.

TESTING CHEMICAL REAGENTS.

It is recommended—

(1) That the double distillation method (B including C) for the determination of nitrogen in crude caustic soda, the modified lead method for the determination of molybdenum trioxid in molybdic acid, and the method for the determination of solids in ethyl ether non-volatile at 100°C., as given in this year's report, be studied further with the view of incorporating them in the official methods for nitrogen, phosphoric acid, and crude fat or ether extract, respectively.

Approved.

(2) That the direct titration method and the Tillmans and Heublein method for the determination of sodium hydroxid and sodium carbonate in crude caustic soda and the method for the determination of P_2O_5 in molybdic acid, as given in this year's report, be studied coöperatively this coming year, and that the effect of the presence of sulphates on the determination of molybdenum trioxid in molybdic acid, together with the cause of the variations in the determination of molybdenum trioxid in molybdic acid by the modified lead method, be studied further.

Approved.

(3) That the study of the nature and amount of the impurities in commercial citric acid be discontinued for the present.

Left for the next referee to decide.

TANNIN.

No report or recommendations.

MEDICINAL PLANTS AND DRUGS.

It is recommended—

(1) That the work on medicated soft drinks, especially the methods for estimating phosphoric acid and glycerin, be continued.

Approved.

(2) That the method for the determination of caffen and acetanilid, as given in this year's report, be adopted as a provisional method.

Approved by the committee on recommendations of referees for final action as provisional in 1915.

(3) That the method for the determination of caffen and antipyrin, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(4) That the method for the determination of caffen and acetphenetidin (U. S. Bur. Chem. Bull. 162, pp. 193-194), as slightly modified in this year's report, be adopted as an official method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as official in 1915.

(5) That the method for the determination of acetanilid and acetphenetidin, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(6) That the method for the determination of acetanilid and sodium salicylate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(7) That the method for the determination of acetanilid and quinin sulphate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(8) That the method for the determination of acetphenetidin and salol, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(9) That the method for the determination of caffeine, acetanilid, and sodium bicarbonate (U. S. Bur. Chem. Bull. 132, pp. 197-198), as slightly modified in this year's report, be adopted as an official method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as official in 1915.

(10) That the method for the determination of caffeine, acetanilid, sodium bicarbonate, and sodium bromid, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(11) That the method for the determination of caffeine, acetanilid, sodium bicarbonate, and sodium salicylate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(12) That the method for the determination of caffeine, acetanilid, sodium bicarbonate, and codein sulphate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(13) That the method for the determination of caffeine, acetanilid, sodium bicarbonate, and quinin sulphate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(14) That the method for the determination of caffeine, acetanilid, sodium bicarbonate, sodium salicylate, and codein sulphate as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(15) That the method for the determination of caffeine, acetanilid, and sodium salicylate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(16) That the method for the determination of alcohol, caffeine, acetanilid, and sodium salicylate (U. S. Bur. Chem. Bull. 152, pp. 236-239), as slightly modified in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(17) That the method for the determination of caffeine, acetanilid, and codein sulphate (U. S. Bur. Chem. Bull. 162, pp. 195-196), as slightly modified in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(18) That the method for the determination of caffeine, acetanilid, and antipyrin, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(19) That the method for the determination of acetanilid, sodium salicylate, and codein sulphate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(20) That the method for the determination of antipyrin, acetphenetidin, and codein sulphate (U. S. Bur. Chem. Bull. 162, pp. 197-198), as slightly modified in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(21) That the method for the determination of acetanilid, quinin sulphate, and morphin sulphate, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(22) That the method for the determination of caffeine, acetanilid, quinin sulphate, and morphin sulphate (U. S. Bur. Chem. Bull. 162, pp. 200-201), as slightly modified in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(23) That the method for the determination of acetphenetidin and heroin, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(24) That the method for the determination of caffen, acetphenetidin and codein sulphate (U. S. Bur. Chem. Bull. 152, pp. 239-240), as slightly modified in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(25) That the method for the determination of antipyrin and acetanilid (U. S. Bur. Chem. Bull. 152, pp. 240-241), as slightly modified in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(26) That the method for the detection of thujone in liqueurs, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

(27) That the method for the determination of santonin in Levant wormseed, as given in this year's report, be adopted as a provisional method.

Approved by the committees on recommendations of referees and editing methods of analysis for final action as provisional in 1915.

REPORT OF COMMITTEE C ON RECOMMENDATIONS OF REFEREES.

By H. E. BARNARD (State Board of Health, Indianapolis, Ind.), *Chairman*,¹
(Food adulteration.)

COLORS.

It is recommended—

(1) That the investigation of methods for the separation and identification of the important natural coloring matters be continued.

Approved.

SACCHARINE PRODUCTS.

It is recommended—

(1) That the work on the detection of artificial invert sugar in honey be continued for another year, studying the tests which proved the most satisfactory in the hands of all the collaborators, namely, (1) the original Fiehe test allowing the color to develop for twenty-four hours; (2) Hartmann's modification of the Fiehe test, noting the color immediately after the addition of the reagent; (3) Bryan's modification of the Fiehe test, changing the word "vigorously" to "gently"; (4) Feder's anilin chlorid test; with a view of adopting these methods as provisional in 1915.

Approved.

¹ Presented by H. E. Bishop.

FRUIT PRODUCTS.

It is recommended—

(1) That the proposed methods for the determination of malic and citric acids, as given in this year's report, be given further study.

Approved.

WINE.

It is recommended—

(1) That the proposed method for the determination of total tartaric acid, as given in the 1913 report, be studied further with regard to its applicability to red wines.

Approved.

(2) That the suggested use of Rochelle salt addition instead of tartaric acid, as provided in the Hartmann and Eoff method, be further studied.

Approved.

BEER.

It is recommended—

(1) That Method 3 for the determination of phosphoric acid in beer by the addition of calcium acetate, and subsequent ashing, as given in the 1913 report, be adopted as a provisional method in place of the direct volumetric determination with uranium acetate.

Adopted, final action.

DISTILLED LIQUORS.

It is recommended—

(1) That the Allen-Marquardt method for fusel oil determination (U. S. Bur. Chem. Bull. 107, Rev., p. 98 (b)), be modified as follows: In the fourth line after "is collected" add "Whenever aldehydes are present in excess of 15 parts per 100,000, add to the distillate 0.5 gram of metaphenylene diamin hydrochlorid, reflux for an hour, distil 100 cc., add 25 cc. of water and continue distillation until an additional 25 cc. is collected."

Approved for further study with collaboration.

(2) That the associate referee be directed to confer with chemists engaged in spirits analysis, to do such collaborative work as is necessary, and to submit at the next meeting a revised Allen-Marquardt method containing such changes in details and manipulation as experience justifies.

Approved.

VINEGAR.

It is recommended—

(1) That Methods 6, 11, 15, 16, and 17, as given in U. S. Bur. Chem. Bull. 152, pp. 126 and 127, be adopted provisionally.

Approved for final action as provisional in 1915.

(2) That Methods 10 and 20, as given in U. S. Bur. Chem. Bull. 152, pp. 126 and 127, be given further study following the suggestions in U. S. Bur. of Chem. Bull. 162, p. 81.

Approved.

FLAVORING EXTRACTS.

It is recommended—

(1) That the saponification method of Hortvet and West for winter-green extract, as described in the *J. Ind. Eng. Chem.* 1909, **1**: pp. 84-95, and slightly modified in U. S. Bur. Chem. Bull. 152, p. 141, be adopted as a provisional method.

Approved for final action as provisional in 1915.

(2) That the method of Hortvet and West for anise and nutmeg extracts, as described in the *J. Ind. Eng. Chem.* 1909, **1**: pp. 84-95, be given further study.

Approved.

(3) That the slight modification of the Howard-Mitchell method for peppermint extract, spearmint extract, and other extracts, be given further study.

Approved.

SPICES.

It is recommended—

(1) That final action regarding the lactic and citric acid methods be withheld pending further study and collection of data.

Approved.

(2) That the methods for determining insoluble solids and sand, as applied to ketchup (U. S. Bur. Chem. Bull. 162, pp. 128-129), be adopted as provisional methods.

Approved for final action as provisional in 1915.

MEAT AND FISH.

It is recommended—

(1) That the recommendations of 1913 be continued.

Approved.

FATS AND OILS.

It is recommended—

(1) That the Bureau of Animal Industry method for the detection of phytosterol in fats (U. S. B. A. I. Cir. 212) be adopted as a provisional method.

Approved for final action as provisional in 1915.

(2) That the digitonin method, as described in this year's report, be adopted as a provisional method.

Approved for final action as provisional in 1915.

DAIRY PRODUCTS.

It is recommended—

(1) That the method proposed in 1911 (U. S. Bur. Chem. Bull. 152, p. 101; Cir. 90, p. 10) as applied to evaporated milk, sweetened condensed milk, thin cream, and ice cream, and recommended in 1913 for final adoption as provisional in 1914, be given further study.

Approved.

(2) That special attention be given to the Roesse-Gottlieb method with a view of correcting its deficiencies, if such exist, particularly with reference to its use in sweetened and unsweetened condensed milk.

Approved.

(3) That the modification for rich cream (U. S. Bur. Chem. Circ. 108, p. 12) be given further study with a view of its final adoption as provisional in 1914.

Recommended in 1913 and unconsidered in 1914.

CEREAL PRODUCTS.

It is recommended—

(1) That the recommendations of 1913 be continued.

Approved.

VEGETABLES.

It is recommended—

(1) That the recommendation of 1913 be continued.

Approved.

COCOA AND COCOA PRODUCTS.

It is recommended—

(1) That the associate referee for the coming year make a study of the manufacture of milk chocolate with the view of finding out whether or not the casein is rendered insoluble in the reagents by different methods of manufacture.

Approved.

TEA AND COFFEE.

It is recommended—

(1) That the Fuller method for the determination of caffeine in tea and coffee be studied further with a view of improving the method of extraction and filtration.

Approved.

(2) That the Görtter method be studied further with a view of purifying the caffeine with sodium carbonate solution for direct weighing.

Approved.

(3) That the combination method, using the Fuller and Görtter methods, as proposed by Hanson in this year's report, be studied.

Approved.

(4) That the modified Stahl Schmidt method be given another trial for the determination of thein in tea.

Approved.

PRESERVATIVES.

It is recommended—

(1) That the Fincke method for formic acid (*Biochem. Z.*, 1913, **51**: 253) be adopted as a provisional method.

Approved for final action as provisional in 1915.

(2) That the natural occurrences of formic acid in food products be further investigated.

Approved.

(3) That the Wegner method (*Z. anal. Chem.*, 1903, **42**: 427) be submitted to trial as a confirmatory test and that steps be taken to secure a reliable quantitative method for the determination of saccharin in foods.

Approved.

HEAVY METALS IN FOODS.

It is recommended—

(1) That the methods for the determination of lead in baking powder and baking powder materials be made the subject of further study, with the coöperation, if possible, of the associate referee on baking powder.

Approved.

(2) That further study be made of the Gutzeit determination for arsenic as carried on this year, especially as to the proper strength of acid to be used.

Approved.

(3) That a study of some modification of the Marsh method for the determination of arsenic be made.

Approved.

(4) That the gravimetric tin method and Baker's volumetric tin method be studied further.

Approved.

(5) That the volumetric tin method of Bloomberg and Lourie be studied further.

Approved.

(6) That methods for the determination of copper, zinc, nickel, and aluminium in food products be made the subject of study by the association as soon as possible.

Approved.

BAKING POWDER.

It is recommended—

(1) That a comparative study be made of the Seeker, Exner, and Remington method for the determination of lead.

Approved.

A motion, made by William Frear, that the Chairmen of Committees A, B and C meet to discuss the method of preliminary research and the conduct of the work of the association, was carried.

REPORT OF COMMITTEE ON RESOLUTIONS.

By WILLIAM FREAR (State College, Pa.), *Chairman*.

Resolved, That, by the death, on July 31, 1914, of Prof. Francis H. Storer, Emeritus Professor of Agricultural Chemistry of Bussey Institution, Harvard University, America has lost one of her pioneers in the application of chemistry to the art of agriculture, a teacher whose widely inclusive studies, instructive writings, and earnestness in his chosen field, have done much to inspire the later generation of American agricultural chemists with a realization of the value of their science as applied to agriculture, and to impress upon the minds of the farming public also its importance in the solution of farm problems.

REPORT OF THE AUDITING COMMITTEE.

By R. T. DAVIDSON (Polytechnic Institute, Blacksburg, Va.), *Chairman*.

The following report by the secretary-treasurer was examined by the auditing committee and found to be correct:

REPORT OF THE SECRETARY-TREASURER FOR THE YEAR ENDING NOVEMBER 15, 1914.

Receipts:

Balance November 15, 1913.....	\$120.02
Dues for the year 1912 to 1913 received from 3 States, (North Dakota, Tennessee, and Vermont) after the report for that year was submitted.....	6.00
Dues for the year ending November 15, 1914, from 58 Federal and State organizations.....	116.00
Total receipts.....	\$242.02

Expenditures:

Telephone calls, Raleigh Hotel, 1913 meeting.....	\$0.65
Tips, Raleigh Hotel, 1913 meeting.....	3.00
January 2, 1914. Stamps.....	2.00
January 5, 1914. 500 Circulars, 300 one-cent envelopes, plain.....	12.85
January 14, 1914. 1 rubber stamp.....	0.70
June 15, 1914. 300 two-cent envelopes.....	6.47
June 29, 1914. 350 letter circulars.....	3.75
July 21, 1914. 500 stamped envelopes.....	11.00
July 30, 1914. 800 announcements.....	16.75
Expressage on badges for 1914 meeting.....	0.42
300 badges from Bastian Brothers Co.....	23.00
Total expenditures.....	\$80.59
Balance November 16, 1914.....	\$161.43

Respectfully submitted,
(Signed) C. L. ALSBERG,
Secretary-Treasurer.

Examined and found correct:

(Signed) R. J. DAVIDSON, *Chairman, Auditing Committee.*

H. C. Lythgoe made a motion, which was carried, that the constitution be so amended that municipal chemists be eligible to membership in the association, and that the matter be referred to the proper committee.

REPORT OF COMMITTEE ON NOMINATIONS.

By W. A. WITHERS (College of Agriculture, Raleigh, N. C.), *Chairman*.

The committee submitted the following nominations for officers for the year ending November 1915: President, C. H. Jones, of Vermont; Vice-President, R. N. Brackett, of South Carolina; Secretary-Treasurer, C. L. Alsberg, of Washington, D. C.; additional members of the executive committee, Julius Hortvet of Minnesota, and E. F. Ladd of North Dakota.

The secretary was instructed to cast the unanimous ballot of the association for these officers.

No report was made by the referee on tannin.

The subject of tannin was dropped from further study by the association.

The association adjourned at 12.30 P.M. to reassemble at 1.30 P.M.

THIRD DAY.

WEDNESDAY—AFTERNOON SESSION.

REPORT ON MEDICINAL PLANTS AND DRUGS.

By W. O. EMERY (Bureau of Chemistry, Washington, D. C.), *Referee*.

RECOMMENDATIONS.

It is recommended—

(1) That the work on medicated soft drinks, especially the methods for estimating phosphoric acid and glycerin, be continued.

(2) That the following method for the estimation of caffein and acetanilid in mixtures, be adopted as official:

CAFFEIN AND ACETANILID.

(W. O. EMERY METHOD.)

Preliminary Treatment.

I. If the sample is already in powder form, reduce to the finest possible condition and uniformity by rubbing in a mortar, then keep in a tightly corked tube or flask until required for analysis. Powders in paper, cachet or capsule containers are frequently of such fineness as to require little further trituration except to produce a more uniform product, in which event the above treatment should be performed carefully prior to the weighing of the sample. On a small tared filter (5.5 cm.) weigh out 0.3-0.5 gram of the product thus prepared, or, if preferred, an amount equal to, or a multiple of, the average unit dose (previously ascertained by weighing collectively 20 or more such doses), wash with successive small portions of residue-free chloroform, 30-50 cc., sufficient at least to insure complete extraction. Collect the solvent in a 200 cc. Erlenmeyer, then distil by gentle heat to about 10 cc., using in this and similar operations, if available, a small spray trap, substantially like that shown in U. S. Bur. Chem. Bull. 152, p. 239. Add 10 cc. of dilute sulphuric acid (1 to 10), remove to a steam-bath and proceed as directed under caffein.

II. If the caffein is present in the citrated form, or the composition of the mixture is such as to preclude complete extraction by the filter method, weigh out on a metal scoop or watch glass the desired amount of the prepared sample, transfer to a separatory funnel of the Squibb type, add 50 cc. of chloroform and 20 cc. of water, shake vigorously and after clearing, draw off the lower layer through a small dry filter into a 200 cc. Erlenmeyer. In the case of coated tablets and pills, take one or more for each determination, first crushing them in a mortar before treatment in a separator. Repeat the extraction twice with two 50 cc. portions of chloroform, on the completion of which any caffein-acetanilid mixture observable about the apex of the delivery tube of the separator or edge of the filter and tip of the funnel should be recovered very carefully by judicious washing with chloroform, such washings being

subsequently united with the main portion. Distil the combined chloroformic portions to about 10 cc., then proceed as directed under caffeine.

III. If the product under examination be an aqueous-alcoholic solution, evaporate a measured quantity on the steam-bath (or take an aliquot of the residue from an alcohol determination) until most of the alcohol has been expelled, transfer to a separatory funnel by pouring and rinsing with a minimum of water so that the final volume does not greatly exceed 20 cc., then, in order to regenerate any acetanilid possibly hydrolyzed during evaporation, add a little solid sodium bicarbonate, 1 drop of acetic anhydrid and 50 cc. of chloroform, shake vigorously and, after clearing, draw off the solvent through a filter into a 200 cc. Erlenmeyer. If the preparation contains alkaloids, acidify with a few drops of dilute sulphuric acid immediately after the above prescribed acetylation and prior to treatment with chloroform, in order to insure retention of such basic material in aqueous solution. Repeat the extraction twice with two 50 cc. portions of chloroform and treat the solvent substantially as in the two preceding cases preliminary to hydrolysis as outlined under caffeine.

A gross separation of the active ingredients from vehicular matter having been effected in accordance with one of the above procedures, the next step consists in the conversion of acetanilid into acetic acid and anilin sulphate, from which caffeine is readily separable by means of chloroform, the anilin salt being under the conditions of the experiment insoluble in this medium. The acid hydrolysis of acetanilid leaves caffeine intact.

Determination.

Caffein ($C_8H_{10}N_4O_2$), *melting point about 235°C.*—Digest on the steam-bath the aqueous acid mixture containing caffeine and acetanilid until the contents of the flask are reduced to 5 cc., add 10 cc. of water and continue the digestion until the liquid is again reduced to 5 cc., cool, pour and rinse into a separatory funnel with a minimum of water, so that the final volume does not greatly exceed 20 cc. Add 50 cc. of chloroform, extract in the usual way and after clearing withdraw the lower layer through a small dry filter into a 200 cc. Erlenmeyer. Repeat the extraction with two 50 cc. portions of chloroform. On completion of the third and final extraction, recover carefully all traces of caffeine observable about the tip of the delivery tube and the edge of the filter, then distil the combined solvent to about 10 cc., finally transfer the residual liquid, by pouring and washing with chloroform, to a tared beaker or crystallizing dish, and allow the solution to evaporate spontaneously or by gentle heat and blast to apparent dryness. Cool and weigh until constant.

Acetanilid ($C_8H_8NHCOCH_3$), *melting point 113°C.*—Draw off into the Erlenmeyer the aqueous-acid solution of anilin sulphate remaining in the separator used in effecting hydrolysis, rinse several times with 5 cc. portions of water to insure complete removal of former contents, and then heat the liquid for 10 minutes on the steam-bath to expel all traces of chloroform. Wash the filter, used in the preceding operation to dry the chloroformic solution of caffeine, once with 5 cc. of water, and add later to the main solution of anilin sulphate. Add 10 cc. of concentrated hydrochloric acid, then run in a standard solution of bromin (1 cc. of which is equivalent to about 5 or 10 mg. of acetanilid) until a faint yellow coloration persists. While adding this reagent, or after each small addition thereof, rotate the flask sufficiently to agglomerate the precipitated tribromoanilin, thus clarifying the supernatant liquid and rendering the state of titration more apparent to the operator. Multiply the number of cc. required to complete the precipitation by the value of 1 cc. in terms of acetanilid in order to ascertain the quantity of this substance originally present in the sample taken.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

Tared filters for use in this and similar work may be conveniently prepared as follows: Fold, adjust dry in a short-stemmed funnel of about 1.5 inches in diameter, moisten and fit carefully so that the entire rim of filter closes snugly onto the glass. After drying, keep in a balance case or other convenient receptacle and when needed weigh on a small glass tripod, an appliance easily constructed in the laboratory.

It is self-evident that all manipulations with tared filters at the balance should be performed under comparable conditions. If, for example, the first weight is taken when the hygrometer indicates a humidity of 50 per cent, the second weighing should clearly be made only when the moisture content of the air is approximately the same. This is indispensable when it is desired to estimate the quantity of chloroform-insoluble residue (as sodium bicarbonate in acetanilid compound) or excipient (starch, sugar, talc, etc.) employed, as in tablets containing but one active ingredient. Experience has shown, however, that a variation in humidity up to 5 per cent between the 2 weighings would not be productive of material error.

It not infrequently happens that certain brands of compressed tablets, when treated on the filter according to Procedure I do not yield to chloroform all the drug actually present in the mass. Such failure in apparently simple combinations to give up all their active principles to a suitable solvent is unquestionably due to the nature of excipient employed in the process of granulation. Starch paste, sirup, dextrin, gum, etc., as ordinarily applied, may easily and doubtless do dissolve and on drying hold or occlude appreciable quantities of any substance possessing a fair degree of solubility in aqueous media. In support of this assumption may be mentioned the fact that, in the case of certain caffein-acetanilid tablets, the chloroform-insoluble residue even after exhaustive treatment with the solvent still showed traces of the active principles in question, while in tablets containing the more insoluble acetphenetidin, no such resistance to extraction was observed. One should therefore not be surprised, in applying the first or filter treatment to caffein-acetanilid mixtures, to discover appreciable amounts of these drugs persisting in the chloroform-insoluble residue. While the quantity thus occluded is usually so small as to be almost negligible, the careful analyst will undoubtedly desire to check the accuracy of the first procedure by applying the second also, involving the use of a separatory funnel. In operating, therefore, according to Procedure I it is wise to apply the isocyanid test to the chloroform-insoluble residue, in order to prove the absence of even traces of acetanilid, which may be present in the occluded state in the powdered mass. As a means of control or in very exact work, it is recommended that the sample be subjected to both Procedures I and II.

Before making an extraction in a separatory funnel, the valve should be "locked" with a drop of water in order to reduce the tendency to loss from capillarity. All chloroform used in drug analysis should be carefully redistilled, unless it is positively known to be a residue-free product. It should also be neutral in reaction. All cork stoppers employed in connection with the distillation of chloroform should have been previously digested with this solvent a sufficient length of time to remove all extractives, which would otherwise contaminate the substances in course of estimation, and thus vitiate the results.

In the operation of withdrawing chloroform from a separatory funnel on completion of an extraction, it will frequently be found advantageous to free the solvent as much as possible from mechanically suspended water, which would otherwise be carried along and in the end contaminate, by virtue of dissolved salts, the substance

it is desired to estimate. The chloroform is accordingly made to pass first, through a small, dry pledget of chloroform-washed cotton, pushed up into the delivery tube of separator, and second, a small (5.5 cm.) dry filter prior to reception in an Erlenmeyer from which distillation is effected. After each withdrawal of chloroform, the cotton is removed to the separator by means of a wire, and, on completion of the following extraction, a fresh pledget substituted in its place. The filter is allowed to dry completely between extractions.

Preparations containing, in addition to caffein and acetanilid, powdered cinnamon, celery seed, ginger or other vegetable products, yield to chloroform certain oils, fats, waxes, resins, pigments and other extractives, which in due course appear either in suspension or solution after the caffein-acetanilid mixture has been subjected to digestion, and finally also contaminate the caffein as first isolated. Any suspended impurities may be eliminated readily by passing the aqueous-acid liquid through a small moistened filter tightly packed in a funnel, prior to extraction with chloroform. Should the caffein, as first recovered and weighed, be deeply colored or apparently contaminated with foreign matter, it may be purified in the following way: Dissolve in very dilute sulphuric acid, pass, if necessary, through a moistened filter to eliminate suspended impurities, add to the clear solution 15-20 cc. of Wagner's reagent, sufficient at least to render the supernatant liquid distinctly claret in color, stir, then allow to stand an hour. Filter and wash the peroxid with a few cc. of iodine solution, transfer both filter and precipitate to a separatory funnel, using for this purpose not more than 20 cc. of water, decolorize with a crystal of thiosulphate, then extract with three 50 cc. portions of chloroform and proceed as directed under caffein.

To prepare a standard solution of bromine, add bromine in slight excess to a concentrated aqueous solution of caustic potash (50 grams), dilute with water to dissolve any separated salts, boil to expel excess of bromine, and finally dilute to 1 liter. Standardize this solution with weighed amounts of pure aniline, or better with recrystallized and dried acetanilid, and finally adjust by proper dilution until 1 cc. is equivalent to 5 or 10 mg. of acetanilid.

Caffeine and acetanilid are the two principal ingredients of the preparation known as "acetanilid compound", a further constituent being sodium bicarbonate. The last named substance appears as the chloroform-insoluble residue and may be accurately determined by titrating such residue, or one obtained by igniting a portion of the original mixture, with standard acid in the presence of congo red as indicator. The bicarbonate may likewise be determined by igniting the original sample, or the chloroform-insoluble residue, with sulphuric acid and weighing the resulting sodium sulphate.

Should the "acetanilid compound" be combined with sodium bromide, the latter substance, in the absence of other halides, is easily estimated volumetrically by means of standard silver nitrate in combination with potassium thiocyanide and ferric ammonium sulphate.

(3) That the following method for the estimation of caffeine and acetphenetidine (phenacetin) in mixtures be adopted as provisional:

CAFFEIN AND ACETPHENETIDIN.

(W. O. EMERY METHOD.)

Preliminary Treatment.

With preparations containing acetphenetidin (phenacetin) instead of acetanilid, but otherwise identical, a gross separation of the caffein-acetphenetidin mixture is effected by practically the same preliminary treatment as that accorded caffein-acetanilid. Only in the matter of subsequent hydrolysis and regeneration of acetphenetidin is it necessary to observe the most careful circumspection as regards minor details involved in the various operations.

Caffein.—After distilling the chloroformic solution of caffein-acetphenetidin to about 10 cc., add 10 cc. of dilute sulphuric acid (1 to 10), then remove to the steam-bath and digest until the liquid is reduced to about 5 cc. Dilute with 10 cc. of water and continue the digestion until the volume is again reduced to 5 cc., then add a second and final 10 cc. of water and continue heating until the residual liquid amounts to 8–10 cc. If, during the process of digestion, particles of acetphenetidin persist on the sides of the flask, add, from time to time, a few drops of chloroform in order to bring such particles more completely within the radius of hydrolytic action. Cool and transfer to a separatory funnel by pouring and rinsing with water, until the final volume of liquid does not greatly exceed 20 cc. Extract, by means of vigorous shaking, with three 50 cc. portions of chloroform, manipulating the solvent and caffein substantially as under caffein-acetanilid treatment.

Acetphenetidin ($C_9H_9O_2N$, $C_6H_5NHCOCH_3$), *melting point* $135^\circ C$.—Wash the filter used to dry the chloroform in the preceding operation once with 5 cc. of water, receiving the latter in the separatory funnel containing the aqueous-acid solution of phenetidin sulphate. Treat with successive small portions of solid sodium bicarbonate until, after complete neutralization of free acid, an excess of the former persists at the bottom of the mixture. Then add 50 cc. of chloroform and, for every 100 mg. of acetphenetidin known or believed to have been present, 5 drops of acetic anhydrid; shake vigorously for some time, allow to clear, then withdraw the chloroform into a second separator containing 5 cc. of water. Shake this mixture and, after clearing, pass the solvent through a small dry filter into a 200 cc. Erlenmeyer. Distil over about 40 cc. of the chloroform, make up to 50 cc. with fresh solvent and extract again. Withdraw to the second separator, wash and distil as before, about 50 cc., which are applied on the third and final extraction. Then distil the chloroform to about 10 cc., transfer the residue to a 50 cc. tared crystallizing dish or beaker, by pouring and rinsing with sufficient fresh solvent, evaporate on the steam-bath to apparent dryness, finally removing any considerable excess of acetic anhydrid by repeated additions of 1 cc. of fresh chloroform, to which has been added a drop of alcohol. The regenerated acetphenetidin should finally appear as a whitish, crystalline mass with a faint acetous odor. The latter will completely disappear on standing some hours in the open, or in a vacuum desiccator over lime. Weigh from time to time until the final weight differs from the preceding not more than 0.5 mg.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

Acetphenetidin, owing to its greater insolubility in aqueous-acid media, is correspondingly more difficult of hydrolysis than acetanilid. In order therefore to insure complete conversion to phenetidin sulphate, special attention should be directed to the end that any crystalline deposits (acetphenetidin), which may appear in the beginning of the operation on the sides of the flask just above the liquid, should be forced back into the hot liquid by an occasional gentle rotation of the

flask or, better, by the occasional addition of a few drops of chloroform. It is self-evident that, if hydrolysis is incomplete, subsequent extraction with chloroform will remove not only caffeine but also any acetphenetidin that may have escaped hydrolytic action, and thus vitiate the results for caffeine. Great care too should be given the degree of evaporation. Should the aqueous-acid solution and suspension of caffeine-acetphenetidin be concentrated far beyond the limits indicated in the method, more or less phenetidin sulphonate is likely to be formed, which later resists acetylation and conversion to acetphenetidin.

(4) That the method for the estimation of caffeine and antipyrin in mixtures,¹ as given in this year's report, be adopted as provisional:

CAFFEIN AND ANTIPYRIN.

(W. O. EMERY AND S. PALKIN METHOD.)

Antipyrin ($C_{11}H_{12}N_2O$), melting point $113^{\circ}C$.—Transfer about 0.25 gram of the caffeine-antipyrin mixture to a 150 cc. separatory funnel by means of two 5 cc. portions of alcohol-free chloroform followed by 10 cc. of water, add 1 gram of sodium bicarbonate and 10–15 cc. of fifth-normal iodine (or double the quantity of tenth-normal iodine), this reagent being applied in successive small portions and the mixture shaken vigorously 15–20 seconds after each addition. When all the iodine has been thus added, a decided excess of this reagent should be apparent in the liquid after a final vigorous shaking of 1 minute, in which event all the antipyrin will have been converted into the moniodo derivative. If, however, all the iodine previously applied should appear to have been expended, a little more must be added to insure an excess, and the mixture again shaken. Now discharge the uncombined iodine by means of a small crystal of sodium thiosulphate, add 15 cc. of redistilled U. S. P. chloroform, shaking vigorously for 1 minute. After clearing, wash the solvent in a second separator with 5 cc. of water, finally passing through a small dry filter into a tared 50 cc. beaker and evaporating to apparent dryness on the steam-bath, accelerating this operation with an air blast. Repeat the extraction with two (three, in case tenth-normal iodine is used) 25 cc. portions of chloroform, and wash, filter and evaporate each portion in rotation as in the first instance. Recover any crystalline product separating about the tips of the delivery tube and the funnel, and the edge of the filter, by judicious washing with chloroform. Dry the nearly colorless crystalline residue of caffeine and iodoantipyrin for 30 minutes at $105^{\circ}C$., then cool and weigh. Designate this weight *A*.

Dissolve the composite residue in 5 cc. of glacial acetic acid, add 10 cc. of saturated aqueous sulphur dioxide solution, then transfer the resulting liquid by diluting, pouring and rinsing with hot water to a 400–500 cc. beaker until the final volume amounts to about 200 cc. Add aqueous silver nitrate solution (about 0.3 gram of silver nitrate), sufficient at least to precipitate all the halogen, follow with a few drops of nitric acid, then heat nearly to boiling, stirring the while in order to agglomerate the silver iodide. Add 15 cc. of concentrated nitric acid, cover the beaker with a watch glass and boil gently for 5 minutes. Filter by decantation through a tared Gooch, washing the precipitate once with a little alcohol, then with two 100 cc. portions of boiling water, and finally transfer the iodide completely to the crucible. Wash several times with hot water and again with alcohol to remove traces of organic matter, prior to drying for 30 minutes in the air-bath at $110^{\circ}C$. Cool and weigh. The weight of silver iodide multiplied by the factor 0.8012 yields the quantity of antipyrin present in the mixture.

¹ *J. Ind. Eng. Chem.*, 1915, 7: 519.

Caffein.—Ascertain the quantity of caffein by subtracting the product obtained by multiplying the weight of silver iodid by the factor 1.3374 from the weight of residue A.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

In practice, the preliminary or gross separation of the caffein-antipyrin mixture from the usual excipients of tablet and pill combinations is most conveniently effected by extraction of the finely powdered mass with chloroform on a tared filter, this solvent, together with its attendant alcohol, being subsequently expelled completely, first by gentle distillation and finally by evaporation on the steam-bath, prior to proceeding with the separation proper. Likewise in the case of alcoholic preparations containing caffein and antipyrin as essential ingredients, the recovery of these two substances is best made after evaporation of the alcohol on the steam-bath. The residual menstruum is thereupon exhausted with chloroform in the usual way, the resulting solution of caffein and antipyrin being treated precisely like the one obtained from tablet or pill combinations.

The use of alcohol-free chloroform in connection with the halogenation of antipyrin is necessary in order to preclude the possible formation of iodoform, the presence of which in the composite residue A would naturally vitiate the analytical findings.

The application of iodine in small portions appears to favor the production of a purer iodoantipyrin than when this reagent is added all at one time, as evidenced by the color of the caffein-antipyrin residue. In order to free the latter from all contaminating volatile products under the conditions outlined in the method, particular attention should be given to actual working conditions of the drying oven,¹ since incomplete or improper desiccation must lead necessarily to widely divergent caffein values, while heating at temperatures materially higher than 105°C. is found to exert an unfavorable influence on the recovery of both caffein and antipyrin.

The color changes following the addition of strong nitric acid are quite characteristic, passing from colorless through deep red to pale yellow in the course of 5 minutes' boiling. In view of this somewhat radical treatment to which the caffein is likewise subjected, any direct determination of this substance becomes impractical. The quantity of caffein is ascertained indirectly therefore with a reasonable degree of accuracy by subtracting the weight of iodoantipyrin (calculated) from that of the composite residue A.

In the analysis of a mixture containing caffein, antipyrin, acetanilid and sodium salicylate, the following essential steps are suggested in effecting a separation:

(1) Extraction of caffein, acetanilid and antipyrin from the aqueous soda solution.

(2) Hydrolytic treatment of the three substances eliminated with sulphuric acid preliminary to the isolation of caffein and antipyrin, which are thereupon subjected to the procedure just described.

(5) That the following method for the estimation of acetanilid and acetphenetidin (phenacetin) in admixture,² be adopted as provisional:

¹ *J. Ind. Eng. Chem.*, 1914, 6: 585.

² *Ibid.*, 665.

ACETANILID AND ACETPHENETIDIN.

(W. O. EMERY METHOD.)

Acetphenetidin.—Introduce 0.2 gram of the acetphenetidin-acetanilid mixture into a small (50 cc.) lipped Erlenmeyer flask, add 2 cc. of glacial acetic acid, heat gently over wire gauze to complete solution, then dilute with 40 cc. of water, previously warmed to 70°C. Transfer the clear acetous liquid to a glass stoppered, graduated 100 cc. flask, by pouring and careful washing of the flask with two 10 cc. portions of warm (40°C.) water into which have been run previously from a burette 25 cc. of standard iodine of a strength slightly above N/5 and warmed to 40°C. Rotate the resulting menstruum to uniformity, closing the flask meanwhile, then add 3 cc. of concentrated hydrochloric acid, close the flask anew and continue the rotation until copious crystallization is apparent, then set aside to cool. If the ratio of acetphenetidin to acetanilid is equal to or greater than 1, the formation of crystalline scales will be almost immediate on the addition of acid. As the proportion of acetanilid increases, however, the periodid is not only more inclined to maintain the liquid state, with the result that crystallization becomes proportionately slower, but its separation also from the menstruum itself is in a measure apparently retarded. In such cases, gentle agitation of the liquid or rotation of the flask in water warmed to 40°C. or less tends to promote the formation of crystals. When the contents of the flask have assumed the room temperature, fill with water to within 2-3 cc. of the mark, rotate to uniformity and allow to stand overnight. Fill to the mark with water, mix thoroughly, then after standing for 30 minutes withdraw a 50 cc. aliquot of the clear liquid by passing through a small (5.5 cm.) dry, closely fitted filter into a graduated 50 cc. flask, rejecting, however, about 15 cc. of the first runnings, the latter being received in any convenient container for eventual later use, along with the additional filtrate, for the recovery of acetanilid. Transfer the 50 cc. aliquot to a 200 cc. Erlenmeyer flask by pouring and washing and titrate with tenth-normal sodium thiosulphate.

If reference is had to the composition of the insoluble addition product, constituting the basis for the foregoing separation,



it will be noted that, for every molecule of acetphenetidin involved, 2 atoms of iodine are required, hence from a titrimetric standpoint 1 atom of iodine is equivalent to one-half molecule of acetphenetidin. If, therefore, the quantity of iodine expended in the formation of insoluble periodid is ascertained as the result of such titration, the quantity of acetphenetidin thereby involved is readily calculated from the expression,

$$\text{acetphenetidin} = I(0.008890 \times N),$$

in which 0.008890 represents the quantity of acetphenetidin in 1 cc. of a tenth-normal solution of this substance, N the normality of a standard thiosulphate employed, while I represents the number of cc. of such thiosulphate corresponding to the iodine entering into combination with phenacetin isolated as periodid.

The gravimetric determination of acetphenetidin may, if desired, be effected substantially as follows: Filter off the periodid, collect it on the filter and wash with 10-15 cc. of standard iodine solution, preferably by suction, then transfer, together with filter (likewise any particles of precipitate eventually remaining in the graduated flask), to a separatory funnel, using not over 50 cc. of water. After discharging both free and added iodine with a few small crystals of sodium sulphite, extract the liquid with three 50 cc. portions of chloroform, subsequently washing each portion

in a second separatory funnel with 5 cc. of water. After washing and clearing, withdraw the solvent through a small (5.5 cm.) dry filter into a 200 cc. Erlenmeyer flask, distil off most of the chloroform, transfer the residual 5-10 cc. to a small tared beaker or crystallizing dish by pouring and washing with fresh solvent. Evaporate to dryness on the steam-bath, cool, and weigh.

Acetanilid.—Should the combined weight of the acetphenetidin-acetanilid mixture be known, that of the latter constituent can be determined by difference or, if necessary, estimated directly from a second aliquot of the filtrate from the acetphenetidin periodid.

To this end, transfer 25-30 cc. of the clear liquid to a separatory funnel by means of a pipette, decolorize with sufficient solid sodium sulphite, and solid sodium bicarbonate in slight excess, follow with 1 or 2 drops of acetic anhydrid, then extract with three 60 cc. portions of chloroform, passing the solvent when cleared through a small, dry filter into a 200 cc. Erlenmeyer, from which the chloroform is distilled, by the aid of gentle heat, to about 20 cc. Then add 10 cc. of dilute sulphuric acid (1 to 10) and digest the product on the steam-bath until the aqueous residue has been reduced one half, add 20 cc. of water and continue the digestion for an hour, add a second 20 cc. portion of water and 10 cc. of concentrated hydrochloric acid, then titrate very slowly, drop by drop, with standard potassium bromid-bromate (1 cc. of which is equivalent to 5-10 mg. of acetanilid), until a faint yellow coloration persists. While adding this reagent, the flask should be rotated sufficiently to agglomerate the precipitated tribromoanilin and thus clarify the supernatant liquid. The number of cc. of standard bromid solution required to complete the precipitation, multiplied by the value of 1 cc. in terms of acetanilid, will give the quantity of this substance present in the aliquot taken.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

The preliminary or gross separation of acetphenetidin and acetanilid from complex mixtures is materially lengthened, if the preparation contains, in addition to those substances, caffein or antipyrin, or both, in which event it would be necessary to subject the mixture of four ingredients to hot digestion with dilute sulphuric acid in order to convert acetphenetidin and acetanilid to phenetidin and anilin sulphates, respectively, from which caffein and antipyrin could be easily separated by means of chloroform, after which acetphenetidin and acetanilid should be regenerated by treating the aqueous-acid solution of the corresponding sulphates with solid sodium bicarbonate in slight excess, thereupon with a few drops of acetic anhydrid, followed by extraction with chloroform.¹

In the operation of transferring the acetous solution of acetphenetidin-acetanilid mixture to the graduated flask containing standard iodine, great care must be exercised that none of the dissolved substances crystallize out as such during or after the transfer, either in the liquid or about the neck of the Erlenmeyer, since any undissolved acetphenetidin introduced into the iodine reagent would fail in obtaining its full complement of iodine, thus vitiating the determination. The necessary transfer is effected most conveniently, and indeed without loss, by the use of an Erlenmeyer provided with a lip, a form easily made in the laboratory from the ordinary type by heating the neck of the flask in a moderate blast flame, then by the aid of a file or other suitable instrument pulling down the rim to the desired pitch. Thus modified, the flask delivers aqueous solutions with little or no tendency to run down on the outside of the neck.

¹ U. S. Bur. Chem. Bull. 162, p. 197.

The standard solution of iodine employed in the foregoing separation and the one giving the best results has a strength slightly above fifth-normal; that is, a solution containing 30 grams of iodine and 40 grams of potassium iodide to the liter. To prepare, dissolve the potassium iodide in the least possible quantity of water, add the iodine and, after complete solution, dilute to 1 liter. Twenty-five cc. of this reagent, the volume taken for each determination, are standardized with a solution of sodium thiosulphate containing 30 grams to the liter, the value of which has been ascertained in turn by titration with very carefully purified iodine. The end point is best observed by adding to the liquid 1 or 2 drops of freshly prepared starch solution toward the close of the titration. In measuring off the standard iodine, make the readings by the aid of transmitted light. This is easily done by holding an electric bulb just back of the burette, thus bringing the lower meniscus into sharp relief.

For this and similar iodometric operations, very pure iodine was prepared by dissolving the commercial resublimed product in concentrated aqueous potassium iodide, pouring the clear liquid into a large volume of water, filtering and washing the finely precipitated iodine on a porous plate several times with water, then drying first in the air and finally in a desiccator over sulphuric acid, where it was kept for future use in a glass stoppered weighing tube. To standardize the sodium thiosulphate, weigh about 0.3 gram of pure iodine in a small glass capsule (about $\frac{1}{2}$ inch high and $\frac{5}{8}$ inch diameter), provided with a closely fitting glass cap or stopper, which, together with capsule and cover, is transferred to a 200 cc. Erlenmeyer, containing 0.5 gram of potassium iodide dissolved in 10 cc. of water. After complete solution, titrate the iodine with sodium thiosulphate, using 1 or 2 drops of starch solution as indicator.

In the qualitative examination of preparations or mixtures, of which acetphenetidine alone is a known or declared ingredient, suitable tests, such as are prescribed in the U. S. Pharmacopoeia or in Allen's Commercial Organic Analysis, should be applied in order to verify the presence or absence of acetanilide. For the identification of acetphenetidine, either alone or in admixture with acetanilide, the following test, in addition to those ordinarily employed for this substance, will be found of value: To 1-2 mg. of the sample in a test tube add a drop of acetic acid, 0.5 cc. of water and 1 cc. of tenth-normal iodine, warm the mixture to about 40°C., then add a drop of concentrated hydrochloric acid. Almost immediately if acetphenetidine alone is present, or on cooling and agitating the liquid, if the sample consists in large part of acetanilide, the iodine addition product of acetphenetidine separates in the form of reddish brown leaflets or needle-like crystals. In the presence of considerable acetanilide, the periodide first separates as minute oily globules, which on vigorous shaking gradually become crystalline aggregates. This test is so delicate that as little as 0.5 mg. of acetphenetidine may be detected, if alone, in the form of its characteristic periodide.

(6) That the following method for the estimation of acetphenetidine (phenacetin) and salol in mixtures,¹ be adopted as provisional:

ACETPHENETIDIN (PHENACETIN) AND SALOL.

(W. O. EMERY, G. C. SPENCER AND C. C. LEFEBVRE METHOD.)

The separation proper follows two distinct lines of procedure, involving, on the one hand, acid hydrolysis of acetphenetidine, on the other, alkaline hydrolysis of salol. In the first instance, a direct estimation of acetphenetidine only is contemplated.

¹ *J. Ind. Eng. Chem.*, 1915, **7**: 681.

plated, the salol being determined by difference in all cases where the combined weight of both constituents is known or can be ascertained conveniently. In this procedure, the acetphenetidin is converted temporarily into phenetidin sulphate and acetic acid, while in the case of salol partial volatilization results during the process of digestion. The phenetidin salt, by virtue of its insolubility in chloroform, is freed readily from further contaminating salol by treatment with that solvent, then by means of acetic anhydrid and sodium bicarbonate reconverted into acetphenetidin, the latter being isolated subsequently and weighed.

In the case of alkaline hydrolysis, the salol is converted first into sodium salicylate and phenolate, from which the acetphenetidin is separated easily and subsequently recovered by extraction with chloroform. The salol is thereupon treated in rotation with standard bromin solution in excess, hydrochloric acid, potassium iodid and sodium thiosulphate, with the net result that 12 atoms of bromin are expended on every molecule of salol. Titration of the iodine liberated by unexpended bromine affords sufficient data for determining the salol content.

Acid Hydrolysis.

Acetphenetidin ($C_2H_5OC_6H_4NHCOCH_3$), *melting point* $135^\circ C$.—Weigh, on a small (5.5 cm.) tared filter, an amount of the sample equal to, or a multiple of, the average weight of a unit dose, wash with successive small portions of chloroform (about 40 cc.), sufficient at least to insure complete extraction of all acetphenetidin and salol present in the mixture. Collect the solvent in a 100 cc. tared beaker, and evaporate by means of a blast to apparent dryness. During this operation, the beaker may with advantage be allowed to stand on a warm plate (50° – $60^\circ C$.) without undue loss of solid substance. Permit the beaker to remain 24 hours in the open at the ordinary temperature, weigh several times until practically constant, then transfer the crystalline residue to a 50 cc. lipped Erlenmeyer by dissolving in and washing with sufficient chloroform, evaporate the solvent by means of a blast and gentle heat, add 10 cc. of dilute sulphuric acid (1 to 10) and digest at full steam-bath heat until the liquid is reduced one half. Add 10 cc. of water and continue the digestion as before, adding finally a second 10 cc. of water and evaporating to 5 cc. as in the two preceding cases. Transfer the residual liquid to a small separatory funnel by pouring and washing with about 20 cc. of water and extract in rotation with 15, 10 and 5 cc. of chloroform, washing each portion as obtained with 5 cc. of water in a second separator, in order to recover traces of phenetidin sulphate possibly taken up by the chloroform, finally rejecting the latter since it contains all the salol not previously eliminated during the process of digestion.

Treat the aqueous-acid solution of phenetidin sulphate in the first separatory funnel, to which likewise the wash water used in the second separator has been transferred, with sodium bicarbonate preliminary to regenerating acetphenetidin, substantially as prescribed in the method for the estimation of caffeine and acetphenetidin in mixtures.

Salol ($HOC_6H_4CO_2C_6H_5$), *melting point* $42^\circ C$.—Ascertain the quantity of salol present in the sample by subtracting the weight of acetphenetidin, as found above, from the combined weight of the two ingredients determined in the preliminary or gross separation.

Alkaline Hydrolysis.

Acetphenetidin.—Weigh an amount of the sample containing not more than 100 mg. of salol on a small tared filter, exhaust with chloroform as in acid hydrolysis, collecting the solvent in a small lipped Erlenmeyer flask. Add 10 cc. of 2.5 per cent sodium hydroxid solution and heat for 5 minutes on a steam or vapor-bath at the

temperature of boiling water. At the end of this period, remove from the bath and cool quickly to room temperature in running water in order to reduce to a minimum any tendency of the acetphenetidin to undergo partial hydrolysis. Transfer the liquid to a separatory funnel by pouring and washing with a minimum of water, finally rinsing out the flask with the first 20 cc. portion of chloroform used in extraction. Extract the aqueous-alkaline solution with three 20 cc. portions of chloroform, washing consecutively each portion as obtained in a second separator with 5 cc. of water, prior to passing the solvent through a small, dry filter into a 200 cc. Erlenmeyer, from which the combined chloroformic portions are distilled to a residue of about 5 cc. Transfer the latter to a small tared beaker or crystallizing dish, by pouring and washing with sufficient fresh chloroform, evaporate on the steam-bath with the additional aid of a blast if available, cool and weigh the residual acetphenetidin at intervals until constant.

Salol.—Unite the aqueous-alkaline solutions from both separatory funnels in a suitable (500 cc.) glass-stoppered bottle, dilute with water to about 200 cc., run in from a burette an excess (about 45 cc.) of seventh-normal potassium bromid-bromate, follow with 10 cc. of concentrated hydrochloric acid, close the flask and shake for 1 minute, then at intervals over 30 minutes. At the end of this time, add 10 cc. of 15 per cent potassium iodid solution, agitating the closed flask at intervals for 15 minutes. Titrate the free iodine with standard thiosulphate (preferably seventh-normal), previously adjusted to the standard bromine solution, 1 cc. of which is equivalent to 0.002548 gram of salol. From the number of cc. of standard bromine solution expended, calculate the salol on the basis of 12 atoms of bromine to 1 molecule of salol.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

In order to facilitate the work and minimize errors due to recharging burettes, seventh-normal solutions are advocated so that the volume of thiosulphate required may not exceed the capacity of a 50 cc. burette. The thiosulphate is standardized against pure iodine. The strength of the bromine solution may also be determined by the aid of pure acetanilid.¹

(7) That the following method for the estimation of acetanilid and sodium salicylate in mixtures, be adopted as provisional:

ACETANILID AND SODIUM SALICYLATE.

(W. O. EMERY METHOD.)

Weigh on a metal scoop or watch glass an amount of the powdered sample equal to, or a multiple of, an average unit dose, transfer to a separatory funnel containing 10 cc. of water and, for every unit dose, 100 mg. of solid sodium bicarbonate. In the case of coated tablets and pills, take one or more for each determination, first crushing in a mortar prior to treatment in the separator. In the examination of alcoholic preparations, concentrate a measured volume on the steam-bath until the alcohol is expelled, transfer to a separatory funnel with a minimum of water, add sufficient solid sodium bicarbonate (0.5-1.0 gram) and proceed as follows:

Acetanilid ($C_6H_5NHCOCH_3$), *melting point* 113°C.—Extract with three 50 cc. portions of chloroform, wash each in rotation with 5 cc. of water in a second separator, and collect the solvent without previous drying in a 200 cc. Erlenmeyer. Distil the chloroform very gently to about 5 cc., add sufficient dilute sulphuric acid

¹ U. S. Bur. Chem. Bull. 132, p. 198, footnote b.

and digest on the steam-bath to complete hydrolysis, titrating the anilin sulphate substantially as outlined under estimation of caffein and acetanilid in mixtures.

Sodium salicylate ($HO.C_6H_4.CO_2Na$).—Acidify the aqueous solution of sodium salicylate with a few drops of concentrated hydrochloric acid and extract with three to five 25 cc. portions of chloroform, sufficient at least to exhaust the salicylic acid present in the mixture. Treat each portion in rotation in a second separator with 20 cc. of water containing, for every 100 mg. of salicylic acid known or believed to be present in the sample under analysis, 1 gram of anhydrous sodium carbonate. Shake vigorously and, after clearing, wash each portion again in a second separator with 5 cc. of water, which on completion of this operation is united with the main aqueous-soda solution of sodium salicylate. Dilute to a known volume, withdraw an aliquot, representing about 100 mg. of salicylic acid, to a 200 cc. Erlenmeyer, make up to 100 cc., heat nearly to boiling, then run in from a burette 25–40 cc. of strong (about fifth-normal) iodine solution, sufficient at least to insure an excess of this reagent during digestion of the product for an hour on the steam-bath. Discharge the free iodine with a few drops of thiosulphate solution, decant the clear liquid through a tared Gooch, care being taken that most of the precipitate $(C_6H_7I_2O)_2$, remains in the flask. To the latter add 50 cc. of boiling water, digest for 10 minutes on the steam-bath, filter, gradually washing all the reddish substance into a Gooch, using for this purpose and final washing about 200 cc. of hot water. Dry to constant weight in an air-bath at $100^\circ C$. Multiply the weight of the precipitate by the factor 0.4658 to obtain the quantity of sodium salicylate present in the aliquot taken.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

Should the combination contain caffein or antipyrin, or both, these substances will appear with acetanilid in the first chloroformic extraction and require a treatment substantially as outlined on pages 64–65. In the event of replacement of acetanilid by acetphenetidin, the general procedure would not be materially altered, this substance being weighed directly after recovery from its washed chloroformic solution as separated from sodium salicylate. If, instead of sodium salicylate, the combination contains the free acid or its ammonium salt, the analyst should have no difficulty in meeting such conditions by appropriate modifications in detail; for example, the addition of a larger quantity of sodium bicarbonate to insure an excess of this reagent above that required for the production of sodium salicylate prior to extraction with chloroform.

In the analysis of a mixture of caffein, acetanilid, sodium salicylate and codein sulphate, the following essential operations would be appropriate:

- (1) Extraction of caffein, acetanilid and salicylic acid from the acidified solution.
- (2) Washing the chloroformic solution with aqueous soda solution for the recovery of salicylic acid, preliminary to its treatment with iodine solution.
- (3) Separation of caffein and acetanilid as outlined on page 60.
- (4) Recovery of codein from the solution of its sulphate after treatment with sodium bicarbonate and chloroform.

(8) That the following method for the estimation of caffein, acetanilid and quinin sulphate in mixtures, be adopted as provisional:

CAFFEIN, ACETANILID AND QUININ SULPHATE.

(W. O. EMERY METHOD.)

The separation of caffein and acetanilid from quinin sulphate is based on the fact that the bisulphate of this alkaloid in aqueous media is practically insoluble in U. S. P. chloroform, while the first named substances are readily taken up by this solvent. The gross separation, therefore, resolves itself into the following main operations:

Transfer to a separatory funnel one or more average unit doses of the powdered (crushed, in the case of pills and coated tablets) sample, follow with 20 cc. of water and 50 cc. of chloroform, add 10 drops of dilute sulphuric acid, then extract in the usual way. After clearing, wash the solvent in a second separator with 5 cc. of water prior to reception in a 200 cc. Erlenmeyer. Repeat the foregoing operations with two 50 cc. portions of chloroform, finally distilling the united solvent by gentle heat to about 10 cc., preliminary to the hydrolytic treatment of the caffein-acetanilid mixture, and subsequent separation and estimation of these two drugs in accordance with the procedure outlined on page 60.

Quinin sulphate, $(C_{20}H_{24}N_2O_2)_2 \cdot H_2SO_4 + 7H_2O$.—Unite the wash water used in the second separator with the aqueous-acid solution of quinin bisulphate, add solid sodium bicarbonate in slight excess, then extract with three 50 cc. portions of chloroform, washing each portion in rotation with 5 cc. of water in a second separator, prior to passing through a dry filter into a 200 cc. Erlenmeyer and distillation therefrom to about 5 cc. Evaporate the residual liquid on the steam-bath to apparent dryness, dissolve the amorphous alkaloid in 5 cc. of neutral alcohol and titrate with fiftieth-normal hydrochloric acid (using 2 drops of methyl red solution as indicator) until the yellow color changes to faint red. Remove to steam-bath and heat until most of the alcohol has been expelled. Should the color have reverted to yellow as the result of such treatment, add sufficient acid to just restore the red, note the number of cc. of acid expended and multiply same by the factor 8.66, the number of mg. of quinin sulphate equivalent to 1 cc. of fiftieth-normal hydrochloric acid, to ascertain the quantity of quinin sulphate present in the mixture.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

Since quinin sulphate readily loses a portion of its water of crystallization when exposed to dry air, it follows that the commercial product employed in compounding medicinal preparations may contain an appreciably greater quantity of the alkaloid than is represented by the formula and, finally, that the quantity of the drug as ascertained by analysis may be appreciably in excess of the declaration.

In the event that the mixture contains acetphenetidin in place of acetanilid, the separation of the several ingredients differs in no material way from the procedure just outlined.

(9) That the following method for the estimation of caffein, acetanilid, and codein sulphate in mixtures, be adopted as provisional:

CAFFEIN, ACETANILID AND CODEIN SULPHATE.

(W. O. EMERY METHOD.)

The entire treatment is similar in every way to that prescribed for caffein, acetanilid and quinin sulphate, except that in the case of codein direct weighing of the alkaloid may be resorted to advantageously prior to titration.

Codein sulphate, $(C_{18}H_{21}NO_3)_2 \cdot H_2SO_4 + 5H_2O$.—Transfer the chloroformic solution of codein to a small tared beaker by pouring and rinsing with sufficient solvent, evaporate to apparent dryness on the steam-bath, add a few drops of alcohol to the amorphous residue, then a like amount of water and evaporate again, cool finally and weigh the usually crystalline product until constant. The weight of this residue multiplied by the factor 1.3144 will give the quantity of codein sulphate originally present.

In order to check this result volumetrically, dissolve the codein in 3-5 cc. of neutral alcohol and titrate with fiftieth-normal sulphuric acid (using methyl red as indicator), until the yellow color is replaced by a faint red. Multiply the number of cc. of standard acid thus expended by the factor 7.80, the number of mg. of codein sulphate equivalent to 1 cc. of fiftieth-normal acid. The quantity of codein sulphate as found by weight will usually be slightly greater than that determined by titration.

(10) That the following method for the estimation of caffein, acetanilid, quinin sulphate and morphin sulphate, be adopted as provisional:

CAFFEIN, ACETANILID, QUININ AND MORPHIN SULPHATE.

(W. O. EMERY METHOD.)

As in the combination, caffein, acetanilid and quinin sulphate, so also in one containing the additional constituent morphin sulphate, the alkaloidal content is readily separated from the two first-named substances by means of chloroform, while the separation of the alkaloids themselves is based on the ability of morphin to yield a morphinate insoluble in chloroform with a metallic hydroxid. The procedure thus resolves itself into the following particulars:

Caffein and acetanilid.—Transfer to a separatory funnel an amount of the powdered sample equal to, or a multiple of, a unit dose (the quantity of material taken for an individual analysis should not contain less than one-half grain of morphin), add 20 cc. of water and 10 drops of dilute sulphuric acid, then extract with three 50 cc. portions of alcohol-free chloroform, washing each in rotation in a second separator with 5 cc. of water, which is subsequently united with the alkaloidal solution in the first separator. Treat the chloroformic solution of caffein and acetanilid as outlined on page 60.

Quinin sulphate.—Add 4-5 cc. of aqueous sodium hydroxid (5 grams of pure sodium hydroxid in 50 cc. of water), to the solution of quinin and morphin sulphates and extract with four 40 cc. portions of chloroform (U. S. P.), wash each in rotation with 5 cc. of water and pass the clear solvent through a small dry filter into a 200 cc. Erlenmeyer, from which the solvent is later removed by gentle distillation, the residual quinin being titrated with fiftieth-normal hydrochloric acid as already outlined on page 72.

Morphin sulphate, $(C_{17}H_{19}NO_3)_2 \cdot H_2SO_4 + 5H_2O$.—Wash the filter employed in the preceding operation with 5 cc. of water, and unite with the aqueous-alkaline solution of the morphinate. Then add 0.5 gram of ammonium chlorid (or an amount slightly in excess of that required to free the morphin as well as convert all of the sodium hydroxid to sodium chlorid) and to the resulting ammoniacal solution add 45 cc. of U. S. P. chloroform and 5 cc. of alcohol, then extract in the usual way, washing the solvent in a second separator with 5 cc. of water. After clearing, pass the chloroform through a small dry filter (as well as a pledget of cotton if deemed necessary) into a 200 cc. Erlenmeyer. Repeat the extraction with three 40 cc. portions

of U. S. P. chloroform, wash and filter as before, finally collect all the solvent in an Erlenmeyer and distil therefrom to about 10 cc. Transfer to a small tared beaker by pouring and washing with additional chloroform, evaporate to apparent dryness, add 0.5 cc. each of water and neutral alcohol, induce crystallization by stirring with a glass rod and final evaporation to dryness. Cool and weigh until constant. Check the weight of morphin thus determined by titration with fiftieth-normal sulphuric acid, using a drop of methyl red as indicator. To this end dissolve the alkaloid in 1-2 cc. of warm neutral alcohol, then after solution is complete, add the acid until the color passes from yellow to red. Evaporate most of the alcohol on the steam-bath, and in the event that the color reverts to yellow add just sufficient acid to restore the red. Note volume of acid expended, multiplying the number of cc. by 7.53 (the number of mg. of morphin sulphate equivalent to 1 cc. of fiftieth-normal acid) to ascertain the quantity of morphin sulphate present in the mixture.

COMMENTS AND SUGGESTIONS BY THE REFEREE.

For the purpose in question, alcohol-free chloroform is readily prepared by washing the U. S. P. product repeatedly with water.

In the various operations involving fixation and subsequent liberation of morphin by means of fixed alkali and ammonium chlorid, the most careful attention should be paid to the manner of adding these reagents, since any undue excess of either might nullify the entire procedure. Any large excess of sodium hydroxid would naturally require for its reduction a correspondingly large amount of ammonium chlorid, the latter in turn yielding its pro rata of hydroxid, relatively large quantities of which through interaction with sodium chlorid tend to inhibit any permanent liberation of alkaloid and thus prevent complete extraction. Furthermore, ammonium chlorid in large amount operates retentively on the morphin in solution, due in part possibly to the formation of an alkaloidal hydrochlorid.

Despite all precautions looking to the exclusion of impurities from the morphin as weighed, the amount of this substance thus determined will usually be greater than that found volumetrically. In order to insure the greatest possible accuracy in volumetric operations on alkaloidal residues like quinin, morphin, and codein, it is suggested that whenever possible the strength of the standard acid used be checked by titration against the pure alkaloid under examination.

(11) That the following method for the evaluation of gum tragacanth,¹ be adopted as provisional:

TRAGACANTH.

(W. O. EMERY METHOD.)

A knowledge of the quantity of volatile (acetic) acidity developed in the acid hydrolysis of gum tragacanth (*Astragalus gummiifer*) affords a valuable index of the purity of this commodity when compared with results obtained by similar treatment of so-called "Indian gum" (*Cochlospermum gossypium* and *Sterculia urens*), a not uncommon substitute and adulterant of tragacanth. The term "volatile acidity", as here used is the number of cc. of tenth-normal potassium or sodium hydroxid required to neutralize the volatile (acetic) acid obtained by subjecting the products of the action of boiling aqueous phosphoric acid on 1 gram of the gum to distillation with steam. As the result of such treatment of numerous authentic samples

¹ *J. Ind. Eng. Chem.*, 1912, 4: 374.

of whole and powdered tragacanth, it was found that 3.6-3.7 cc. (averages obtained with 35 samples whole and 21 samples powdered gum, respectively) tenth-normal alkali represents the volatile acidity of this gum, corresponding to 2.15-2.20 per cent of acetic acid, while in the case of Indian gum 26.7-26.5 cc. of tenth-normal alkali (corresponding to 15.97-15.79 per cent of acetic acid) were obtained as averages of 9 samples of whole and 14 samples of powdered gum respectively.

In determining the volatile acidity, the following procedure was used:

Treat 1 gram of the whole or powdered sample in a 700 cc. round-bottomed flask, provided with a long neck, for several hours in the cold with 100 cc. of distilled water and 5 cc. of sirupy phosphoric acid until the gum is completely swollen. Boil gently for 2 hours in connection with a reflux condenser, whereby a nearly clear, colorless solution is effected. A very small amount of cellulose substance will remain undissolved. Then subject the hydrolyzed product to slow distillation in a vigorous current of steam until the distillate amounts to 600 cc. and the acid residue to about 20 cc. This should not be driven too far, however, otherwise there may be danger of scorching the non-volatile, organic degradation products, with consequent possible contamination of the distillate. It has been found that a spray trap of the form shown in U. S. Bur. Chem. Circ. 94, p. 4, if used in connection with the flask containing the hydrolyzed gum, is effective in preventing traces of phosphoric acid being carried over into the distillate. Titrate with tenth-normal potassium hydroxid in connection with 10 drops of phenolphthalein solution, finally boiling the liquid under examination until a faint pink color persists. Run a control on the same amount of distillate obtained by a parallel operation, with the omission of the gum, but using like quantities of other ingredients and observe the same conditions as in the test.

It should be noted in this connection that while tragacanth yields a practically colorless solution when boiled with aqueous phosphoric acid, Indian gum, on the other hand, gives a pink or rose colored solution, a sure indication in an unknown sample that the last named product is present.

REPORT ON SYNTHETIC PRODUCTS.

By W. O. EMERY (Bureau of Chemistry, Washington, D. C.), *Associate Referee.*

During the past year, coöperative work on synthetic products has been restricted to mixtures containing two drugs, caffeine and antipyrin. On account of similarity in behavior towards the more common immiscible solvents, a quantitative separation depending on any simple extraction method was considered impracticable. Accordingly, recourse was had to a procedure based on the fact that, when an aqueous solution of iodine and sodium bicarbonate is permitted to react with antipyrin, either alone or in the presence of caffeine, mono-iodoantipyrin is formed. This latter substance, together with caffeine, is thereupon extracted with chloroform, the weight of extractives ascertained and the iodine therein estimated as silver iodide, from which the antipyrin can be calculated by means of the appropriate factor. This procedure was very carefully tested under a

variety of conditions prior to submission to the collaborating chemists, hence there was every reason to expect satisfactory results from workers of average skill in the art.

The following instructions were sent to collaborators:

COÖPERATIVE WORK.

CAFFEIN AND ANTIPYRIN IN ADMIXTURE.

PREPARATION OF SAMPLE.

Powder No. 16.—Prepared by triturating anhydrous caffein and antipyrin (1 to 4) in a mortar and subsequent sifting.

DETERMINATION.

Weigh on a metal scoop or watch glass 0.25 gram of the sample, transfer to a 150 cc. separatory funnel with 5 cc. of alcohol-free chloroform and 10 cc. of water, add 0.5 gram of sodium bicarbonate and about 10 cc. of fifth-normal iodine (or double the quantity of tenth-normal iodine), then shake vigorously for 1 minute. After this operation a slight excess of iodine should still be apparent in the liquid menstruum, provided all the antipyrin has been converted to iodoantipyrin. If, however, all the iodine has been thus expended, a little more should be added and the mixture again shaken. Then discharge the uncombined iodine by means of a small crystal of sodium thiosulphate, add 20 cc. of U. S. P. chloroform and shake. After clearing, draw off the solvent into a second separator containing 5 cc. of water, shake and after clearing pass the chloroform through a small, dry filter into a 50 cc. tared beaker, evaporate to apparent dryness on the steam-bath, accelerating this operation by means of an air blast, if available. Repeat the extraction with two (three, in case tenth-normal iodine were used) 25 cc. portions of chloroform, washing, filtering and evaporating each portion in rotation substantially as directed for the first portion. Recover all traces of crystalline products separating about the tips of funnels and edge of the filter by judicious washing with chloroform. Dry the final colorless, crystalline residue of caffein and iodoantipyrin for 30 minutes at 100°C., then cool and weigh. Designate this weight *A*.

Dissolve this residue in 5 cc. of glacial acetic acid, add 10 cc. of saturated aqueous solution of sulphur dioxide, then transfer the resultant liquid to a 400–500 cc. beaker by pouring and rinsing with hot water until the final volume amounts to about 200 cc. Add aqueous silver nitrate solution (containing about 0.25 gram of silver nitrate) and follow with a few drops of nitric acid, then heat nearly to boiling, stirring the while in order to agglomerate the silver iodide. Add about 15 cc. of concentrated nitric acid, cover the beaker with a watch glass and boil gently for 5 minutes. Filter by decantation through a tared Gooch, wash the precipitate twice with boiling water, finally transferring the silver iodide completely to the Gooch, washing several times with hot water and finally with alcohol. Dry for 30 minutes in an air-bath at 110°C., cool and weigh.

The weight of silver iodide multiplied by the factor 0.8012 yields the quantity of antipyrin present in the sample taken. The quantity of caffein present in the sample is ascertained by subtracting the product, obtained by multiplying the weight of silver iodide by the factor 1.3374, from the weight designated *A*.

PREPARATION OF SAMPLE.

Liquid No. 17.—Obtained by dissolving 5 grams of anhydrous caffein and 25 grams of antipyrin in water and diluting to 1 liter at 25°C. The liter flask employed was standardized at 20°C.

DETERMINATION.

By means of a pipette standardized at 20°C., transfer 10 cc. of the solution, previously warmed or adjusted to 25°C., to a 150 cc. separatory funnel containing 0.5 gram of sodium bicarbonate. Add about 15 cc. of fifth-normal iodine (or double the quantity of tenth-normal iodine), then shake vigorously for 1 minute. All succeeding operations are identical with those outlined for the powder mixture.

Report recoveries in mg. and parts per 100.

The foregoing procedure is based on the fact that, when an aqueous solution of iodine and sodium bicarbonate is permitted to react with antipyrine, whether alone or with other substances, such as caffeine, monoiodoantipyrine is formed. This latter substance, together with caffeine, is thereupon extracted with chloroform, the weight of such mixture ascertained and the iodine therein estimated as silver iodide, from which the antipyrine is calculated by means of the appropriate factor. In view of the somewhat radical treatment with dilute nitric acid, to which the caffeine is necessarily subjected incidental to the precipitation of silver iodide, a direct determination of that substance is impracticable. The quantity of caffeine is, however, ascertained by subtracting the amount of iodoantipyrine, as calculated from the silver iodide, from the combined weight of caffeine and iodoantipyrine.

The use of alcohol-free chloroform is specified in order to preclude the possible formation of iodoform, the presence of which would necessarily render the method valueless.

The final washing of the silver iodide with alcohol should be thorough in order to eliminate certain unidentified organic substances arising from the action of the nitric acid on the antipyrine complex.

DISCUSSION OF RESULTS.

Examination of the following tabulated data reveals the fact that a majority of the collaborators were able to obtain quite satisfactory results. A few, however, report values so divergent from the theory as to indicate that somewhere in the procedure the directions may not have been sufficiently clear to insure uniformity in manipulation. Wherever such discrepancy is material, low antipyrine and high caffeine values were found almost without exception, unquestionably due for the most part to incomplete iodation. Assuming all apparatus used to have been strictly clean and chemicals pure, further contributory causes for antipyrine shortage and caffeine excess may have been due to the following causes: (a) Failure to properly desiccate or remove all volatile substances from the composite residue A; (b) incomplete removal of alcohol-soluble material associated with the silver iodide; or, (c), faulty technique.

Coöperative results on the recovery of caffein and antipyrin in admixtures.

ANALYST	POWDER, NO. 16		LIQUID, NO. 17	
	Antipyrin 80.00 per cent	Caffein 20.00 per cent	Antipyrin 83.33 per cent	Caffein 16.67 per cent
J. M. Bartlett, Orono, Me.....	70.88	28.72	81.47	17.03
	70.36	27.56	78.80	21.43
L. A. Brown, Lexington, Ky.....	79.35	22.44	82.10	23.63
	78.96	22.96	83.37	25.87
E. O. Eaton, San Francisco, Cal.....	80.92	18.96	82.80	17.30
	83.00	16.87
W. O. Emery, Washington, D. C.....	79.96	19.96	82.97	16.77
	79.76	20.28	83.27	16.73
	79.96	20.04	82.93	17.17
C. O. Ewing, Washington, D. C.....	83.43	16.40
	83.10	16.77
	83.37	16.37
J. F. Darling, New York, N. Y.....	73.04	28.04	82.33	22.73
	79.56	15.44	81.17	18.50
	79.99	20.12	82.87	17.80
	82.13	18.67
	83.97	16.93
H. Engelhardt, Baltimore, Md.....	79.80	19.24	83.50	20.00
	79.80	19.24	83.60	18.96
	79.16	21.48	82.27	20.84
	79.56	21.36	82.80	20.56
H. C. Fuller, Washington, D. C.....	79.44	22.44	82.60	17.85
	79.16	22.56	82.63	17.53
C. K. Glycart, Chicago, Ill.....	78.44	19.76	82.53	17.37
	78.76	21.08	82.67	17.07
	79.04	20.64
A. W. Hansen, Honolulu, Hawaii.....	73.32	26.60	81.27	20.33
	77.84	22.56	80.50	20.73
R. Hertwig, Honolulu, Hawaii.....	73.00	26.12	78.50	21.10
	77.24	24.04
	79.04	22.24
	79.00	21.12
	79.08	21.60
	79.08	23.20
H. B. Mead, Philadelphia, Pa.....	79.64	21.52
	79.08	21.40	83.60	17.93
	80.48	19.84	83.50	17.57
	79.88	27.04	83.57	19.13
C. B. Morison, New Haven, Conn.....	80.16	21.32	82.70	18.60
	77.32	23.32	83.00	16.73
	77.68	22.84
	79.09	22.76
	79.40	22.08

Coöperative results on the recovery of caffein and antipyrin in admixtures.—Continued.

ANALYST	POWDER, NO. 16		LIQUID, NO. 17	
	Antipyrin 80.00 per cent	Caffein 20.00 per cent	Antipyrin 83.33 per cent	Caffein 16.67 per cent
S. Palkin, Washington, D. C.....	80.12	19.88	83.33	17.10
	79.76	20.56	83.17	16.97
	83.23	16.43
G. C. Spencer, Washington, D. C.....	79.16	20.63	83.03	17.33
	79.28	21.06	82.56	17.93
	79.40	20.60	82.50	18.06
A. R. Todd, Lansing, Mich.....	74.67	28.40	81.27	20.20
	74.12	29.00	81.17	19.87
	73.90	28.16	80.97	20.27
	80.33	18.00
	80.50	18.37
C. D. Wright, Washington, D. C.....	79.00	21.40	82.90	17.00
	79.40	21.20	82.97	17.17
	79.40	20.80

COMMENTS.

L. A. Brown suggested a greater number of extractions with chloroform since in his opinion caffein and iodoantipyrin continue to be recovered even up to the eighth or ninth extraction.

J. F. Darling recommended that in order to insure a more complete desiccation of the composite residue *A*, the time of heating be extended one-half hour.

H. Engelhardt facilitated the final expulsion of chloroform from this residue by the addition of a little ether prior to heating in the air-bath.

H. C. Fuller suggested the employment of a third separator for the assembling of all three (or more) chloroformic portions prior to passing through cotton into the tared beaker.

R. Hertwig and *A. W. Hansen* recommended washing the silver iodid once by decantation with alcohol after first decanting off the hot aqueous solution.

The referee suggests the following procedure if it seems desirable to recover most of the chloroform:

Withdraw the several portions into a 200 cc. Erlenmeyer instead of the tared beaker, distil most of the solvent by gentle heat, then transfer the residue to the tared beaker by pouring and washing with sufficient fresh chloroform for treatment as specified in the method.

The following committee on revision of the constitution was appointed: William Frear, of Pennsylvania; R. J. Davidson, of Virginia; and E. W. Magruder, of Virginia.

DETERMINATION OF SANTONIN IN LEVANT WORMSEED.

By E. K. NELSON (Bureau of Chemistry, Washington, D. C.).

In view of certain difficulties reported in the determination of santonin, an investigation of the various methods in use was undertaken, experiments being conducted with the Thaeter method, the Katz method, and Fromme's modification of the Katz method.

The Katz and Katz-Fromme methods necessitate the use of a solubility correction factor which is avoided in the Thaeter method. With the latter method, however, all of the santonin was not recovered, and as it is altogether cumbersome and tedious it was dropped from further consideration.

The chief difficulty found with the Katz-Fromme method was that the santonin residues were not free from resinous impurities, and were consequently discolored and returned a santonin value in excess of the truth. Accurate checks were not obtained on account of the incomplete elimination of impurities.

With certain precautions in its use the Katz method, with slight modifications, was found to give concordant and evidently accurate results. The santonin residues were of good color, and the method is very simple and easy to operate.

SUGGESTED MODIFICATION OF THE KATZ-FROMME METHOD.

Extract 10 grams of medium ground Levant wormseed in a Soxhlet extraction apparatus for 3 hours with chloroform. Distil off the chloroform till 7-8 grams remain, add 100 grams of a 5 per cent solution of barium hydroxid, and heat on the steam-bath till the odor of chloroform has disappeared. Boil for 5 minutes, cool and pass carbon dioxid (washed through sodium bicarbonate solution to free it from traces of acid which might be carried over mechanically) till saturated. An excess of carbon dioxid will do no harm. Filter by suction, preferably on a small Büchner funnel, and wash twice with 10 cc. of water. To the filtrate, heated on a steam-bath, add 5 cc. of 25 per cent hydrochloric acid and warm for 5 minutes. Cool till lukewarm and extract with 20, 15 and 15 cc. of chloroform, passing the solvent through a small filter into a flask. Evaporate to dryness, being careful to see that the last traces of chloroform are removed. Add 7.5 grams of absolute alcohol to the residue, and dissolve (by gentle warming if necessary). Then add 42.5 grams of water heated to 60°-70°C., stopper the flask and allow to cool. (At this point it is advisable to see that crystallization starts, which can be done by scratching the side of the beaker or adding a *minute* crystal of santonin. Samples put aside in a cool place for 24 hours, and which contained santonin in liberal amount have been found in a supersaturated condition where this precaution was not observed.) Keep the flask at a temperature of 15°-17°C. for 24 hours. Filter and wash with 2 portions of 10 cc. of 15 per cent alcohol by weight. Dry the flask and filter at 100°C., dissolve the santonin left in the flask in chloroform and filter into a tared beaker. Wash the santonin out of the flask and filter thoroughly with chloroform, evaporate the filtrate and dry at 100°C. till all traces of chloroform are gone. To the weight found add 0.04 gram for the santonin dissolved in the dilute alcohol, and multiply the total by 10 for the per cent of santonin.

(The solubility factor was found correct for a temperature of 15°-17°C. At 30°-32°C., 0.054 gram of santonin was left in solution.)

The above method was submitted to a number of chemists for co-operative work and criticism, with a uniform sample of Levant wormseed. The results obtained are as follows:

Results of coöperative work on santonin.

ANALYST	SAN- TONIN	ANALYST	SAN- TONIN
	<i>per cent</i>		<i>per cent</i>
C. K. Glycart, Chicago, Ill.....	2.41	J. G. Roberts, Philadelphia, Pa. ¹	2.96
	2.44		2.91
	2.52		
J. F. Darling, New York, N. Y....	2.77	H. C. Fuller, Washington, D. C. .	2.81
			3.11
P. Rothberg, New York, N. Y....	2.74	J. M. Francis, Detroit, Mich.....	2.52
			2.75
J. Ehrlich, New York, N. Y.....	2.65		
H. Runkel, St. Louis, Mo.....	3.02	H. Engelhardt, Baltimore, Md....	2.79
	2.89		2.83
			2.77
B. H. St. John, Washington, D. C.	2.85	E. K. Nelson, Washington, D. C..	2.87
			2.66
H. M. Rhea, Washington, D. C....	2.62		2.61
	2.58		2.69
	2.62		2.58
	2.48		2.73
Average			2.72
Maximum			3.11
Minimum			2.44

¹ The solubility factor used was 0.054. When the factor 0.04 was used these results became 2.82 per cent and 2.77 per cent.

COMMENTS BY ANALYSTS.

J. G. Roberts, of Smith, Kline & French: Although somewhat elaborate, the method submitted is satisfactory. It gives concordant results and yields santonin in a crystalline state, apparently free from resin and having only a little color. However, there is one feature that may operate against the general adoption of the method and that is the requirement that crystallization shall be conducted at 15°-17°C. for 24 hours. We consider that 2°C. is a rather limited range to maintain for 24 hours. We consider also that the method would be improved if instructions were given to have the diluted alcohol used for washing the crystals at the same temperature as the solution containing the crystals of santonin. In order to eliminate as much error as possible we filtered and washed the santonin crystals at 30°C. and used the correction of 0.054 gram given in the method for that temperature.

H. C. Fuller: In both determinations by the Nelson method the identical procedure was used, but the second assay contained some resin and was darker than the santonin in the first sample.

J. F. Darling: The chloroform extraction appears to be more efficient than the extraction with ether, and the residues obtained by the modified method are much cleaner than in the method of J. Katz which we have hitherto used.

J. M. Francis: We do not think that 3 hours' extraction in a Soxhlet is sufficient for the thorough extraction of the drug. In the first assay the extraction was continued to 3½ hours, the distillation of the chloroform being moderately rapid throughout the process. In the second case the extraction was allowed to proceed for about an hour and the heat was then shut off for the night, the drug being left well covered with chloroform in the Soxhlet extractor. In the morning the chloroform in the Soxhlet was very deeply colored, and the extraction was continued for 3 hours longer in the usual way.

We are of the opinion that there is a distinct advantage in Fromme's original process of maceration with occasional shaking for 12 hours or more, and then of removing an aliquot portion of the chloroformic solution for the assay.

A 5 per cent solution of barium hydroxid is directed. Owing to the limited solubility a 5 per cent solution can only be obtained by warming to a temperature of 30°-40°C. If this strength is desired, therefore, specific instructions should be included, directing that the solution be warmed.

On the whole, we consider that the process works very well, and doubtless with a little practice results would be obtained which would check better than those indicated above.

H. Engelhardt: The santonin obtained was by no means pure and colorless, the color varying from light brown to dark brown. Some time ago I made quite a number of assays of samples of Levant wormseed and I have found that by the Katz-Fromme method (which is rather convenient inasmuch as no Soxhlet is used for extracting the drug) reliable and uniform results could be obtained, the santonin being in most cases very pure. I applied this method to the powder submitted and obtained 2.67 per cent and 2.70 per cent, respectively, which correspond fairly well with those obtained by the other method. The santonin was of a creamy white, thus showing that only a very small percentage of impurities was present.

H. Runkel: I wish to state that these residues were exceptionally pure considering the fact that there is a large amount of resinous material, etc., which must be removed. I find, however, that boiling with barium hydrate for a full 5 minutes is quite essential.

No report was presented by the associate referee on medicated soft drinks.

A paper on the "Comparison of the Physical Properties of Some Official Volatile Oils with Special Reference to the Requirements of the U. S. P., 1910," by Frank Rabak of the Bureau of Plant Industry, was read by Mr. Rabak. This article has since been published in the *J. Am. Pharm. Assoc.*, 1914, 3: 670.

No report was made by the associate referee on alkaloids.

REPORT ON BALSAMS AND GUM RESINS.

By E. C. MERRILL (Bureau of Chemistry, Washington, D. C.), *Associate Referee.*

The object of this work was to secure data which would indicate the extent to which the lead number method for asafoetida would check when used by independent chemists working on samples of known character. It was further desired to verify the efficacy of certain color tests for foreign resins which are later bulletined in the method.

PREPARATION OF SAMPLES.

For these purposes a large sample of good asafoetida, about 10-12 pounds, was procured. This was run through a power sausage grinder several times in order to thoroughly mix the samples. The friction of the sample passing through the machine warmed the product to a temperature of about 110°F. when it became a semi-fluid mass, easily stirred, and homogenous. This sample was divided into two portions, *A* and *B*. Sample *A* was transferred to $\frac{1}{4}$ pound tins.

Two and ten-hundredths pounds of powdered rosin were added to the remainder of the sample, 5.25 pounds, and the latter passed through the grinder several times to insure a uniform mixture. The amount of adulterant in this case was about 28.5 per cent. This mixture was put up similarly in quarter pound tins and marked *B*.

Sets of Samples *A* and *B* accompanied the instructions sent to collaborators.

ASAFOETIDA.

LEAD NUMBER.

(1) Preparation of Resin.

Use a sample (about 20 grams) sufficient to furnish between 5 and 10 grams of the ether purified resin. Determine the alcohol insoluble material in the usual manner. Transfer the first 2 filtrates, representing the major part of the sample, to a casserole or a flat-bottomed porcelain dish, and evaporate the alcohol on the steam-bath. Treat the resinous mass with ether (sometimes it is necessary to warm gently to facilitate solution of the resin). Filter the ethereal solution into a separatory funnel and wash with water until the aqueous layer separates without any milkiness. (If the ether solutions persist in remaining turbid, more ether may be added, or it may become necessary to dry the ether solution by shaking with sodium chlorid in the separatory funnel after as much as possible of the aqueous solution has been removed.) Then filter the ethereal solution through a folded filter paper, moistened with ether, into a flask or beaker, and evaporate the solvent on the steam-bath.

(2) Determination.

(The residual ether purified resin from the above preparation is now in a state where it can usually be broken up when cold, and powdered.)

Into a small tared beaker (about 75 cc. capacity) weigh roughly about 1.1-1.2 grams of the resin prepared in (1) and dry for 5 hours in the air-bath at 110°C. Place in a desiccator, cool and weigh. (The weight noted is to be used in subsequent calculations.) Dissolve in 20 cc. of 95 per cent alcohol, boil gently until the resin is in solution, transfer to a 100 cc. graduated flask, wash the beaker with hot 95 per cent alcohol, care being taken that the final volume does not exceed 70 cc. Add 25 cc. of the alcoholic lead acetate (*A*), agitate and allow to stand overnight. Make up to the mark with 95 per cent alcohol, shake well, allow to stand a few minutes, and then filter through a fluted paper and pipette an aliquot of 25 cc. from the filtrate into a beaker. Add 10 cc. of water and evaporate to 10 cc. Add 5 cc. of 10 per cent sulphuric acid and then 100 cc. of alcohol, stirring vigorously to dissolve

any separated resin, and heat if necessary. Allow to stand for an hour, then filter the lead sulphate on a tared Gooch, wash with alcohol and finally with ether, dry at 100°C. to a constant weight and weigh.

Run a blank on the alcoholic lead acetate solution, and calculate the amount of metallic lead absorbed by 1 gram of the dried resin. The mg. of lead per gram of sample represent the lead number. The factor for the conversion of lead sulphate to metallic lead equals 0.6830.¹

QUALITATIVE TESTS FOR PURITY.

(1) *Copper Acetate.*

Transfer about 40 cc. of the alcoholic solution (obtained by determining the alcohol insoluble) to a separatory funnel, add 30 cc. of petroleum ether, mix thoroughly, then add 50 cc. of water and shake for 30 seconds. Allow to separate and discard the aqueous milky layer. Wash the petroleum ether layer with water until the wash water and petroleum ether are both clear. Add 30 cc. of $\frac{1}{2}$ per cent copper acetate solution and shake well. No green color should appear in the petroleum ether. The appearance of a green color indicates the presence of foreign resins.

(2) *Concentrated Hydrochloric Acid.*

Transfer 3-5 cc. of the alcoholic solution (obtained by determining the alcohol insoluble) to a small beaker, add concentrated hydrochloric acid, drop by drop, until a faint turbidity appears on agitation, then add sufficient alcohol to clear up the turbidity. A green color results which fades within 10 minutes. The appearance of a red or violet color indicates the presence of galbanum or other foreign resins. (The temperature at which this reaction takes place should be kept below 35°C.)

(3) *Ferric Chlorid.*

To 1 or 2 cc. of the alcoholic solution diluted to 5 cc. with 95 per cent alcohol, add a few drops of ferric chlorid solution (10 per cent). A clear, green color results. The appearance of a red tinge or a dark red or a brownish color indicates sophistication with foreign resins.

REMARKS AND COMMENTS BY COLLABORATORS.

Earl B. Putt: When the acid is first added in the hydrochloric acid test the green coloration is very pronounced, but this color soon fades and a variety of color shades are produced when alcohol is added in varying amounts. Therefore, this test does not appear to be especially indicative. The ferric chlorid test is quite satisfactory and seems to require a rather strong solution of ferric chlorid. The test is improved rather than interfered with by an excess of ferric chlorid.

The copper acetate test is very satisfactory. In giving directions for the above qualitative tests it might be well to state in each case the approximate weight of the sample represented in the volume of alcoholic solution taken. Different analysts will use different quantities of alcohol in determining the alcohol soluble portion, hence concentration of the alcoholic solution will vary.

H. Runkel: The concentrated hydrochloric acid test with *B* does not fade like *A*.

¹ For the preparation of alcoholic lead acetate dissolve 5 grams of normal lead acetate in 20 cc. of water and add 80 cc. of 95 per cent alcohol. A turbidity generally results, due to the precipitation of lead carbonate caused by carbon dioxide in the alcohol. Allow the solution to stand overnight. The clear, supernatant liquid can then be used without filtering for the determination of the lead number.

The blank on 25 cc. of alcoholic lead acetate solution should be equivalent to at least 1 gram, calculated as $PbSO_4$.

Lead number and color tests for foreign resins obtained on Samples A and B.

ANALYST ¹	LEAD NUMBER		COPPER ACETATE TEST A	CONCENTRATED HYDROCHLORIC ACID TEST A	FERRIC CHLORID TEST A
	A	B			
E. O. Eaton, San Francisco, Cal.	200.0	109.0	O.K.	O.K.	O.K.
H. Kleuter, Madison, Wis.....	233.2	160.0	Light green	Violet	Brownish
E. C. Merrill, Washington, D. C.	240.4	151.1	Negative	Green, fading gradually	Green
E. B. Putt, New York, N. Y....	238.0	165.0	Negative	Green, changing to blue-green	Green
S. Palkin, Washington, D. C...	245.6	158.8	O.K.	O.K.	O.K.
P. Rothberg, New York, N. Y.	241.0	164.0	Negative	Green to pinkish brown	Yellowish green
J. K. Rippetoe, New York, N. Y.	220.1	144.9	Negative	Blue green	Yellow green
N. Smith, New York, N. Y.....	213.1	140.1
H. Runkel, St. Louis, Mo.....	201.0	147.9	Negative	Green fading in 3 minutes	Clear green
H. A. Seil, New York, N. Y....	240.1	154.7	Negative	Green to light pink	Green
A. E. Taylor, San Juan, P. R...	207.0	166.1	Negative	Green, fading to very light pink	Clear olive solution
J. F. Darling, New York, N. Y.	237.7	143.2	Negative	Faintly positive	Negative
E. Clark, Boston, Mass.....	237.5	164.7	Negative	Reddish brown to violet tinge	Positive
F. M. Boyles, Baltimore, Md...	184.0	104.0	Negative
J. B. Luther, New York, N. Y.	245.6	151.6	Negative	Green, fading to faint red	Green
H. B. Mead, Philadelphia, Pa...	222.6	141.1	Faint green

¹The associate referee desires to acknowledge his indebtedness to the analysts whose names appear in this column for the assistance rendered in this work.

Color tests obtained on Sample B.

ANALYST ¹	COPPER ACETATE TEST	CONCENTRATED HYDROCHLORIC ACID TEST	FERRIC CHLORID TEST
E. O. Eaton, San Francisco, Cal...	Resins present	Resins present	Resins present
H. Kleuter, Madison, Wis.....	Deep green	Not red or violet	Brownish
E. C. Merrill, Washington, D. C....	Intense green	Green, quickly changing to dark yellow	Brownish yellow
E. B. Putt, New York, N. Y.....	Bright green	Green	Red
S. Palkin, Washington, D. C.....	Resin indicated (green)	Positive	Positive
P. Rothberg, New York, N. Y.....	Strongly positive	Green to brown	Brown
J. K. Rippetoe, New York, N. Y....	Green	Grayish	Reddish brown
H. Runkel, St. Louis, Mo.....	Decided green	Faint green	Decided red tinge
H. A. Seil, New York, N. Y.....	Strongly positive (green)	Green to dark yellow	Brownish
A. E. Taylor, San Juan, P. R.....	Green	Green, fading to yellow	Brown solution, slightly turbid
J. F. Darling, New York, N. Y.....	Strongly positive	Negative	Positive
E. Clark, Boston, Mass.....	Positive	Negative	Positive
F. M. Boyles, Baltimore, Md.....	Decided green
J. B. Luther, New York, N. Y.....	Intense green	Green, fading to colorless	Yellowish brown
H. B. Mead, Philadelphia, Pa.....	Strong green

¹ The associate referee desires to acknowledge his indebtedness to the analysts whose names appear in this column for the assistance rendered in this work.

COMMENTS BY THE ASSOCIATE REFEREE.

It is believed, in view of the results obtained by the other collaborators that the lead numbers by Messrs. Eaton and Boyles are abnormally low and are probably due to misinterpretation of the method at some stage in the process. The figures obtained by Messrs. Runkel and Taylor on Sample A also appear somewhat below the average. In the knowledge of the associate referee, Messrs. Eaton, Putt, Rothberg, Seil, Darling and Clark were previously familiar with this method and there appears to be a general concordance in the results obtained by them.

The general average of the lead numbers on A, excepting the 4 previously mentioned, which appear to be abnormal, is 234.2. The widest variation from this

figure is roughly about 10 per cent in one instance, while the remainder of the figures differs by about 5 per cent or less. The general average of the lead number for *B*, not taking into account 2 results which appear abnormal, is 154.0, with a maximum variation from this figure of less than 5 per cent.

The copper acetate test for colophony appears satisfactory.

The concentrated hydrochloric acid test appears of doubtful value as an index for foreign resins.

The ferric chlorid test in the hands of different chemists leads to varying conclusions and is therefore doubtful.

SUMMARY OF RECOMMENDATIONS.

It is recommended—

(1) That the lead number test for asafoetida be adopted unofficially by the association, and that the method be further studied during the coming year.

(2) That further work be done to establish more satisfactory color tests for foreign resins.

(3) That the iodine number of cinnamoin in Peru balsam as an index to true or artificial Peru balsam be studied during the coming year with a view to later collaboration.

REPORT OF COMMITTEE ON REVISION OF CONSTITUTION.

By WM. FREAR (State College, Pa.), *Chairman*.

The resolution introduced by Mr. Lythgoe having been considered, it is recommended that Paragraph 2 be amended by the insertion after the first sentence of the following:

Analytical chemists connected with municipal laboratories that perform work upon any of the subjects specified in Paragraph 1 hereof, shall be entitled ex officio to associate membership, with the privilege of discussion but without those of entering motions, voting or becoming eligible for office.

Approved.

REPORT OF COMMITTEE ON RECOMMENDATIONS OF REFEREES.¹

By P. F. TROWBRIDGE (Experiment Station, Columbia, Mo.), *Chairman*.

It is believed that the present system of referees and the assigning of special subjects to associate referees for investigation is the best policy for the association.

¹ Presented by B. B. Ross.

The committee therefore recommends that the executive committee be authorized to appoint additional associate referees to investigate special problems, when in their opinion there is need for such special investigation.

Adopted.

Mr. Frear moved that a vote of thanks be given the President, which was unanimously carried.

The association adjourned.

OFFICERS, REFEREES AND COMMITTEES OF THE ASSOCIATION OF OFFICIAL AGRICULTURAL CHEMISTS,
FOR THE YEAR ENDING NOVEMBER, 1915.

Honorary President.

H. W. WILEY, Washington, D. C.

President.

C. H. JONES, Burlington, Vt.

Vice-President.

R. N. BRACKETT, Clemson College, S. C.

Secretary-Treasurer.

C. L. ALSBERG, Washington, D. C.

Additional Members of the Executive Committee.

JULIUS HORTVET, St. Paul, Minn.

E. F. LADD, Fargo, N. Dak.

Referees.

Phosphoric Acid: L. S. Walker, Amherst, Mass.

Nitrogen:

Determination: R. N. Brackett, Clemson College, S. C.

Separation of nitrogenous substances: A. W. Bosworth, Geneva, N. Y.

Polash: E. E. Vanatta, Columbia, Mo.

Soils: J. W. Ames, Wooster, Ohio.

Dairy Products: L. I. Nurenberg, Boston, Mass.

Feeds and feeding stuffs: G. L. Bidwell, Washington, D. C.

Food adulteration: Julius Hortvet, St. Paul, Minn.

Sugar: C. A. Browne, New York, N. Y.

Insecticides: R. C. Roark, Washington, D. C.

Inorganic plant constituents: A. J. Patten, East Lansing, Mich.

Medicinal plants and drugs: W. O. Emery, Washington, D. C.

Water: W. W. Skinner, Washington, D. C.

Testing chemical reagents: C. O. Ewing, Washington, D. C.

Water in foods and feeding stuffs: W. J. McGee, Washington, D. C.

Organic and inorganic phosphorus in foods, feeding stuffs, and drugs: F. M. Beegle, Wooster, Ohio.

Associate Referees.

Phosphoric acid: W. J. Jones, Jr., Lafayette, Ind.

Nitrogen:

Determination: H. D. Haskins, Amherst, Mass.

Special study of Kjeldahl method: I. K. Phelps, Washington, D. C.

Separation of nitrogenous substances:

Milk and cheese: Leroy S. Palmer, Columbia, Mo.

Meat proteins: P. F. Trowbridge, Columbia, Mo.

Potash:

Determination: T. D. Jarrell, College Park, Md.

Soils:

Nitrogenous compounds: C. B. Lipman, Berkeley, Calif.

Dairy products: J. T. Keister, Washington, D. C.

Feeds and feeding stuffs: A. C. Summers, Columbia, S. C.

Feed adulteration: Carleton Cutler, W. Lafayette, Ind.

Crude fiber: C. K. Francis, Stillwater, Okla.

Food adulteration:

Colors: W. E. Mathewson, New York, N. Y.

Saccharine products: F. L. Shannon, Lansing, Mich.

Fruit products: P. B. Dunbar, Washington, D. C.

Wine: B. G. Hartmann, Chicago, Ill.

Beer: H. S. Paine, Washington, D. C.

Distilled liquors: A. B. Adams, Washington, D. C.

Vinegar: C. M. Bradbury, Richmond, Va.

Flavoring extracts: A. E. Paul, Chicago, Ill.

Spices: H. E. Sindall, Philadelphia, Pa.

Baking powder: H. E. Patten, Washington, D. C.

Meat and fish: E. D. Clark, Philadelphia, Pa.

Fats and oils: R. H. Kerr, Washington, D. C.

Dairy products: Julius Hortvet, St. Paul, Minn.

Cereal products: L. A. Fitz, Manhattan, Kans.

Vegetables: W. D. Bigelow, Washington, D. C.

Cocoa and cocoa products: H. C. Lythgoe, Boston, Mass.

Tea and coffee: J. M. Bartlett, Orono, Me.

Preservatives: A. F. Seeker, New York, N. Y.

Heavy metals in foods: E. L. P. Treuthardt, Washington, D. C.

Sugar: M. N. Straughn, Washington, D. C.

Insecticides: O. B. Winter, East Lansing, Mich.

Inorganic plant constituents: R. W. Thatcher, St. Paul, Minn.

Medicinal plants and drugs:

Synthetic products: W. O. Emery, Washington, D. C.

Medicated soft drinks: W. F. Sudro, Fargo, N. Dak.

Medicinal plants: Frank Rabak, Washington, D. C.

Alkaloids: H. C. Fuller, Washington, D. C.

Balsams and gum resins: E. C. Merrill, Washington, D. C.

Pepsin: V. K. Chestnut, Washington, D. C.

Water: H. P. Corson, Urbana, Ill.

Testing chemical reagents: C. S. Lykes, Clemson College, S. C.

PERMANENT COMMITTEES.

Coöperation with Other Committees on Food Definitions.

William Frear, State College, Pa., *Chairman.*

Julius Hortvet, St. Paul, Minn.

J. P. Street, New Haven, Conn.

Recommendations of Referees and Revision of Methods.

(Figures in parentheses refer to year in which appointment expires.)

P. F. Trowbridge, *Chairman.*

SUBCOMMITTEE A: A. J. Patten (1918), W. W. Skinner (1916), *Chairman, Bureau of Chemistry, Washington, D. C.*, B. B. Ross (1920). (Phosphoric acid, nitrogen, potash, soils, inorganic plant constituents, insecticides, water.)

SUBCOMMITTEE B: R. E. Stallings (1918), *Chairman, Department of Agriculture, Atlanta, Ga.*, P. F. Trowbridge (1916), H. C. Lythgoe (1920). (Dairy products, feeds and feeding stuffs, sugar, water in foods and feeding stuffs, organic and inorganic phosphorus in foods, feeding stuffs, and drugs, separation of nitrogenous substances, testing chemical reagents, medicinal plants and drugs.)

SUBCOMMITTEE C: L. M. Tolman (1918), H. E. Barnard (1916), *Chairman, State Board of Health, Indianapolis, Ind.*, R. E. Doolittle (1920). (Food adulteration.)

SPECIAL COMMITTEES.

Editing Methods of Analysis (U. S. Bur. Chem. Bull. 107, Rev.).

R. E. Doolittle, New York, N. Y., *Chairman.*

W. A. Withers, Raleigh, N. C.

J. P. Street, New Haven, Conn.

A. F. Seeker, New York, N. Y.

G. W. Hoover, Chicago, Ill.

B. L. Hartwell, Kingston, R. I.

Proposed Journal of Agricultural Chemistry.

C. H. Jones, Burlington, Vt., *Chairman.*

R. N. Brackett, Clemson College, S. C.

C. L. Alsberg, Washington, D. C.

Julius Hortvet, St. Paul, Minn.

E. F. Ladd, Fargo, N. Dak.

Availability of Phosphoric Acid in Basic Slag.

C. B. Williams, West Raleigh, N. C., *Chairman.*

C. G. Hopkins, Urbana, Ill.

H. D. Haskins, Amherst, Mass.

B. L. Hartwell, Kingston, R. I.

J. A. Bizzell, Ithaca, N. Y.

Review of the Analysis of Lime-Sulphur Solutions.

R. J. Davidson, Blacksburg, Va., *Chairman.*

C. S. Cathcart, New Brunswick, N. J.

H. H. Hanson, Orono, Me.

OFFICERS, REFEREES, ASSOCIATE REFEREES AND COMMITTEES OF THE ASSOCIATION OF OFFICIAL AGRICULTURAL CHEMISTS, FOR THE YEAR ENDING NOVEMBER, 1916.

Honorary President.

H. W. WILEY, Woodward Building, Washington, D. C.

President.

R. N. BRACKETT, Clemson College, S. C.

Vice President.

J. K. HAYWOOD, Bureau of Chemistry, Washington, D. C.

Secretary-Treasurer.

C. L. ALSBERG, Bureau of Chemistry, Washington, D. C.

Additional Members of the Executive Committee.

W. J. JONES, Jr., La Fayette, Ind.

E. B. HOLLAND, Amherst, Mass.

Referees.

Phosphoric acid: W. J. Jones, Jr., La Fayette, Ind.

Nitrogen:

Determination: H. D. Haskins, Amherst, Mass.

Separation of nitrogenous substances: L. L. Van Slyke, Geneva, N. Y.

Potash: E. E. Vanatta, Columbia, Mo.

Soils: J. W. Ames, Wooster, Ohio.

Dairy products: Harry Klueter, Dairy and Food Department, Madison, Wis.

Foods and feeding stuffs: A. C. Summers, Department of Agriculture, Commerce and Industries, Columbia, S. C.

Food adulteration: Julius Hortvet, Dairy and Food Commission, St. Paul, Minn.

Sugar: C. A. Browne, Sugar Trade Laboratory, New York, N. Y.

Insecticides: R. C. Roark, Bureau of Chemistry, Washington, D. C.

Inorganic plant constituents: A. J. Patten, East Lansing, Mich.

Medicinal plants and drugs: W. O. Emery, Bureau of Chemistry, Washington, D. C.

Water: W. W. Skinner, Bureau of Chemistry, Washington, D. C.

Water in foods and feeding stuffs: W. J. McGee, U. S. Custom House, Savannah, Ga.

Testing chemical reagents: C. O. Ewing, Bureau of Chemistry, Washington, D. C.

Organic and inorganic phosphorus in foods, feeding stuffs and drugs: E. B. Forbes, Wooster, Ohio.

Lime requirements of soils: F. P. Veitch, Bureau of Chemistry, Washington, D. C.

Associate Referees.

Phosphoric acid: C. S. Lykes, Clemson College, S. C.

Nitrogen:

Determination: R. B. Deemer, La Fayette, Ind.

Special study of Kjeldahl method: I. K. Phelps, Bureau of Chemistry, Washington, D. C.

Separation of nitrogenous substances:

Milk and cheese: Leroy S. Palmer, Columbia, Mo.

Meat products: P. F. Trowbridge, Columbia, Mo.

Potash:

Determination: T. D. Jarrell, College Park, Md.

Soils:

Nitrogenous compounds: C. B. Lipman, Berkeley, Cal.

Dairy products: J. T. Keister, Bureau of Chemistry, Washington, D. C.

Foods and feeding stuffs: P. H. Smith, Amherst, Mass.

Feed adulteration: Carleton Cutler, W. La Fayette, Ind.

Crude fiber: C. K. Francis, Stillwater, Okla.

Food adulteration:

Colors: W. E. Mathewson, Bureau of Chemistry, Washington, D. C.

Saccharine products: (not appointed).

Fruit products: P. B. Dunbar, Bureau of Chemistry, Washington, D. C.

Wine: B. G. Hartmann, Transportation Building, Chicago, Ill.

Beer: H. S. Paine, Bureau of Chemistry, Washington, D. C.

Distilled liquors: A. B. Adams, Bureau of Internal Revenue, Washington, D. C.

Vinegar: E. H. Goodnow, Federal Office Building, Minneapolis, Minn.

Flavoring extracts: A. E. Paul, Transportation Building, Chicago, Ill.

Spices: H. E. Sindall, Weikel & Smith Spice Co., Philadelphia, Pa.

Baking powder: H. E. Patten, Bureau of Chemistry, Washington, D. C.

Meat and fish: E. D. Clark, 1833 Chestnut Street, Philadelphia, Pa.

Fats and oils: R. H. Kerr, Bureau of Animal Industry, Washington, D. C.

Dairy products: Julius Hortvet, Dairy and Food Department, St. Paul, Minn.

Cereal products: J. A. LeClerc, Bureau of Chemistry, Washington, D. C.

Vegetables: W. D. Bigelow, National Cannery Association, Washington, D. C.

Cocoa and cocoa products: E. Bloomberg, Federal Building, Buffalo, N. Y.

Tea and coffee: H. M. Loomis, Eastport, Me.

Preservatives: A. F. Seeker, U. S. Appraiser's Stores, New York, N. Y.

Metals in foods: David Klein, State Food Commission, Chicago, Ill.

Sugar: M. N. Straughn, Bureau of Chemistry, Washington, D. C.

Insecticides: O. B. Winter, East Lansing, Mich.

Inorganic plant constituents: R. W. Thatcher, University Farm, St. Paul, Minn.

Medicinal plants and drugs:

Synthetic products: W. O. Emery, Bureau of Chemistry, Washington, D. C.

Medicated soft drinks: W. W. Skinner, Bureau of Chemistry, Washington, D. C.

Medicinal plants: A. Viehoever, Bureau of Chemistry, Washington, D. C.

Alkaloids: H. C. Fuller, Institute of Industrial Research, Washington, D. C.

Balsams and gum resins: E. C. Merrill, Bureau of Chemistry, Washington, D. C.

Pepsin and papain: V. K. Chesnut, Bureau of Chemistry, Washington, D. C.

Water: H. P. Corson, Grove City, Pa.

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PRESIDENT'S ADDRESS.¹

THE STORAGE OF SUGAR IN THE MAPLE TREE.

By C. H. JONES (Agricultural Experiment Station,
Burlington, Vt.), *President*.

Maple sugar is obtained mainly from the species of maple known to botanists as *Acer saccharum*. Census returns indicate a total yearly production in the United States of the equivalent of some 45,000,000 pounds. The more northern sections contribute most largely to the supply. The yield is subject to extreme variations, being dependent on certain daily weather conditions during a few weeks in early spring. Although insignificant in amount when compared with the world's production of cane and beet sugars, yet as a table luxury maple products have long held an honored place among our food products.

ECONOMIC INVESTIGATIONS.

Economic investigations connected with the production of maple sugar and sirup have occupied the attention of many students of the subject and their observations have been recorded in various publications of a scientific and popular nature. They include data on the history of the maple industry, composition and flow of maple sap, the action of micro-organisms on sap and their effect on the finished product, together with a vast amount of strictly chemical data directed toward a proper standardization of the marketable product and the detection of adulteration therein.

Chemically speaking, the sugar from the maple has been shown to be identical in composition with the ordinary granulated sugar secured from the cane and beet. The importance it holds as a delicacy is due to a characteristic and pleasing flavor associated with it which, however, is destroyed if the usual methods of sugar refining are applied to the crude product.

The nature of the considerations about to be presented is fundamental rather than economic; fundamental in that they deal with the tree itself and seek to indicate the various changes taking place within the trunk at different seasons of the year and their visible manifestations as indicated by pressure and sap flow. Before proceeding further a brief description of maple wood will be given. No attempt is made to describe the several structures in detail but attention is directed to such tissues as seem most directly concerned with our general subject.

¹ Presented Tuesday, November 16, as special order of business for 11.30 o'clock.

GENERAL STRUCTURE.

In general structure, the maple trunk or stem (*xylem*) may be divided into a brown heart wood (*duramen*) and a white sap wood (*alburnum*). The sap wood contains the living cells and is the main conducting medium for water and food. The heart wood is relatively lifeless and functions largely as a mechanical support for the crown and as a reservoir in which to store water during the winter months.

The stem is composed of three kinds of cells, viz., wood fibers, wood parenchyma and vessels, without intercellular spaces. At right angles to these cells, extending from the circumference toward the heart, are the *medullary rays*. These are *living cells*. They vary greatly in width and length and serve as the main centers for the storage of starch. Measurements of the medullary rays in the maple have shown that they occupy about 18% by volume of the outer or recently formed sap wood.

The starch grains found in the maple are angular, not uniform in shape, and vary in size from 1.4-5.7 microns in diameter. The characteristic blue color is obtained by treatment with iodine solution. Starch is stored not only in the medullary rays but also in the wood (parenchyma) which in some cases forms pockets, visible to the naked eye. These as a rule extend at right angles to and border on the medullary rays. Starch may also be found in the rows of cells (from three to many) formed in the summer wood of the past few years.

Between the medullary rays are longitudinal cells often of great length containing water and food ready for transference throughout the tree. They are known as *tracheae*. Associated with these are similar but shorter cells known as *tracheids*. Both *tracheae* and *tracheids* are *dead cells*. They comprise about 7% by volume of the sap wood.

CHEMICAL PHASES OF SUGAR STORAGE.

A consideration of the more strictly chemical phases of sugar storage in the maple will indicate conditions existing within the tree during the year. The specimens used in determining this matter were second growth maples, quite uniform as to size, age and leaf area, which grew near each other, under similar general conditions of soil formation and exposure. Differences due to these causes were thus minimized and as samples were taken monthly for 29 months a period is represented including 3 successive sugar seasons, 2 full periods of growth and 3 periods of rest. To further confirm these observations precautions were taken during the second 12 months to secure 2 trees on the dates which the first year's work indicated to be what may be termed critical points in the yearly cycle.

The several trees were felled, measured and immediately brought to

the laboratory where suitable sections representing the base (outer and inner wood) and the top wood were reduced to sawdust by means of a circular saw. The samples thus prepared were subjected to chemical analysis directed primarily to ascertain the content of moisture, sugar, and starch, which are assumed to be the so-called readily available stored foods. The last transformation of the sugar molecule used for plant food purposes is outside a strictly chemical province, being rather a problem for the plant physiologist.

While variations in the results obtained for corresponding months in different years were noted, and individual differences were occasioned by samples and natural fluctuations in soil moisture, sunlight and temperature conditions, yet the same general trend of all results was plainly manifested during successive seasons.

Instead of burdening you with a mass of data representing monthly conditions for 29 months the results have been summarized in a brief table where the months of the year are divided into 4 groups, each group representing a distinct epoch in the life functions of the tree.

The first division includes the months of December, January and February, the winter months, during which interval the tree is generally considered dormant as regards development. The second division comprises the months of March and April. The sap flow occurs during this

TABLE 1.

Percentage of available carbohydrate food in maple wood.

DETERMINATION	DECEMBER, JANUARY AND FEBRUARY	MARCH AND APRIL	MAY, JUNE, JULY AND AUGUST	SEPTEMBER, OCTOBER AND NOVEMBER
OUTER BASE WOOD:				
Water.....	35.01	32.86	30.30	29.80
Sucrose.....	0.94	0.80	0.36	0.72
Reducing sugars.....	1.40	0.53	0.13	0.41
Starch.....	0.78	0.96	1.01	1.43
INNER BASE WOOD:				
Water.....	30.28	29.64	26.87	29.05
Sucrose.....	0.49	0.52	0.18	0.41
Reducing sugars.....	0.65	0.33	0.09	0.26
Starch.....	0.43	0.49	0.62	0.51
TOP WOOD (outer and inner):				
Water.....	33.85	31.36	32.46	32.14
Sucrose.....	1.12	1.04	0.41	0.83
Reducing sugars.....	1.79	0.67	0.23	0.59
Starch.....	1.09	1.33	1.23	1.63
ROOTS:				
Water.....	33.07	35.46	31.00	30.23
Sucrose.....	1.52	0.93	0.66	1.41
Reducing sugars.....	1.00	0.79	0.42	0.32
Starch.....	3.92	2.25	3.41	5.76

period which, in northern latitudes, is designated the sugar season. The tree is preparing for its yearly work of growth and reproduction. The third period, May, June, July and August, represents the time of bud and leaf development, the formation of the annual ring and the storage of reserve food. The fourth division, September, October and November, is a period of gradual preparation for the cold of winter. Early in this period, as the leaves fall, reserve foods are deposited largely in the form of starch which later may be changed in part to sugar.

SUCROSE IN TRACHEAE AND TRACHEIDS.

When fresh maple wood is taken from a tree, for example during March or April, suitably pulverized, and treated with water, the resulting solution is found to contain both reducing sugars and sucrose. Maple sap as it flows from the tree has been shown repeatedly to be practically free from reducing sugars, but to contain sucrose. This would indicate that there is a separation of the two sugar solutions in the tree.

By taking a suitable section of maple wood and subjecting it to a water pressure of about 30 pounds, it was found that only sucrose was removed, no reducing sugars being washed out. The amount of sucrose thus obtained depended to some extent on the quantity of water forced through the stem. Three trials on different samples using 1250, 900 and 2500 cc. of water removed 28, 51 and 67% respectively of the total sucrose present. It would seem, therefore, that sucrose must be stored chiefly in the *tracheae*, the long longitudinal tubes, to which mention has been made, while the reducing sugars must be contained in the cell sap of the living cells.

CONSIDERATION OF RESULTS.

From a study of Table 1 it is clear that marked variations exist in the water, sugar and starch content of wood in different parts of the maple tree at similar periods and also that the composition is not constant during the year. Further, that the top wood is supplied more abundantly with sugars and starch than either the outer or inner base wood.

Considering the results obtained in the periods indicated, it is noted that the amount of water present is greatest in the outer, inner and top wood sections during the cold winter months, December to February. Similarly, the sugars are also higher. The starch, on the other hand, is less at this time than at other intervals during the year. The percentage of water is lowest in the inner wood but if account is taken of the total bulk of wood represented a large excess of water would be found in the inner, over that contained in the outer, base tissues.

The March-April period shows a slight reduction in the water content of the several sections. The sugar percentages continue to decrease as warmer weather approaches, while the starch content increases.

The third period, representing the season of growth, indicates that the amount of water in the tree is at its minimum, large amounts having been given off by transpiration. The percentage amount of sugars present is small but it remains quite uniform during the growing season. The starch content is practically unchanged in the outer and top wood, while the inner tissues show an increase. The small amount of sugars present during this period of active photosynthesis is explained first, by the fact that these constituents have been drawn on to feed the developing buds and leaves and, secondly, by the fact that the elaborated food is used immediately in the development of the annual ring.

The last period, September to November, finds the water content of the outer and top wood remaining constant while a decided gain in moisture is observed in the inner section. The sugars show an increase of some 130% over amounts present during the growing season. In like manner the starch percentages have become augmented in the outer and top sections.

The roots exhibit the highest water content in March and April, the highest sugar content from December to February and the lowest between May and August. The starch content of the roots greatly exceeds that found in the trunk wood regardless of season. It is most abundant during the autumn months, September to November, when an average percentage of 5.76 was obtained. This gradually decreases during the cold winter months and is lowest during the period just preceding the renewal of spring activities when but 2.25% was indicated.

RELATION OF SUCROSE TO REDUCING SUGARS.

There is a condition shown by the results in Table 1 to which special attention should be directed and that is the relation between the percentages of sucrose and reducing sugars and the relation of both to the starch content. Reducing sugars are greater in amount than is sucrose during only one period of the year, December to January. Sucrose predominates throughout the remaining months. The change in relativity begins just previous to the sugar season, March and April. Coincident with the decrease in reducing sugars an increase in starch percentage is observed. The roots throughout the year show a predominance of sucrose over reducing sugars, particularly in the autumn months of September, October and November. Mention already has been made of the large starch content.

DENSITY OF SAP SOLUTIONS.

Investigators are most familiar with the composition of maple sap, secured during March and April by the ordinary tapping process, from that part of the tree designated in Table 1 as outer base wood. It is

commonly known that maple sap is essentially a dilute solution of sucrose containing small amounts of mineral matter and that its sugar content varies from 1-7% with an average of approximately 3%. Reference to Table 1 shows that but 0.80% of sucrose was present in the basewood. This amount, however, must have been held in solution by the water in the tree trunk, as were the reducing sugars, and a simple calculation will show the sucrose content of the sap to have been 2.34%. This represents an average for several young trees during 3 successive sugar seasons.

The grouping of averages by periods in Table 1 does not emphasize the high concentrations obtained on certain dates. Thus, when the individual tree data are consulted, instances are not lacking which show that this concentration during the sugar season ran as high as 5% of sucrose and that even a total sugar concentration of 12% occurred during extremely cold weather.

In order that the density of the sugar solutions found in the tree may be readily compared the results already tabulated have been calculated to a concentration or density basis.

TABLE 2.
Percentage density of sugar solutions in maple wood.

DETERMINATION	DECEMBER, JANUARY AND FEBRUARY	MARCH AND APRIL	MAY, JUNE, JULY AND AUGUST	SEPTEMBER, OCTOBER AND NOVEMBER
OUTER BASE WOOD:				
Sucrose.....	2.52	2.34	1.17	2.33
Reducing sugars.....	3.75	1.55	0.42	1.33
INNER BASE WOOD:				
Sucrose.....	1.56	1.71	0.66	1.38
Reducing sugars.....	2.07	1.08	0.33	0.87
TOP WOOD (outer and inner):				
Sucrose.....	3.05	3.14	1.24	2.47
Reducing sugars.....	4.87	2.03	0.69	1.76
ROOTS:				
Sucrose.....	4.27	2.50	2.06	4.41
Reducing sugars.....	2.81	2.12	1.31	1.00

The same general variations already discussed are again in evidence but these figures more nearly represent the density of the sucrose and reducing sugar solutions found in the cells and tissues composing the tree structure. Starch, being insoluble in water, is not included in this calculation although one would be justified in so including it for it is well known that starch is one of the reserve foods stored by the maple and that it must be transformed into soluble sugars before it can be transported.

DENSITY OF SAP INCREASES FROM BASE TO TOP OF TREE.

The ascent of water and food to the tops of tall trees has long occupied the attention of scientists. No explanation, entirely satisfactory, thus far has been presented although many theories have been advanced. Data as to the amounts of soluble sugars and water in different parts of the tree undoubtedly have a bearing on this question. The increased carbohydrate concentration in the top wood of the maple over that in the base section has been noted. For several reasons, particularly when the structure of the wood and region of consumption of food are considered, it seems logical to assume that this increase in concentration of the sugars, together with the higher starch content, should be a gradual one, as we ascend the tree.

A small maple tree cut early in May was examined in sections of 4 feet each, from the base to the top, with the following results:

TABLE 3.
Percentage density of sugar solutions.

SECTION	WATER	STARCH	REDUCING SUGARS	SUCROSE	TOTAL SUGARS
1st (base).....	29.07	0.83	0.10	1.82	1.92
2nd.....	33.15	0.81	0.21	1.78	1.99
3rd.....	28.97	0.92	0.37	2.26	2.63
4th.....	26.85	1.06	0.32	2.92	3.24
5th (top).....	25.84	1.10	0.30	3.10	3.40

Table 3 would seem to indicate that the sugar concentration of both active cell, and *tracheae*, sap gradually increases from the lower portions of the tree trunk upward. Starch also shows a similar increase. On the other hand, water, in this instance, decreases in amount as we ascend.

INTERPRETATION OF RESULTS.

It is a relatively easy matter to present for consideration data secured by physical and chemical operations in the field and laboratory but the significance of such data when applied to the solution of nature's processes is not so readily formulated. A lengthy interpretation of the results bearing on the phenomena of sugar formation and sap flow will not be attempted but a few general deductions enumerated.

The work of numerous investigators has shown clearly that sunlight, leaf area and soil moisture are factors influencing the abundant storage of starch and sugars. A correlation table, showing relationships between stored carbohydrates on the one hand, and sunlight, leaf area and rainfall on the other, undoubtedly would give most suggestive results. It is a different matter to prove the relationship from a practical or economic standpoint as weather conditions at the time of sap flow seriously affect

results. In other words, one can determine with reasonable certainty that an abundance of carbohydrates has been stored in the tree but the best of prophecies fail when attempts are made to predict the amount of sugar likely to be secured the following spring.

The yearly life cycle of the maple tree has for its objects nutrition and reproduction. Green leaves manufacture starch daily in sunlight. At night it is transformed to soluble carbohydrates and carried down the inner bark and cambium layer where it is again deposited in the medullary rays as starch or remains in solution in the tree tissues. The excess only is thus deposited, what is necessary being used for the formation of the annual ring.

When the leaves fall in the autumn a protective enzyme action may take place in the medullary rays, and sucrose is ultimately formed and distributed through the *tracheae*. The increased concentration of the sugar solutions in the maple trunk during the cold weather period appears to have for its purpose the protection of the tree tissue from freezing, so that an early transference of abundant water and food materials to the leaf-bearing sections of the tree may be established. The maple is the earliest of northern trees to develop its buds and leaves in spring.

The rate of movement of sap in the tapped tree has been measured by injecting lithium chlorid and noting the time of its appearance in the sap from holes at known distances above and below the point of insertion. A rate of 2-6 inches per minute was observed. Since this flow is an artificial one, stimulated nevertheless by conditions within the tree, it does not follow that sap movement up and down within the untapped tree trunk, before leaf formation has started, parallels these rates. Doubtless the water movement is far less rapid, although there is warrant for asserting that changes in sugar concentration and water content in different sections of the tree are taking place constantly.

Considerable work has already been done touching the matter of direction of sap flow and pressure under the artificial conditions created by the ordinary tapping process. It has been shown that during certain weather conditions in early spring, characterized by a sharp frost at night and a rise in temperature to a few degrees above the freezing point the following day, a state of pressure is manifested in the tree trunk and sap is exuded at the tap hole or point of least resistance. This pressure may vary from 1-25 pounds and while the flow of sap is not necessarily in direct proportion to the pressure, there is strong evidence nevertheless that some relation exists. During the greater part of the year a state of negative pressure or suction is exerted by the tree.

The simple determination of the water content of maple wood at different seasons of the year is not without interest. It is more than a coincidence and doubtless for a definite purpose that the highest water

content always precedes the leaf formation and transpiration period and that the lowest amount is present when the leaves have ceased their activities.

Beginning in December, the reducing sugars exceed the sucrose content of maple wood until March when a decided decrease occurs in the percentage of reducing sugars. A relatively high percentage of sucrose obtains until May. A predominance of sucrose over reducing sugars, though in reduced amounts, is maintained through the summer and fall, until, in December, the reducing sugars again take the lead. Thus a high sugar content is found during the coldest period, reducing sugars predominating over sucrose, the ratio of reducing sugars, starch and sucrose being 1.5 : 0.8 : 1. As warm weather approaches the total sugar content decreases and a most pronounced increase of sucrose over reducing sugars occurs, the above ratio becoming 0.7 : 1.2 : 1.

Warm weather favors the deposition of starch from soluble carbohydrates. Continued cold weather causes the insoluble starch to decrease in amount, transforming it to soluble carbohydrates, thus increasing the total sugar content during the winter months.

From the chemical evidence presented it is plain that changes are taking place within the tree throughout the year. A fall deposition of starch occurs. Later, as cold weather approaches, the leaf-manufactured food that has been deposited as starch is changed in the living protoplasmic cells to reducing sugars and sucrose which, held in a water solution, fill the vessels throughout the tree. These sugars are most abundant in the young living tissues of the trunk nearest the cambium layer.

Reducing sugars and starch seem to be most intimately associated with the living protoplasmic cells, while a considerable portion at least of the elaborated sucrose fills the dead vessels. These channels, it is well established, serve as quick carriers for water and sucrose to upper and distant parts of the tree, and are in direct communication with the roots or water-gathering system. The uniformity of sap concentration during the four months comprising the growing season indicates that the leaf elaborated carbohydrates are used, either directly or indirectly, for the development of the annual ring. Until that duty is performed no marked increase in soluble carbohydrate storage is noted.

OUR JOURNAL.

In conclusion I desire to say a few words on the subject of our new JOURNAL. The problems confronting us and the ways and means for dealing with them were ably considered by my predecessor in his annual address one year ago. As you all know this association now has a quarterly publication and our deliberations, printed in a most attractive form,

will soon be placed promptly at our disposal. This publication is not a one man volume and every member should be allowed and encouraged to do his share. It is needless for me to urge the members to subscribe but I do want to ask particularly that the younger members not only subscribe but also read and study the JOURNAL with a view toward detecting shortcomings and rectifying them in the future.

It is too early to predict the ultimate outcome of our venture. We have men enough, brains enough and, I hope, a loyalty to our chosen profession sufficient to carry on this project.

PROCEEDINGS OF THE THIRTY-SECOND ANNUAL CONVENTION OF THE ASSOCIATION OF OFFICIAL AGRICULTURAL CHEMISTS, 1915.

FIRST DAY.

MONDAY—MORNING SESSION.

The thirty-second annual convention of the Association of Official Agricultural Chemists was called to order by the President, C. H. Jones, of Burlington, Vt., on the morning of November 15, 1915, at the Raleigh Hotel, Washington, D. C.

REPORT ON WATER.

THE SEPARATION AND DETERMINATION OF CALCIUM AND STRONTIUM.

By W. W. SKINNER (Bureau of Chemistry, Washington, D. C.), *Referee*.

As a result of the experimental work reported by the referee last year, the Stromeyer-Rose method for the separation and determination of calcium and strontium was modified by substituting for cold water heretofore used a 1% solution of ammonium oxalate for washing the precipitate of the mixed oxalates. As shown by Holleman, Kohlrausch, Richards and others, the solubility of calcium oxalate in cold water and also in hot water is so low that it may be disregarded as a factor of accuracy in ordinary analytical work. The error in the determination of calcium, due to the solubility of the oxalate, should not be in excess of 1 mg. of calcium when the quantity of precipitate is such that 200 cc. or less of wash water are required.

Strontium oxalate, however, as shown by the experimental work reported last year, is quite soluble in hot water and sufficiently soluble in cold water to cause serious error in the determination of strontium. This is particularly true when relatively small amounts of strontium are to be separated from large amounts of calcium, requiring a greater amount of washing of the precipitate. The method was further modified by providing for 2 extractions of the mixed nitrates with the ether-alcohol mixture, which increased the quantity of solvent used.

The method as modified is as follows:

DETERMINATION OF CALCIUM AND STRONTIUM.

Calcium.

Combine the 2 filtrates obtained from the precipitates of iron and aluminium, concentrate to 150–200 cc. and to this solution, containing not more than 0.6 gram of calcium, calculated as calcium oxid, or 1 gram of magnesium, calculated as magnesium pyrophosphate, add 1–2 grams of oxalic acid and sufficient hydrochloric acid to clear the solution. Heat to boiling and neutralize with ammonium hydroxid, stirring constantly. Add ammonium hydroxid in slight excess and allow to stand for 3 hours in a warm place. Filter off the supernatant liquid and wash the precipitate once or twice by decantation with 1% ammonium oxalate solution. Dissolve the precipitate in hydrochloric acid, dilute to 100–200 cc., add a little oxalic acid, and precipitate as above. After standing for 3 hours, filter, wash with the ammonium oxalate solution as before, dry, ignite, heat over a blast lamp, and weigh as calcium and strontium oxids. Subtract from this weight the weight of strontium oxid equivalent to the strontium sulphate found. The difference is the weight of calcium oxid. Calculate to calcium.

As a check on the calcium oxid, evaporate to dryness the combined filtrates from the strontium nitrate, dissolve the calcium nitrate in water, precipitate as oxalate, filter, wash, ignite and weigh as calcium oxid.

Strontium.

Dissolve in dilute nitric acid the oxids obtained in the determination of calcium and test with the spectroscope for strontium. If strontium is present, transfer the nitric acid solution to a small Erlenmeyer flask. Evaporate nearly to dryness over a low flame and heat in an air bath at 150°–160°C. for 1 or 2 hours after the water is evaporated. Break up the dried material with a stirring rod, add 10–15 cc. of a mixture of equal parts of absolute alcohol and ether to dissolve the calcium nitrate. Cork the flask and allow to stand with frequent shaking for 2 hours or longer. Decant the solution through a 5.5 cm. filter, preserving the filtrate. Wash the residue several times by decantation with small portions of ether-alcohol solution. Dry the residue and the filter paper and wash the filter paper repeatedly with small portions of hot water, collecting the filtrate in the flask containing the main portion of the strontium nitrate residue. Add 1 or 2 drops of dilute nitric acid, evaporate, dry, pulverize, and treat with 10–15 cc. of ether-alcohol mixture as before. Cork the flask and let stand for about 12 hours with occasional shaking. Filter, wash with ether-alcohol mixture until a few drops of the filtrate evaporated on a watch glass leave practically no residue. Dry the paper and precipitate. Dissolve the strontium nitrate in a few cc. of hot water. Add a few drops of sulphuric acid, then a volume of alcohol equal to the volume of the solution and allow to stand for 12 hours. Filter, ignite, weigh as strontium sulphate and calculate to strontium. Test spectroscopically for absence of calcium.

A copy of this method, together with the sample specially prepared for the purpose, was sent to 10 chemists who had signified a desire to undertake the coöperative work and reports have been received from 6 of them.

The solution, prepared for the coöperative work contained in the aliquot taken for examination 203 mg. of calcium and 21.8 mg. of strontium.

Results of coöperative work on calcium and strontium.

ANALYST	CALCIUM BY DIFFERENCE	CALCIUM DIRECT	STRONTIUM
	<i>Mg. in 10 cc.</i>	<i>Mg. in 10 cc.</i>	<i>Mg. in 10 cc.</i>
H. P. Corson, Illinois State Water Survey, Urbana, Ill.	200.9	198.6	21.9
O. B. Winter, Agricultural Experi- ment Station, East Lansing, Mich.	204.6 203.9	197.1 196.1	17.1 16.9
E. L. Griffin, Department of Agri- culture, Washington, D. C.	203.2 203.5 203.4	201.5 200.1 203.2	18.1 18.1 17.9
P. L. Hibbard, Agricultural Experi- ment Station, Berkeley, Cal.	203.2 202.3 202.8 202.9	lost lost lost 201.8	20.0 21.9 21.0 21.0
J. W. Sale, Bureau of Chemistry, Washington, D. C.	203.9 203.8 202.8 203.7	202.6 203.2 202.9 202.7	19.1 18.8 19.0 19.6
W. F. Baughman, Bureau of Chem- istry, Washington, D. C.	204.1 203.7 204.3	202.1 202.1 201.9	19.3 20.1 20.3
Maximum.....	204.6	203.2	21.9
Minimum.....	200.9	196.1	16.9
Average.....	203.3	201.1	19.4
Theory.....	203.0	203.0	21.8

By referring to the table it will be noted that the average of the 17 results reported for calcium by difference varies from theory by only 0.3 mg. or 0.15%; while the difference between the highest and the lowest result is only 3.7 mg. These results are as good as could be expected and confirm the observations of the referee upon the results reported in 1913 to the effect that the method for the separation of calcium from strontium and the determination of calcium by the indirect method leaves little to be desired. The calcium direct is slightly low, the average for the 14 results reported being 1.9 mg. or approximately 0.93% below theory. The cause of the low results may be due to an imperfect extraction by the ether-alcohol mixture of the calcium nitrate from the mixed calcium and strontium nitrates but, if so, it apparently does not affect the final result for strontium.

The results reported for strontium while uniformly slightly lower than theory are fairly satisfactory. The results reported by the 6 analysts at this time are fairly concordant and much superior to the results reported in 1913. The work this year and that of previous years shows conclusively that the separation of the calcium and strontium nitrates

by the 2 or more treatments with the ether-alcohol mixture is quite satisfactory, and also that the modified method, substituting a 1% ammonium oxalate solution for cold water in the washing of the mixed oxalates, has eliminated the error in the determination of strontium due to the solubility of strontium oxalate in washing. And while it must be admitted the method gives slightly low results for strontium, due probably to the fact that strontium does not completely precipitate as oxalate,—considerably less so than does calcium,—the results of the investigation and study of the method extending over a period of 4 years are such as to warrant the Association in adopting the method as official and it is so recommended by the referee. The method is presented for its first reading for final adoption as provided in the By-Laws of the Association.

DISCUSSION.

Mr. Skinner: The Committee on Editing Methods of Analysis, Mr. Doolittle, Chairman, early in the year asked the referee to review the methods for the analysis of water. This review showed our methods to be incomplete in that certain standard methods in general use had never been proposed for coöperative work by any referee and had therefore never been adopted as official or provisional methods by the Association, and furthermore, that certain new methods had been substituted for older methods that were not at the same time repealed. It appeared advisable, since the methods were to be printed in the JOURNAL, that such standard methods be included for the sake of completeness, even though they had not been adopted by the Association. These methods were inserted therefore and described as “tentative” to distinguish them from the official and provisional methods. The determinations involved are: turbidity, color, odor, oxygen required (when the chlorine content of the sample is high), dissolved oxygen, specific gravity, hydrogen sulphid, free carbon dioxid, temporary hardness, total hardness, permanent or non-carbonate hardness.

It seemed equally advisable to omit from the JOURNAL those methods which have been superseded, but which have not been officially repealed by the Association. The methods involved are the Pettenkofer method for carbon dioxid and the ammonium sulphate method for barium and strontium.

Under the direction of the referee some time has been devoted to a study of methods for the determination of the radioactivity of waters. The referee has asked Mr. Furber of the Water Laboratory of the U. S. Bureau of Chemistry to present a brief statement of the method and the work.

REPORT ON THE DETERMINATION OF RADIOACTIVITY.

By F. B. FURBER (Bureau of Chemistry, Washington, D. C.).

The Water Laboratory of the U. S. Bureau of Chemistry has been concerned for some time with the examination of substances claimed to be radioactive, chiefly in connection with the enforcement of the Federal

Food and Drugs Act. The samples have been the so-called "radioactive" bottled mineral waters and various therapeutic preparations.

This paper is based partly on work reported to the U. S. Bureau of Chemistry, January 1, 1915, by Mr. W. D. Collins. Radioactive measurements made by the writer since that time have confirmed Mr. Collins' results and have also established the value of certain other methods. This brief account of the experience of the laboratory is presented merely to bring the matter to the attention of the Association with the hope that eventually uniform procedures for the measurement of radioactivity may be adopted by the Association.

In connection with the routine work, various methods of measurement have been tried and different forms of apparatus used.

The fontactoscope, an apparatus which comprises a gold leaf system and a very large detachable discharging chamber to hold samples of water from which the emanation is separated by shaking, would seem to be unsuited for accurate laboratory measurements, but in the field it is considered serviceable by many¹. The U. S. Bureau of Chemistry has not found field experiments necessary as yet.

The gamma ray electroscope, which is used in measuring quantities of radium as large as a milligram, has not been needed.

An alpha ray electroscope² used by this laboratory for measurement of the radioactivity of substances in solid form makes possible a very rapid and fairly accurate examination of certain drugs and minerals.

An active solid introduced into the discharging chamber gives off chiefly alpha particles which ionize the enclosed air, thus allowing the charge imposed on a gold leaf to escape rapidly. The rate of fall of the leaf is noted just as with an emanation electroscope.

The electroscopes most used in this laboratory are of the emanation type and resemble one made by Charles W. Cook (Manchester, England) after the pattern of Rutherford³. They were made according to specifications which allowed for some modifications of the original. The comparatively small number of samples to be examined made it unnecessary to use an interchangeable electroscope of the kind described by Lind⁴. Also, since in many samples no radioactivity could be detected, it seemed better to have the measuring apparatus in complete fixed units.

For making accurate measurements of low radioactivity the Water Laboratory has followed the generally used method, which is to separate the radium emanation from the substance to be examined and then, by

¹ Engler, Sieveking and Koenig. *Chem. Ztg.*, 1914, **38**: 425-7, 446-50; *Physik. Z.*, 1914, **15**: 441-7.

² Rutherford. *Radioactive Transformations*. 1906, p. 28.

³ Rutherford. *Radioactive Substances and Their Radiations*. 1913, p. 90.

⁴ *J. Ind. Eng. Chem.*, 1915, **7**: 406-10.

means of an electroscope, to compare the effect of this with the effect of emanation from a known amount of radium.

If the sample to be tested is a mineral water or solution, but little treatment is needed before separating the emanation by boiling. Soluble solids are dissolved in water or acids. Insoluble substances are fused with mixed sodium and potassium carbonates. This fusion is treated with dilute sodium carbonate solution and the residue remaining after filtration is dissolved in acid. This acid solution and the alkaline extract, which is now acidified, are placed in separate flasks, boiled and sealed. After about 4 days, half the emanation has formed and the gases containing it are collected. Boltwood's apparatus¹ for separating and collecting the emanation has been found very satisfactory. If the solution is boiled for about 20 minutes, the emanation appears to be separated completely.

The Schlundt and Moore apparatus² for collecting emanation by boiling it out, does not work properly unless given very close attention. It is possible also to extract emanation from solutions by bubbling air through while heating³ and this method is often more simple and convenient than others. The gases obtained by boiling the solution are kept for 10 minutes, to allow for the decay of any thorium emanation present, and then transferred through a calcium chlorid drying tube to the discharging chamber of an emanation electroscope. The rate of fall of the charged gold leaf is observed about 3 hours after this. The electroscope is calibrated by measuring the rate of discharge caused by a known amount of emanation formed in a standard radium solution. The solution of radium barium bromid in hydrochloric acid (1.10 sp. gr.) used in the U. S. Bureau of Chemistry was furnished by the U. S. Bureau of Standards.

The electroscopes have been calibrated also by the use of pitchblende, the uranium content of which was determined as described by Brearley⁴. A sample of the ore is dissolved in nitric acid and the emanation collected and passed into the electroscope by Boltwood's method. Correction is made for the emanation lost spontaneously by sealing up a quantity of the ore in a glass tube and at the end of a month transferring the collected emanation to an electroscope by means of a stream of air. The temperature at which the sample is kept has an appreciable effect on the amount of emanation given off by the ore. If time permits, this correction may be eliminated by using the method of solution described by Lind and Whittemore⁵ in which the ore is sealed up for a month in the flask where it is to be dissolved. Then, when the emanation in equilibrium

¹ Phil. Mag., 1905, 6th ser., **9**: 599-613; Am. J. Sci., 1904, 4th ser., **18**: 378-87.

² J. Phys. Chem. 1905, **9**: 320-32.

³ Curie. *Traité de Radioactivité*. 1910, **1**: 284.

⁴ Brearley. *The Analytical Chemistry of Uranium*. 1903; see also U. S. Bur. Mines Bull. 70, and J. Am. Chem. Soc., 1914, **36**: 2075-8.

⁵ J. Am. Chem. Soc. 1914, **36**: 2071-2.

with the ore is all confined, solution in acid gives the total emanation corresponding to the radium present.

In many cases, to save time and manipulation, departure has been made from approved methods given in the literature but a detailed discussion is impossible here. For example, in most of the Water Laboratory's work the extra flask of Boltwood's collecting apparatus has been found unnecessary and a plain 100 cc. Erlenmeyer flask has been used for dissolving certain minerals and boiling strongly active solutions.

There has not been, as yet, occasion for making an extended investigation of thorium preparations. Thorium emanation has so short a life that it must be measured by passing it in a continuous stream through an electroscope. The method of Mache and Bamberger¹ seems to be the most satisfactory of any in the literature.

In dealing with mineral waters and most medicinal preparations this laboratory reports radioactivity in millimicrocuries (billionths (10^{-9}) of a curie)² per liter of solution or gram of substance. Many results in the literature have been expressed in mache or electrostatic units, but since so many investigators have not allowed for corrections (chiefly the Duane³ correction) it is hard to place absolute reliance on their results or to make comparisons between results stated in the two different systems. For most purposes, however, the mache unit equals 0.37×10^{-9} curies or the electrostatic unit equals 0.37×10^{-6} curies is accepted as a satisfactory conversion ratio. The Water Laboratory accepts also the uranium-radium ratio of Heinmann and Marekwald⁴. (One gram of uranium equals 332.8×10^{-9} grams of radium.)

In regard to the magnitude of amounts of emanation determined, it may be said that the emanation electroscopes of the Water Laboratory have a limit of sensibility corresponding to 0.005×10^{-9} grams of radium, and will measure easily amounts of emanation corresponding to 20×10^{-9} grams of radium.

Some samples of artificial waters and medicinal preparations have been found to be as radioactive as claimed on the label. Most natural waters, when bottled, lose all their radium emanation after 30 days, because they do not contain dissolved radium salts or insoluble suspended radioactive matter to make them permanently active. Such waters, therefore, and also a number of artificial products, have been found so slightly radioactive at the time of sale that, in comparison with actual doses of radium salts used in medicine, and with the waters of natural springs known for their radioactivity at source, the radioactivity is wholly negligible.

¹ Sitzb. kais. akad. Wiss. Wien. Abt., 1914, **123** (IIa): 334-45.

² One curie is the radioactivity corresponding to one gram of radium element.

³ Compt. rend., 1905, **140**: 581-3; 1910, **150**: 1421-3.

⁴ Physik. Z., 1913, **14**: 303-5.

WATER IN FOODS AND FEEDING STUFFS.

By W. J. McGEE (Food and Drug Inspection Laboratory,
U. S. Custom House, Savannah, Ga.), *Referee*.

The principal object of this year's work was the study of the method of drying without heat in a vacuum over sulphuric acid. The following outline of the method was sent out for trial:

DRYING IN VACUO WITHOUT HEAT.—OPTIONAL.

(This method was first suggested by Mr. Trowbridge at the meeting of the Association in 1909; approved in 1913 for final action as an optional official method in 1914; final action not yet taken.)

Mix the sample thoroughly and weigh about 2 grams by difference from a stoppered weighing bottle into tared crucibles provided with covers which are tared with crucibles. Place 200 cc. of fresh C. P. sulphuric acid in a good 6 inch vacuum desiccator. Put triplicate samples in the desiccator, smear the edges of the latter and the stop-cock with lubricant (a mixture of 3 parts of hard paraffin and 5 of vaseline) and exhaust by means of a vacuum pump. If a pump is not available, place 10 cc. of ether contained in a small beaker in the desiccator and exhaust with a water filter pump. It will be found convenient to interpose between the pump and the desiccator an empty bottle next to the desiccator and a bottle of water following this. Draw the air from the desiccator through the water and turn the desiccator stop-cock at just the instant when the water begins to rise in the tube leading from the empty bottle. Gently rotate the desiccator 4 or 5 times during the first 12 hours to mix the sulphuric acid with the water which has collected as an upper layer. At the end of 24 hours open the desiccator, forcing the incoming air to bubble through C. P. sulphuric acid. If a good vacuum has been maintained the samples are ready for the first weighing. After weighing place in a desiccator with fresh C. P. sulphuric acid and exhaust as before. Rotate the desiccator once or twice during the interval and weigh again at the end of 24 hours, repeating this process of drying in vacuo over sulphuric acid until the weight is constant.

REPORTS OF COLLABORATORS.

W. D. Richardson: Sausage meat (vacuum method).—Constant weight in about 208 hours with a loss in weight of about 1% less than by the heating methods.

D. B. Bisbee: Cheese (vacuum method).—Weight nearly constant eighth day. Loss of weight about 0.4% less than by heating methods.

J. O. Clarke: Cottonseed meal (vacuum method).—Weight practically constant in 9-15 days with loss about the same as in heating methods.

Corn meal.—In vacuum desiccator dried to approximately constant weight in 4 days and the loss was about 0.4% less than by heating methods.

Tomato ketchup.—With water content of 83.11% as indicated by heating at 100°C. in water oven, lost 80.15% in vacuo over sulphuric acid in 5 hours, and 81.21% in 5 days. In another experiment lost 80.53% in 18 hours and 80.72% in 42 hours. Both of these experiments were terminated by accident before constant weight was attained.

Hamburger steak.—In vacuo over sulphuric acid lost 70.42% of its weight in 4 days; by heating methods, 71.09%.

Lean meat.—Ground fine lost 65.56% in 44 hours and 67.10% by heating methods.

Apple jelly.—Lost 21.66% in 48 hours and 27.45% in vacuum oven at 70°C.

Cattle Food Laboratory, U. S. Bureau of Chemistry: Comparison of this method with drying at 100°C. at the house vacuum (average 86.4) gave the following results:

Mixed alfalfa, molasses, corn feed at 100°C.—Average pressure 86.4 mm. Moisture, 3.60%.

At 27°C.—Average pressure 7.5 mm. Moisture 1.89%.

Bran at 100°C.—Average pressure 86.4 mm. Moisture 10.79%.

At 27°C.—Average pressure 7.5 mm. Moisture 10.43%.

Linseed meal at 100°C.—Average pressure 86.4 mm. Moisture 10.86%.

At 27°C.—Average pressure 7.5 mm. Moisture 10.71%.

TABLE 1.

Moisture determinations under varying conditions.

(J. H. Roop, Indiana.)

SUBSTANCE	DRIED 5 HOURS IN HYDROGEN AT 100° C.	DRIED 5 HOURS IN VACUO AT 75° C.	DRIED IN 6 INCH VACUUM DESICCA- TOR WITH SUL- PHURIC ACID AVERAGE
	<i>Per cent loss</i>	<i>Per cent loss</i>	<i>Per cent loss</i>
Cottonseed meal.....	8.75	8.10	7.55
Wheat bran.....	11.65	11.27	11.04
Corn germ meal.....	4.82	4.45	4.36
Linseed meal.....	9.97	9.57	9.15
Corn and oats.....	13.01	12.42	12.68
Distillers' grains.....	8.90	7.95	7.64

C. O. Swanson: This method has been in use at the Kansas Agricultural College for several years and is considered very satisfactory. For flour the drying is continued for from 3-5 days. Very moist samples are allowed to remain in the desiccator for 3 weeks. A shaking machine is employed to keep the acid stirred up.

COMMENTS BY THE REFEREE.

It has been found at the U. S. Food and Drug Inspection Laboratory at Savannah that the desiccator acid containing ether and water can be recovered by digesting for about 4 hours over a free flame in a Kjeldahl flask with a drop of mercury. A study of this method from the work of this and previous years seems to show that it can be relied upon when heating the sample is inadmissible; and that it is valuable also when subsequent fat extraction is contemplated and heating would tend to harden the sample too much. For work of this kind in which time is not a factor it may be very convenient to have an official method.

TABLE 2.

Comparison of dehydrating powers of reagents used in desiccators at room temperature of 20°-33°C. and at atmospheric pressure.

SUBSTANCE	TIME	SULPHURIC ACID	CALCIUM CARBID	SODIUM METAL	CALCIUM OXID	CALCIUM CHLORID	SODIUM HYDROXID
	Days	Per cent loss	Per cent loss	Per cent loss	Per cent loss	Per cent loss	Per cent loss
Corn meal ¹ :	1	10.18	10.06	9.54	9.22	7.76
	2	11.13	10.85	10.69	9.84	8.72
	3	11.49	11.22	11.09	10.05	9.21
	4	11.60	11.29	11.03	10.02	9.06
	5	11.74	11.43	11.20	10.10	9.19
	6	11.82	11.54	11.22	10.14	9.28
	7	11.87	11.53	11.30	10.14	9.23
Lean meat, finely ground ² :	1	43.70	36.20	45.23	28.77
	2	65.15	63.22	62.69	56.41
	3	66.04	65.50	63.53	64.08
	4	66.40	65.97	63.89	64.75
	5	66.56	66.10	63.98	64.90
	6	66.65	66.27	64.13	65.12
	7	66.78	66.38	64.23	65.25
	10	66.98	66.62	64.40	65.50
Hamburger steak ³ :	Hours						
	3	9.00	11.93	8.73
	Days						
	1	59.26	62.17	51.74
	2	69.50	69.40	69.15
	3	70.16	69.97	69.82
	4	70.47	70.24	70.14
Tomato paste ⁴ :	1	77.51	40.16	77.10	77.32
	2	77.67	76.40	77.35	77.54
	3	77.98	77.10	77.50	77.64
Glucose and apple jelly ⁵ :	1	18.91	19.59	17.84	16.43
	2	19.62	18.07	16.57
Pure apple jelly ⁶ :	1	19.29	20.44	18.90
	2	20.17	20.84	19.78
	3	21.11	20.19

¹ Vacuum oven 12.60%; in hydrogen 13.51%.

² Loss at 100°C., 67.10%.

³ Loss at 100°C., 71.09%.

⁴ Vacuum oven at 70°C., 78.4%.

⁵ Vacuum oven at 70°C., 23.83%.

⁶ Vacuum oven at 70°C., 27.45%.

It appears from this work and from the experience of previous years, particularly as reported at the meeting in 1913, that calcium carbide is a desiccator reagent nearly as efficient as sulphuric acid and one that is portable. This reagent is good until a large percentage of the lumps have loosened up into powder.

It will be best, however, to continue using sulphuric acid, lime or calcium chloride for precipitates of copper suboxide.

Leaving phosphorus pentoxide out of the question on account of its high cost and its rapid decrease in efficiency, it appears that the 3 best desiccator reagents are sulphuric acid, calcium carbide and calcium oxide, in the order named.

RECOMMENDATIONS.

It is recommended—

(1) That further work be done in comparing the drying of various food and feeding products over sulphuric acid, calcium carbide and calcium oxide, with any other reagents thought desirable, at atmospheric pressure and in a partial vacuum.

(2) That the method of drying without heat over sulphuric acid as outlined above be finally made an official optional method.

DISCUSSION.

P. F. Trowbridge: Mr. Grindley of the University of Illinois has used the method with good results—much better than we have been able to obtain at our Missouri Laboratory. Some materials present difficulties because they dry to a horn-like substance. The referee has not sufficiently emphasized the need for a good vacuum. Seven millimeters of mercury are not enough. A pump is necessary. With a pressure not exceeding 1 mm. it is possible to obtain from 2–10% more moisture from grain than by drying at 102°C.

REPORT ON FOODS AND FEEDING STUFFS.

By G. L. BIDWELL (Bureau of Chemistry, Washington, D. C.), *Referee.*

The coöperative work this year was all in charge of associate referees.

It is recommended—

(1) That the method for the detection of oats bleached with sulphur dioxide¹, be made official.

(2) That the method for determining the acidity of corn², be made official.

(3) That the method for determining the acidity of corn be studied to see if it is applicable to other materials.

(4) That the effect of preliminary drying on the ether extract content of feeds be studied.

(5) That the use of toluol or other suitable substance as a preservative for feed samples be studied.

(6) That the method of Bryan, Given and Straughn³ for the estimation of sugar in feeds be studied and made official.

FEED ADULTERATION.

By C. CUTLER (Agricultural Experiment Station, La Fayette, Ind.),
Associate Referee.

The work during the past year was confined to the following recommendation, approved at the 1914 meeting:

“(1) That samples be sent out for the quantitative determination of

¹ U. S. Bur. Plant Industry Circ. 40.

² Ibid., Bull. 199; U. S. Dept. Agr. Bull. 102.

³ U. S. Bur. Chem. Circ. 71.

adulterants on amounts varying from 5 to 25 grams to determine the smallest amount necessary to work with to get concordant results."

The following instructions were sent with the samples to 9 prospective collaborators:

INSTRUCTIONS FOR COLLABORATIVE WORK.

Quarter the samples, discard alternate quarters until the residue weighs approximately 25 grams. Procure a 10 gram sample in the same manner. A hand lens may be used in picking out small pieces of adulterants. The use of 20, 30 and 40 mesh sieves aids the work.

Sample No. 1 (wheat bran and screenings).—Determine the total amount of foreign material in 10 and 25 gram portions and identify the same.

Sample No. 2 (linseed meal containing cottonseed meal and hulls).—Determine the total amount of cottonseed meal and hulls in 10 and 25 gram portions.

Sample No. 3 (scratch feed).—Determine the total amount of grit in 10 and 25 gram portions; also the total amount of weed seeds present, identifying the principal ones.

Sample No. 4 (scratch feed containing coarse grit).

Sample No. 5 (wheat bran containing screenings).

RESULTS OF COLLABORATIVE WORK.

Per cent of adulterants found.

ANALYST	FOREIGN MATERIAL IN SAMPLE NO. 1		COTTONSEED MEAL AND HULLS IN SAMPLE NO. 2		WEED SEEDS IN SAMPLE NO. 3		GRIT IN SAMPLE NO. 3		GRIT IN SAMPLE NO. 4			SCREENINGS IN SAMPLE NO. 5	
	10 gram sample	25 gram sample	10 gram sample	25 gram sample	10 gram sample	25 gram sample	10 gram sample	25 gram sample	10 gram sample	25 gram sample	50 gram sample	10 gram sample	25 gram sample
A. W. Clark and O. C. McCreary, Agricultural Experiment Station, Geneva, N. Y.	4.2	3.2	5.0	4.1	32.1 ¹	32.9 ¹	22.6	24.0
B. H. Silberberg, Bureau of Chemistry, Washington, D.C.	7.5	7.6	19.1 24.4	21.5
G. W. Hoover and F. G. Smith, Transportation Building, Chicago, Ill.	5.1	4.2	0.58 ²	0.45 ²	33.5 ¹	33.6 ¹	23.6	21.1
P. H. Smith and Geo. H. Chapman, Agricultural Experiment Station, Amherst, Mass.	2.1	2.5	15.4 ³	14.0 ³	21.9	23.1
Carleton Cutler, Agricultural Experiment Station, La Fayette, Ind.	5.8	5.7	6.8	6.6	25.9	24.1	24.3 22.8	23.0 22.4	5.0	7.4	6.0 6.4	2.7	2.7

¹ Includes millet.

² Cottonseed hulls. Estimation of cottonseed meal present based on microscopical examination, 3-5%.

³ Includes timothy, but not shrivelled alfalfa and clover seeds.

The greatest variation in these results is in the determination of grit in Sample No. 3. However, the average of the 10 gram samples compares favorably with the average of the 25 gram samples.

Sample No. 4, scratch feed, contained very coarse limestone grit. Determinations were made on carefully quartered 25 and 50 gram samples. The 2 determinations on the 50 gram sample gave 6.0 and 6.4% grit, showing a variation of only 0.4%; while the 10 gram sample gave 5%, and the 25 gram sample 7.4%. Thus, the larger sample gives more concordant results on feeds containing coarse grit.

The variation in percentage of weed seeds found by different analysts in scratch feed No. 3, is due to the fact that all small seeds, including millet, were determined by 2 of the collaborators reporting. The determinations on the 10 and 25 gram samples agree very closely, the average variation on 3 determinations being only 0.3%.

The determinations of screenings in Sample No. 1, and cottonseed meal and hulls in Sample No. 2, are fairly concordant on 10 gram and 25 gram samples.

The results on Sample No. 5, wheat bran and screenings, made on 10 and 25 gram samples show identical percentages of foreign materials.

RECOMMENDATIONS.

Based upon the facts brought out in this year's results, together with experience obtained during the past several years, it is recommended—

(1) That the following method for the approximate estimation of foreign material, excluding grit, in feeding stuffs, be adopted as a provisional method:

PREPARATION OF SAMPLE.

Thoroughly mix the original unground sample, quarter, discard diagonally opposite quarters until the remaining 2 quarters weigh 10 grams.

DETERMINATION.

Separate the 10 gram sample by means of 20, 30, 40 and 50 mesh sieves. With the aid of a hand lens, or other magnifying instrument, pick out foreign materials; first from the finest portion, and then from the next finest, and so on in order until all portions have been examined. Combine the foreign materials separated and weigh.

(2) That the size of sample of scratch and poultry feeds necessary to get concordant results on quantitative grit determinations be further investigated.

(3) That the following recommendations of 1914 be studied during the coming year:

"3. That methods for the detection of peat dried at high temperatures in feed stuffs be investigated," and "4. That the maximum percentage of foreign materials permissible in mill by-products be investigated".

REPORT ON CRUDE FIBER.

By C. K. FRANCIS (Agricultural Experiment Station, Stillwater, Okla.),
Associate Referee.

The work on crude fiber has been confined to a study of the official method, with linen and asbestos filters, and the one filtration method, using paper as a filter.

Samples of fat-free kafir corn and cottonseed meal were sent to the cooperating chemists with the following instructions:

INSTRUCTIONS TO COLLABORATORS.

Pass the sample through a 20 mesh sieve. Make all determinations in triplicate and report the time required for the analysis under each method.

Weigh portions of 2 grams on hard filter papers and extract 4 times with ether, or use the residue from the crude fat determination.

DETERMINATION OF CRUDE FIBER.

Method 1.—Official¹.

Make the first filtration through linen and the second through asbestos. Report fineness of the linen in number of threads to the inch, counting both ways. Dry and incinerate as directed under Method 2. Make blank determinations to show any loss of weight of the asbestos.

Method 2.—One Filtration Through Paper.

Brush carefully the dry, fat-free residue into an 800 cc. lipless beaker. Add 200 cc. of boiling 1.25% sulphuric acid and boil for 30 minutes, using round-bottomed flasks filled with cold water as condensers placed on top of the beakers. The flasks should fit closely if the beakers are round and have well-formed rims. (Kjeldahl flasks are used in the associate referee's laboratory.) Direct a gentle blast of air into the beaker when the foaming becomes serious. At the end of 30 minutes add 200 cc. of boiling 3.52% sodium hydroxid and continue the boiling for another 30 minutes.

A Büchner funnel, outside diameter 10.5 cm., is prepared with a 9 cm. S. & S. No. 575 hard filter paper. The paper should be moistened with water and pressed well into place, so that there will be no holes uncovered and no open channels left about the edges. It is advisable to place a glass filter plate over the paper, to protect it and to prevent excessive packing of the fiber.

After the 30 minutes alkali boiling, filter rapidly with the aid of suction, wash with boiling water, then with a 1.25% solution of concentrated hydrochloric acid (14 cc. made up to 500 cc. with water), until the washings are acid. Much time may be saved if the liquid is set aside for about 2 minutes to allow the fine material to settle so that the clear liquid may be decanted. Wash with hot water until free from chlorids, and finally wash the filter several times with 80% alcohol. Remove the paper from the funnel. Transfer the residue to a Gooch crucible with the aid of a spatula and a small amount of 80% alcohol. Rub the paper lightly with the finger to detach the last traces of the fiber residue.

Place the crucible in an oven heated to 105°–110°C. for from 2–6 hours, cool and weigh. The material may become dry in 2 hours but when large quantities of fiber

¹ U. S. Bur. Chem. Bull. 107, Rev., p. 56.

are present a longer heating will be necessary. Heat for 30 minutes before weighing the second time.

Partially cover the crucible and incinerate at a low red heat for 15 minutes or until a white or light gray ash is obtained. Cool and weigh. The loss of weight is crude fiber.

RESULTS OF COLLABORATIVE WORK.

TABLE 1.
Determination of crude fiber and time required.

ANALYST	KAFIR CORN				COTTONSEED MEAL				NUMBER OF THREADS TO THE INCH OF FILTER- ING CLOTH
	Official Method		Proposed Method		Official Method		Proposed Method		
	per cent	hours	per cent	hours	per cent	hours	per cent	hours	
O. C. Smith, Agri- cultural Experi- ment Station, Stillwater, Okla.	1.73	3	1.77	2	10.38	3	11.23	2	60 × 57
	1.70	1.69	10.20	10.59	
	1.70	1.64	10.44	10.67	
	Average.....	1.71	1.70	10.34	10.63	
D. G. Morgan, Ag- ricultural Ex- periment Sta- tion, Stillwater, Okla.	1.81	3½	1.62	3	10.32	3½	11.65	3	60 × 57
	1.80	1.78	10.42	10.80	
	1.83	1.64	10.24	10.61	
	Average.....	1.81	1.68	10.33	10.70	
G. W. Roark, Ag- ricultural Experi- ment Station, College Station, Tex.	1.80	1.63	10.11	10.81	6	42 × 42
	1.77	1.10	10.37	
	1.73	1.56	10.25	10.44	
	Average ...	1.76	1.43 ¹	10.24	10.62	
A. S. Wells, Dairy and Food Com- mission, Port- land, Ore.	1.75	8	1.90	7	9.57	10	11.01	16	92 × 92
	1.80	1.89	9.60	
	1.75	1.94	9.80	10.82	
	Average.....	1.76	1.91	9.65 ¹	10.91	
P. F. Trowbridge, Agricultural Ex- periment Sta- tion, Columbia, Mo.	1.52	10	1.64	7	10.05	10	11.16	7	68 × 71
	1.53	1.85	10.14	11.01	
	1.75	1.81	10.18	11.25	
	Average.....	1.60	1.76	10.12	11.14	
G. P. Walton, Bu- reau of Chemis- try, Washington, D. C.	1.70	4½	1.54	9	10.87	4½	10.17	9	42 × 46
	1.70	1.50	10.41	9.41	
	1.79	1.53	10.33	10.07	
	Average.....	1.73	1.52	10.53	9.88 ¹	

¹ Omitted from corrected general average.

TABLE 1.—Continued.

ANALYST	KAFIR CORN				COTTONSEED MEAL				NUMBER OF THREADS TO THE INCH OF FILTER- ING CLOTH
	Official Method		Proposed Method		Official Method		Proposed Method		
	per cent	hours	per cent	hours	per cent	hours	per cent	hours	
Jas. W. Kellogg, Department of Agriculture, Har- risburg, Pa.	1.62	3.06	9.59	14.07	75 × 75
	1.57			9.59			
	1.60			9.63	12.65	
Average	1.59	3.06 ¹	9.60 ¹	13.36 ¹	
W. D. Richardson, Swift and Com- pany, Chicago, Ill.	1.47	3	1.55	2	9.13	4	10.03	4-5	87 × 87
	1.60	1.58	9.48	10.37	
	1.56	1.40	9.78	10.55	
Average.....	1.54	1.51	9.46 ¹	10.31	
F. G. Smith, Trans- portation Build- ing, Chicago, Ill.	1.64	1½	1.91	1½	10.64	1½	11.19	4-8	58 × 54
	1.70	1.93	10.75	11.30	
	1.74	1.96	11.01	11.99	
Average.....	1.69	1.93	10.80	11.24 ¹	
D. T. Evans, Fort Worth, Tex.	1.69	1	1½	10.10	1½	2	80 × 100
	1.67	9.95	
		9.42	
Average ..	1.68	9.82 ¹	
J. H. Roop, Agri- cultural Experi- ment Station, La Fayette, Ind.	1.65	3	1.82	5	9.87	3	10.32	5	75 × 76
	1.63	1.82	9.81	9.92	
	1.60	1.75	9.96	10.30	
Average ..	1.63	1.80	9.88 ¹	10.18 ¹	
General average ...	1.68	1.83	10.07	10.90	
Corrected general average	1.68	1.72	10.39	10.72	

¹ Omitted from corrected general average.

COMMENTS OF ANALYSTS.

G. W. Roark: Method 2 was shorter and easier than the regular (Official) method but there was difficulty in filtering through paper; also, it is believed, there is some loss around the edges of the paper.

A. S. Wells: Fine material passes through the linen during the first filtering. This is the main objection to the official method. The proposed method worked fairly well with the sample of kafir corn, but the sample of cottonseed meal packed so hard that it could not be removed without taking off some of the filter paper.

P. F. Trowbridge: After trying a great number of samples of muslin and linen we secured a very fine grade of muslin, 98 x 102 threads per inch. The best grade

of linen we were able to secure ran 6S x 71 threads to the inch. Furthermore, the particles of fiber adhere much more to the linen threads than to the muslin threads.

Considerable difficulty was experienced in filtering through asbestos. The proposed method was more rapid than the official method, but the filtering was slow and there was difficulty in transferring the fiber from the paper to the Gooch crucible. Whenever any modification of the Sweeney method has been tried with single filtration higher results have always been obtained.

G. P. Walton: As pointed out in last year's report by this laboratory on crude fiber, this one filtration method so modifies the regular method as to make it extremely improbable that results from the 2 methods can be reconciled. Besides the question of the redissolving of substances precipitated, on making the acid liquid alkaline, there is the real objection to digesting with twice the volume of 1.25% sodium hydroxid solution specified in the official method.

The transfer of the fiber residuum is a tedious operation, and difficulty was experienced in obtaining all of it without including fiber from the filter paper.

Directions to wash finally with strong alcohol should be incorporated in the official method.

J. W. Kellogg: Proposed method entirely unsatisfactory when compared with the official method.

W. D. Richardson: With such samples as cottonseed meal it was found quite difficult to pass 400 cc. through the same filter, as the pores soon became clogged and made the filtration and washing unsatisfactory. Kafr corn gives good results by this method and is easy to handle.

F. G. Smith: Method 2 required from 4-8 hours to make the digestion and filtration, in one case it being practically impossible to complete the filtering.

D. T. Evans: The filtration through paper was so difficult that asbestos was substituted.

J. H. Roop: Great difficulty was encountered in filtering the alkaline solution through the paper and in transferring the fiber from the filter paper without including some fiber of the filter paper.

EXPERIMENTS BY THE ASSOCIATE REFEREE.

It may be of interest to mention some additional work on crude fiber. The data presented in Table 2 show the results obtained with 3 kinds of filters when but one filtration was made.

TABLE 2.

Determination of crude fiber in cottonseed meal by the one filtration method using different filters.

FILTERS	SEPARATE DETERMINATIONS			AVERAGE
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
Linen.....	8.57	8.58	8.54	8.56
Hard paper.....	9.54	9.53	9.37	9.48
Asbestos.....	10.87	10.81	10.84

Portions of cottonseed meal from the same sample were examined in a similar manner, observing the directions of the official method.

TABLE 3.

Determination of crude fiber in cottonseed meal by the official method using different filters.

FILTER USED		SEPARATE DETERMINATION			AVERAGE	REMARKS
After acid digestion	After alkali digestion	per cent	per cent	per cent		
Linen	Linen	8.44	8.16	7.96	8.19	No suction; residue washed into a Gooch crucible.
Linen	Asbestos	8.56	8.82	8.69	Asbestos filter into a Gooch crucible, i.e., small amount asbestos used.
Linen	Asbestos	8.96	8.79	9.02	8.92	Asbestos filter in a Hirsch funnel, i.e., large amount asbestos.
Paper	Paper	9.63	9.53	9.51	9.56	S. & S. No. 575 paper in a Büchner funnel.

It will be observed from the data in Tables 2 and 3 that the determinations with a given filter, when considered alone, appear satisfactory, but when comparisons of the groups are made, the results do not agree.

Determinations in addition to those here reported have shown that the loss with linen depends upon the number of times the mixture is filtered through it, thus demonstrating that there is a loss of crude fiber.

The crude fiber determinations in which filter papers were used appear to be the most uniform for both methods. The paper filters certainly do not permit any solid matter to pass into the filtrate, and they offer a uniform filter which may be easily obtained and from which the residue can be readily transferred.

Some determinations were made in which asbestos was used for both filtrations according to the official method, but the large quantity of material added to the second filter, in this way, caused the filtration to proceed very slowly.

Portions of filter paper, S. & S. No. 597, were ground so as to pass a 40 mesh sieve. This material was intended to be used for testing the efficiency of different filters, but the residue obtained, after passing through the entire official method for crude fiber, showed so great a loss, that it became necessary to ascertain where the loss occurred. The final procedure involved digestion of separate samples in 1.25% sulphuric acid and in 1.25% sodium hydroxid, and in both reagents as outlined in the method for one filtration.

The results of this test are presented in Table 4.

TABLE 4.

Action of crude fiber reagents on pure filter paper.

PROCEDURE	FIBER RECOVERED BY USE OF PAPER FILTERS		FIBER RECOVERED BY USE OF ASBESTOS FILTERS	
	Original basis	Ash plus water-free basis	Original basis	Ash plus water-free basis
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
Acid digestion only:	93.90	99.42	101.15	107.14
	95.03	100.62	102.53	108.56
	94.56	100.11	97.38	103.10
Average.....	94.49	100.05	100.35	106.27
Alkali digestion only:	91.33	96.70	98.06	103.86
	92.27	97.72	98.52	104.38
	91.78	97.17	98.74	104.59
Average.....	91.79	97.20	98.44	104.28
Acid and alkali, 1 filtration:	64.91	68.75	71.50	75.71
	64.67	68.47	74.71	79.13
			73.00	77.30
Average.....	64.79	68.61	73.07	77.38
Acid and alkali, 2 filtrations:	65.02	68.85		
	65.58	69.18		
	64.26	68.05		
Average.....	64.95	68.69

The action of the acid is apparently negligible, and the alkali does not seem to dissolve a very large quantity. However, the material digested in both acid and alkali showed a very large loss, approximately 31% by both methods. The asbestos filter gave higher results in all tests.

CONCLUSIONS.

The results reported this year indicate that hard filter paper, used in the manner suggested, is not satisfactory. All analysts obtained good results for kafir corn by the official method and the returns on the same sample tested by the one filtration method were encouraging. However, the showing for the sample of cottonseed meal was not so good and indicates a need for further study of crude fiber methods. A number of the chemists did not report the loss of weight due to the asbestos, but those reporting indicated that an error of from 0.05-0.20% plus, may be introduced. Results showing much higher losses were reported by several chemists.

THE DETERMINATION OF CRUDE FIBER¹.

By C. E. MANGELS AND P. F. TROWBRIDGE (Agricultural Experiment Station, Columbia, Mo.).

In laboratories where a great many crude fiber determinations are made, it is important that the method used be as free as possible from tedious manipulations. As in all empirical determinations, however, it is important that the methods followed should be the same as in other laboratories until it has been shown that slight modifications do not affect the accuracy of the results.

In the collaborative work, the referee's samples were tested by methods which have been used in the Missouri Agricultural Experiment Station. The method in use this year and which seems to give the most satisfactory results is as follows:

Transfer the residue from the ether extraction to a lipless beaker, add 200 cc. of cold 1.25% sulphuric acid and make a mark on the beaker to indicate the height of the liquid. Heat rapidly to boiling the contents of the beaker and continue the boiling for exactly 30 minutes. Filter rapidly the contents of the beaker through a muslin filter and wash the fiber free from acid with hot water. Place the original beaker under a funnel and transfer the fiber from the muslin to the beaker, using a spatula and a stream of 1.25% sodium hydroxid from a wash bottle. Then fill the beaker to the mark with 1.25% sodium hydroxid, heat rapidly to boiling the contents of the beaker and boil for exactly 30 minutes. Filter the hot alkaline solution through the same muslin filter, wash the beaker twice with hot water and transfer the washings to the filter. Then wash the fiber on the filter once with 1.25% hydrochloric acid and then thoroughly with hot water to remove all acid and chlorids. Transfer the bulk of the fiber from the muslin to a prepared Gooch crucible, spreading the fiber on the sides of the crucible so as to avoid clogging the filtration. Place the original beaker under the funnel and wash the last traces of the fiber from the muslin into the beaker by means of a stream of hot water. Connect the Gooch crucible with suction and transfer the fiber in the beaker to that in the crucible, by the aid of hot water and a policeman. Wash finally the fiber in the Gooch crucible with alcohol and dry to constant weight at 102°C., then ignite and weigh again to determine the fiber by difference.

The writers have tested a great many samples of linen and muslin to secure a satisfactory medium for filtering and have finally selected and secured a quantity, sufficient for several years, of close-woven muslin having 98 x 102 threads to the inch. The best linen which could be secured had 68 x 71 threads to the inch.

The method just described differs from the regular official method in that both the acid and alkali solutions are added to the sample cold, thereby making the contact with the sample 30 minutes in addition to the time required to heat the 200 cc. of solution to the boiling point. This, if anything, should give lower results than by the regular official method.

¹ Presented by P. F. Trowbridge.

The results are shown under Method C in the following table and are slightly higher than results obtained by the regular official method (A). The writers have at different times tried the Sweeney method (single filtration) with various modifications, hoping to get a rapid method which was at the same time comparable with the regular official method.

In this method, the results of which are reported under (D) in the following table, 200 cc. of boiling 1.25% sulphuric acid were added to the fiber and the boiling continued for exactly 30 minutes. Two hundred cc. of boiling 3.52% sodium hydroxid were added immediately to this solution and the boiling continued for another 30 minutes. Rapid filtration was then made through the muslin filter. The beaker was washed twice with hot water, the washings being transferred to the muslin filter. The fiber was then washed once on the filter with cold 1.25% hydrochloric acid, and the washing completed with hot water. The transfer of the fiber and the completion of the determination were conducted exactly as described in the previous method. In length of time required this method is found to be only slightly shorter than the double filtration through muslin. The results obtained are seen to be higher in each case than those obtained by any of the other methods.

The results obtained by Mr. Francis' single filtration method through hardened filter are higher than either of the double filtration methods. For example, one of the single filtration methods gives 1.640%, on Sample No. 1, which is lower than any of the results obtained on this sample with our double filtration method and also lower than one of the results by the regular official method filtering once through linen and once through asbestos. On Sample No. 2 one of the determinations, 11.010, is lower than either of the results given by our double filtration method. These results would seem to indicate that the longer heating, caused by starting with the cold acid and alkali solutions, does not reduce the amount of crude fiber. The other modification in the method, by washing once with 1.25% hydrochloric acid, greatly facilitates the removal of the alkali and the chlorids and since it does not appear to affect the final amount of crude fiber its use is to be recommended.

Per cent of crude fiber obtained.

PROCEDURE	SAMPLE 1	SAMPLE 2	REMARKS
(A) Official method, first filtration through linen and second filtration through asbestos:	1.520	10.045	8-10 hours, filtration difficult.
	1.525	10.140	
	1.750	10.180	
Average.....	1.598	10.122	

Per cent of crude fiber obtained.—Continued.

PROCEDURE	SAMPLE 1	SAMPLE 2	REMARKS
(B) Smith-Francis method of filtration through hardened filter on Büchner funnel:	1.640 1.845 1.805	11.160 11.010 11.245	6-7 hours, filtration and transfer difficult.
Average.....	1.763	11.138	
(C) Missouri Station method, 2 filtrations through muslin, starting with cold 1.25% acid and alkali:	1.715 1.780 1.740	Lost 11.105 11.060	About 4½ hours, muslin used 98 x 102 threads per inch.
Average.....	1.745	11.083	
(D) Sweeney method, one filtration through muslin:	1.845 1.845 1.840	11.475 11.215 11.560	3½-4 hours.
Average.....	1.843	11.417	

REPORT ON SUGAR.

By C. A. BROWNE (Sugar Trade Laboratory, New York, N. Y.),
Referee.

Since the report of Mr. Cross in 1913, the work of the Association upon sugar has been interrupted, no referee having been appointed for the year 1913-1914.

In resuming the work which was so efficiently carried out by his predecessor, it occurred to the present referee that this Association during the next few years might in a measure retrieve a part of the damage which has resulted to the progress of sugar analysis as a consequence of the European war. Two international congresses had been scheduled this year to report upon sugar, that of the International Congress of Applied Chemistry, which was to meet in Petrograd in August, and that of the International Commission upon Uniform Methods of Sugar Analysis, which was to meet in Amsterdam in September. Both of these meetings have been indefinitely postponed and many years must elapse before the important questions which they were to consider are reported.

At the last meeting of the International Commission upon Uniform Methods of Sugar Analysis in New York in 1912, a committee was appointed to study the inversion constant of the Clerget method. As the present referee, in connection with his duties upon this committee, is conducting a series of researches along this line, he has selected the pres-

ent occasion as an opportune one in which to report upon certain modifications of the Clerget method.

A STUDY OF CERTAIN MODIFICATIONS OF THE CLERGET METHOD

The volume to which the solution is made up for invert polarization which was formerly used by this Association and which is still employed by many sugar chemists, is the one originally proposed by Clerget. In this procedure 50 cc. of the clarified solution used for direct polarization were transferred to a flask graduated at 50 cc. and 55 cc. Concentrated hydrochloric acid was then added to the 55 cc. mark, the solution was mixed, a thermometer was inserted and the inversion was carried out according to the regulations prescribed for temperature and time of heating. After cooling, the solution was read upon the polariscope in a 200 mm. tube and the observed reading was increased by one-tenth to correct for the dilution from 50-55 cc. This corrected reading and the temperature of the solution were then substituted in the Clerget formula for calculating the percentage of sucrose.

One great advantage of diluting the solution from 50-55 cc. is that any errors in reading the inverted solution are increased by only one-tenth, whereas in the modifications which dilute from 50-100 cc. these errors are multiplied by 2. But notwithstanding this advantage, the experience of chemists has shown a certain unreliableness to exist in the old Clerget procedure so that in later years this method of making up to volume has been largely discontinued.

The referee believes that one great cause of the difficulty, which has been experienced with the old Clerget procedure, has been the neglect of a very serious error, viz: the diminution in volume which takes place in the 55 cc. of solution during inversion. This diminution of volume is due to three causes:

(1) The contraction in volume which all sucrose solutions undergo during inversion and which for 13 grams of sucrose in 55 cc. is about one-fourth of a cubic centimeter.

(2) The elevation in temperature produced by the addition of the hydrochloric acid. This elevation, for 5 cc. of concentrated hydrochloric acid to 50 cc. of sugar solution, is about 3°C., the cooling of the solution from 23°C. at the beginning to 20°C. at the end of the inversion produces a further slight contraction.

(3) The evaporation of water from the neck of the flask during inversion, the amount of such evaporation depending upon the diameter of the neck of the flask, and the time and temperature of inversion.

The combined influence of these 3 factors causes the volume of the 55 cc. of solution at the end of inversion to be about one-third of a cubic centimeter too small for the half normal weight of 13 grams.

It is essential to the accuracy of any method of double polarization that the volume of the solution after inversion be fixed with the utmost accuracy. The sources of error just mentioned are easily overcome if the volume of the inverted solution be brought to 55 cc. at the end of inversion and not at the beginning. If this precaution be followed more accurate results can be secured by the process of making the inverted solution up to 55 cc. than by any of the modifications which dilute to 100 cc. The control of temperature by this procedure is easily carried out by means of a thermometer placed in a control flask containing 55 cc. of blank solution. There is no danger of irregular mixing of the acid with the sugar solution, as the diffusion of the hydrochloric acid through the body of the liquid takes place quickly without shaking.

In employing this method the referee makes all his inversions at room temperature; 5 cc. of concentrated hydrochloric acid were added to the 50 cc. of sugar solution in a 50-55 cc. flask and after standing overnight the volume was completed to exactly 55 cc. at 20°C., after gently tapping the walls of the flask to detach any air bubbles which might have accumulated. The solution is then mixed and read in a polariscope which is also at a temperature of 20°C. The invert reading is then corrected by adding the necessary one-tenth.

The formula for calculating the sucrose by this method of inversion, when a normal weight of 26 grams of pure sucrose is taken, is

$$S = \frac{100 (A - B)}{144.9 - \frac{t}{2}}$$

The above formula, corrected for the differences in specific rotation of invert sugar due to the varying concentration, when less than 26 grams of sucrose are dissolved to 100 cc., is

$$S = \frac{100 (A - B)}{144.9 - \frac{t}{2} - 0.01 \left[144.9 - \frac{t}{2} - (A - B) \right]} \quad \text{in which}$$

S = per cent of sucrose;

A = direct polarization;

B = corrected invert polarization;

t = temperature of the invert solution.

If the invert polarization be made at exactly 20°C. the above formula becomes

$$S = \frac{100 (A - B)}{134.9 - 0.01 [134.9 - (A - B)]}$$

The determinations of Table 1 give an idea of the comparative accuracy of this method when applied to solutions of pure sucrose.

TABLE 1.

Application of Clerget modified procedure to solutions of pure sucrose.

NUMBER	DIRECT POLARIZATION (A)	CORRECTED INVERT POLARIZATION (B)	TEMPERATURE (t)	SUCROSE (S)	
				Found	Taken
			°C.	per cent	per cent
1	99.90	-35.00	19.8	99.92	99.90
2	76.95	-26.79	19.6	76.97	76.92
3	57.70	-19.69	20.2	57.66	57.69
4	38.45	-13.04	20.2	38.44	38.46
5	19.20	- 6.49	20.0	19.15	19.23
6	7.70	- 2.53	20.0	7.65	7.69

TABLE 2.

Application of Clerget modified procedure to solutions of sucrose and dextrose.

NUMBER	DEXTROSE	DIRECT POLARIZATION (A)	CORRECTED INVERT POLARIZATION (B)	TEMPERATURE (t)	SUCROSE (S)	
					Found	Taken
	grams per 100 cc.			°C.	per cent	per cent
1	4.8	91.70	-11.88	20	76.96	76.92
2	4.8	72.45	- 5.06	20	57.70	57.69
3	4.8	53.15	+ 1.60	20	38.45	38.46
4	4.8	33.90	+ 8.20	20	19.23	19.23

The importance of using a formula, which corrects for the differences in specific rotation of invert sugar due to varying concentration, may be illustrated by taking the case of determination No. 3 in Table 2. This determination, using the uncorrected formula, gives 38.21 per cent sucrose, which is a quarter of a per cent too low.

In this connection the referee would very urgently recommend that the Association adopt a modified Herzfeld formula, or the Herzfeld table of factors, in place of the uncorrected formula¹. The correction of the Herzfeld factor 142.66, for concentration has been done in several ways.

According to one method the corrected Herzfeld factor equals $141.78 + 0.0676 C$, C being the grams of sucrose in 100 cc. The objection to this method is that a preliminary calculation with the uncorrected factor 142.66 is necessary in order to obtain an approximate value of C for determining the true factor. The work of calculation is thus doubled.

According to another method the corrected Herzfeld factor equals $141.84 + 0.05 n$, in which n is the scale reading of the inverted solution. The objection to this second method is that it applies to inverted solutions of pure sucrose and is of no value when other optically active substances are present.

¹ U. S. Bur. Chem. Bull. 107, Rev., p. 41.

The two objections named are overcome if the correction for concentration be based upon the difference between the direct and invert polarization. The Herzfeld formula as thus modified becomes

$$S = \frac{100 (A - B)}{142.66 - \frac{t}{2} - 0.0065 \left[142.66 - \frac{t}{2} - (A - B) \right]}$$

which for 20°C. is simplified to

$$S = \frac{100 (A - B)}{132.66 - 0.0065 [132.66 - (A - B)]}$$

The above modified Herzfeld formula is the one which the referee would recommend to the Association for adoption.

A great deal of work has been done by different investigators during the past few years upon methods of double polarization in the effort to make them better adapted to the analysis of sugar mixtures. In the ordinary methods of procedure any levulose, or other sugar, whose optical activity is changed by the addition of the inverting acid, has a different rotation before and after inversion so that a considerable error may be introduced in the calculation of sucrose. The possible magnitude of this error may be seen from the determinations given in Table 3 which were made upon known mixtures of sucrose and invert sugar. The modified procedure of the original Clerget method was used.

TABLE 3.

Application of Clerget modified procedure to solutions of sucrose in invert sugar.

NUMBER	INVERT SUGAR	DIRECT POLARIZATION (A)	CORRECTED INVERT POLARIZATION (B)	TEMPERATURE (t)	SUCROSE (S)	
					Found	Taken
	<i>grams per 100 cc.</i>			<i>°C.</i>	<i>per cent</i>	<i>per cent</i>
1	1.10	94.60	-35.20	19.8	96.18	96.15
2	2.74	73.35	-30.42	20.0	77.10	76.92
3	5.48	50.70	-27.28	20.4	58.14	57.69
4	8.21	28.15	-24.04	20.4	38.99	38.46
5	9.86	6.90	-19.58	20.4	19.82	19.23

It is seen that as the amount of invert sugar increases there is a corresponding increase in the sucrose error. In case of honeys and other products containing a high percentage of levulose the error in the sucrose determination due to this cause may exceed 1%.

In mixtures of sucrose with invert sugar alone, it is possible to correct this error in the determination of sucrose by means of an empirical factor. Such a method of correction is not adapted, however, to the analysis of complex mixtures of optically active substances. The efforts of chemists

have been directed, therefore, towards devising a method by which the direct and invert polarizations could be made under similar conditions.

Of the various methods which the referee has tested in this connection, the one which has given the most satisfactory results was that by means of invertase.

The use of invertase from yeast has long been recognized by chemists as the method of inversion which is most perfect in theory, but the difficulty of preparing satisfactory solutions of invertase has been a great obstacle against the introduction of the method. In 1909 Mr. Hudson devised several improvements for preparing invertase from yeast and in April 1910 published a paper upon the use of his invertase solution in the quantitative estimation of sucrose. In Hudson's method 50 cc. of the clarified solution used for the direct polarization are transferred to a 100 cc. flask, faintly acidified with acetic acid and then treated with 5 cc. of stock invertase solution. The volume is completed to 100 cc. and after inversion the solution read in the usual way, the invert reading being corrected for the optical activity of the invertase solution. The formula proposed by Mr. Hudson for this modification of the Clerget process is

$$S = \frac{100 (A - B)}{141.7 - \frac{t}{2}}$$

The assertion has been made that the influence of concentration upon the Clerget factor would practically disappear, provided the inverting agent was without influence upon the rotation of the invert sugar. This, however, is not the case and cannot be the case, since the specific rotation of invert sugar necessarily increases with the concentration. The referee has subjected the invertase method to a most careful examination and finds that the influence of concentration is just as pronounced as with the methods which employ hydrochloric acid.

The invertase solution employed by the referee was prepared according to the original procedure of Mr. Hudson and had the following properties:

Sp. gr. at $\frac{20^{\circ}\text{C.}}{20^{\circ}} = 1.00363$;

Refractive index = 1.33468;

Solids in 10 cc. solution = 0.1027 grams;

Ash in 10 cc. solution = 0.0040 grams;

Rotation of solution in 200 mm. tube = $+0.65^{\circ}$ Ventzke.

In using this invertase solution for the Clerget determination the referee followed the method proposed by Mr. Hudson except that the 50 cc. of sugar solution were treated with 10 cc. of invertase reagent in-

stead of 5 cc. and the solution was warmed to 50°C., to hasten the inversion. The solution after standing overnight was made up to 100 cc. at 20°C. and polarized at this temperature. The invert polarization was then increased by 0.0065 to correct for the dextro-rotatory effect of the 10 cc. of invertase solution and then multiplied by 2 to correct for the dilution to 100 cc. The formula for calculating the percentage of sucrose by this method, as calculated from the average of many determinations upon sugar solutions of different concentration, was found to be

$$S = \frac{100 (A - B)}{142 - \frac{t}{2} - 0.0065 \left[142 - \frac{t}{2} - (A - B) \right]}$$

It will be noted that the concentration factor 0.0065 is the same as that found for the Herzfeld method, which follows the same method of diluting the 13 grams of inverted product to 100 cc.

The application of the above formula to the determination of sucrose in various mixtures with invert sugar is given in Table 4.

TABLE 4.
Application of invertase method to solutions of sucrose and invert sugar.

NUMBER	INVERT SUGAR	DIRECT POLARIZATION (A)	CORRECTED INVERT POLARIZATION (B)	TEMPERATURE (t)	SUCROSE (S)	
					Found	Taken
	<i>grams per 100 cc.</i>			<i>°C.</i>	<i>per cent</i>	<i>per cent</i>
1	0.00	99.80	-31.83	20	99.72	99.80
2	0.18	95.85	-30.83	20	95.99	95.96
3	0.43	89.75	-29.33	20	90.27	90.20
4	0.67	85.60	-28.23	20	86.31	86.37
5	0.92	79.50	-26.73	20	80.58	80.61
6	2.44	47.00	-18.63	20	49.89	49.90
7	2.44	27.90	-12.43	20	30.69	30.71

The results show that the invertase method gives far more accurate results than the method of acid inversion in the analysis of complex mixtures which contain sucrose and levulose.

The referee does not believe that the invertase method will immediately displace the method of acid inversion for purposes of commercial analysis. The preparation of the invertase solution is somewhat troublesome and the gradual deterioration of the reagent involves the risk of incomplete inversion. The process is one which requires care and watchfulness. A blank analysis upon a weighed amount of pure sucrose should be carried out with every set of determinations. The invertase method is invaluable, however, as a control upon the accuracy of the method of acid inversion and the referee would recommend that it be adopted as a provisional method by the Association.

RECOMMENDATIONS.

In planning the work for next year the referee believes that attention should be devoted to those matters which are of most pressing importance to sugar analysis. He would, therefore, recommend—

(1) That the study of the modifications of the Clerget method be continued with special reference to the accurate determination of sucrose in complex mixtures of carbohydrates.

(2) That efforts be made towards the adoption of an accurate method for determining small quantities of invert sugar in the presence of large amounts of sucrose. In this connection the colorimetric methods for estimating invert sugar should be examined.

(3) That next year's referee in collaboration with the U. S. Bureau of Chemistry establish a table of reduction factors for the more common reducing sugars.

There are a number of improvements and changes, which the progress of sugar analysis, during the past 10 years, requires to be made in the official and provisional methods of the Association. The referee has gone over these methods with several of the members of the Committee on Editing Methods of Analysis and it was considered advisable to present the following recommendations:

(4) That the official text of the methods of the International Commission be substituted for the present text¹ and that these methods of the International Commission be brought up to date.

(5) That the following method for preparing a salt-free alumina cream be adopted in place of the present method²:

Precipitate concentrated alum solution with a slight excess of ammonium hydroxid and wash the precipitate by decantation with water until the solution is free from sulphates. Pour off the excess of water and store the residual cream in a stoppered bottle.

(6) That at the end of section (c)³ the following be inserted:

For concentrations of sucrose of less than 13 grams to 100 cc. of invert solution the following general formula should be used.

$$S = \frac{100 (P - I)}{142.66 - \frac{t}{2} - 0.0065 \left[142.66 - \frac{t}{2} - (P - I) \right]}$$

The above formula is applicable to the determination of sucrose in the presence of dextrose, commercial glucose, and all other substances whose optical activity is not affected by the inverting acid. With materials which contain much levulose, such as honey, fruit products, etc., the method gives too high results.

¹ U. S. Bur. Chem. Bull. 107, Rev., pp. 39-40.

² Ibid., p. 40.

³ Ibid., p. 41.

(7) That at the end of section (d)¹ the following formula be substituted for the present formula for calculating the per cent of sucrose in the presence of raffinose:

$$S = \frac{0.5124 P - I}{0.839}$$

The above formula supposes that the polarizations be made at exactly 20°C. If the temperature (T) be other than 20°C., the following formula should be used:

$$S = \frac{P (0.4724 + 0.002 T) - I}{0.899 - 0.003 T}$$

Having calculated S,

$$R = \frac{P - S}{1.852}$$

(8) That after section (d)¹ the following be inserted:

DETERMINATION OF SUCROSE BY MEANS OF INVERTASE.—PROVISIONAL.

Preparation of Invertase Solution, Hudson Method.

Break up 5 pounds of pressed yeast, which may be either baker's or brewer's yeast, add 30 cc. of chloroform to it in a closed flask and allow to stand at room temperature (20°C.) overnight. By morning the solid mass will have become fluid and it should then be filtered through filter paper allowing several hours for draining. To the filtrate add neutral lead acetate until no further precipitate forms and again filter. Precipitate the excess of lead from the filtrate with potassium oxalate and filter. To this filtrate add 25 cc. of toluene and dialyze the mixture in a pig's bladder for 2 or 3 days against clear running tap water. The dialyzed solution should be colorless, perfectly clear after filtration, and neutral to litmus; it should be preserved in an ice-box with the addition of a little toluene to prevent the growth of micro-organisms. The optical activity of the invertase solution is noted and a correction for this, according to the amount of solution used, must be applied to the invert reading.

Determination.

Dissolve the normal weight (26 grams) of substance in water, clarify, make up to volume, and take the direct polarization (P) as under section (d). Remove the excess of lead from the filtrate, if lead has been used as a clarifying agent, with anhydrous sodium carbonate or potassium oxalate, and filter. To 50 cc. of the filtrate in a 100 cc. flask add acetic acid by drops until the reaction is acid to litmus, add 10 cc. of the stock invertase solution and let stand in a warm place (about 40°C.) overnight. Cool and make up to 100 cc. at 20°C. Polarize at 20°C. in a 200 mm. tube. Allow the solution to remain in the tube for an hour and repeat the polarization. If there is no change from the previous reading the inversion is complete, when the reading and temperature of the solution are carefully noted. The reading is corrected for the optical activity of the invertase solution and then multiplied by 2. The percentage of sucrose is then calculated by the following formula:

¹ U. S. Bur. Chem. Bull. 107, Rev., p. 41.

$$S = \frac{100 (P - I)}{142 - \frac{t}{2} - 0.0065 \left[142 - \frac{t}{2} - (P - I) \right]}$$

S = per cent of sucrose;

P = direct reading;

I = invert reading;

t = temperature at which invert reading is made.

(9) That before section (c)¹ the following be inserted:

Reducing sugars other than dextrose may be determined, using Allihn's modification of Fehling's solution, by means of the above table and method by use of the following factors:

Arabinose	= Glucose \times 0.969;
Xylose	= Glucose \times 1.017;
Levulose	= Glucose \times 1.093;
Invert sugar	= Glucose \times 1.044;
Galactose	= Glucose \times 1.114.

(10) That before section (3. Ash²) the table for specific gravity and total solids of sucrose solutions at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ used by the U. S. Bureau of Standards and Reichsanstalt of Germany be inserted.

(11) That in the Munson and Walker table³ for calculating lactose the column for lactose with one-half molecule of water be omitted.

No report was made by the referee on the general subject of the separation of nitrogenous substances.

REPORT ON THE SEPARATION OF NITROGENOUS SUBSTANCES (MEAT PRODUCTS).

By P. F. TROWBRIDGE (Agricultural Experiment Station, Columbia, Mo.), *Associate Referee*.

No collaborative work has been undertaken. In connection with other work, a number of carefully prepared samples have been secured upon which it is planned to study the products of dissociation.

It is recommended that the referee for next year attempt to determine the relative amounts of some of the dissociation products in water-soluble and water-insoluble meat proteins.

¹ U. S. Bur. Chem. Bull. 107, Rev., p. 51.

² *Ibid.*, p. 67.

³ *Ibid.*, pp. 243-51.

REPORT ON THE SEPARATION OF NITROGENOUS SUBSTANCES (MILK AND CHEESE).

BY LEROY S. PALMER (Agricultural Experiment Station, Columbia, Mo.),
Associate Referee.

The recommendation adopted for this work at the last meeting of the Association called for studies leading to the adoption of methods for the determination of the non-casein proteins and the products of protein decomposition in milk. Your referee begs to report that a serious illness last spring prevented his giving this work the attention originally planned. Some progress has been made, however, and although the results secured to date are too indefinite for detailed report, the work is being continued.

In the report made last year attention was called to the fact that the combined official method of the Association for casein, and the provisional method for the so-called albumin does not remove all the substances from milk that are of true protein character. This is particularly true in old milk that has undergone protein decomposition; but the writer wishes to point out that even the very freshest milk will usually show a non-casein-non-heat-coagulable-protein percentage approximately equal to the percentage of heat-coagulable protein obtained by the present provisional method. The following table shows the analyses of six 10 cc. portions of the same skim milk¹, the analyses being started about 3 hours after the milk was drawn.

Per cent of protein in milk by different methods.

METHOD	SAMPLE 1	SAMPLE 2	SAMPLE 3	SAMPLE 4	SAMPLE 5	SAMPLE 6
Total protein (official).....	3.80	3.80	3.80	3.80	3.80	3.80
Casein (official).....	2.89	2.89	2.89	2.84	2.84	2.88
Heat-coagulable proteins (provisional)....	0.526	0.502	0.451	0.529	0.544	0.510
Residual proteins (precipitated by Almén's reagent).....	0.280	0.280	0.288	0.273	0.322	0.274
Residual nitrogen (as protein not precipi- tated by Almén's reagent).....	0.160	0.205	0.301	0.232	Lost	0.205

Other figures could be presented showing relatively the same results for other samples of milk. The preceding figures show approximately 8% of the total true protein of fresh milk not recovered by acid precipitation or heat coagulation, a percentage sufficiently great to demand attention. Unless further study shows that these residual proteins, at least those precipitated by Almén's tannic acid reagent, are formed as the result of the present methods of analysis for casein and heat-coagulable protein, it would seem that the presence of these simpler protein substances in fresh milk should be recognized, and suitable methods incorporated in

¹ Mixed milk from the University herd.

the official methods for their determination. Careful distinction should be made at the same time between these substances present in fresh milk, and the products of protein decomposition, some of which would be determined by the same method. The data at least serve to call attention to the need of a careful study of the origin and exact character of these protein bodies present in fresh milk, which would ordinarily be called proteoses and peptones.

RECOMMENDATION.

It is recommended—

(1) That the studies be continued leading to the adoption of methods for the determination of the non-casein proteins and the products of protein decomposition in milk.

REPORT ON DAIRY PRODUCTS¹.

By LEWIS I. NURENBERG (State Department of Health, Boston, Mass.)
Referee.

A study, and compilation of figures, has been made of the sour serum method for the detection of added water in milk. In this work the referee has had the coöperation of Messrs. J. T. Keister, associate referee, and L. W. Ferris, of the Bureau of Chemistry, Washington, D. C., and George B. Taylor, J. R. Keeney, and J. S. Slack, of the State Board of Health, New Orleans, La.

The referee has obtained figures of over 660 samples of known purity milk from individual cows, and 49 samples from herds. These samples are representative of practically all breeds of cows mentioned in the 1914 report and of such influencing conditions as variation in season and lactation period. More than one-half of the total number represent low-grade milk.

The methods of preparation and analysis of the serum are as follows:

PREPARATION OF THE SOUR SERUM².

Allow the milk to sour spontaneously, and filter.

REFRACTION OF SOUR SERUM.

Determine the index of refraction of the clear serum at 20°C. by means of the Zeiss immersion refractometer. A refraction below 38.3 indicates added water.

Transfer 25 cc. of the serum to a flat-bottomed platinum dish and evaporate to dryness over the water bath. Then heat the contents of the dish over the small flame (to avoid sputtering) until charred. Place the dish in an electric muffle, with pyrometer connected, and ash at a heat not greater than 500°C., or 900°F. Cool and weigh. Express result in grams per 100 cc. An ash below 0.730 indicates added water.

¹ Presented by H. C. Lythgoe.

² Matthes and Müller. Z. offic. Chem., 1903, 9: 173-8.

In the original article as described by A. Burr and F. M. Berberich¹, the authors have outlined the following method:

ASH OF SOUR SERUM¹.

Measure 50 cc. of the serum into a platinum dish, evaporate to dryness, and carbonize over a low flame. Extract the char with hot water, burn the insoluble residue, add the solution to this ash, evaporate to dryness, ignite at a low temperature and weigh.

Experience has shown, however, with the exercise of care and an accurate pyrometer that the former stated method is as accurate as the latter, and involves less manipulation.

Table 1 contains the refractions of the sour serum of 660 samples of known purity milk from individual cows and 49 samples from herds. In the case of the individual cows the variation in the readings extends from 38.3-45.9. The greatest percentage of samples (25.2%) refracts between 41 and 42. There appears to be a certain uniformity each side of this medial point. The percentage of samples refracting between 40 and 41, and 42 and 43 is practically the same (from 18-20%); between 39 and 40, and 43 and 44 the percentage is from 12-13.5%, while from 38.3-39 and 44-45 the percentage is 4-5%.

In the herds the largest percentage of samples (75%) refracts between 41 and 43. Between 40 and 41 and 43 and 44 the percentage is from 10-12. As would be expected, no samples were found refracting at the maximum (45.9) or minimum (38.3) limits.

TABLE 1.
Variation in refractive indices of sour serum of known purity milk.
(660 samples from individual cows; 49 samples from herds.)

INDIVIDUAL COWS			HERDS		
Reading at 20°C.	Number of samples examined	Per cent of samples examined	Reading at 20°C.	Number of samples examined	Per cent of samples examined
38.3-38.9	34	5.2	38.3-38.9	0	0
39-39.9	81	12.3	39-39.9	2	4.1
40-40.9	121	18.3	40-40.9	6	12.3
41-41.9	167	25.2	41-41.9	18	36.7
42-42.9	133	20.2	42-42.9	18	36.7
43-43.9	90	13.6	43-43.9	5	10.2
44-44.9	27	4.1	44-44.9	0	0
45-45.9	7	1.1	45-45.9	0	0

Following is the graphic representation of the figures given in Table 1. The curves cross at the 20% line with a refractive index of 40.8 and again at the 15% line with a refractive index of 43.3.

¹ Chem. Ztg., 1908, 32: 617-S.

VARIATION IN REFRACTIVE INDICES OF SOUR SERUM OF KNOWN PURITY MILK.

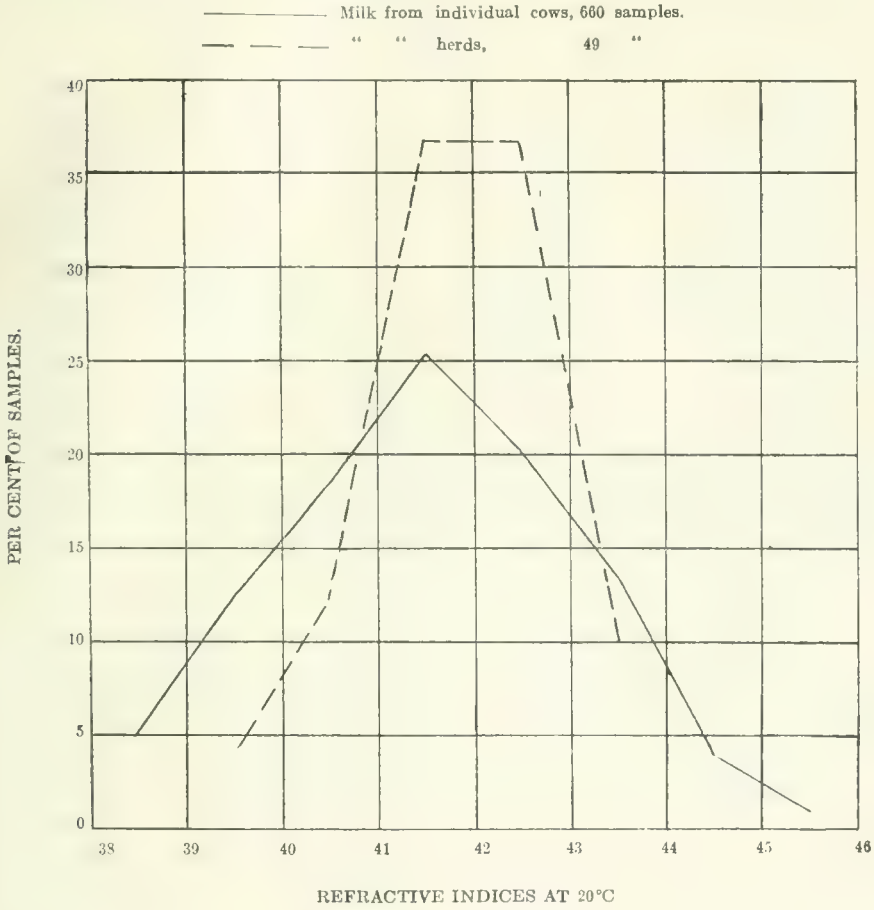


TABLE 2.
Variation in ash of sour serum of known purity milk.
 (504 samples from individual cows; 35 samples from herds.)

INDIVIDUAL COWS			INDIVIDUAL COWS			HERDS		
Ash gram per 100 cc.	Number of samples examined	Per cent of samples examined	Ash gram per 100 cc.	Number of samples examined	Per cent of samples examined	Ash gram per 100 cc.	Number of samples examined	Per cent of samples examined
0.730-0.739	41	8.2	0.840-0.849	17	0.730-0.739
0.740-0.749	54	0.850-0.859	7	4.8	0.740-0.749	1	2.9
0.750-0.759	34	17.3	0.860-0.869	16	0.750-0.759	2
0.760-0.769	51	0.870-0.879	7	4.6	0.760-0.769	4	17.1
0.770-0.779	38	17.7	0.880-0.889	7	0.770-0.779	2
0.780-0.789	52	0.890-0.899	4	2.2	0.780-0.789	6	22.8
0.790-0.799	45	19.3	0.900-0.909	4	0.790-0.799	6
0.800-0.809	51	0.910-0.919	0.8	0.800-0.809	9	42.9
0.810-0.819	37	17.3	0.920-0.929	1	0.810-0.819	4
0.820-0.829	26	0.930-0.939	1	0.4	0.820-0.829	1	14.3
0.830-0.839	9	7.0	0.940-0.949	2	0.4	0.830-0.949

Table 2 presents the sour serum ash figures of 504 samples of known purity milk from individual cows and 35 samples from herds. In the individual cows only 8.2% of the ash figures extended between the minimum limit 0.730 and 0.740, while 71.6% were between 0.740 and 0.820; 16.4% of the figures were between 0.820-0.880, and 3.8% between 0.880-0.950. In the milk from herds no samples were found with ash figures from the minimum limit 0.730-0.739, or from 0.830 to the maximum limit 0.949; 2.9% of the ash figures were between 0.740-0.749; 82.8% were between 0.750 and 0.810; and 14.3% were between 0.810 and 0.830.

The data from Table 2 are shown in the following plot. The curves cross at the 17.5% line with an ash percentage of 0.761.

VARIATION IN ASH OF SOUR SERUM OF KNOWN PURITY MILK.

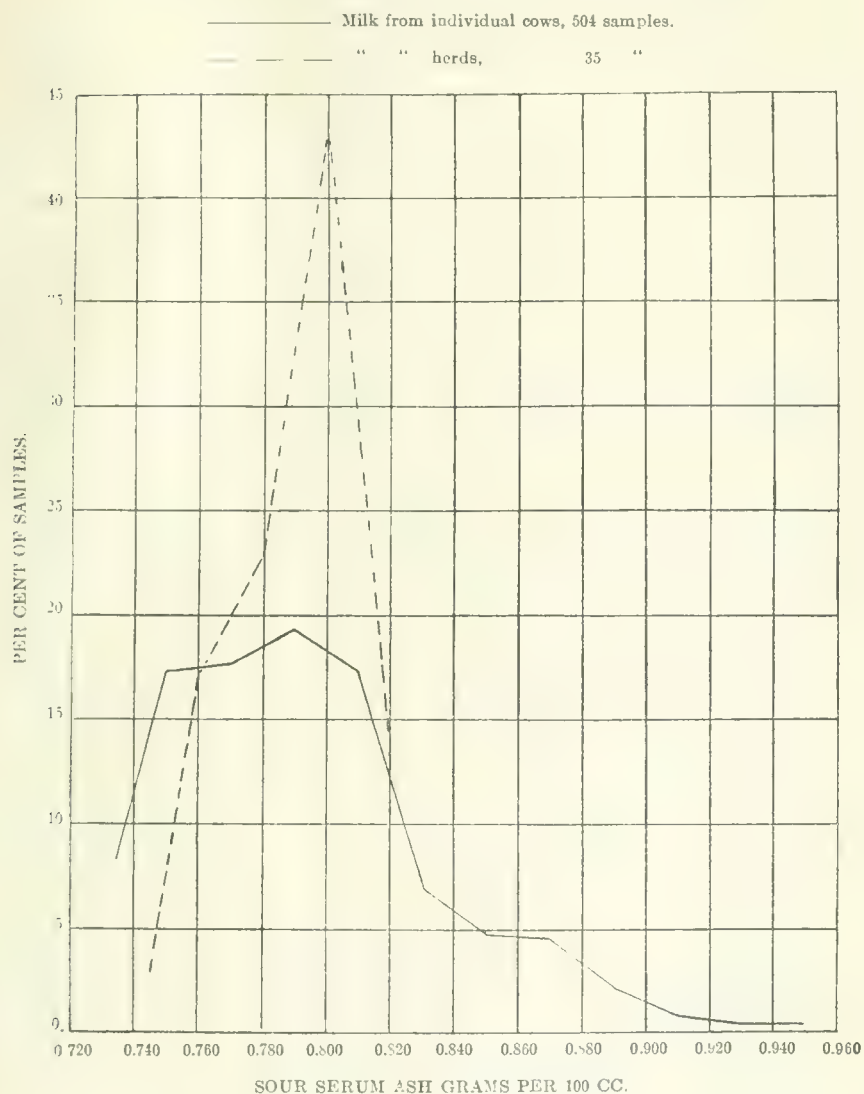


TABLE 3.

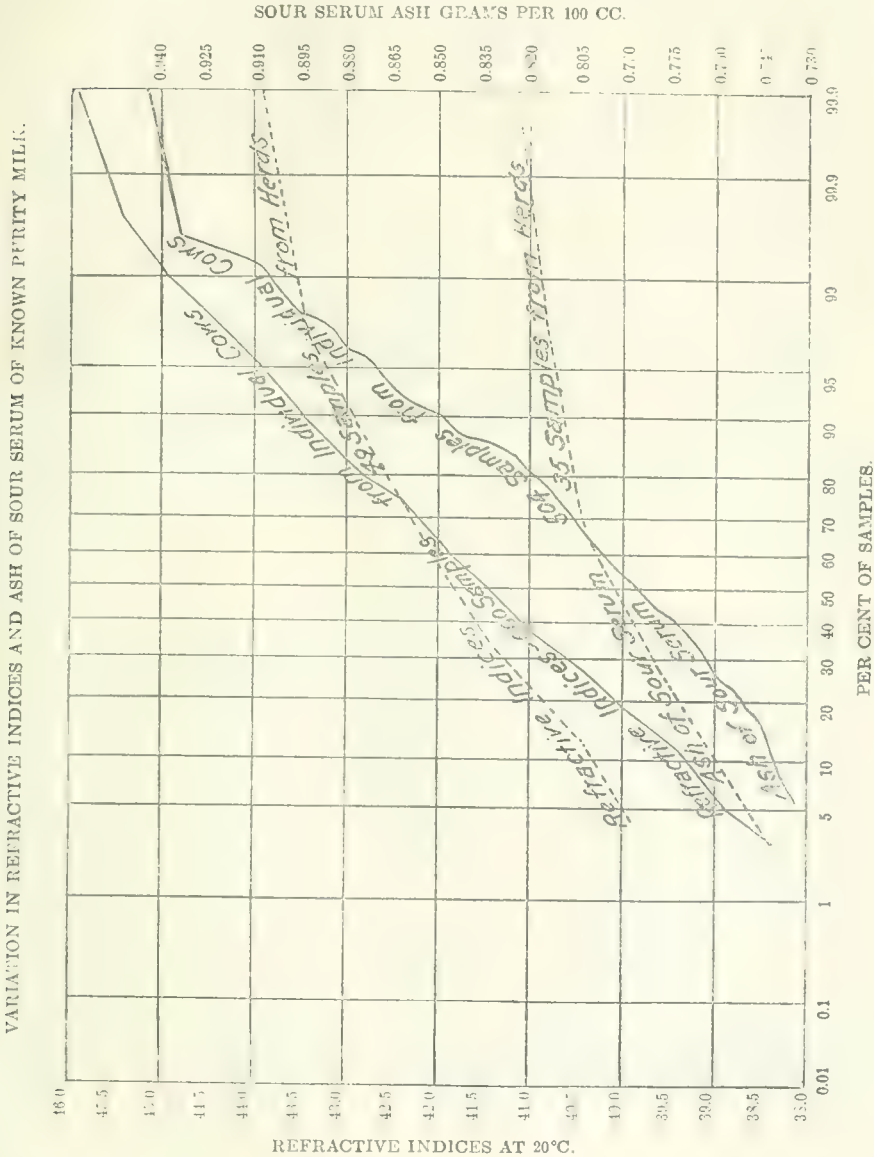
Variation in refractive indices and ash of sour serum of known purity milk.
From individual cows.

REFRACTIVE INDICES	NUMBER OF SAMPLES EXAMINED	PER CENT OF SAMPLES EXAMINED	ASH GRAM PER 100 CC.	NUMBER OF SAMPLES EXAMINED	PER CENT OF SAMPLES EXAMINED	ASH GRAM PER 100 CC.	NUMBER OF SAMPLES EXAMINED	PER CENT OF SAMPLES EXAMINED
38.0-38.4	20	3.0	0.730-0.734	30	5.9	0.840-0.844	8	1.6
38.5-38.9	14	2.1	0.735-0.739	11	2.2	0.845-0.849	9	1.8
39.0-39.4	45	6.8	0.740-0.744	33	6.5	0.850-0.854	3	0.6
39.5-39.9	36	5.5	0.745-0.749	21	4.1	0.855-0.859	4	0.8
40.0-40.4	60	9.1	0.750-0.754	19	3.8	0.860-0.864	8	1.6
40.5-40.9	61	9.2	0.755-0.759	15	3.0	0.865-0.869	8	1.6
41.0-41.4	87	13.2	0.760-0.764	33	6.5	0.870-0.874	5	1.0
41.5-41.9	80	12.1	0.765-0.769	18	3.6	0.875-0.879	2	0.4
42.0-42.4	87	13.2	0.770-0.774	26	5.2	0.880-0.884	4	0.8
42.5-42.9	46	7.0	0.775-0.779	12	2.4	0.885-0.889	3	0.6
43.0-43.4	52	7.9	0.780-0.784	31	6.1	0.890-0.894	1	0.2
43.5-43.9	38	5.8	0.785-0.789	21	4.1	0.895-0.899	3	0.6
44.0-44.4	18	2.7	0.790-0.794	25	4.9	0.900-0.904	3	0.6
44.5-44.9	9	1.4	0.795-0.799	20	4.0	0.905-0.909	1	0.2
45.0-45.4	5	0.7	0.800-0.804	32	6.3	0.910-0.914
45.5-45.9	2	0.3	0.805-0.809	19	3.8	0.915-0.919
Total.....	660	100.0	0.810-0.814	27	5.4	0.920-0.924	1	0.2
			0.815-0.819	10	2.0	0.925-0.929
			0.820-0.824	20	4.0	0.930-0.934	1	0.2
			0.825-0.829	6	1.2	0.935-0.939
			0.830-0.834	7	1.4	0.940-0.944	2	0.4
			0.835-0.839	2	0.4	Total...	504	100.0

From herds.

REFRACTIVE INDICES	NUMBER OF SAMPLES EXAMINED	PERCENT OF SAMPLES EXAMINED	ASH GRAM PER 100 CC.	NUMBER OF SAMPLES EXAMINED	PERCENT OF SAMPLES EXAMINED
38.0-38.4	0.730-0.734
38.5-38.9	0.735-0.739
39.0-39.4	0.740-0.744	1	2.9
39.5-39.9	2	4.1	0.750-0.754	2	5.7
40.0-40.4	4	8.2	0.760-0.764	2	5.7
40.5-40.9	2	4.1	0.765-0.769	2	5.7
41.0-41.4	8	16.3	0.770-0.774	1	2.9
41.5-41.9	10	20.4	0.775-0.779	1	2.9
42.0-42.4	11	22.4	0.780-0.784	2	5.7
42.5-42.9	7	14.3	0.785-0.789	4	11.4
43.0-43.4	4	8.2	0.790-0.794	4	11.4
43.5-43.9	1	2.0	0.795-0.799	2	5.7
Total.....	49	100.0	0.800-0.804	4	11.4
			0.805-0.809	5	14.3
			0.810-0.814	4	11.4
			0.820-0.824	1	2.9
			Total.....	35	100.0

Table 3 contains the refractive indices and sour serum ash figures arranged for the arithmetic probability curve, which follows.



If the samples were fairly representative, and if the variable follows the law of probability, the resulting curve will be a straight line. Although these lines are not straight, yet they are so nearly straight that

it can be said that sufficiently representative samples have been taken to draw definite conclusions.

The medial line (50% line) shows that 50% of the samples had refractive indices below 41.4 from individual cows, and 41.8 from herds. The curves for the refractive indices of the herds and individual cows cross at the 73% point with refractive index of 42.4. The medial line also shows that in 50% of the samples the sour serum ash was below 0.784 from individual cows and 0.793 from herds. The curves for the sour serum ash of the herds and individual cows cross at the 68% line with the sour serum ash of 0.802.

Table 4 contains the results submitted by Mr. L. W. Ferris of the U. S. Bureau of Chemistry. His experimental work has shown conclusively that the time of spontaneous souring affects the sour serum ash. In the cases of partial souring the sour serum ash figure is lower than the acetic serum ash which, of course, should not be. Unless the sample is sufficiently sour, some of the calcium salt is precipitated with the casein and the sour serum ash is very liable to be low, as shown in Table 4. The minimum sour serum ash figure, reported as 0.671, on longer souring was raised to 0.722 which corresponds to the figure obtained by the acetic serum after being multiplied by 1.02.

COMMENTS BY ANALYSTS.

All samples except the first 8 represent single milkings from individual cows.

The acetic acid serum was obtained by adding 2 cc. of 25% acetic acid to 100 cc. of milk. The results were multiplied by 1.02 to make them comparable with the results on the spontaneous serum. The results obtained by both methods are about the same. The degree of souring of the spontaneous serum appears to make no difference in either the ash or refraction, although clearer serums were obtained by allowing the sample to stand until the whey and curd had separated. In some cases it was impossible to get a clear serum. The spontaneous souring involves less work than the acetic acid method and in most cases the spontaneous serum is about as clear as that obtained by the acetic acid method.

One sample gave a refraction of 37.9 (cow No. 15) on the spontaneous serum and samples from 4 different cows (Nos. 2, 5, 6 and 7) gave an ash of serum below 0.73.

Table 5 contains the results submitted by Mr. J. T. Keister, associate referee. In this table there are 3 abnormal sour serum ash figures. The referee is of the opinion that in each case the milk was not thoroughly sour. If the sample is thoroughly sour, a clear serum will be obtained which will contain all the mineral matter.

TABLE 1.
Data for detection of added water to milk. (September-October, 1915.)
 (Leslie W. Ferris, Washington, D. C.)

LABO- RATORY NUM- BER	NUM- BER OF COW	BREED	AGE OF COW	TIME SINCE CALVING	AMOUNT OF MILK	TIME OF MILKING	SPONTANEOUS SERUM			ACETIC ACID SERUM		
							Refraction at 20°C.	Ash	Lactic acid	Refraction at 20°C.	Ash $\times 1.02$	Lactic acid
			years	months	pounds		reading	gram per 100 cc.	gram per 100 cc.	reading	gram per 100 cc.	gram per 100 cc.
9420 ¹		Market milk					41.6	0.798		42.3	0.788	0.727
9421		Market milk					42.0	0.766		42.3	0.796	0.691
9422		Market milk					41.9	0.773				
		Same sample, longer souring					41.5	0.785				
9423		Market milk					40.7	0.782		42.0	0.780	0.657
		Same sample, longer souring					41.1	0.781				
9423		Market milk					41.5	0.775		41.7	0.783	0.695
		Same sample, longer souring					40.7	0.776				
9428	1	Holstein	4 $\frac{1}{2}$	6	20	1 p.m.	39.7	0.802	0.639	40.2	0.804	0.634
		Same sample, longer souring					39.9	0.802	0.675			
9434	1	Holstein	4 $\frac{1}{2}$	6	20	1 p.m.	39.7	0.801	0.497	39.4	0.814	0.733
		Same sample, longer souring					39.9	0.802	0.675			
		Same sample, longer souring					39.4	0.803	0.695			
9436	1	Holstein	4 $\frac{1}{2}$	6	12.5	7.30 p.m.	38.7	0.796	0.727	39.7	0.801	0.790
		Same sample, longer souring					39.1	0.811	0.713			
9438	1	Holstein	4 $\frac{1}{2}$	6	18	4.45 a.m.	39.7	0.812	0.675	40.2	0.815	0.760
9439	2	Holstein	2 $\frac{1}{2}$	5	19.2	1 p.m.	Cloudy ¹	0.717	0.581	41.8	0.722	0.733
		Same sample, longer souring					41.0	0.717	0.648			

TABLE 4.—Continued.

LABORATORY NUMBER	NAME OF COW	BREED	AGE OF COW	TIME SINCE CALVING	AMOUNT OF MILK	TIME OF MILKING	SPONTANEOUS SERUM		ACETIC ACID SERUM	
							Refractum at 20°C.	Ash gram per 100 cc.	Reduction at 20°C.	Ash × 1.02 gram per 100 cc. Lactic acid
9435	2	Holstein	2½	5 months	19.2 pounds	1 p.m.	reading	gram per 100 cc.	reading	gram per 100 cc.
							40.7	0.709	41.2	0.779
9437	2	Holstein	2½	5 months	12.8	7 30 p.m.	40.7	0.711		
							40.7	0.702		
9439	3	Holstein	2½	5 months	19.0	4 45 a.m.	40.7	0.671	40.7	0.751
							41.7	0.687		
9443	9	Grade Holstein	8-9	Nearly dry	6.5	p.m.	40.2	0.722		
							40.2	0.713		
9445	5	Holstein	5	4 months	21.5	p.m.	41.7	0.639	42.0	
							40.7	0.757	41.2	
9447	10	Grade Holstein	8-9	Nearly dry	13.2	p.m.	41.2	0.757	41.6	
							41.4	0.715	42.0	
9449	5	Holstein	5	4 months	21.5	p.m.	40.7	0.757	41.7	
							40.1	0.732	41.1	
9451	7	Holstein	6	8 months	8.6	p.m.	41.3	0.724	41.7	
							40.9	0.755	41.5	
9453	6	Holstein	6	7 months	12.3	p.m.	42.5	0.733	41.2	
							39.4	0.721	40.1	
9455	6	Holstein	6	7 months	15.5	p.m.	41.0	0.709	39.8	
							41.0	0.769	41.9	
9457	6	Holstein	6	7 months	15.5	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	
9459	7	Holstein	7	8 months	8.6	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	
9461	7	Holstein	7	8 months	8.6	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	
9463	9	Grade Holstein	8-9	Nearly dry	6.5	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	
9465	9	Grade Holstein	8-9	Nearly dry	6.5	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	
9467	10	Grade Holstein	8-9	Nearly dry	6.5	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	
9469	10	Grade Holstein	8-9	Nearly dry	6.5	p.m.	41.0	0.769	41.9	
							41.0	0.769	41.9	

9446	11	Grade Holstein	8-9	10	4.3	a.m.	42.7	0.807	0.675	42.7	0.811
9451	12	Grade Holstein	6	10	16.8	a.m.	38.5	0.794	0.517	38.7	0.796
9455	12	Grade Holstein	6	10	17.7	p.m.	41.4	0.816	0.734	39.2	0.818
9459	13	Grade Holstein	8	5.4	p.m.	41.4	0.924 ³	0.637	Cloudy ⁴	0.877
9460	13	Grade Holstein	8	8.0	p.m.	42.6	0.862	0.716	43.7	0.864
9464	14	Grade Holstein	6	$\frac{3}{4}$	19.0	a.m.	40.3	0.822	0.733	42.7	0.816
9465	15	Grade Holstein	9	4	7.0	p.m.	39.1	0.776	0.619	38.0	0.775
9466	15	Grade Holstein	9	4	10.0	a.m.	39.2	0.797	0.657	38.8	0.783
9472	15	Grade Holstein	9	4	7.8	p.m.	37.9	0.766	0.517	37.8	0.769
9467	16	Grade Holstein	2	11	6	p.m.	39.9	0.744	0.601	40.5	0.741
9468	16	Grade Holstein	2	11	8.2	a.m.	40.0	0.767	0.657	40.0	0.745
9473	16	Grade Holstein	2	11	7.4	p.m.	40.5	0.745	0.657	40.9	0.753
9441	17	Durham	5	2	13.0	p.m.	40.7	0.774	0.742	41.2	0.775
9442	18	Durham	2 $\frac{1}{2}$	1	p.m.	41.0	0.751	0.704	41.3	0.756
9470	19	Durham	6-7	$\frac{1}{2}$	17.4	p.m.	41.9	0.756	0.751	42.5	0.736
9471	20	Durham	5	$\frac{1}{2}$	17.4	p.m.	41.5	0.803	0.704	41.8	0.785
9440	21	Black Polled	2 $\frac{1}{2}$	$\frac{1}{2}$	13.0	p.m.	39.7	0.763	0.666	40.1	0.786
9445	22	Black Polled	2	$\frac{1}{2}$	13.0	a.m.	41.7	0.762	0.733	42.2	0.814
9469	23	Grade Guernsey	3	1	13.0	p.m.	41.4	0.772	0.779	42.1	0.755
Minimum		37.9	0.671	38.0	0.709
Maximum		42.7	0.924	43.7	0.877
Mean		40.6	0.765	41.0	0.769

¹ Could not get spontaneous serum clear enough to read.² Approximately.³ Extra high results may have been due to slow filtering of very small amount of serum.⁴ Could not get filtrate clear enough to read.

TABLE 5.
Analysis of milks from individual cows. (September-October, 1915.)
 (J. T. Keister, Washington, D. C.)

LABORATORY NUMBER	NUM-BER OF COW	BREED	AGE OF COW	TIME SINCE CALVING	AMOUNT OF MILK	TIME OF MILKING	FAT (ROSE-GOTTLIEB METHOD)	FAT (BARCOCK METHOD)	SOLIDS NOT FAT (GRAVIMETRICALLY)	SOLIDS NOT FAT (CALCULATED)	SOUR SERUM		ASH
											Refraction at 20° C.	Ash	
			years	months	pounds		per cent	per cent	per cent	per cent	reading	gram per 100 cc.	per cent
9434	1 ¹	Holstein	4½	6	15.2	1 p.m.	3.82	3.85	7.96	7.86	39.2	0.748	0.75
9436	1	Holstein	4½	6	12.5	7.30 p.m.	3.71	3.80	7.78	7.78	39.0	0.790	0.74
9438	1	Holstein	4½	6	18.0	4.45 a.m.	2.79	2.80	8.09	8.00	40.0	0.773	0.76
9435	2 ¹	Holstein	2½	5	16.4	4.22 p.m.	4.22	4.30	8.18	8.12	40.9	0.678	0.657
9437	2	Holstein	2½	5	12.8	7.30 p.m.	3.88	3.95	8.12	8.00	40.3	0.668	0.66
9439	2	Holstein	2½	5	19.0	4.50 a.m.	2.55	2.70	8.52	8.35	40.8	0.701	0.669
9448	3	Holstein	5	3	20.7	p.m.	3.02	3.10	8.33	8.48	40.9	0.753	0.73
9453	3	Holstein	5	3	23.2	a.m.	3.01	3.10	8.44	8.33	41.3	0.761	0.72
9450	4	Holstein	9	9	11.8	p.m.	3.14	3.15	8.74	8.53	41.1	0.731	0.73
9454	4	Holstein	9	9	16.8	a.m.	2.48	2.50	8.76	8.50	40.8	0.756	0.71
9475	4	Holstein	9	9	13.0	3.40 p.m.	2.80	2.80	8.78	8.46	41.1	0.745	0.68
9449	5	Holstein	5	4	21.5	p.m.	3.07	3.15	8.35	8.41	40.4	0.769	0.73
9456	5	Holstein	5	4	27.5	a.m.	3.02	3.15	8.36	8.45	40.9	0.743	0.70
9457	6	Holstein	6	7	12.3	p.m.	3.77	3.90	8.64	8.40	41.3	0.787	0.706
9458	6	Holstein	6	7	19.6	a.m.	3.30	3.05	8.13	7.96	39.2	0.740	0.68
9474	6	Holstein	6	7	15.5	p.m.	4.20	8.05	38.0	0.731	0.664
9461	7	Holstein	6	8	8.6	p.m.	3.31	3.30	8.76	8.56	41.9	0.770	0.69
9462	7	Holstein	6	8	10.0	a.m.	3.21	3.30	8.53	8.31	40.4	0.731	0.74
9476	8	Holstein	4	2	20.0	p.m.	3.75	8.10	39.7	0.729	0.71
9443	9	Grade Holstein	8 or 9	8 or 9	8.7	p.m.	3.47	3.20	8.56	40.9	0.853	0.76
9444	9	Grade Holstein	8 or 9	8 or 9	8.7	p.m.	3.72	3.50	8.63	0.73
9447	10	Grade Holstein	3	3	17.4	p.m.	4.00	8.26	40.4	0.731
9452	10	Grade Holstein	3	17.4	a.m.	3.60	8.59	42.7	0.739
9446	11	Grade Holstein	8 or 9	8 or 10	4.3	a.m.	7.70	8.99	43.2	0.808

9451	12	Grade Holstein	6	10	16.8	p.m.	3.40	3.45	7.49	7.54	41.8	0.801	0.75
9455	12	Grade Holstein	6	10	17.7	a.m.	2.84	2.95	7.65	7.71	41.7	0.813	0.73
9459	13	Grade Holstein	8	5.4	p.m.	4.80	9.30	41.8	0.882
9460	13	Grade Holstein	8	8.0	a.m.	4.40	9.08	42.6	0.895
9464	14	Grade Holstein	6	3	19.0	a.m.	3.10	8.97	41.1	0.832
9465 ²	15	Grade Holstein	9	4	7.0	p.m.	(1.10) ³	(1.10)	7.48	7.37	40.2	0.781	0.68
9466	15	Grade Holstein	9	4	10.0	a.m.	3.81	3.80	7.79	7.58	40.9	0.806	0.70
9472	15	Grade Holstein	9	4	7.8	p.m.	3.09	3.10	7.60	7.51	38.4	0.790	0.72
9467	16	Grade Holstein	2	11	6.0	p.m.	4.20	8.09	40.0	0.760
9468	16	Grade Holstein	2	11	8.2	a.m.	3.54	3.45	8.22	8.21	41.4	0.751	0.68
9473	16	Grade Holstein	2	11	7.4	p.m.	3.37	3.45	8.47	8.36	41.1	0.770	0.686
9441	17	Grade Durham	5	2	13.0	p.m.	3.75	8.46	40.8	0.782
9442	18	Grade Durham	2½	1	10.9	p.m.	3.50	8.55	41.0	0.745
9470	18	Grade Durham	6	3	17.4	p.m.	3.65	8.98	41.8	0.811
9471	20	Grade Durham	5	3	17.4	p.m.	3.80	8.55	41.8	0.761
9440	21	Black Polled	2½	2	13.0	p.m.	3.18	3.25	8.35	8.26	40.4	0.791	0.767
9445	21	Black Polled	2½	2	13.0	a.m.	3.10	3.20	8.86	8.68	41.9	0.814	0.79
9469	22	Grade Guernsey	3	1	13.0	p.m.	3.85	8.68	41.1	0.779
9478	23	Grade Jersey	20(about)	3	8.7	p.m.	2.83	2.80	8.33	40.4	0.806
9479	24	Grade Jersey	10	1	13.0	p.m.	4.80	7.81	40.1	0.816	0.75
9480	25	Grade Durham	5	1	10.9	p.m.	6.60 ³	8.57	40.6	0.869
9481	26	Grade Durham	10(about)	1	10.9	p.m.	7.90 ³	8.85	43.0	0.832
9482 ⁴	Herd	p.m.	4.60	8.42	40.7	0.790

¹ Cows milked 3 times a day.
² Sample taken by milker, probably represents first portion of the milking.
³ Only 8 hours since previous milking, which probably partly accounts for high fat figure.
⁴ Herd test of 10 cows (Nos. 13, 17, 18, 19, 20, 21, 23, 24, 25, 26).

TABLE 6.

Composition of milk from Fagot Dairy, Metairie Ridge, Jefferson Parish.
(G. B. Taylor, J. R. Reeny and J. S. Slack, Louisiana.)

NAME	AGE	BREED	AMOUNT OF MILK	TIME SINCE CALV- ING	SP. GR. AT 15° C.	TOTAL SOLIDS ¹	FAT	SOUR SERUM	
								Ash	Refrac- tion
	<i>years</i>		<i>pounds</i>	<i>months</i>		<i>per cent</i>	<i>per cent</i>	<i>gram per 100 cc.</i>	<i>reading</i>
Brown.....	8	Grade Jersey	3	9	1.0354	14.88	5.00	0.898	45.2
Nigger.....	8	Brown Grade Jersey	8	6	1.0344	15.11	5.40	0.876	43.6
Grandma.....	10	Holstein	8	$\frac{1}{2}$	1.0317	15.16	6.00	0.805	44.2
Hazel.....	3	Grade Jersey	4 $\frac{1}{2}$	2	1.0328	14.95	5.60	0.788	43.4
Spot.....	5	Grade Jersey	7	5	1.0315	15.71	6.50	0.744	43.1
Rabbit.....	7	Grade Jersey	8	2	1.0325	14.63	5.40	0.787	43.9
Silver.....	3	Grade Jersey	4 $\frac{1}{2}$	2	1.0302	14.89	6.10	0.707	42.3
Baby.....	6	Jersey	7	2	1.0260	...	8.20	0.880	43.1
Goldie.....	12	Jersey	5	6	1.0298	15.64	6.80	0.840	43.4
Red.....	8	Grade Jersey	5	6	1.0295	13.15	4.80	0.842	41.5
Brownie.....	5	Brown Grade Jersey	9	5	1.0298	15.40	6.60	0.844	44.4
Rose.....	10	Red Grade Jersey	8 $\frac{1}{2}$	8	1.0293	13.82	5.40	0.874	43.6
Jessie.....	6	Jersey	12 $\frac{1}{2}$	2	1.0315	14.14	5.20	0.744	42.0
Mollie.....	9	Holstein	6	10	1.0316	11.75	3.20	0.813	42.0
Marie.....	7	Jersey	5	10	1.0321	...	7.40	0.902	43.6
White.....	8	Grade Jersey	8	5	1.0351	14.57	4.80	0.866	43.6
Cherry.....	8	Grade Durham	9	4	1.0329	14.01	4.80	0.795	44.2
Crip.....	5	Grade Jersey	6	2	1.0322	14.55	5.40	0.820	41.7
Juliet.....	8	Grade Jersey	5 $\frac{1}{2}$	4	1.0345	11.52	2.40	0.708	42.8
Composite ²			29		1.0302	15.01	6.20	0.810	43.4
Herd ³			129 $\frac{1}{2}$		1.0315	13.89	5.00	0.805	42.8

¹ U. S. Bur. Animal Industry Bull. 134, p. 23.

² Composite of 6 Grade Jerseys.

³ Four Jerseys; 12 Grade Jerseys; 2 Holsteins; 1 Durham.

TABLE 7.
Composition of 8 samples of herd milk.
(Arranged in order of Total Solids.)

TOTAL SOLIDS	FAT	SOLIDS NOT FAT	PRO-TEIN	SUGAR	COPPER SERUM		SOUR SERUM		
					Ash	Refrac- tion	Ash	Refrac- tion	
<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>gram per 100 cc.</i>	<i>reading</i>	<i>gram per 100 cc.</i>	<i>reading</i>	
13.02	4.60	8.42	0.790	40.7	
12.95	4.10	8.85	3.29	4.74	0.70	37.4	0.774	40.4	
12.62	3.40	9.22	3.04	5.01	0.76	38.5	0.816	41.0	
12.60	3.35	9.25	2.33	5.04	0.78	38.7	0.832	41.9	
12.57	4.00	8.57	3.18	4.82	0.68	37.2	0.766	41.5	
12.24	3.55	8.69	3.34	4.65	0.74	36.7	0.808	
12.13	3.35	8.78	2.96	4.72	0.80	36.8	41.3	
11.67	3.40	8.27	2.84	4.51	0.77	36.3	0.804	39.1	
Maximum:	13.02	4.60	9.25	3.34	5.04	0.80	38.7	0.832	41.9
Minimum:	11.67	3.35	8.27	2.33	4.51	0.68	36.3	0.766	39.1
Average:	12.48	3.72	8.76	3.00	4.78	0.75	37.4	0.799	40.8
Average of 67 samp'les from herds reported in 1914.									
12.83	4.06	8.77	37.9	0.798	41.7	

There is no relation between the refraction of the sour serum and the sour serum ash, since these figures depend upon different milk constituents. When both of these figures fall below the lowest limits established for pure milk (38.3 and 0.730) it is absolute proof of the presence of added water, and all possibility of the sample being abnormal milk from a sick cow is removed.

In the determination of the ash of sour serum there are less than 2 grams of organic matter to be burned, and the influence of combustion upon the 190 mg. of ash is very slight.

The refractive index of the sour serum occurs half way between the copper and acetic indices. Where a milk has partially soured the copper serum reading will be higher, and the acetic serum lower than would be given by the same milk before souring. In such cases the sour serum is the only reliable reading to be made.

Six years' experience with the refractive index and 4 with the sour serum ash have shown these methods to be invaluable. In all doubtful cases the sour serum ash has served as a court of last resort.

RECOMMENDATIONS.

It is recommended—

- (1) That the following be adopted as auxiliary provisional methods.

DETECTION OF ADDED WATER.

*Sour serum*¹.

Allow the milk to sour spontaneously, filter and determine the index of refraction of the clear serum by means of the Zeiss immersion refractometer. A reading below 38.3 indicates added water.

*Ash of sour serum*².

Allow the milk to sour spontaneously and filter. Transfer 25 cc. of this serum to a flat-bottomed platinum dish and evaporate to dryness over the water bath. Heat the contents of the dish over a small flame (to avoid sputtering) until charred. Place the dish in an electric muffle, with pyrometer connected, and ash at a heat not greater than 500°C., or 900°F. Cool and weigh. Express result in grams per 100 cc. An ash below 0.730 indicates added water. (A white ash is invariably obtained after one leaching.)

Ash of acetic serum.

Transfer 25 cc. of the serum to a flat-bottomed platinum dish and proceed as directed under "Ash of Sour Serum". An ash figure below 0.715 gram per 100 cc. indicates added water.

(2) That in conjunction with the copper, acetic or sour serum refraction method, the ash of the sour serum or of the acetic serum be determined in all cases where the indices of refraction fall below the minimum limit. The acetic serum ash multiplied by the factor 1.021 equals the sour serum ash (dilution of the acetic serum being 2%).

No report was made by the referee on the general subject of food adulteration.

REPORT ON COLORS.

By W. E. MATHEWSON (Food and Drug Inspection Laboratory, U. S. Appraiser's Stores, New York, N. Y.), *Associate Referee*.

Two samples of commercial caramel and sealed samples of high grade commercial amaranth, orange I, naphthol yellow S, and light green S F yellowish were sent out for collaborative work. The collaborating analysts were asked to examine the caramels in 0.2% solution by those methods for the detection of this substance that they consider best. In the coal tar dyes they were asked to determine the amount of color by the titanium trichlorid reduction method of Knecht and Hibbert which has the merit of using a comparatively simple reagent, stable if protected from the air, and which has given good results in the New York Food and Drug Inspection Laboratory of the U. S. Bureau of Chemistry when used under

¹ Matthes and Müller. Z. öffent. Chem., 1903, **9**: 173.

² Burr and Berberich. Chem. Ztg., 1908, **32**: 617-8.

the proper precautions and with a thorough consciousness of the sensitiveness of the reagent to air. With the samples was sent a statement of the precautions to be followed in applying the method to each of the permitted dyes. These are embodied in the suggested provisional methods which come before the Association this year and need not be given here.

The collaborators reported upon the caramel samples as follows:

RESULTS OF COLLABORATIVE WORK.

C. F. Jablon: The samples of caramel under observation did not behave alike to the Amthor test. In solution of 0.2%, one gave a slight precipitate, while the other gave a pronounced precipitate. In stronger solutions (1%) a slight turbidity was observed in the first, while the other showed a pronounced precipitate.

The Woodman-Newhall test does not appear to give any more reliable results. In weaker solutions (0.2%) it appears to be less satisfactory, while a 1% solution gave positive reactions. To sum up the series of experiments, it is safe to state that although strong caramel solutions gave positive tests, ordinary concentrations, such as may be found in food analysis, do not give very satisfactory results.

Ray W. Clough: A few qualitative tests on the caramel samples gave the following results:

Crampton and Simons fuller's earth method¹, all color extracted from a 0.2% solution; Amthor test, negative; Woodman-Newhall method, good results (especially heavy precipitate with the phenylhydrazin reagent).

C. R. Smith: The Woodman-Newhall method was applied to the samples of caramel. It has been found that they, as well as other samples of commercial caramel and caramel prepared in the laboratory, precipitate out well in the Woodman-Newhall method. It is doubtful if the same holds good in fruit juices and other food products. In testing black currant juice of high acidity no precipitate was produced, using the usual proportion of reagents after neutralizing with potash.

By adding a large excess of zinc chlorid (8 cc.) and potash sufficient to produce a good precipitate a large amount of brownish black coloring matter was carried down, which, however, did not precipitate by the paraldehyde test. The Woodman-Newhall test applied with special reference to each substance may be useful if carefully studied and developed.

The results obtained by the collaborators in the determination of the amount of color in the samples of commercial anilin colors are given in Table 1.

¹ U. S. Bur. Chem. Bull. 107, Rev., p. 199.

TABLE 1.

Determinations of the amount of pure color in samples of anilin colors by titration with titanium trichlorid.

ANALYST	NAPHTHOL YELLOW S	ORANGE I	AMARANTH	LIGHT GREEN S F YELLOWISH
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
C. L. Black, Food and Drug Inspection Laboratory, U. S. Appraiser's Stores, Philadelphia, Pa.....	93.28	90.82	88.39	66.44
A. L. Burns, Food and Drug Inspection Laboratory, U. S. Appraiser's Stores, New York, N. Y.....	92.9	91.7	88.6	66.8
C. F. Jablon, Food and Drug Inspection Laboratory, U. S. Appraiser's Stores, New York, N. Y.....	92.7	91.0	88.7	67.1
Ray W. Clough, Arcade Annex Building, Seattle, Wash.....	93.4	93.8	88.8	66.1
C. R. Smith, Food and Drug Inspection Laboratory, U. S. Appraiser's Stores, New York, N. Y.....	93.1	93.2	89.1	67.9

THE SEPARATION OF PONCEAU 3R FROM OTHER DYES, ESPECIALLY
NAPHTHOL YELLOW S BY THE USE OF A
SOLUBLE BARIUM SALT.

(C. F. Jablon, New York.)

As the quantitative separation and estimation of ponceau 3R and naphthol yellow S is, under ordinary conditions, rather slow, a method was sought whereby the more or less tedious amyl alcohol extraction would be eliminated.

Inasmuch as ponceau 3R forms a very difficultly soluble barium compound¹ a series of experiments were performed using barium and strontium salts as precipitants in solutions containing the dyes in various concentrations. The best results were obtained as follows:

Dissolve the dye mixture in a definite amount of water. Transfer an aliquot containing about 0.1-0.3 gram of naphthol yellow S to a 250 cc. volumetric flask and dilute with water to about 225 cc. Add 2.0 cc. of 10% barium acetate and 1 cc. of glacial acetic acid, shake thoroughly and make up to the mark with water. Then pass through a dry filter and titrate an aliquot with standard titanium trichlorid solution in sodium tartrate solution, using light green S F yellowish as indicator. Titrating an equivalent part of the original solution gives the total color, and subtracting the yellow, the difference gives the value for ponceau 3R.

Table 2 gives some results obtained.

¹ Heumann. Die Anilinfarben und ihre Fabrikation. 1903, 4 (1): 856.

TABLE 2.

Determinations by the barium method of naphthol yellow S in presence of ponceau 3R.
(C. F. Jablon, New York.)

SAMPLE	WEIGHT PONCEAU TAKEN	WEIGHT YELLOW TAKEN	WEIGHT YELLOW FOUND	YELLOW RECOVERED
	<i>gram</i>	<i>gram</i>	<i>gram</i>	<i>per cent</i>
1	0.100	0.2491	0.2496	100.2
2	0.200	0.2076	0.2050	98.7
3	0.300	0.2030	0.2028	99.9
4	0.200	0.1833	0.1819	99.2
5	0.100	0.1538	0.1555	101.2
6	0.300	0.1005	0.0998	99.0
7	0.200	0.0240	0.0258	107.6
8	0.300	0.0000	0.0013

The filtrate from the precipitated ponceau 3R was somewhat redder in shade, so it was deemed advisable to determine colorimetrically with a Koenig-Martens instrument the amount of ponceau remaining in solution. Following are the amounts found, using various concentrations of ponceau 3R:

Weight of ponceau taken	Ponceau in solution
<i>gram</i>	<i>per cent</i>
0.05	2.25
0.125	1.4
0.250	0.5
0.500	0.2
1.000	0.1

The incomplete precipitation is probably due to the fact that the ponceau 3R used was not a chemically uniform product. It was the ordinary grade of dye used for food products which contains more or less of the lower homologues derived from xyloidin, etc., and also small amounts of monosulphonated color such as sodium trimethyl-benzene-azo-2-naphthol-6-sulphonate.

Mixtures of ponceau 3R with the other permitted dyes in amounts varying from 0.1–0.3 gram of each dye when treated by the procedure just described showed that erythrosine is precipitated almost completely, and the indigo somewhat less so with ponceau. Amaranth and orange I are carried down with the ponceau precipitate only slightly, if at all. Light green S F yellowish appears to remain entirely in the filtrate.

Because of the high relative oxidizing power of naphthol yellow S as shown towards titanium trichlorid, the error caused by the presence of a small and varying amount of ponceau in the filtrate does not greatly affect the determination of the yellow with mixtures containing not too small amounts of the latter. With the other dyes this is not the case, but the results stated may be applied to the analysis of mixtures containing ponceau and naphthol yellow with the others. Here the total dye

in the filtrate is determined with titanium, the amaranth, green, etc., estimated directly by some suitable method, and correction made to obtain the value for the yellow. With lesser accuracy, the procedure may be used for the direct separation of ponceau from orange, amaranth and green. The quantitative determination of indigo carmine in admixture with amaranth by reduction with titanium trichlorid, reoxidation with air, and estimation of the blue colorimetrically seems to give good results.

Ponceau and yellow being separated with some difficulty by other methods and often used in admixture with each other, the procedure just given will be found useful in examining mixtures.

THE USE OF THE SPECTROPHOTOMETER FOR THE EXAMINATION OF FOOD COLORING MATTER.

(W. E. Mathewson, New York.)

No other methods have the wide range of applicability of those depending on exact measurements under suitable conditions of the opacity of solutions to light of various wave lengths. The principles and methods of spectrophotometry have been developed by numerous investigators and are discussed in an extensive literature¹. These methods are in limited use, probably because the necessary instruments are so expensive that few laboratories are equipped with them. Hence no collaborative work could be done this year. In this part of the report some experimental work of the referee is reported which was designed to learn to what extent spectrophotometric methods might be used in the analysis of food colors. As each of the various optical systems that have been devised and each of the light sources available has its advantages and disadvantages, such apparatus as seemed most suitable was selected.

APPARATUS AND PROCEDURE.

The spectrophotometer used was a Koenig-Martens instrument with Rutherford prism, made by Schmidt and Haensch. Some experience with a spectrophotometer of the Vierort principle has convinced the writer that those forms of apparatus in which an eye-piece is employed (forming a system focused on the collimator slit) are much more difficult to use in work where it is very frequently necessary to take readings in parts of the spectrum in which the absorption varies greatly for small variations in the wave length. As a light source a 1500-3000 c. p. projection lamp with automatic arc and condensing lenses was employed. A Nernst

¹ For a résumé compare especially Kayser. *Handbuch der Spectroscopic*. 1905, 3; and G. and H. Kruss. *Kolorimetrie und Quantitative Spektrolana yse*. 2d ed., 1909.

lamp although steadier can scarcely be used for measurements in the blue and violet. Monochromatic light sources suitable for all regions of the spectrum would be much more preferable, but would add greatly to the cost of the equipment. An absorption cell of the Vierort type was used, 11 mm. in width and containing the Schultz glass cube exactly 1 cm. in thickness. This form of cell is rather objectionable for measurements in non-homogeneous light, as all the rays passing into the instrument must go through 1 mm. of the solution; but the laboratory is not provided with the better apparatus devised by Martens and Grünbaum¹. The cell was placed before the instrument, the glass cube first in front of one slit, then the other. The ocular slit or screen in the telescope is rather troublesome to get at and adjust in the Koenig-Martens instrument, and chiefly for this reason was set at a width corresponding to 100 (smallest) divisions on the scale carrying the telescope (at the mercury line 546.1), and kept at this width for all the measurements. To correspond, the telescope was moved 100 scale divisions between the successive regions. The jaws of the divided slit before the collimating lens were set as close as possible, still giving a field of sufficient brightness, and the observations made with the glass cube before each half of the slit after each resetting of the latter. The necessary opening of the collimator slit corresponded to a maximum overlapping in the field of view of rays differing by about $2.5 \mu\mu$ in the indigo and violet to $0.1 \mu\mu$ in the greenish yellow. In all cases the width of the slit was noted in taking measurements, though for Table 3, containing averaged figures, the statements just made are sufficient. For each reading the rotating nicol prism was set 2 or 3 times and the mean value recorded. The results were calculated to "extinction coefficients" e , by the formula

$$e = \log \tan a_1 + \log \cot a_2$$

a_1 and a_2 representing respectively the angular readings with the Schulz body before the right and left parts of the slit. For the loss of light by reflection at the surfaces of the Schulz body the correction for water was found to be 0.02, this amount to be added to the observed "extinction coefficient". In Table 3, this correction has not been made, however.

EXPERIMENTAL RESULTS.

Extinction coefficients of the permitted dyes.—In Table 3 are given the observed "specific extinction coefficients" obtained with solutions of the 7 permitted dyes, corresponding to the values for 0.001% solutions. Solutions of each color were made containing 0.001%, 0.004%, 0.008%, etc., of pure dye as determined by analysis, the strongest solution being

¹ Ann. Physik., 1903, 4th ser., 12: 991-2.

from 0.064–0.512%. The extinction coefficients for each solution were determined in those regions where the values ranged from 0.030–1.50. From these figures the e values for 0.001% solutions through the whole spectrum were calculated, it being assumed that the observed figures of value equal to about 0.60, if not more accurate, were at least more suitable for subsequent application.

Light green S F yellowish being quite sensitive to alkali, dilute solutions in pure water if kept in glass soon became paler and in 24 hours may show a conversion of 50% or more of the dye into the colorless carbinol form. Orange I and erythrosine are also sensitive indicators and may contain as impurities small amounts of sodium carbonate, free color acid, etc. This source of difficulty was practically eliminated by adding to each solution a little of a mixture of acetic acid and secondary sodium phosphate. A stock solution was made containing in each liter 71.4 grams of crystallized secondary sodium phosphate and 9.6 cc. of glacial acetic acid. Five cc. of this solution were used in making up each 100 cc. of the dye solutions, giving these latter a disodium phosphate normality of 0.01, and an acetic acid normality of 0.008 plus. This amphoteric mixture is faintly acid to litmus, faintly alkaline to alizarin and does not perceptibly change the absorption of the solutions of the colors other than light green S F yellowish, when added to their absolutely neutral solutions in not too large quantities.

The extinction coefficients as determined are not exactly proportional to concentration, being affected by a number of factors. The molecular condition of both electrolytes and colloids varies with changes in concentration, and the use of non-homogeneous light is open to various objections. However, the figures serve as a guide, indicating especially where are located those regions of maximum absorption best adapted for measurements, and the values to be expected. The accurate estimation of a dye must be made by determining the approximate concentration of its solution by the use of Table 3, changing the concentration, if necessary, so that the values for e will be from 0.6–1.00 in the region of maximum absorption, making a standard solution of the same calculated concentration from a pure color and determining the absorption constants for each under exactly the same conditions. The values obtained for e will not differ greatly and it may be assumed under these conditions that the differences are proportional to concentration.

Analysis of mixtures.—The analysis of mixtures must be carried out in a similar way. After quantitative examination of the constituents by chemical, spectroscopic or other means, measurements are made in those regions where the constituent dyes show the maximum absorption. The concentration is calculated, using the figures in Table 3, and from this a similar standard solution is prepared with colors of known purity. This is

TABLE 3.

Specific extinction coefficients of the 7 permitted food dyes.

(0.01 gram per liter is taken as unit concentration; i.e., the numbers given represent the coefficients of 0.001% solutions.)

WAVE LENGTH OF LIGHT IN REGION MEASURED	SCALE READ- INGS	NAPHTHOL YELLOW S	ORANGE I	PONCEAU 3R (FROM COMMER- CIAL PSEUDO- CUMIDIN)	ERY- THROSINE	AMA- RANTH	INDIGO- CARMINE	LIGHT GREEN SF YELLOW- ISH
418.8-422.6	22	0.46	0.30	0.13	0.027	0.093	0.029	0.18
422.6-426.5	23	0.46	0.32	0.13	0.028	0.095	0.029	0.18
426.5-430.7	24	0.46	0.38	0.13	0.028	0.105	0.030	0.18
430.7-435.1	25	0.45	0.45	0.13	0.029	0.11	0.031	0.17
435.1-440.0	26	0.45	0.52	0.14	0.030	0.115	0.032	0.15
440.0-444.5	27	0.44	0.58	0.155	0.034	0.125	0.033	0.13
444.5-449.6	28	0.40	0.67	0.17	0.040	0.13	0.034	0.105
449.6-455.0	29	0.34	0.73	0.19	0.052	0.14	0.034	0.077
455.0-460.7	30	0.29	0.80	0.23	0.068	0.165	0.034	0.056
460.7-466.7	31	0.23	0.87	0.265	0.085	0.19	0.034	0.034
466.7-473.1	32	0.16	0.91	0.30	0.12	0.215	0.034	0.023
473.1-480.0	33	0.105	0.93	0.34	0.17	0.25	0.034	0.015
480.0-487.1	34	0.060	0.90	0.39	0.24	0.29	0.035	0.011
487.1-494.8	35	0.031	0.84	0.43	0.32	0.33	0.038	0.011
494.8-503.0	36	0.014	0.71	0.48	0.37	0.37	0.043	0.013
503.0-511.8	37	0.0051	0.59	0.48	0.48	0.40	0.053	0.017
511.8-521.2	38	0.0024	0.43	0.46	0.76	0.41	0.066	0.024
521.2-531.4	39	0.00092	0.27	0.425	0.97	0.41	0.090	0.036
531.4-542.3	40	0.00031	0.17	0.37	0.59	0.38	0.125	0.055
542.3-554.2	41	0.00005	0.092	0.24	0.16	0.32	0.17	0.086
554.2-567.1	42	Less	0.043	0.11	0.032	0.24	0.23	0.14
567.1-581.1	43	0.019	0.045	0.010	0.14	0.30	0.23
581.1-596.4	44	0.0072	0.017	0.0023	0.054	0.37	0.36
596.4-613.3	45	0.0024	0.0056	0.0004	0.016	0.46	0.53
613.3-631.9	46	0.0008	0.0018	0.0001	0.0046	0.41	0.81
631.9-652.5	47	0.0003	0.0004	0.0013	0.023	0.77
652.5-675.6	48	0.0001	0.0001	0.0003	0.075	0.30
675.6-701.4	49	0.0002	0.022	0.069
701.4-730.6	50	0.007	0.010
622.4-642.0	46.5	0.88

examined under the same conditions. If, as should be the case, the e values do not differ more than 10%, the figures given in Table 3 are shown to be sufficiently correct under the applied conditions to be used for calculating the corrections to be made, these being deduced from the figures in the table and the differences between the e values for the known and the unknown solutions. With mixtures, especially, the use of non-homogeneous light is a cause of differences and useful results can hardly be expected without comparative solutions. The figures given for light in the violet, of wave length less than 430μ , are quite inaccurate, and for analytical purposes it would be scarcely advisable to make measurements beyond 440μ .

Most binary mixtures may be analyzed with fair ease and accuracy. With more complicated ones, it has been found that usually one or more

of the constituents may be determined with advantage in this way, but that the analysis is best made partly by other methods.

In Table 4 are given the results of 5 consecutive test analyses of mixtures of 2 components.

TABLE 4.
Spectrophotometric analysis of mixtures of 2 dyes.
(W. E. Mathewson, New York.)

DYES	CONCENTRATION GRAM PER 100 CC.	PERCENT OF TRUE VALUE FOUND BY FIRST CALCU- LATION (USING FIGURES OF TA- BLE 3)	PERCENT OF TRUE VALUE FOUND AFTER MAKING COMPARISON SO- LUTIONS AND DE- TERMINING COR- RECTION
Naphthol yellow S.....	0.005625	101.0	101.7
Light green S F yellowish.....	0.002375	95.6	100.4
Indigo carmine.....	0.001800	92.6	99.1
Amaranth.....	0.002200	101.0	100.7
Orange I.....	0.002000	106.0	102.0
Ponceau 3R.....	0.001440	94.4	96.7
Orange I.....	0.000800	99.0	101.0
Naphthol yellow S.....	0.007200	100.1	99.0
Amaranth.....	0.002000	97.8	98.4
Erythrosine.....	0.001200	102.3	100.0

Maximum error 3.3%; average error 1.3%.

The calculations with binary mixtures may be made more easily graphically than algebraically.

Determination of coal tar dyes other than those permitted.—The optical method permits a good quantitative estimation with extremely small amounts of color, and has been found to be well suited for the determination of such non-permitted dyes as orange II, acid yellow G, tartrazine and rose bengal in food products.

Determination of natural coloring matters used for food products.—Very few of these coloring matters can be estimated even approximately by chemical methods. Even where the coloring principle has never been isolated in purity, the method still allows an expression of the amount of color in terms of the extinction coefficients and permits standards to be set for such food coloring products as caramel, archil, etc.

A number of samples of saffron examined in the New York Food and Drug Inspection Laboratory of the U. S. Bureau of Chemistry have shown a wide variation in tinctorial power. One gram of the drug was macerated with occasional shaking for 24 hours with 200 cc. of alcohol, exactly 50% by volume. Two cc. of this solution were measured off, diluted to 100 cc. with 50% alcohol, and the values for e determined. The value of the extinction coefficient at 461–467 μ varied from 0.72–1.73.

A few analyses have indicated that curcumine in turmeric root is readily determined by extraction of the powdered drug with 95% alcohol in a Soxhlet apparatus, dilution of the extract so that the ratio to the weight of the turmeric taken is about 10,000:2 and determination of the extinction coefficients of the solution. These may be compared with the values obtained with pure crystallized curcumine under the same conditions.

REPORT ON SACCHARINE PRODUCTS.

DETECTION OF ARTIFICIAL INVERT SUGAR IN HONEY.

By F. L. SHANNON (Dairy and Food Department, Lansing, Mich.),
Associate Referee.

The work this year was a continuation of work previously inaugurated. Last year the referee selected 9 of the tests the literature showed to be most commonly used. These 9 tests were submitted to the collaborators along with samples and each test studied. From this work the referee for 1914 recommended "That four of the tests which proved the most satisfactory in the hands of all the collaborators be further studied". Accordingly instructions for these 4 tests were sent with the samples to the collaborators.

PREPARATION OF SAMPLES.

Seven samples were prepared as follows:

Sample A.—Pure honey plus 3% commercial invert sugar¹.

Sample B.—Pure honey plus 5% commercial invert sugar.

Sample C.—Pure honey plus 10% commercial invert sugar.

Sample D.—Pure honey.

Sample E.—Pure honey plus 3% Neumoline².

Sample F.—Pure honey plus 5% Neumoline.

Sample G.—Pure honey plus 10% Neumoline.

INSTRUCTIONS TO COLLABORATORS.

It is requested that the following tests be applied to the samples submitted:

FIEHE'S ORIGINAL TEST³.

Reagent.—Dissolve 1 gram of resorcinol in 100 cc. of HCl (1.19). Redistilled ether.

Manipulation.—Rub 1 gram of honey in a mortar with ether. Filter off the ether and evaporate at room temperature. Moisten the residue with a drop of the reagent.

Results.—In the presence of artificial invert sugar an orange-red color is developed, changing to cherry-red and then to brown-red. Pure honeys sometimes give a pink coloration. Note color after standing 10 minutes and again after standing 24 hours.

¹ Obtained on the market and sold as such.

² A commercial product put on the market as a non-crystallizable sirup and also a substitute for honey.

³ Analyst, 1908, 33: 397.

HARTMANN'S MODIFICATION OF FIEHE'S TEST¹.

Reagent.—Same as original Fiehe Test.

Manipulation.—Add 2 drops of the reagent directly to 1 gram of the honey in a porcelain dish.

Results.—If artificial invert sugar is present, a cherry-red color appears as in the original test. Natural honeys give the reaction after standing about 45 minutes.

BRYAN'S MODIFICATION OF FIEHE'S TEST².

Reagent.—Same as original Fiehe Test.

Manipulation.—Place 10 cc. of a 50% honey solution in a test tube and run in 5 cc. of ether on top. Shake contents gently and allow to stand for some time until the ether layer is perfectly clear; transfer 2 cc. of this clear ether solution to a small test tube and add a large sized drop of the resorcin solution. Shake and note immediately the color.

Results.—In the presence of artificial invert sugar the drop of added acid in the bottom assumes immediately an orange-red color, turning to a dark red.

FEDER'S ANILIN CHLORID TEST³.

Reagent (freshly prepared).—To 100 cc. of C. P. anilin add 30 cc. of 25% hydrochloric acid.

Manipulation.—Mix directly 5 grams of the honey in a porcelain dish with 2.5 cc. of the anilin reagent.

Results.—Bright red color indicates the presence of artificial invert sugar. The intensity of the color is proportional to the amount present.

Give your opinion, based on the foregoing tests, as to which samples are adulterated, if any, and which are pure, if any. Report results on each test on each sample as positive, negative or doubtful.

TABLE 1.
Results of collaborators on Sample A.
(Pure honey plus 3% commercial invert sugar.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICA- TION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood, Bureau of Chemistry, Wash- ington, D. C.....	Very strong trace	Very strong trace	Trace	Verydark cherry- red	Trace	Very slight trace
J. O. Clark, Depart- ment of Agriculture, Atlanta, Ga.....	Positive	Positive	Positive	Positive	Positive	Positive
W. L. Scovill, Dairy and Food Department, Lansing, Mich.....	Positive	Positive	Positive	Positive	Positive	Positive
N. A. Childs, Dairy and Food Department, Lansing, Mich.....	Positive	Positive	Positive	Positive	Positive	Positive
F. L. Shannon, Dairy and Food Depart- ment, Lansing, Mich.	Positive	Positive	Positive	Positive	Positive	Positive
C. G. Sutton, Dairy and Food Department, St. Paul, Minn.....	Positive	Positive	Positive	Positive	Positive	Positive

¹ Z. Nahr. Genussm, 1911, 21: 374.

² U. S. Bur. Chem. Bull. 154, p. 15.

³ Analyst, 1911, 36: 586.

TABLE 2.

Results of collaborators on Sample B.
(Pure honey plus 5% commercial invert sugar.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICA- TION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood.....	Very strong trace	Very strong trace	Strong trace	Very dark cherry- red	Strong trace	Trace
J. O. Clarke.....	Positive	Positive	Positive	Positive	Positive	Positive
W. L. Scovill.....	Positive	Positive	Positive	Positive	Positive	Positive
N. A. Childs.....	Positive	Positive	Positive	Positive	Positive	Positive
F. L. Shannon.....	Positive	Positive	Positive	Positive	Positive	Positive
C. G. Sutton.....	Positive	Positive	Positive	Positive	Positive	Positive

TABLE 3.

Results of collaborators on Sample C.
(Pure honey plus 10% commercial invert sugar.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICA- TION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood.....	Very strong trace	Very strong trace	Very strong trace	Very dark red	Very strong trace	Very strong trace
J. O. Clarke.....	Positive	Positive	Positive	Positive	Positive	Positive
W. L. Scovill.....	Positive	Positive	Positive	Positive	Positive	Positive
N. A. Childs.....	Positive	Positive	Positive	Positive	Positive	Positive
F. L. Shannon.....	Positive	Positive	Positive	Positive	Positive	Positive
C. G. Sutton.....	Positive	Positive	Positive	Positive	Positive	Positive

TABLE 4.

Results of collaborators on Sample D.
(Pure honey.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICA- TION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood....	Negative	Doubtful	Negative	Very dark red	Negative	Negative
J. O. Clarke.....	Negative	Negative	Slight pink Doubtful	Positive	Negative	Negative
W. L. Scovill.....	Negative	Positive	Negative	Positive	Negative	Negative
N. A. Childs.....	Negative	Negative	Doubtful	Positive	Negative	Negative
F. L. Shannon.....	Negative	Negative	Negative	Positive	Negative	Negative
C. G. Sutton.....	Negative	Negative	Positive Faint pink	Positive	Negative	Negative

TABLE 5.
Results of collaborators on Sample E.
(Pure honey plus 3% Neumoline.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICATION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood....	Negative	Doubtful	Very slight trace	Very dark color	Very slight trace. Several minutes to develop color	Negative
J. O. Clarke.....	Positive	Positive	Positive	Positive	Positive	Positive
W. L. Scovill.....	Doubtful	Positive	Doubtful	Positive	Positive	Positive
N. A. Childs.....	Doubtful	Negative	Doubtful	Positive	Doubtful	Doubtful
F. L. Shannon.....	Negative	Positive	Negative	Positive	Positive	Doubtful
C. G. Sutton.....	Positive	Positive	Positive	Positive	Positive	Positive
	Very faint pink					Light pink

TABLE 6.
Results of collaborators on Sample F.
(Pure honey plus 5% Neumoline.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICATION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood....	Doubtful	Trace	Very slight trace	Very dark red	Very slight trace	Doubtful
J. O. Clarke.....	Positive	Positive	Positive	Positive	Positive	Positive
W. L. Scovill.....	Positive	Positive	Positive	Positive	Positive	Positive
N. A. Childs.....	Positive	Positive	Positive	Positive	Positive	Positive
F. L. Shannon.....	Positive	Positive	Positive	Positive	Positive	Positive
C. G. Sutton.....	Positive	Positive	Positive	Positive	Positive	Positive
	Faint pink					Pink

TABLE 7.
Results of collaborators on Sample G.
(Pure honey plus 10% Neumoline.)

ANALYST	FIEHE'S ORIGINAL TEST		HARTMANN'S MODIFICATION OF FIEHE'S TEST		BRYAN'S MODIFICATION OF FIEHE'S TEST	FEDER'S ANILIN CHLORID TEST
	10 minutes	24 hours	10 minutes	24 hours		
S. F. Sherwood.....	Trace	Strong trace	Trace	Very dark red	Trace	Trace
J. O. Clarke.....	Positive	Positive	Positive	Positive	Positive	Positive
W. L. Scovill.....	Positive	Positive	Positive	Positive	Positive	Positive
N. A. Childs.....	Positive	Positive	Positive	Positive	Positive	Positive
F. L. Shannon.....	Positive	Positive	Positive	Positive	Positive	Positive
C. G. Sutton.....	Positive	Positive	Positive	Positive	Positive	Positive

TABLE 8.
Conclusion as to purity of samples.

ANALYST	SAMPLE A	SAMPLE B	SAMPLE C	SAMPLE D	SAMPLE E	SAMPLE F	SAMPLE G
F. Sherwood ...	Adulterated	Adulterated	Adulterated	Pure	Doubtful	Doubtful	Adulterated
O. Clarke	Adulterated	Adulterated	Adulterated	Pure	Doubtful	Adulterated	Adulterated
L. Scovill. . .	Adulterated	Adulterated	Adulterated	Pure	Doubtful	Adulterated	Adulterated
A. Childs. . .	Adulterated	Adulterated	Adulterated	Pure	Doubtful	Adulterated	Adulterated
L. Shannon. . . .	Adulterated	Adulterated	Adulterated	Pure	Pure	Adulterated	Adulterated
G. Sutton. . . .	Adulterated	Adulterated	Adulterated	Pure	Adulterated	Adulterated	Adulterated

COMMENTS OF COLLABORATORS.

S. F. Sherwood: The color developed in Feder's test is very uncertain. Hartmann's modification is liable to give the color reaction in pure honeys and hence be misleading. Bryan's modification is preferable to Fiehe's original test, as a more thorough extraction of the honey is obtained and less time required. The color should be developed at once. Bryan's modification is preferable to any of the others. It has been used by the writer frequently and with satisfactory results.

J. O. Clarke: All reactions on Sample No. 5 were somewhat faint, but sufficient to be classed as "positive".

W. L. Scovill: In Bryan's modification of Fiehe's test it is preferable to add 2 drops of the reagent instead of 1. The 24 hour test with the original Fiehe reaction and Hartmann's modification is not characteristic, as a pure honey will give a positive reaction.

The original Fiehe test and Hartmann's modification give slight positive reactions with pure honey.

F. L. Shannon: There is no advantage in either Fiehe's original or in Hartmann's modification in allowing them to stand 24 hours and then drawing conclusions. The color is well developed in 2 minutes and the best conclusion can be drawn at the end of that time.

DISCUSSION.

The results of the collaborators on these 4 tests indicate that it is not possible in all cases to detect artificial invert sugar in honey when the adulteration is less than 5%. It is also evident that the possibility of detecting artificial invert sugar in honey depends upon the nature of the invert sugar present. None of the collaborators hesitated to pronounce Samples A, B and C adulterated although Sample A contained but 3% adulterant, while in the case of Sample E, which contained but 3% adulterant, all were doubtful as to its purity, although not quite positive that it was adulterated. Sample A contained 3% of one kind of invert sugar and Sample E 3% of another.

All of these reactions depend upon furfural or its derivatives and the intensity of the color developed depends upon the amount of furfural present. This may be large or small, depending upon the manner in which the artificial invert sugar has been made. No accurate conclusions

can be drawn, therefore, from any of the tests as to the amount of the adulterant present.

Nothing is gained by allowing any of the tests to stand for any length of time before drawing conclusions. The color in all instances is developed at once when the adulteration is perceptible.

RECOMMENDATIONS.

It is recommended—

(1) That Bryan's modification of Fiehe's test, as given on page 150, be adopted as provisional.

(2) That Fiehe's test as given on page 149, adopted as provisional, except that the sentence "Note color after standing 10 minutes and again after standing 24 hours", be stricken out.

(3) That Feder's anilin chlorid test, as given on page 150, be adopted as provisional, except that the sentence "The intensity of the color is proportional to the amount present", be stricken out.

The Association adjourned at 11.55 to reassemble at 1 p.m.

MONDAY—AFTERNOON SESSION.

REPORT ON FRUIT PRODUCTS.

By P. B. DUNBAR, *Associate Referee*, AND H. A. LEPPER (Bureau of Chemistry, Washington, D. C.).

The work on fruit products during the past year has been directed principally to a further study of methods for the determination of malic and citric acids. The associate referee for 1914, Mr. H. C. Gore, recommended a continued study of his proposed methods for the determination of these acids, and this recommendation has been followed in the case of the malic-acid method. The present referee has been unable to secure any details of the proposed method for citric acid and consequently no work on this method has been undertaken. A study has been made, however, of the modification of Stahre's method¹ for the determination of citric acid, recently published by Kunz.² The "Methods for the analyses of fruits and fruit products,"³ have also been reviewed, and a critical study of the modified Schmidt-Hiepe method for tartaric, citric, and malic acids⁴ has been made. It has not seemed advisable to send out collaborative samples during the present year, as the details of the methods under study have not been completely worked out.

CITRIC ACID.

A somewhat detailed abstract of the method proposed by Kunz for the determination of citric acid has already been published.⁵ The quantitative method described is applicable only to the determination of citric acid in wines presumably dry. Methods for its qualitative detection in milk, marmalade, and fruit sirup are also described.

The quantitative method as applied to wine consists in treating the concentrated sample with sulphuric acid and bromin water, followed by a solution of potassium bromid. The solution is then oxidized with potassium permanganate, when insoluble pentabromacetone is precipitated quantitatively. While the method appears promising for the determination of citric acid in dry wines and other materials containing small amounts of permanganate reducing substances, it cannot be applied directly

¹ Nordisk Tidsskrift (1895), **2**: 141; Z. anal. Chem. (1897), **36**: 195.

² Arch. Chem. Mikros (1914), **7**: 285-299.

³ U. S. Bur. Chem. Bul. 107 (rev.), pp. 77-82.

⁴ Ibid., pp. 80-81.

⁵ Chem. Abstracts (1915), **9**: 687.

to fruit juices on account of the presence of large amounts of sugar. It, therefore, becomes necessary in order to apply the method to adopt some procedure for the precipitation of citric acid and its removal from most of the other permanganate reducing bodies. Precipitation as barium citrate from alcoholic solution has been advocated by many investigators, and seems to offer the most practical method of procedure.

The method as finally adopted for study differs little, except for the fact that the citric acid is previously precipitated as barium citrate, from that recommended by Kunz for the determination of citric acid in wine. The procedure used for precipitating the acid is essentially that recommended for malic acid by the previous referee, Mr. Gore, but a solution of barium hydroxid is used instead of the solid substance, and the pectins are not removed before precipitation of barium citrate.

The method is as follows:

Determine the total acidity of the sample. It is convenient to use 1% azolitmin (made by dissolving 1 gram azolitmin in 80 cc. hot water, diluting to 100 cc. with 85% alcohol, and filtering) as an indicator in conjunction with a porcelain spot plate.

Weigh out 25 grams of the sample and transfer to a 600 cc. beaker with 95% alcohol; add an amount of approximately 0.25 N/1 barium hydroxid solution just sufficient to neutralize the acidity of the sample and 5 drops of 50% barium acetate solution, to insure an excess of barium. Dilute to a total volume of 375 cc. with 95% alcohol, stir well and allow to settle. After the precipitates of pectins and barium salts have settled well, decant the supernatant liquid into another beaker. Filter the decanted liquid through a folded filter to remove any small portions of precipitate which it may contain and wash the beaker with 95% alcohol. Filter the remainder of the pectin and barium salt precipitate on the same paper and wash the beaker and paper once with 95% alcohol. Transfer the precipitate quantitatively from the paper to the original beaker with hot water, boil until no more alcohol can be detected by odor, add enough sulphuric acid (1:5) to precipitate all of the barium originally added and to allow 2 cc. of the acid in excess, evaporate by careful boiling to between 60 and 70 cc., cool and add 5 cc. of freshly prepared saturated bromin water; the solution should show an excess of bromin. Transfer quantitatively to a 100 cc. graduated flask and dilute to the mark at room temperature; allow the precipitate of barium sulphate and that caused by the addition of bromin to settle and filter. The precipitate may be separated by centrifuging, and the supernatant liquid decanted if necessary. Now pipette an aliquot of the filtrate, containing not more than 400 mg. of citric acid, calculated from the total acidity of the sample, into a 300 cc. Erlenmeyer flask. The amount of citric acid in the aliquot should, if possible, exceed 50 mg. Add 10 cc. sulphuric acid (1:1) and 5 cc. of potassium bromid solution (15 grams KBr in 40 cc. water). After shaking, warm the flask in a water bath to 48-50°C. and allow it to remain in the bath for five minutes. After removing from the bath, add 25 cc. of 5% KMnO_4 from a burette in rapid drops with frequent interruptions and constant vigorous shaking, care being taken that the temperature during oxidation does not exceed 55°. Set the flask aside until the hydrated peroxid of manganese begins to settle. (The supernatant liquid should be dark brown in color, showing an excess of KMnO_4 . If an excess is not indicated, more KMnO_4 must be added.) Then shake and again set

aside to settle, and repeat this operation until the precipitate takes on a yellow color and most of it has dissolved. Finally, while the solution is still warm, remove the last undissolved portion of the hydrated peroxid of manganese precipitate and also the excess of bromin by adding drop by drop a clear concentrated solution of ferrous sulphate acidified with H_2SO_4 . Allow the solution to cool, with occasional shaking. If the operations have been properly carried out, a heavy white precipitate of pentabromacetone is obtained. This becomes crystalline on occasional shaking and in this condition is entirely insoluble in water. After the precipitate has become crystalline, preferably after standing overnight, collect it by means of gentle suction on a porcelain Gooch crucible provided with a thin pad of asbestos, previously dried over H_2SO_4 in a vacuum desiccator, and wash with distilled water slightly acidified with H_2SO_4 and, finally, twice with distilled water (if the precipitate has a tendency to pass through the filter, 1% H_2SO_4 should be used for washing). Dry the precipitate to constant weight in a vacuum desiccator over H_2SO_4 protected from strong light. The weight of pentabromacetone multiplied by the factor 0.464 equals citric acid plus 1 molecule of water of crystallization. It sometimes happens that the pentabromacetone is obtained in the form of oily droplets. These also become crystalline on standing, or on cooling, but are usually discolored by negligible traces of manganese or iron. There appears to be no difference in the results obtained whether the precipitate is originally in the form of crystals or oily droplets, which later become crystalline.

The accuracy of this method has been tested by determinations on water solutions of citric acid containing varying amounts of sucrose, and on diluted apple sirup and raspberry sirup containing added known amounts of citric acid, and in some cases added malic and tartaric acids. The results obtained are given in Table 1.

According to Partheil and Hübner,¹ 100 grams of alcohol of specific gravity 0.8092 dissolves 0.00578 gram of barium citrate $(\text{C}_6\text{H}_5\text{O}_7)_2 \text{Ba}_3 + 7\text{H}_2\text{O}$ at 25°C.

In determining small percentages of citric acid, the solubility of barium citrate in the volume of alcohol used will give rise to a relatively large error in the percentage recovered. Considerable time has been given to attempts to obviate this difficulty by determining the citric acid without previous precipitation, but these have been unsuccessful. It is hoped that the difficulty may be met by the use of a solubility correction. It is hardly necessary to point out, however, that a relatively small variation in the actual weight of citric acid found will give a marked variation in the percentage recovered and that comparisons should be made between the actual weights, in grams per 100 cc., of citric acid present and found. From the results on sirups containing added malic and tartaric acids, it appears that these acids do not interfere with the determination. This has been further shown by applying the method to 50 cc. portions of 1% solutions of malic, tartaric, and succinic acids. In no case was a precipitate obtained.

¹ Arch. der Pharm., **241**: 412-435; Chem. Centr. (1903), **74**: (2), 1026.

This method seems of sufficient promise to warrant further study, and it is recommended that the referee for the following year be instructed to take up this work and, if possible, to send out collaborative samples.

TABLE 1.

Determination of citric acid by Kunz modification of Stahre's method.

CITRIC ACID IN AQUEOUS SOLUTION.

NO PRELIMINARY PRECIPITATION AS BARIUM CITRATE.

CITRIC ACID PRESENT		CITRIC ACID RECOVERED		
In 50 cc. sample	Per 100 cc.	gram	gram per 100 cc.	per cent
<i>gram</i>	<i>gram</i>	<i>gram</i>		
0.0184	0.04	0.013	0.03	75.0
0.0908	0.18	0.086	0.17	95.2
0.0923	0.18	0.089	0.18	100.0
0.1845	0.37	0.181	0.36	97.3
0.1853	0.37	0.180	0.36	97.3
0.2270	0.45	0.223	0.45	98.3
0.2779	0.56	0.275	0.55	98.2
0.3706	0.74	0.370	0.74	100.0
0.4632	0.93	0.461	0.92	98.9

PRECIPITATION AS BARIUM CITRATE INCLUDED.

CITRIC ACID PRESENT		APPROXIMATE AMOUNT SUCROSE ADDED PER 100 CC.	CITRIC ACID RECOVERED		
In 25 cc. sample	Per 100 cc.		grams	grams per 100 cc.	per cent
<i>gram</i>	<i>grams</i>	<i>grams</i>	<i>grams</i>		
0.4610	1.84	0	0.4503	1.80	97.68
0.4610	1.84	0.5	0.4491	1.80	97.42
0.4610	1.84	1.0	0.4493	1.80	97.47
0.4610	1.84	2.0	0.4487	1.79	97.33
0.4610	1.84	5.0	0.4465	1.79	96.85
0.4610	1.84	10.0	0.4448	1.78	96.50
0.4610	1.84	20.0	0.4455	1.78	96.64

ADDED CITRIC ACID IN DILUTED APPLE SIRUP.

CITRIC ACID PRESENT		APPROXIMATE MALIC ACID PRESENT	CITRIC ACID RECOVERED		
In 25 cc. sample	Per 100 cc.		gram	grams per 100 cc.	per cent
<i>gram</i>	<i>grams</i>	<i>per cent</i>	<i>gram</i>		
0	0	0.51	0	0	0
0.0369	0.15	0.03	0.029	0.12	80.0
0.0484	0.19	0.53	0.038	0.15	78.9
0.0554	0.22	0.03	0.049	0.20	90.9
0.0727	0.29	0.53	0.059	0.24	82.8
0.0923	0.37	0.03	0.081	0.32	86.5
¹ 0.0969	0.39	0.53	0.082	0.33	84.6
² 0.0969	0.39	0.53	0.082	0.33	84.6
0.1211	0.48	0.53	0.109	0.44	91.7
0.1845	0.74	0.03	0.178	0.71	96.0
0.2707	1.08	0.51	0.243	0.97	89.8
0.4060	1.62	0.51	0.406	1.62	100.0

¹ 2.5 grams sucrose added to 25 cc. sample.

² 5 grams sucrose added to 25 cc. sample.

TABLE 1.—*Continued.*

ADDED CITRIC ACID IN RASPBERRY SIRUP.

CITRIC ACID PRESENT		APPROXIMATE MALIC ACID ADDED	APPROXIMATE TARTARIC ACID ADDED	CITRIC ACID RECOVERED ²		
In 25 cc. sample	Per 100 cc.					
gram	gram	per cent	per cent	gram	gram per 100 cc.	per cent
⁴ 0.1373	0.55	0	0	0.155	0.62	112.7
⁴ 0.1373	0.55	0	0	0.154	0.62	112.7
⁵ 0.0992	0.40	0	0	0.097	0.39	97.5
⁵ 0.0992	0.40	0.39	0	0.094	0.38	95.0
⁵ 0.0992	0.40	0	0.40	0.097	0.39	97.5
⁵ 0.0992	0.40	0.19	0.20	0.093	0.37	92.5

² The last four determinations are corrected for the amount of citric acid found in the original sirup by precipitation.

⁴ Calculated from titration of original sirup.

⁵ Grams of citric acid added in excess of that originally present.

MALIC ACID.

The previous referee, Mr. Gore, has given considerable attention to a study of methods for the determination of malic acid during the past three years. In his report for the year 1912¹ he gives results obtained by eight collaborators, using the polariscopic method proposed by Dunbar and Bacon.² This method depends on the change in the rotation of malic-acid solutions when treated with uranyl acetate. The sample of cider analyzed contained 0.50% of total acidity calculated as malic. The results reported varied from 0.468 to 0.531%, and averaged 0.49%. During the following years, Mr. Gore paid considerable attention to the effects, on the rotations of uranium-malic acid solutions, of various factors. He further investigated the possibility of developing a method based on the change in rotation of malic-acid solutions when treated with ammonium heptomolybdate. As a result of this work he suggested, in his report for 1914, two polariscopic methods for the determination of malic acid, both of which call for a preliminary precipitation of the acid as barium malate.

The uranyl-acetate method suggested differs from the Dunbar-Bacon method, first sent out by Mr. Gore for collaborative work, in that precipitation of malic acid is suggested, a solution of uranyl acetate is used in place of the solid salt, and the amounts of malic acid are determined by referring the polariscopic reading to a table, rather than by the use of a constant factor. The method of Dunbar and Bacon is not applicable in the presence of tartaric acid nor in the case of highly colored liquids which cannot be readily polarized. A method whereby the malic acid may be precipitated and separated from interfering substances is therefore highly desirable. The use of a solution of uranyl acetate in

¹ U. S. Bur. Chem. Bul. 162, p. 63.

² U. S. Bur. Chem. Circ. 76.

place of the solid salt is also advantageous in that it obviates the indefinite error of dilution, due to the solution of unknown amounts of uranyl acetate. In the case of products containing small amounts of malic acid, however, the dilution of the solution with an equal volume of uranyl-acetate solution produces a corresponding decrease in the polarization and thereby increases the relative percentage error due to inaccuracies in reading the polariscope.

The 1914 report on fruit products contained results by only two collaborators using the proposed methods for malic acid suggested by Mr. Gore. These, unfortunately, do not agree very satisfactorily, and it seemed advisable, therefore, to carry out further experimental work with solutions containing known amounts of malic acid. Numerous difficulties were encountered in precipitating the acid; and, although considerable time was given to the work, the results so far obtained are not such as to justify a final recommendation regarding the method. It was found

TABLE 2.

Determination of malic acid by the uranyl-acetate and molybdate methods in aqueous solutions containing tartaric acid.

MALIC ACID PRESENT		TARTARIC ACID PRESENT IN 25 CC. SAMPLE	MALIC ACID RECOVERED					
In 25 cc. sample	Per 100 cc.		By uranyl-acetate method			By molybdate method		
gram	gram	gram	gram	gram per 100 cc.	per cent	gram	gram per 100 cc.	per cent
0.0938	0.38	0.0997	0.092	0.37	97.4	0.102	0.41	107.9
0.0938	0.38	0.1496	0.088	0.35	92.1	0.100	0.40	105.2
0.1407	0.56	0.0997	0.130	0.52	92.9	0.157	0.63	112.5
0.1876	0.75	0.0499	0.165	0.66	98.0	0.223	0.89	118.6

to be difficult, if not impossible, to dissolve the solid barium hydroxid which is added to precipitate the malic acid, and a strong standardized barium-hydroxid solution was therefore substituted. The error due to the solubility of barium malate does not seem to be serious. It was found, however, that in the case of solutions containing malic and tartaric acids, a sufficient amount of barium tartrate was dissolved to reduce the negative polarization somewhat in the uranyl-acetate method and increase it in the molybdate method. The irregularities of results due apparently to this cause are shown in Table 2. Comparison of the actual weights of malic acid present with those found, however, show that the error in grams per 100 cc. is not large. In the case of determinations of malic acid in fruit juices, much of the color of the original sample is carried down with the barium precipitate and again dissolved when this precipitate is taken up in water.

It thus appears that the method suggested for the precipitation of barium malate does not entirely obviate the difficulties in the original method when interfering color is present, but is of promise in determining

malic acid in the presence of tartaric when the color of the solution is not excessive. In the absence of tartaric acid and color the precipitation method introduces a number of time-consuming operations without materially affecting the results.

The work which was undertaken on citric acid prevented an extended study of the malic-acid method. From the results of the collaborative work reported by Mr. Gore in 1912 and those obtained by the present referee and reported in U. S. Bureau of Chemistry Circular 76, it would appear that the malic-acid method as originally described in that circular can undoubtedly be applied with accuracy to solutions which can be polarized readily and which contain no optically active acid other than malic. Where the amount of malic acid present is sufficient, the use of a solution of uranyl acetate is apparently to be preferred to the solid salt. When it is possible to make readings without difficulty, rapid determination may be made with reasonable accuracy without previous neutralization or clarification by polarizing before and after treatment with uranyl acetate. In the presence of tartaric acid, preliminary precipitation as the barium salt is advantageous.

REVIEW OF METHODS FOR THE ANALYSES OF FRUITS AND FRUIT PRODUCTS.

A number of recommendations for changes in the methods for the analyses of fruits and fruit products have been forwarded to the chairman of the committee on the revision of methods. These recommendations will doubtless be included in the report of the committee.

TABLE 3.

*Experiments with the Schmidt-Hiepe method for tartaric, citric, and malic acids.*¹

NO.	TARTARIC ACID			CITRIC ACID			MALIC ACID		
	Added		Recovered	Added		Recovered	Added		Recovered
	gram	gram	per cent	gram	gram	per cent	gram	gram	per cent
1	0.4896	0.4834	98.73	0.4601	0.3955	85.96	0.4856	0.0304
2	0.4896	0.4884	99.76	0.0019	0.4856	0.4000	82.37
3	0.0195	0.4462	0.2957	66.27	0.4856	0.5832	120.1
4	0.3917	0.3856	98.44	0.3570	0.1760	49.30	0.3885	0.4536	116.8
5	0.1469	0.0706	48.06	0.1339	0.0803	59.97	0.0586
6	0.1469	0.0702	47.78	0.0025	0.1457	0.1286	88.29
7	0.0075	0.1339	0.0141	10.53	0.1457	0.1631	112.0
8	0.0979	0.0957	97.75	0.0920	0.0118	12.83	0.1003	0.0141	13.06
9	0.0490	0.0342	69.80	0.0460	0.0109	23.69	lost
10	0.0490	0	0	0.0102	0.0502	0	0
11	0.0045	0.0460	0.0048	10.43	0.0502	lost
12	0.0490	0.0305	62.24	0.0460	0	0	0.0502	lost

¹ U. S. Bur. Chem. Bul. 107 (rev.), p. 80.

In 3, 7, and 11, where a recovery is recorded for tartaric acid when none was present, the titer was probably caused by calcium acetate retained, due to the limitation to the washing.

The amount of tartaric acid recovered in 3, 7, and 11 is the amount actually titrated, the 0.0286 gram dissolved in the 100 cc. filtrate not being added, as no tartaric was present.

The amount of malic recovered was always corrected for the tartaric (0.0286) present when it was known to be there, due to original addition.

Table 3 contains the results of a study of the modified Schmidt-Hiepe method for the determination of tartaric, citric, and malic acids. From these results it appears that the method is unreliable, and should be dropped as one of the official methods.

RECOMMENDATIONS.

It is recommended—

- (1) That the Kunz modification of Stahre's method for the determination of citric acid be further studied.
- (2) That the uranyl-acetate and ammonium heptomolybdate methods for the determination of malic acid be further studied.
- (3) That the method for "Tartaric, citric, and malic acids (Schmidt-Hiepe method modified) official"¹ be dropped.

REPORT ON WINE.

By B. G. HARTMANN (Bureau of Chemistry Food and Drug Inspection Laboratory, Chicago, Ill.), *Associate Referee*.²

The provisional method for the determination of total tartaric acid content in wines,³ is inaccurate and unreliable. In a paper read before the association in 1912, Hartmann and Eoff called attention to this fact, presenting proof in support of their criticisms and offering for trial a modification of this method. This paper gave the initiative for a series of coöperative investigations extending over a period of three years, conducted for the purpose of determining the relative merits of the two methods as to their accuracy and adaptability to varying conditions. In the course of these investigations the methods were tried on wines, grape juices, and synthetic solutions containing varying amounts of tartaric and phosphoric acids.

The results of this work proved conclusively that the provisional method is far from satisfactory, whereas the proposed method was found to be accurate and reliable.

During the present year five samples of grape juice, with and without the addition of tartaric acid and phosphoric acid, were sent to collaborators with the following instructions:

INSTRUCTIONS TO COLLABORATORS.

The samples were filtered and no precipitation is expected. However, as a precautionary measure, shake the samples thoroughly just before measuring the various portions.

¹ U. S. Bur. Chem. Bul. 107 (rev.), pp. 80-81.

² Presented by M. J. Ingle.

³ U. S. Bur. Chem. Bul. 107 (rev.), p. 86.

ACIDITY OF THE JUICE.

Measure 10 cc. of the sample into a 250 cc. beaker, add 20 cc. distilled water and titrate with tenth-normal sodium hydroxid until two drops of the solution when mixed on a porcelain tile with several drops of a neutral azolitmin solution give no red tint. Prepare the azolitmin solution by dissolving 0.5 gram pure azolitmin in a liter of distilled water, neutralizing with either acid or alkali, as the case may be.

DETERMINATION OF TOTAL TARTARIC ACID.

Provisional method.—Determine as described in U. S. Bureau of Chemistry Bulletin 107 (revised), page 86, using 50 cc. of sample, and dilute with 50 cc. distilled water. Use 20 cc. of alcohol instead of 15 cc. Collect the cream of tartar crystals on a double filter paper in a Büchner funnel (see U. S. Bureau of Chemistry Bulletin 162, p. 73) and titrate with N/10 alkali, using phenolphthalein as indicator. Multiply by 2 to obtain grams per 100 cc. Do not stir longer than 1 minute and do not place in an ice box for the 15 hours prescribed, but allow to remain at room temperature.

Proposed method (Hartmann and Eoff).—Transfer 50 cc. of the sample to a 250 cc. beaker and neutralize with sodium hydroxid. The amount of normal alkali required is calculated by multiplying the cc. N/10 sodium hydroxid required to titrate the 10 cc. of the sample (under 1) by 0.5. To the neutralized solution, add enough water to make 100 cc. and add tartaric acid. The tartaric acid used should be of the purest obtainable and should be powdered and dried for about two hours at the temperature of boiling water. Determine the purity of the tartaric acid by titrating 0.30 gram with N/10 alkali, using phenolphthalein as indicator. Keep the tartaric acid in a desiccator. The amount of tartaric acid to be added is calculated by multiplying the number of cc. N/1 NaOH required to neutralize the 50 cc. sample by 0.075. Weigh the amount accurately.

After the tartaric acid has gone into solution, add 15 grams powdered potassium chlorid and stir until the salt has dissolved. Now add 2 cc. glacial acetic acid and 20 cc. of 95% alcohol. Stir until precipitation has started and place in an ice box for at least 15 hours. Collect the cream of tartar crystals on a double filter paper in a Büchner funnel and titrate with N/10 alkali, using phenolphthalein as indicator. Calculate the total tartaric acid and subtract the tartaric added, making proper correction for purity of the latter. This will give the tartaric acid in 50 cc. Multiply by 2 to obtain grams per 100 cc.

The basis of the five samples was a pure Concord grape juice. In order to prevent precipitation of potassium acid tartrate, the filtered juice was diluted with 25% of distilled water. The samples were made up as follows:

No. 1.—The diluted juice.

No. 2.—The diluted juice with the addition of 0.569 gram of tartaric acid per 100 cc.

No. 3.—The diluted juice with the addition of 0.06 gram of phosphoric acid per 100 cc.

No. 4.—The diluted juice with the addition of 0.30 gram of phosphoric acid per 100 cc.

No. 5.—The diluted juice with the addition of 0.086 gram sodium carbonate to neutralize the free tartaric acid.

¹ U. S. Bur. Chem. Bul. 162, p. 72.

All samples were sterilized.

The following results were obtained by the collaborators and the associate referee.

Total tartaric acid (grams per 100 cc.).

ANALYST	SAMPLE									
	1		2		3		4		5	
	No. 1	No. 2	No. 1	No. 2	No. 1	No. 2	No. 1	No. 2	No. 1	No. 2
J. R. Eoff, Jr.	0.528	0.614	0.912	1.210	0.504	0.614	0.414	0.628	0.555	0.624
M. J. Ingle.	0.534	0.608	0.924	1.208	0.507	0.620	0.408	0.620	0.558	0.620
F. D. Merrill	0.558	0.627	0.969	1.205	0.542	0.636	0.433	0.638	0.590	0.637
E. H. Berry	0.578	0.628	1.024	1.213	0.579	0.586	0.382	0.586	0.562	0.602
T. G. Gleason	0.536	0.626	0.960	1.200	0.544	0.613	0.444	0.627	0.582	0.615
Edw. F. Higgins	0.564	0.629	0.964	1.205	0.550	0.625	0.450	0.633	0.585	0.615
B. G. Hartmann	0.538	0.618	0.952	1.200	0.526	0.636	0.435	0.644	0.584	0.632
Average	0.554	0.622	0.966	1.203	0.538	0.618	0.427	0.624	0.576	0.619

No. 1, provisional method; No. 2, proposed method.

No comments on the two methods were offered by the collaborators. M. J. Ingle submitted the results which he obtained on the five samples by the Rochelle-salt method, as follows:

The following are the results obtained by using Rochelle salt on the five samples submitted. I did not add any alkali to these determinations, merely a quantity of Rochelle salt calculated by multiplying the number of cc. of N/1 alkali required for complete neutralization by the factor 0.141. The portions of the juice used were measured on July 19 and had to be thoroughly shaken to mix in the precipitated tartar. I then added 50 cc. of water, 2 cc. acetic, 15 grams KCl, and 20 cc. of 95% alcohol, and placed in the ice box over night. I ran these determinations in duplicate and obtained good checks. I found out that one cannot be certain of the state of hydration of these salt crystals, particularly if grinding is resorted to. It will therefore be necessary to run a blank in duplicate on 1-gram samples of the salt as used. The tartaric acid recovered will represent the factor for obtaining the percentage of acid added from the weight of Rochelle salt used.

The procedure in the case of the blank consisted in merely diluting to 100 cc. and adding the acetic and other constituents, omitting, as above, the potassium acetate.

Total tartaric acid (grams per 100 cc.).

	Rochelle-salt method	Proposed method
No. 1	0.624	0.627
No. 2	1.170	1.205
No. 3	0.624	0.636
No. 4	0.620	0.638
No. 5	0.640	0.637

DISCUSSION OF RESULTS.

The general average of the results obtained by the collaborators for the two methods are:

Total tartaric acid (grams per 100 cc.).

	Provisional method	Proposed method
No. 1	0.554	0.622
No. 2	0.966	1.203
No. 3	0.538	0.618
No. 4	0.427	0.624
No. 5	0.576	0.619

With the exception of sample No. 2, the total tartaric-acid content of the five samples submitted is identical. Taking as a basis the general average (0.621) of the results obtained on these four samples by the proposed method, the recoveries by the provisional method are:

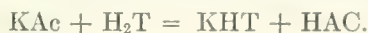
	Per cent
No. 1 (containing free tartaric acid).....	89
No. 3 (containing 0.06 gram phosphoric acid).....	87
No. 4 (containing 0.30 gram phosphoric acid).....	70
No. 5 (containing no free tartaric acid).....	93

Sample No. 2 had an addition of 0.569 gram of tartaric acid per 100 cc. Of this amount the proposed method recovered 102% and the provisional method 61%.

From these results it is evident that the provisional method is unreliable and inaccurate and that the proposed method gives very satisfactory results. Regarding the provisional method the results show—

- (1) That it fails to determine all the free tartaric acid.
- (2) That the addition of potassium acetate has a decomposing effect upon potassium-acid tartrate.
- (3) That the presence of a free mineral acid such as phosphoric acid partially prevents the formation of potassium-acid tartrate.

The first two points, the presence or absence of free tartaric acid, are the two conditions occurring in natural wines and grape juices, whereas the presence of added phosphoric acid is very often met with in artificial grape products, such as grape-juice sirups and other soda-fountain beverages. Accordingly, the provisional method is not applicable to any of the conditions which occur in wines or grape juices, unless it is assumed that a relative amount of free tartaric acid retards the decomposing action of the potassium acetate upon potassium-acid tartrate:



In conclusion it should be said that the deductions made from the results obtained in this investigation are similar to those made in the three previous reports on this subject.

In making a recommendation for the adoption of the proposed method, it seems desirable to call attention to the fact that many results have been published on authentic samples of wines and grape juices of known purity examined by the provisional method. Results obtained by the proposed method would not, therefore, be comparable with previously published figures. It does not seem, however, that a method found to be entirely unsatisfactory should be retained by the association.

RECOMMENDATION.

It is recommended—

That the provisional method be dropped by the association and the proposed method be adopted for the determination of total tartaric acid in wines, grape juice, and soda-fountain sirups.

The following is the method for wines:

Neutralize 100 cc. wine with normal sodium hydroxid. The amount of alkali necessary for neutralization is calculated from the acidity of the wine. If the volume of the solution is increased more than 10% by the addition of the alkali, evaporate the solution to approximately 100 cc. Add tartaric acid to the neutralized solution. For each cubic centimeter normal alkali used add 0.075 gram tartaric acid.¹ After the tartaric acid has dissolved, add 2 cc. glacial acetic acid and 15 grams potassium chlorid. After the potassium chlorid has dissolved, add 15 cc. of 95% alcohol, stir vigorously until the cream of tartar starts to precipitate, and let stand for at least 15 hours in an ice box. Decant the liquid from the separated acid-potassium tartrate on a Gooch prepared with a very thin film of asbestos or on filter paper in a Büchner funnel. Wash the precipitate and filter three times with a small amount of mixture of 15 grams of potassium chlorid, 20 cc. of 95% alcohol, and 100 cc. of water, using not more than 20 cc. of the wash solution in all. Transfer the asbestos or paper and precipitate to the beaker in which the precipitation took place, wash out the Gooch or Büchner funnel with hot water, add about 50 cc. of hot water, heat to boiling, and titrate the hot solution with tenth normal sodium hydroxid, using phenolphthalein as indicator. Increase the number of cubic centimeters of N/10 alkali required by 1.5 cc. on account of solubility of the precipitate. One cubic centimeter of N/10 alkali is equivalent to 0.015 gram tartaric acid. To ascertain the tartaric acid originally present in the wine, subtract the amount of tartaric acid added. This will give the grams of total tartaric acid per 100 cc. of wine.

¹ The tartaric acid used should be of the purest obtainable and should be powdered and dried for about two hours at the temperature of boiling water.

In the case of grape juice or sirups, use 50 cc. of the sample, neutralize with normal sodium hydroxid, and add distilled water to 100 cc. volume. Proceed as under wine, adding 20 cc. of alcohol instead of 15 cc. After subtracting the tartaric acid added, multiply by 2 to obtain grams total tartaric acid in 100 cc.

REPORT ON BEER.

By H. S. PAINE (Bureau of Chemistry, Washington, D. C.),
*Associate Referee.*¹

No plan for further work was submitted in the last report of the associate referee on this subject and, owing to a combination of circumstances, a plan for work which was deemed to be sufficiently profitable was devised too late for submittal to the collaborators.

Briefly, the matter may be stated as follows: Comparison of the analyses of various malt beverages, correlated with a knowledge of the conditions of the mash, leads to the conclusion that the methods employed for the determination of maltose (or "reducing sugars") and dextrin do not yield accurate results in all cases—the degree of accuracy being variable—that is, fairly acceptable in some cases and quite unsatisfactory in others. This variation is probably dependent upon the proportion of maltose and dextrin, and this in turn depends, of course, upon the mash conditions.

In the method which is based upon polarization there is a grave question of the accuracy of the assumed rotatory power of dextrin. There are various dextrans, of course, and it is a question whether the assumed value is always representative.

In the reduction method, which is more reliable, the factors which must be considered are somewhat as follows:

(1) The effect of dextrin and other constituents upon the reducing action of maltose in the reduction before hydrolysis, i.e., the question of applicability of the maltose tables to these conditions.

(2) The possibility of some reducing action by the lower dextrans.

(3) The possible effect of other sugars which might be present.

(4) The accuracy of the factors employed.

Various other considerations have a possible effect.

It is impossible in the absence of some preliminary experimental work to say just what the effect of these various factors may be and which will be of the most importance. The point to be emphasized, however, is that the variations in the composition of the mash and conditions of mashing are the controlling factors which cause a variable degree of accuracy in the above determinations. The other methods employed in beer analysis are, on the whole, satisfactory.

It is believed that the carbohydrate determinations are most in need of attention at the present time, and it is recommended that the methods for the determination of maltose and dextrin in beer be made the subject of study for the coming year.

No report was made by the associate referee on distilled liquors.

¹ Presented by M. J. Ingle.

A STUDY OF THE FUSEL OIL AND ESTERS IN
DISTILLED LIQUORS.By LOUIS KATZ (Bureau of Chemistry Food and Drug Inspection
Laboratory, New York).¹

The examination of brandies and other distilled liquors for fusel oil is ordinarily limited to an estimation of the quantity of fusel oil present. No attempt is made to gain an insight into the composition of the fusel oil. Considering the importance of the fusel oil constituent of distilled liquors as a basis of judgment to be pronounced by the chemist as to their genuineness, and further considering the comparative ease with which the recognized minimum quantity of fusel oil could be added to neutral spirits, it seemed quite obvious that a mere quantitative determination of fusel oil is a somewhat weak link in the chain of evidence the chemist gathers in the examination of liquors. At best it enables the chemist to pronounce a liquor sophisticated only in that limited number of cases where not enough of a so-called bona-fier or too much neutral spirits have been added to keep the quantity of fusel oil above the required minimum. A qualitative examination of the fusel oil of distilled liquors, supplementary to its quantitative determination, seemed highly desirable and was accordingly undertaken at the suggestion of Mr. A. F. Seeker. The property of the volatile acids of the acetic acid series, when fractionally distilled from an aqueous solution, to pass over at a rate which is definite and characteristic for each acid,² was made the basis of the study. This property of the individual acids causes any definite mixture of them to distil over at a rate which is definite and characteristic for that particular acid mixture. In the regular course of analysis of distilled liquors, the so-called fusel oil, or the sum total of the higher alcohols, is determined quantitatively by oxidizing these alcohols to their corresponding acids and titrating the acid mixture with alkali. Now this acid mixture, after having been titrated and thus having served the purpose of quantitative estimation of the fusel oil, was again acidified with sulphuric acid and submitted to fractional distillation. As was expected, the rate of distillation differed in the case of acid mixtures obtained from the fusel oils of different samples of liquors, such differences being obviously due to differences in the composition of the acid mixtures, hence also of the fusel oils in these samples. In other words, the fusel oil of a distilled liquor yields upon oxidation an acid mixture which, when fractionally distilled from an aqueous solution under definite conditions, passes over at a rate which is dependent upon the composi-

¹ Presented by A. F. Seeker.² See method of M. Duclaux of detecting and estimating organic acids. Allen's Commercial Organic Analysis, 1905, 1: 490.

tion of the fusel oil. The curve representing that rate of distillation affords, therefore, a ready means to differentiate between fusel oils of different composition without making an actual analysis of those complex mixtures.

The esters of distilled liquors may also be studied qualitatively by isolating the sum total of their combined acid radicles and submitting the acid mixture to fractional distillation.

The procedure was in all cases as follows:

To 119 cc. of the acid mixture in a 500 cc. distillation flask, 1 cc. of concentrated H_2SO_4 was added, the flask connected to a condenser, and the solution distilled at a moderate rate, the receiver being changed each time 20 cc. passed over. Five fractions of 20 cc. each were distilled over, 20 cc. remaining in the distillation flask. Each fraction was titrated with standard alkali, thus furnishing a set of five numbers, representing the rate of distillation of the mixed acids, from which a curve graphically showing this rate of distillation could be estimated. The numbers were expressed as percentages of acid in each fraction with regard to the total acid that had passed over in the entire 100 cc. distilled. Thus, a rate of distillation represented by the following five numbers: 40, 25, 15, 10, 10, would mean that the first 20 cc. fraction contained 40% of total acid that distilled over in all the five fractions taken together, the second 20 cc. fraction contained only 25%, the third, 15%, the fourth, 10%, and the fifth, 10%.

Tables 1, 2, and 3 represent the results of fractional distillation of acid mixtures resulting from the oxidation of fusel oils derived from various distilled liquors.

TABLE 1.

Fractional distillation of fusel oil—French brandies.

SAMPLE NO.	PER CENT OF ACID IN EACH FRACTION					SAMPLE NO.	PER CENT OF ACID IN EACH FRACTION				
	1	2	3	4	5		1	2	3	4	5
1	41.0	25.9	15.6	9.6	7.8	20	36.2	23.2	15.2	12.4	13.0
2	41.0	24.8	15.4	10.2	8.5	21	38.6	23.9	14.5	11.0	12.0
3	40.6	22.6	15.1	10.9	10.8	22	38.2	23.1	15.6	11.6	11.5
4	40.7	23.2	13.9	11.1	11.1	23	40.6	24.1	14.4	10.7	10.2
5	43.8	21.9	14.4	9.6	10.3	24	41.9	23.9	14.4	10.5	9.3
6	38.3	23.4	17.0	10.7	10.6	25	38.0	23.0	15.6	11.8	11.6
7	42.4	24.3	14.4	10.8	8.1	26	38.3	24.0	15.3	11.2	11.2
8	40.2	23.9	15.2	10.9	9.8	27	40.1	22.9	14.4	10.6	11.9
9	42.7	24.3	15.5	8.8	8.7	28	40.1	23.2	15.0	10.6	11.1
10	45.7	23.1	13.0	9.2	8.9	29	41.2	22.4	13.7	10.7	12.0
11	39.4	24.8	15.2	10.4	10.2	30	40.3	24.3	14.8	10.4	10.2
12	38.9	23.6	15.4	10.9	11.2	31	41.8	24.6	15.6	9.8	8.2
13	41.1	24.3	14.8	10.0	9.8	32	41.9	25.0	14.5	10.2	8.4
14	41.5	22.9	14.6	10.6	10.4	33	38.5	24.8	13.7	11.2	11.8
15	45.5	23.5	13.4	9.0	8.6	34	36.0	23.6	15.6	12.4	12.4
16	46.6	23.3	13.5	8.2	8.4	35	45.2	25.2	14.0	8.3	7.3
17	36.0	23.4	15.1	12.0	13.5	36	41.2	24.3	15.0	10.5	9.0
18	38.0	23.2	15.1	11.4	12.3	37	39.1	24.2	15.2	11.4	10.1
19	38.4	23.5	14.4	11.6	12.1	38	37.3	23.7	15.9	11.9	11.2

Table 4 is a study, on identical lines, of the fractional distillation of acid mixtures representing the acid radicles of the esters. In the last case those acid mixtures were isolated in the following manner: The free acidity of the liquor having been determined in a separate portion, 100 cc. of it was neutralized exactly with the required amount of standard alkali and distilled. The distillate was saponified in the ordinary manner (as in the determination of esters) and again distilled till all alcohol had passed over. The residue in the distillation flask being then diluted with water, acidified with H_2SO_4 and distilled, the residue in the distillation

TABLE 2.
Fractional distillation of fusel oil—miscellaneous brandies.

SAMPLE NO.	PER CENT OF ACID IN EACH FRACTION					SAMPLE NO.	PER CENT OF ACID IN EACH FRACTION				
	1	2	3	4	5		1	2	3	4	5
French brandy						60	42.7	24.4	14.5	9.8	8.6
39	40.8	23.4	15.1	10.6	10.1	61	36.0	24.0	16.7	12.0	11.3
40	41.3	24.0	14.9	10.3	9.5	62	47.6	25.5	13.7	7.4	5.8
41	40.8	24.1	15.4	10.5	9.2	63	38.3	24.7	15.5	11.4	10.1
42	40.3	24.6	15.3	10.9	8.9	Hungarian brandy					
43	40.4	25.7	15.5	10.6	7.8	1	37.8	18.9	16.2	13.5	13.5
44	40.2	23.9	15.0	10.8	10.1	2	35.4	21.0	14.8	13.7	15.1
45	38.9	24.0	14.9	11.1	11.1	3	34.7	22.2	15.3	13.9	13.9
46	41.7	24.2	14.6	10.4	9.1	4	41.6	23.4	14.7	10.5	9.8
47	40.0	24.2	15.3	11.0	9.5	5	38.7	22.9	15.0	11.7	11.7
48	35.0	23.5	16.5	12.5	12.5	Italian brandy					
49	47.2	24.6	13.3	8.3	6.6	1	48.5	24.2	12.6	7.7	7.0
50	41.0	23.9	14.9	10.4	9.8	Greek brandy					
51	44.5	25.3	14.6	8.9	6.7	1	38.1	22.0	14.8	11.9	13.2
52	42.3	24.0	14.7	10.0	9.0	2	46.4	23.7	12.4	8.3	9.2
53	40.2	25.1	15.9	10.5	8.3	3	48.8	24.1	12.7	7.5	6.9
54	35.9	22.5	15.9	13.1	12.6	4	34.8	22.8	15.4	13.1	13.9
55	44.5	25.4	15.0	8.7	6.4	5	47.8	23.8	12.3	7.8	8.3
56	38.8	23.6	15.2	11.9	10.5	6	48.0	23.8	14.5	7.7	6.0
57	42.9	25.2	14.1	9.8	8.0						
58	37.9	24.2	15.5	11.8	10.6						
59	35.9	23.3	16.8	12.6	11.4						

flask was again diluted and again distilled. The combined distillates containing the volatile acid radicles of the esters were submitted to fractional distillation in the manner described above for fusel oil.

Of all the samples examined, only the eight samples marked "Authentic cognac" are known to be genuine cognacs. All the other samples are marked according to the legend on the label, and their history, origin, and mode of manufacture are not known. Handicapped as we are because of this uncertainty as to the origin and peculiarities in the methods of manufacture of almost all of the samples under examination, still it is possible to draw some general conclusions from a study of the tables submitted.

TABLE 3.
Fractional distillation of fusel oil—miscellaneous liquors.

SAMPLE	PER CENT OF ACID IN EACH FRACTION				
	1	2	3	4	5
Spanish brandy:					
No. 1.....	36.7	22.0	15.3	13.0	13.0
No. 2.....	40.4	22.9	15.3	10.7	10.7
Chile brandy.....	41.3	24.0	14.6	10.4	9.7
Batavia arrac.....	27.7	19.7	16.2	16.8	19.6
German plum brandy.....	36.3	23.1	15.6	12.5	12.5
Slivowitz.....	41.3	23.6	14.8	10.7	9.6
Pomace brandy (French):					
No. 1.....	36.6	21.7	14.7	12.6	14.4
No. 2.....	29.7	20.4	15.9	15.3	18.7
No. 3.....	29.9	19.6	15.7	15.6	19.2
Pomace brandy (Italian).....	30.9	21.4	16.4	14.6	16.7
Jamaica rum.....	24.1	20.7	19.3	17.9	18.0
West Indies rum.....	46.9	24.1	12.7	8.2	8.1
St. Croix rum.....	47.0	23.5	12.8	8.4	8.3
Cuba rum.....	35.8	22.7	16.1	13.1	12.3
French rum.....	41.3	23.8	14.8	10.1	10.0
Raisin brandy:					
No. 1.....	46.6	25.7	13.5	8.1	6.1
No. 2.....	39.2	23.7	14.4	11.4	11.3
Brandy:					
No. 1.....	35.7	22.3	15.9	12.7	13.4
No. 2.....	29.4	19.6	16.3	15.7	18.9
No. 3.....	28.0	20.2	16.9	15.2	19.7
No. 4.....	46.4	25.4	13.6	8.2	6.4
Authentic cognac:					
No. 1.....	46.6	24.8	13.0	8.1	7.5
No. 2.....	44.3	23.1	13.5	9.4	9.7
No. 3.....	45.1	24.5	13.1	8.9	8.4
No. 4.....	49.7	25.2	12.4	7.0	5.7
No. 5.....	48.5	24.5	12.4	7.6	7.0
No. 6.....	46.6	24.4	13.0	8.4	7.6
No. 7.....	49.6	23.8	12.2	7.4	7.0
No. 8.....	45.2	24.1	13.7	8.5	8.5
Commercial fusel oil.....	49.9	24.6	12.0	7.0	6.5

(1) The combined acids of the esters in distilled liquors seem to be surprisingly constant in their composition, and their rate of distillation does not, therefore, offer as promising a means for detecting sophistication as is undoubtedly the case with the higher alcohols.

(2) In the case of the acids obtained by oxidation of the higher alcohols of brandies, the greatest variations are shown by the values for the first and fifth fractions, their averages being for—

	Per cent	Per cent
Authentic cognacs.....	47.0	7.7
French brandies.....	40.5	9.9
Pomace brandies.....	31.8	17.2

The values of the fourth fraction are usually not much different from those of the fifth fraction, and the variations in the values of the second and third fractions are rather small.

(3) The rate of fractional distillation seems to offer a promising means of differentiating between cognacs, ordinary brandies, and pomace brandies, and generally between distilled liquors where difference in geographical origin or mode of manufacture has affected the composition of the fusel oil complex.

TABLE 4.
Fractional distillation of esters—miscellaneous liquors.

SAMPLE	PER CENT OF ACID IN EACH FRACTION				
	1	2	3	4	5
French brandy:					
No. 1.....	18.3	16.1	18.0	20.7	27.0
No. 2.....	17.0	16.2	18.2	21.0	27.5
No. 3.....	21.2	15.9	17.1	20.5	25.2
No. 4.....	17.4	17.8	17.8	20.4	26.6
No. 5.....	16.7	16.8	18.3	21.3	26.9
No. 6.....	17.0	17.0	18.1	21.3	26.6
No. 7.....	18.1	16.1	18.0	20.6	27.2
No. 8.....	16.9	16.0	16.5	21.4	29.2
No. 9.....	16.6	16.2	18.2	21.2	27.8
No. 10.....	17.0	16.3	18.1	20.7	27.9
No. 11.....	18.8	16.3	18.0	19.8	27.1
No. 12.....	16.6	16.4	18.2	21.1	27.7
No. 13.....	17.6	16.3	17.2	21.1	27.8
No. 14.....	17.3	16.5	18.4	20.8	27.0
No. 15.....	17.3	16.9	18.5	21.3	26.0
No. 16.....	17.5	16.5	18.1	21.2	26.7
Authentic cognac:					
No. 1.....	21.0	17.3	17.3	19.4	25.0
No. 2.....	19.4	16.4	17.9	20.2	26.1
No. 3.....	20.3	17.0	18.0	19.8	24.9
No. 4.....	20.9	17.7	18.0	19.3	24.2
Pomace brandy:					
No. 1.....	19.4	17.0	17.6	20.7	25.3
No. 2.....	17.6	16.7	18.1	21.1	26.5
No. 3.....	19.0	16.6	18.6	20.0	25.8
Greek brandy:					
No. 1.....	23.4	16.7	17.5	18.5	23.9
No. 2.....	18.2	16.1	18.4	20.9	26.4
No. 3.....	16.0	16.0	18.0	20.9	29.1
No. 4.....	16.6	13.3	19.4	22.0	28.7
Rum (Jamaica).....	19.2	17.3	17.9	20.3	25.3
Rum (French):					
No. 1.....	20.4	15.8	17.3	20.6	25.9
No. 2.....	18.4	16.7	18.1	21.1	25.7
Rum (Martinique).....	19.6	16.7	18.1	19.2	26.4
Rum (St. Croix).....	16.4	16.4	17.5	21.2	28.5
Rum (West Indies).....	15.7	16.7	18.5	21.8	27.3
Apricot liquor.....	18.9	16.8	18.8	20.4	25.1
Batavia arrac.....	15.9	16.7	18.2	21.1	28.1

(4) In one case—that of French brandy (No. 17, Table 1)—which gave upon analysis rather high figures for fusel oil and esters, thus apparently indicating good quality, suspicion as to its genuineness was aroused by the fact that the fractional distillation of the acids from the fusel oil gave the following figures: 36.0, 23.4, 15.1, 12.1, 13.5, indicative of

pomace rather than of ordinary brandy. The sample was thereupon submitted to two expert tasters, both of whom agreed in pronouncing the sample sophisticated. There are a number of brandies marked "French brandies" in Tables 1 and 2 whose fractional distillation curves are more or less similar to the curve mentioned above, and thus point the same way; that is, that they are not genuine brandies.

(5) A comparative organoleptic test of the "Authentic cognacs" seems to point to the fact that the finer the flavor and aroma of the cognac the higher the value of the first fraction in the fractional distillation.

(6) To sum up: The results so far obtained warrant the belief that valuable information relative to the origin and character of distilled liquors can be gained by ascertaining the rate of distillation of the mixed acids obtained by oxidation of their higher alcohols. In order to establish the value and limitations of the method, it will be necessary to work upon a sufficiently large number of samples from various sources the history of which is well authenticated.

APPLICATION OF THE PROCEDURE TO THE ANALYSIS OF A FLAVORING ESSENCE.

Table 5 shows the rate of distillation of formic, acetic, propionic, butyric, and valeric acids and also of simple mixtures of some of these acids when distilled under conditions indicated above in the discussion of fusel oils in distilled liquors.

The figures in the table are averages of two determinations in each case except in the case of formic acid, where an average was taken of four determinations. The separate determinations ordinarily check within 0.5%, but formic acid behaves in a rather erratic manner, its figures varying sometimes as much as 2%. Column 6 of Table 5 gives the percentage relation of the total acid in the total distillate of 100 cc. to the acid originally present in the distillation flask.

It is obvious that use may be made of the fractional distillation for the following purposes:

- (1) Identification of any of the acids under discussion when present in a state of purity.
- (2) Isolation of any of the acids in a pure state from a mixture of two and sometimes more acids for the purpose of identification.
- (3) Testing for purity of any of the above acids.
- (4) Analysis of a simple mixture of two and sometimes even more of the acids.

The above is the basis of M. Duclaux's method (as quoted in Allen's Commercial Organic Analysis) of identification and estimation of the lower acids of the acetic acid series.¹ In this connection it is well to note that

¹ See also article by A. Landolt, Unterscheidung natürlicher und künstlicher Fruchtather, in Chem. Ztg., 1911, 35: 677.

the composition of an acid mixture cannot always be computed from its fractional distillation curve with any satisfactory degree of accuracy, as the acids mutually affect each other during distillation. The curve of the mixture nevertheless gives an approximate idea of its composition, and two or three experiments are generally sufficient to make up a known mixture of the given acids that will yield upon fractional distillation a curve practically identical with the one from the unknown mixture. The total acid in both the known and unknown mixture should be approximately the same in order that the results of their fractional dis-

TABLE 5.
Fractional distillation of acids.

ACID IN DISTILLATION FLASK	MIXTURES	PER CENT OF ACID IN EACH FRACTION					TOTAL ACID IN COMBINED DISTILLATE
		1	2	3	4	5	
<i>cc. N/10</i>							<i>per cent</i>
8.5-17.7	Formic acid.....	11.6	13.9	17.2	22.0	35.3	42.8
11.6-7.5	Acetic acid.....	14.8	17.1	18.9	21.8	27.4	68.1
6.4-12.8	Propionic acid.....	24.5	22.6	20.7	17.8	14.4	92.1
7.3-15.6	Butyric acid ¹	36.1	27.1	18.4	11.8	6.6	94.0
11	Valeric acid ²	54.6	27.2	12.2	4.3	1.7	97.0
30	2 molecules valeric plus 1 molecule acetic.....	45.6	24.8	13.5	8.5	7.6	87.5
21	2 molecules valeric plus 1 molecule propionic..	44.7	25.9	15.0	8.8	5.6	96
21	2 molecules valeric plus $\frac{1}{2}$ molecule acetic plus $\frac{1}{2}$ molecule formic.....	46.5	24.8	13.4	8.1	7.1	84
58	1 molecule valeric (14.5 cc.) plus 3 molecules acetic (43.5 cc.).....	28.2	20.5	16.8	16.1	18.4
.....	1 molecule valeric (11.6 cc.) plus 4 molecules acetic (46.4 cc.).....	25.8	20.0	17.2	17.1	19.9
48	1 molecule valeric (8 cc.) plus 5 molecules acetic (40 cc.).....	24.4	19.4	17.4	17.7	21.1
49	1 molecule valeric (7 cc.) plus 6 molecules acetic (42 cc.).....	23.0	19.2	17.6	18.4	21.8

¹Of uncertain purity.

²Obtained by oxidation of amyl alcohol (Kahlbaum's).

tillation might be comparable, as the fractional distillation curve is somewhat affected by the concentration of the acid to be distilled. The limits of permissible variation in the strength of the acid to be distilled without affecting the distillation curve have not been determined, though, as will be seen from Table 5, a 100% variation in the strength of an approximately N/100 acid does not materially affect its distillation curve.

A complete analysis of a flavoring essence—a so-called banana oil—has been made in this laboratory by a procedure based largely on the above-described facts and considerations involving the fractional dis-

tillation of acids and their mixtures. A detailed description of the procedure followed will now be given as an illustration of the possibilities of the method.

DESCRIPTION OF METHOD.

About 5 grams of the banana oil was saponified with an excess of aqueous KOH by boiling under reflux condenser. The total alcohols in the saponification product were then distilled off and thus two solutions obtained:

(1) The residue in the distillation flask containing the total combined acids from the esters, together with a small amount of acid originally free in the oil and which has been determined in a separate portion of the oil by direct titration; and

(2) The alcoholic distillate containing the total alcohol, free and combined, of the original oil.

The excess of alkali in solution (1) was titrated with sulphuric acid and the total combined and free acid in the oil thus estimated. The neutralized solution was then acidified and an aliquot fractionally distilled. The values obtained for the fractions were as follows: 22.7, 19.1, 17.7, 18.8, 21.7. Ratio of acid in total distillate to acid in distillation flask, 72%.

With a view of isolating the component acids of the mixture, fraction 1 and also the residue in the distillation flask were, upon proper dilution, again fractionally distilled. The latter (the residue in distillation flask) yielded values for the fractions practically identical with those of pure acetic acid. The first fraction, however, yielded a curve indicating a mixture and not a simple acid. The first fraction in this, the second distillation, was again refractionated and the operation repeated until after the fifth fractional distillation of the first fractions values were obtained practically identical with those of a pure valeric acid. Thus was the acid mixture in the saponification product proved to be a mixture of valeric and acetic acids. It was then established by direct experiment that a mixture of one molecule valerie and six molecules acetic acids would yield upon fractional distillation the following values: 23.0, 19.2, 17.6, 18.4, 21.8. Ratio of acids in total distillate to acid in distillation flask, 71.7%.

The above values are practically identical with those obtained from the acid mixture in the saponification product of the banana oil. Thus was the composition of the total acid (free and combined) in the banana oil determined to be as follows:

	cc. N/10 per gram
Free acid (assumed to be acetic).....	2.0
Total combined acids (by saponification).....	67.4
Combined acetic acid (by fractional distillation).....	57.5
Combined valeric acid (by fractional distillation).....	9.9

An aliquot of the solution of the total acids was neutralized, evaporated off, dried, and weighed, and the weight agreed fairly well with the composition indicated above.

The alcoholic distillate from the saponification product containing the total alcohol (free and combined as asters) of the banana oil was oxidized with alkaline permanganate to their corresponding acids.¹ The resulting acids were distilled off, the total acidity of the distillate titrated, and the solution then examined for its com-

¹See method of C. R. Smith and A. S. Mitchell, U. S. Bur. Chem. Bul. 122: "Determination of Fusel Oil by Alkaline Permanganate."

ponent acids qualitatively and quantitatively by a system of fractional distillation, exactly in the same manner as outlined above for the combined and free acids in the oil.

The acid mixture resulting from the oxidation of the total alcohols (free and combined) in the oil proved to be composed as follows:

	cc. N/10 per gram
Valeric acid.....	55
Acetic acid.....	30

thus indicating that the total alcohols were composed as follows:

	cc. N/10 per gram
Amyl alcohol.....	55
Ethyl alcohol.....	30

The above results were checked in the following manner:

The original banana oil was saponified with alcoholic potash, all the alcohol in the saponification product distilled off, and the alcoholic distillate examined for higher alcohols in exactly the same manner as fusel oil is determined in brandy (Allan Marquart fusel-oil method). Titration of the final acid distillate gave 55 cc. N/10 acid per 1 gram oil, and the fractional distillation of the solution gave values for the fractions identical with those of pure valeric acid.

Further examination of the banana oil gave the following data:

Free ethyl alcohol was proved to be absent; the presence of amyl acetate, ethyl acetate, and amyl valerate was indicated by the odor of various fractions obtained from a fractional distillation of the original banana oil; nonvolatile unsaponifiable matter, 1%.

Summing up the results obtained, we arrive at the following composition:

	cc. N/10 per gram
Free acid (as acetic).....	2.0
Combined acetic acid.....	57.5
Combined valeric acid.....	9.9
Combined ethyl alcohol.....	30.0
Combined amyl alcohol.....	37.4
Free amyl alcohol.....	17.6
Nonvolatile unsaponifiable (per cent).....	1

and the rational formula may be expressed as follows:

	cc. N/10 per gram	per cent
Free acid (as acetic).....	2.0	1.2
Amyl valerate.....	9.9	17.0
Ethyl acetate.....	30.0	26.4
Amyl acetate.....	27.5	35.7
Free amyl alcohol.....	17.6	15.5
Nonvolatile unsaponifiable.....		1.0
		96.8

No report was made by the associate referee on vinegar.

REPORT ON SPICES.

By HARRY E. SINDALL (Philadelphia, Pa.), *Associate Referee*.

The work done this year falls under two headings—moisture determinations on whole spices and the total ash of herbs.

MOISTURE.

No recommendations were made by last year's associate referee for further work on spices, but it seemed desirable this year to study a method for determining moisture in whole spices. No samples were sent out, as the nature of the work rendered it unsuitable for coöperative investigation. The work was done on whole cloves and whole black pepper. The method under consideration was the Brown-Duvel distillation method for moisture in grain. But since unsatisfactory results were obtained by using Renown engine oil, or in fact any engine oil, the best grade kerosene oil was substituted and found to give more uniform results.

The method as worked by the associate referee is as follows:

Place 50 grams of whole spice in a distilling flask with 150 cc. kerosene, whirl the flask several times to bring the oil in contact with each particle of spice. Place the flask on an asbestos board. Cut so that the bottom of the flask extends below the surface. A wire gauze with an asbestos center is placed about one-half of an inch below the bottom of the flask. The object is not to bring the flame in direct contact with the flask, and the asbestos board serves to keep the heat uniform. Connect the flask directly with a vertical condenser and collect the distillate in a graduated cylinder or burette. Insert a thermometer through the stopper of the distilling flask, extending down into the oil. Adjust the flame so that about twenty minutes will be required to reach the temperature of 170°C. Now extinguish the flame, after which the thermometer will show a slight gradual increase in temperature. As soon as the water stops dropping from the condenser tube, which usually requires from four to six minutes, the operation is complete. Multiply the volume of the water layer by 2 to obtain the percentage of moisture.

The following table shows results obtained by drying 2 grams of whole cloves and whole black pepper at 110° C. to constant weight; from the resulting loss in weight the amount of volatile ether extract obtained by extracting the whole spices was subtracted.

The samples marked (a) were taken before the cloves went on the mill to be ground; the samples marked (b) represent the same cloves immediately after grinding. The loss according to weight by grinding was 6.26%.

Comparative results on moisture.

BY EXTRACTION			BY DISTILLATION		
	per cent			per cent	
Whole cloves ..	(a) 13.94	(a) 11.2	Ground cloves.	10.16
Do	15.93	12.6	Whole pepper..	6.02	6.0
Do	11.24	9.8	Do	7.95	6.8
Do	10.61	9.5	Ground pepper.	7.87	8.0
Ground cloves.	(b) 6.11	(b) 6.0			

This loss consists chiefly of moisture and some volatile oil, due to the fact that the cloves are more or less heated while passing through a high-speed mill.

The large percentage of moisture in the cloves as shown in the above table under extraction method seems due to the fact that all the volatile oil was not extracted by the ether, as shown in the following table:

Volatile ether extract.

	<i>per cent</i>		<i>per cent</i>
Whole cloves.....	2.88	Ground cloves.....	15.74
Do.....	2.89	Whole pepper.....	0.30
Do.....	3.13	Do.....	0.53
Do.....	3.31	Ground pepper.....	2.13
Ground cloves.....	13.91		

TOTAL ASH DETERMINATION.

The associate referee's attention was called to wide discrepancies in the results of total ash determinations obtained by different chemists on samples taken from the same lot of herbs. For the purpose of studying this matter, large samples of marjoram, sage, savory, and thyme were ground under the associate referee's supervision, and subdivisions of these were sent to collaborators, with instructions to determine the total ash in each sample by the method commonly used by the respective collaborators and describe the method used. Reports were received from eight collaborators:

Total ash.

ANALYST	MARJORAM	SAGE	SAVORY	THYME
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
F. L. Shannon, Lansing, Mich.	{ (a) 14.34 (b) 14.135	(a) 9.265 (b) 9.275	(a) 13.335 (b) 13.29	(a) 12.89 (b) 12.78
C. L. Black, Philadelphia, Pa.	16.16	9.45	14.06	13.14
H. B. Mead, Philadelphia, Pa.	15.84	9.75	14.05	13.42
C. S. Brinton, Philadelphia, Pa.	{ 15.21 15.10	9.39 9.43	13.63 13.72	13.47 13.21
A. P. Coulture, Ottawa, Canada..	{ 14.54 14.61	9.47 9.50	13.68 13.56	12.47 12.59
Paul Rudnick, Chicago, Ill.	{ 13.42 13.44	9.31 11.21	12.60 12.68	13.14 12.48
C. O. Dodge, Washington, D. C.	14.05	8.41	12.41	13.46
J. H. Bornmann, Chicago, Ill.	{ 14.08 14.48	9.80 9.82	13.77 13.85	13.67 13.43

Mr. Dodge also determined nonvolatile matter and ash on a nonvolatile basis, with the following results:

	NONVOLATILE MATTER	ASH ON NON- VOLATILE BASIS
	<i>per cent</i>	<i>per cent</i>
Marjoram.....	92.6	15.17
Sage.....	93.0	9.05
Savory.....	91.6	13.54
Thyme.....	92.9	14.48

METHODS OF COLLABORATORS.

Mr. Shannon made the determinations in duplicate. Sample (a) in each case was ignited in an ordinary platinum milk dish; sample (b) in a fused silica dish of the same size. All were burned first over a Meker burner at a very low red heat and then exposed to a low red heat in a muffle, cooled in a desiccator, and weighed.

Mr. Black used 2-gram portions in platinum dishes which were held about one-fourth inch above bottom of muffle by means of asbestos board supported by small pieces of pipestem. The bottom of the muffle was kept at a barely visible red heat. After ashing was apparently complete, the residue was moistened with alcohol to reveal any unburned carbon and then reheated if necessary until treatment showed no unburned carbon.

Mr. Mead used 2-gram samples in flat-bottom round dishes, dimensions about 3 inches by 1 inch. These were elevated on pieces of pipestems about three-eighths inch above the bottom of the gas muffle heated to a visible red. The dishes were below visible red. The ash was moistened with alcohol and reheated. There was practically no carbon or change after the first heating.

Mr. Brinton used 2-gram portions and made the combustion in small platinum dishes about 1 inch square and approximately three-eighths inch deep. At no time during any of the work were the dishes heated to a red heat. The determinations were made in a gas-heated muffle, the bottom of the muffle at times being more or less red; but the dishes were supported on pieces of asbestos board about one-fourth inch or more above the bottom of the muffle. An air space, therefore, separated the asbestos board from the red-hot bottom of the muffle. The asbestos board was supported by pieces of crucible lid.

Mr. Coulture used 2-gram samples, and charred them in platinum at a heat below redness. This temperature was found to be sufficient to completely burn off all the carbon. He did not use a muffle. After weighing, dish and contents were heated again slightly below red heat for a period of half an hour and the operation repeated to constant weight. After a time there was a slight increase in weight, and the minimum was taken as being the true weight of the ash.

Mr. Rudnick used 2 grams in a small flat-bottom porcelain dish and ignited carefully at a low temperature so as to avoid loss of sample. The ignition was continued at a low red heat and finally at a bright red for a few minutes, then cooled and weighed. The appearance of the ash after weighing was carefully noted to be sure that all carbon had been burned off, also to note whether particles of sand and foreign matter other than true ash were apparent.

Mr. Dodge followed the method given in U. S. Bureau of Chemistry Bulletin 107 (revised), except that the products were dried at 110°C. for four hours and weighed, then dried for one hour and again weighed. The greatest loss during the second drying was less than 1%.

Mr. Bornmann charred 2 grams of the well-mixed substance in a platinum dish over asbestos, using a small flame. The carbon was burned off in a muffle at a temperature below redness.

CONCLUSIONS.

These results are not encouraging. While the samples submitted were known to contain several per cent of sand, it is not believed that the discrepancies can possibly be due entirely to this cause, but that they were attributable, in part at least, to the temperature employed in mak-

ing the combustion. It would seem, at all events, that the subject warrants further investigation, particularly along the line of operating at definite temperatures.

RECOMMENDATIONS.

It is recommended—

(1) That the associate referee's modification of the distillation method for water in spices be given further study.

(2) That the subject of ash determination in herbs be further studied, with particular reference to the influence of the exact temperature employed in the combustion.

NOTE REGARDING THE DETERMINATION OF CRUDE FIBER IN BLACK PEPPER.

By A. E. PAUL (Bureau of Chemistry Food and Drug Inspection
Laboratory, Chicago, Ill.).

Two samples, one with a low fiber content and the other high in this constituent, were sent to various laboratories in this district, with the request that analyses be made. The results follow:

Analyses of black pepper.

SAMPLE NO. 1.

ANALYST	NON-VOLATILE ETHER EXTRACT	VOLATILE ETHER EXTRACT	TOTAL ASH	ASH IN- SOLUBLE IN HCl	CRUDE FIBER	PENTO- SANS
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
J. H. Bornmann, Chicago... {	8.03	0.70	5.09	0.57	10.46
	8.27	0.83	5.41	0.61	11.38
E. H. Berry, Chicago					11.09
J. Feldbaum, Chicago.....	8.10	1.04	5.35	0.62	11.08
H. D. Grigsby, Cincinnati....	8.19	0.55	5.24	0.51	11.99
J. S. McCune, St. Louis.....	8.87	5.16	0.43	10.41	6.1
C. L. Clay, New Orleans.....	8.69	4.62	0.34	11.26	6.0

SAMPLE NO. 2.

	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
J. H. Bornmann, Chicago... {	7.20	1.03	6.48	0.95	16.60
	7.26	1.01	6.61	1.00	17.11
E. H. Berry, Chicago					16.60
J. Feldbaum, Chicago.....	7.15	1.18	6.48	0.92	16.80
H. D. Grigsby, Cincinnati....	7.18	0.57	6.43	0.79	17.41
J. S. McCune, St. Louis.....	7.82	6.37	0.76	15.84	8.40
C. L. Clay, New Orleans.....	7.80	5.48	0.56	17.23	7.90

In view of the very great discrepancy in the various fiber determinations, a new 5-pound sample was taken by an analyst, carefully mixed, quartered to a small bulk, and transferred to a small sample bottle.

The remainder was again carefully mixed, and a small subsample withdrawn as before. Then still a third sample was taken in the same manner. The analyst was requested to make crude-fiber determinations in duplicate on each subdivision.

The remainder of the pepper was then delivered to another analyst, who prepared three subsamples in the same manner as above.

Both analysts were instructed to stir the material in the bottle very carefully with a spoon of approximately 2-grams capacity, to withdraw a spoonful from the interior of the bottle, and carefully transfer the entire contents to the balance pans. It was then left to the option of the analysts to utilize the quantity thus taken for the fiber determination, or else to remove carefully the slight excess over 2 grams, or add sufficient to make that amount. Of the two analysts, Mr. Perry preferred the former detail, while Mr. Bornmann chose to work on the exact 2 grams. Their results were:

ANALYST	SAMPLE NO. 1	SAMPLE NO. 2	SAMPLE NO. 3
	<i>per cent</i>	<i>per cent</i>	<i>per cent</i>
E. H. Berry.....	13.36	12.98	13.46
	13.15	13.00	13.16
J. H. Bornmann.....	13.03	13.15	13.25
	13.25	13.07	13.31

These results are quite acceptable, and show that the trouble is not in the method itself, but rather in the manner of mixing and weighing out the samples.

PRELIMINARY REPORT ON DETERMINATION OF ASH, ESPECIALLY IN SPICES.

By CLEMENT S. BRINTON (Food and Drug Inspection Laboratory, U. S. Appraiser's Stores, Philadelphia, Pa.)

The object of this paper is to emphasize the necessity of carefully following the directions for the determination of ash as given in the Official and Provisional Methods of Analysis of this association (U. S. Bur. Chem. Bul. 107, rev.), particularly where, under "Ash, official" for the analysis of foods and feeding stuffs, the following language is used: "* * * burn until free of carbon at the lowest possible heat" (p. 38). Again, under "Preparation of ash," etc., (p. 238), it is directed to "Conduct the combustion at a comparatively low temperature, never employing a full red heat * * *." Other similar directions could be cited.

The desirability of preparing the material mentioned herein arose through trouble in obtaining closely agreeing results for ash by different

analysts during the examination of spices, especially those having normally a high sand-free ash, such as marjoram with 11 to 16% of ash and thyme with from 10 to 14%.

This lack of agreement was found to be due not to faulty sampling but to lack of uniformity in the analytical method. This was ascertained by having portions of a large sample of marjoram finely ground, about 60 mesh, thoroughly mixed, and analyzed in seven laboratories in the Eastern States according to the procedure usually employed in those laboratories. Determinations were made in platinum dishes in all cases except laboratory 6, where a porcelain crucible was used. Electric muffles were used in laboratories 1 and 4, and probably in laboratory 5; gas muffles were used in laboratories 2, 3, and 7; and laboratory 6 burned in open flame. The results submitted are given in Table 1.

TABLE 1.
Ash determinations on marjoram—first report by collaborators.

LABORATORY	ANALYST	TOTAL ASH	REMARKS
		<i>per cent</i>	
1	W. C. T.....	15.21	{ Muffle below redness; time of burning, 1 hour. Not observed carefully to ascertain when free of carbon.
		15.24	
		15.16	
		15.18	
2	W. J. McG.....	13.47	{ Muffle high red; time of burning not stated. Heated to constant weight.
		13.45	
3	J. B. L.....	15.14	{ Muffle low cherry red; time of burning not stated. Ash stirred with rod from time to time until carbon burned. Analyst J. E. heated longer and found no loss in weight.
	J. E.....	14.90	
4	F. O. W.....	15.11	{ Muffle below redness; time of burning not stated. Moistened with water to observe presence of carbon; again heated and weighed when free of carbon.
		15.13	
		15.11	
		15.09	
5	L. P.....	15.40	{ Muffle below redness; time of burning not stated. Moistened with water after weighing and found free of carbon.
		15.30	
6	H. E. S.....	13.24	{ Open flame with crucible bright red; time of burning, 40 minutes. Ash determined as he usually does in spices.
		13.35	
7	C. L. B.....	15.68	{ Dish not visibly red; time of burning not stated. Ash treated with water after weighing to test for carbon.
		15.70	
	H. B. M.....	16.11	{ Bottom of muffle below visible red heat; time of burning not stated. Moistened and reheated before weighing; weighed after free of carbon.
		16.16	
		16.03	
		15.69	

The writer obtained results ranging from a maximum of 15.8% down to a minimum of 14.89%, according to conditions, all results being obtained when the dish was not at a visible red heat as viewed in the open muffle by daylight. All determinations were made in platinum dishes about 1 inch square, one-half inch deep, in a 5-burner gas Wiesnegg muffle. The results are given in Table 2.

TABLE 2.
Ash determinations on marjoram—results by author.

DATE (1915)	TOTAL ASH		REMARKS
	Dish 1	Dish 2	
	<i>per cent</i>	<i>per cent</i>	
May 24	15.43	15.46	Time of burning, 1½ hours in all. Dishes on bottom of muffle entire time. Heated 1½ hours below visible red heat, then raised to dull red for one-fourth hour.
	15.01	14.89	Muffle just visible red on bottom; time of burning, 35 minutes. Dishes on bottom of muffle entire time and carbon-free when moistened with alcohol.
May 25	15.37	15.75	Not a visible red; time of burning, 35 minutes. Both dishes about one-fourth inch above bottom of muffle on crucible lids. Muffle open. Dish with high result in front of other.
May 26	15.74	15.86	Not a visible red; time of burning, about 2½ hours. These results from same conditions as above except muffle not so hot. After 1½ hours tested but not carbon-free. After weighing and getting these results found carbon free.
June 3	15.06	15.39	Bottom dull red in spots; time of burning, about 2 hours. One dish in front of other; otherwise same as second set May 24. Dishes raised off bottom and not a visible red.
	15.20	15.48	Bottom dull red in spots; time of burning, only 35 minutes in all. Carbon all burned out. One dish in front of other. Dishes raised off bottom.
June 8	15.18	15.10	Bottom dull red in spots; time of burning, only 37 minutes in all. Carbon all burned out. Dishes side by side and supported one-fourth inch above bottom on asbestos board. Neither asbestos board nor dishes a visible red. Hot air could circulate under and over asbestos and dishes.
June 9	15.70	15.65	Time of burning, about 6 hours. Conditions same as on June 8, but lower temperature used. Both ashes moistened after weighing and free of carbon. Dishes side by side.

TABLE 2.—Continued.

DATE (1915)	TOTAL ASH		REMARKS
	Dish 1	Dish 2	
	<i>per cent</i>	<i>per cent</i>	
June 9	23.35	23.87	Muffle slightly red in spots; time of burning, 17 minutes. Much carbon present. Conditions same as preceding.
	16.18	16.20	Muffle slightly red in spots; time of burning, 10 minutes long. Carbon still present in both.
	15.88	15.68	Dishes not visible red; time of burning, additional 10 minutes. Traces of carbon in both; most in higher result.
	¹ 16.37	¹ 16.10	Dishes not visible red. Both moistened, evaporated to dryness, heated in muffle below redness before weighing.
	16.15	15.93	Dishes not visible red. Same as preceding after one-half hour longer in muffle.
	15.75	15.65	Dishes not visible red. Same as preceding after 1 hour longer in muffle; temperature slightly higher.

¹ Note the effect of moistening with water and subsequent long heating required to drive off this water and bring result down to that obtained before moistening.

The results reported on May 26 agreed closely with those obtained by Analyst C. L. B., but are slightly below those reported by Analyst H. B. M. Further attempts to duplicate the latter's results were in vain, as shown by work subsequent to May 26. This later work brought out very emphatically the fact that this sample of marjoram contained mineral matter very susceptible to slight changes in temperature, length of time of burning, etc., and that without exact duplication of temperature and other conditions closely agreeing results at different times could not be determined. See various results in Table 2 on June 3, June 8, June 9, and June 10, where determinations in duplicate in every particular, except that one dish was in front of the other and in slightly cooler part of muffle, show variations of 0.3%, and yet both are free of carbon.

It was found that only when the two dishes were placed side by side could close duplicates be obtained, as shown by results of June 8 and June 9, each set of which agrees closely; but there is a marked difference of about 0.5% because of different temperatures, the dishes in neither set being a visible red, although the bottom of muffle was dull red in spots. The second set of results on June 9 were made at a low temperature and weighed at short intervals to determine if some easily volatile mineral matter was present which was completely volatilized about the time the carbon was completely burned. The tabulated results show that lower results were obtained than reported by Analyst H. B. M., yet some carbon was still present. Just at this point, the writer learned that Analyst H. B. M. had moistened his ash before weighing to uncover unburned car-

bon, then added enough alcohol to make a mixture which would burn, ignited the alcohol and burned it off, and replaced the dish in muffle for a short time, then cooled and weighed as usual. Because of this analyst obtaining higher results than anyone else and higher than the writer could obtain in any other way, the last set in Table 2 when about free of carbon were moistened, treated with alcohol, etc., about the same as did Analyst H. B. M., and it was found that his high results as originally reported could be duplicated, but in no other way; therefore, the conclusion was made that the water treatment before weighing explained his higher results. Later work by the author confirms this conclusion (see Table 4). The work reported in Table 2 convinced the author that for marjoram especially, and probably in other substances high in ash, an accurate determination of the ash required that more care be given to the control of the temperature than is usually given, and it was thought that higher results would be reported by the various analysts if their attention was called to this point. Consequently, the following method was sent to the previous collaborators:

Take 2 grams of the well-mixed sample in a platinum dish and burn at *lowest possible temperature*, below a visible red heat, until free from carbon when examined by the eye; cool, and weigh as usual. Now moisten ash with 95% alcohol, and examine for small specks of carbon. If specks of carbon are found, ignite alcohol, allow to burn off, and then return dish to muffle and allow to heat a little longer at same temperature. Cool and weigh again, repeating this operation until all carbon is burned off.

All results obtained were requested, also an opinion as to the true value for ash in the sample of marjoram sent out, the various results previously submitted by all of them being sent also so they would see the reason for further work. The second set of results are given in Table 3.

TABLE 3.

Ash determinations on marjoram—second report by collaborators.

LABORATORY	ANALYST	TOTAL ASH		REMARKS
		Dish 1	Dish 2	
		<i>per cent</i>	<i>per cent</i>	
1	H. R. S. ¹	15.47	Time of burning not given. Two grams in flat-bottom platinum dish, center cold electric muffle; rheostat set for temperature below red heat and burned till free of carbon.
		15.56	Same as above, except that when entirely charred dish was withdrawn. Ash covered with alcohol, ignited, and again heated in muffle <i>below red heat</i> . After 2 treatments with alcohol carbon-free.

¹ General comment by analyst; "It is evident that prolonged heating at even very low temperature does cause a loss of inorganic matter by volatilization. Also the most practical and reliable method for determining ash in organic products is that outlined in method 3 above."

L. NOTE.—No report was received from laboratories 6 and 7.

TABLE 3.—Continued.

LABORATORY	ANALYST	TOTAL ASH		REMARKS
		Dish 1	Dish 2	
		<i>per cent</i>	<i>per cent</i>	
1	H. R. S.....	16.00	Same as above at start. When charred, removed, cooled, and all soluble ash leached out with hot water and filtered in usual manner. Filter and char burned in dish below red heat. Solution of soluble ash added, evaporated to dryness on steam bath, and finally heated at low temperature and weighed.
		15.04	Same as above at start. Rheostat set for lowest possible temperature; allowed to heat all night (16 hours).
2	W. J. McG..	15.10	Time of burning not given. Heated in gas muffle just below red heat till carbon had disappeared, and after one alcohol treatment. After heating at red heat for 30 minutes, obtained 13.03%, 13.06%, and a further drop to 12.46%, 12.54% after 30 minutes more at red heat.
		15.18		
		15.24		
		14.96		
3	J. F. D.....	16.16	16.46	Visible carbon present.
		15.74	15.87	Do.
		15.63	15.77	Do.
		15.37	15.76	Visible carbon absent.
		15.36	15.77	Total time of burning, 9 hours. Constant weight. Six treatments with alcohol were required. Muffle at all times below visible red heat. These results are about 0.5% higher than previously reported from this laboratory, but analyst does not consider increase in accuracy justifies increase of time.
4	F. O. W....	15.03	Time of burning not given. "The heating was done very slowly at a low temperature with proper precautions until no specks of carbon were left."
		15.06		
		14.95		
5	L. P.....	15.66	15.65	Time of burning not given. Burned in electric muffle at lowest temperature possible. When carbon seemed all gone, weighed, then moistened with alcohol and found carbon-free. Temperature is slightly lower than used for first results.
		14.91	Time of burning not given. Obtained at higher temperature than above, but still a temperature which shows no redness in muffle or in dish.
		14.69	Time of burning not given. Obtained on another determination by higher heat than preceding. Analyst considers last two results worthless.

Comments on Table 3 by author: All laboratories submitting reports obtained higher results by using lower temperatures, etc., than first reported, with the single

exception of laboratory 4. Further, these higher results, with two exceptions, agree very well with those obtained in laboratory 7, and those of the author as given in Table 2. The results of Analyst H. R. S., where he used water to moisten ash, confirm those of Analyst H. B. M. and the author, where water was added. The tenacious combination of this water is remarkable since, as shown by the author in Table 2, about $1\frac{1}{2}$ hours' heating at approximately 400°C . was required to drive it off and bring the ash down to the same value as before its addition.

The author, about this time, suggested similar work to the associate referee on spices. He has already submitted his results.

TABLE 4.

Ash determinations on marjoram and stramonium with temperature control.
MARJORAM.

DATE (1915)	TOTAL ASH	MAXIMUM TEMPERATURE BY PYROMETER	TIME BURNING	ASH AFTER MOISTENING AND REHEATING	TIME REHEATING	MAXIMUM TEMPERATURE WHILE REHEATING	INCREASE DUE TO MOISTENING	REMARKS
	per cent	$^{\circ}\text{C}$	<i>h. m.</i>	per cent	<i>h. m.</i>	$^{\circ}\text{C}$.	per cent	
Oct. 8...	15.69	(1)	(1)	15.94	1	400	0.25	Muffle entirely closed except 1-inch hole in front and three-eighths inch hole in back.
	15.90	400	(1)	16.20	1	400	0.30	Do.
Oct. 12..	15.25	(2)	1 35	15.43	1 51	400	0.18	Quartz dish. Muffle wide open. An increase of about 0.2% after almost 2 hours heating.

STRAMONIUM.

	per cent	$^{\circ}\text{C}$.	<i>h. m.</i>	per cent	<i>h. m.</i>	$^{\circ}\text{C}$.	per cent
Oct. 26..	18.21	400-440	4 30	18.45	1 50	400	0.24
	18.27	400-440	4 30	18.37	1 50	400	0.10

¹ Not noted.

² 440°C . during combustion of carbon, then constant at 400°C .

The data given in Table 4 show results obtained where a pyrometer was used in the open gas muffle. The majority of these results are not satisfactory because of difficulties in maintaining a constant temperature in the muffle. The temperature and results reported are only approximate, but show very plainly that where an accurate result is desired on a substance like marjoram, great care must be observed to have exactly identical conditions, or considerable variations must be expected. The effect of moistening the ash is again plainly brought out in this table.

The drug stramonium has also a high ash, and some little work has been done on one sample (see Table 4).

This paper is a preliminary one intended to emphasize certain phases of the determination of ash in spices, especially the necessity of using a low temperature, *below redness at lowest possible temperature*, as specified in the Official and Provisional Methods of Analysis (p. 162) previously cited. The writer expects to present more data later, together with conclusions as to the composition of the mineral matter which volatilizes so easily at or below redness. Evidence will also be presented to show that with products like marjoram, thyme, etc., it is better to determine ash in an open muffle, allowing plenty of circulation of air, instead of in a closed muffle, where, owing to an absence of oxygen, a longer time is required to burn the carbon.

CONCLUSIONS.

(1) The mineral matter in marjoram, thyme, and other plant products having a high ash is easily volatile to a very marked degree at a temperature just below redness.

(2) Great care is necessary by analysts in ashing these products. Temperatures below visible redness must be used or erroneous and low results will be obtained.

(3) After moistening the ash with water the weight before moistening is not regained at the temperature of combustion, even after a reheating of over one hour.

(4) Further work on this subject is necessary so that the official methods will plainly caution analysts against the troubles mentioned above.

The committee for the suggestion of changes in the constitution and by-laws, previously authorized by the association, was announced as follows: B. B. Ross, chairman; C. L. Alsberg, and H. D. Haskins.

REPORT ON FLAVORING EXTRACTS.

By A. E. PAUL (Bureau of Chemistry Food and Drug Inspection Laboratory, Chicago, Ill.), *Associate Referee*.¹

Three methods were studied last year by collaborators on flavoring extracts:

(1) The saponification method for wintergreen extract, devised by Hortvet and West.

(2) The brine method for anise and nutmeg extracts, as described by Hortvet and West.

(3) The Howard-Mitchell method, slightly modified, for peppermint, spearmint, and wintergreen extracts.

¹ Presented by E. K. Nelson.

In each case the method was recommended for further study by the association. Inasmuch as the results reported last year appeared to be quite satisfactory, these three methods only were submitted this year, in the hope that the results obtained would be such as to warrant their final adoption as provisional in this meeting.

Of each extract two samples were submitted, one of full standard strength, as directed in U. S. Bureau of Chemistry Circular 19, and the other one-fourth that strength. The nutmeg extracts, therefore, contained 2.0% and 0.5% of oil, while all the others contained 3.0% and 0.75%. All were made with 95% alcohol, which gives the test of the methods the maximum of severity. In the case of the weaker extracts this was perhaps more severe than necessary, since the lower grades of extracts are usually made with more or less dilute alcohol.

SAPONIFICATION METHOD FOR WINTERGREEN EXTRACT.

In connection with the saponification method for wintergreen extract, Mr. C. B. Gnadinger suggested titrating instead of weighing the separated salicylic acid. Details for this operation were added to the method as previously studied, and collaborators were requested to express an opinion as to their preference. It seems that the same idea had occurred to other operators, and that essentially the same details have already been in use in other laboratories. The results reported and the comments submitted follow:

Analyses of wintergreen extract.

COLLABORATORS	OIL PRE- SENT	OIL FOUND			
		Gravimetric		Volumetric	
		per cent	per cent	per cent	per cent
C. O. Dodge.....	0.75 3.00	0.78 3.01		0.73 2.90	
C. B. Gnadinger.....	0.75 3.00	0.76 2.99	0.76 2.98	0.75 2.94	0.75 2.94
C. F. Jablonski.....	0.75 3.00	0.74 2.96	0.74 2.97	0.73 2.93	0.73 2.94
H. L. Lourie.....	0.75 3.00			0.79 2.98	0.77 2.98
J. P. Street.....	0.75 3.00	0.80 3.01	0.74 2.97		
C. F. Sutton.....	0.75 3.00	0.76 2.99	0.74 2.99	0.76 2.98	0.74 2.98
A. R. Todd.....	0.75 3.00	0.90 3.06		0.80 2.97	
P. B. Yost.....	0.75 3.00	0.77 3.02	0.80 3.05	0.78 3.06	
Averages.....	0.75 3.00	0.77 3.00		0.76 2.96	

COMMENTS BY COLLABORATORS.

C. B. Gnadinger: Method very satisfactory. Little choice between gravimetric and volumetric modifications. I prefer the latter because it is slightly shorter.

Chas. F. Jablonski: Suggest combining the two methods by titrating the weighed residue.

C. E. Morrison (J. P. Street): The salicylic acid was determined gravimetrically, which method is preferred in this laboratory.

Clarence F. Sutton: Titrated the weighed salicylic acid.

A. R. Todd: The method appears to be very satisfactory. It seems to me that the amount of water used in transferring the solution to a separatory funnel could be cut down to 15 cc.

P. B. Yost: I prefer the original method of weighing, as I find end point in the titration to be rather indefinite.

Analyses of anise and nutmeg extracts.

COLLABORATORS	ANISE			NUTMEG		
	Oil present		Oil found	Oil present		Oil found
	per cent	per cent		per cent	per cent	
E. H. Berry.....	0.75	0.8	0.8	0.5	0.5	0.5
	3.00	3.2	3.2	2.0	2.0	2.0
C. L. Black.....	0.75	0.75	0.8	0.5	0.5	0.6
	3.00	3.1	3.1	2.0	2.0	2.0
E. Bloomberg.....	0.75	(¹)	(¹)	0.5	(¹)	(¹)
	3.00	3.4	3.4	2.0	2.0	2.0
Jacob Feldbaum.....	0.75	0.8	0.8	0.5	0.6	0.5
	3.00	3.0	3.2	2.0	1.8	2.0
C. B. Gnadinger.....	0.75	0.7	0.7	0.5	0.5	0.4
	3.00	3.0	2.9	2.0	1.9	1.9
E. J. Munch.....	0.75	1.0		0.5	0.8	
	3.00	3.2		2.0	2.2	
L. Patton.....	0.75	(¹)	(¹)	0.5	(¹)	(¹)
	3.00	3.4	3.4	2.0	2.0	2.0
E. W. Thornton.....	0.75	0.9		0.5	0.2	
	3.00	3.2		2.0	2.0	
G. W. Trainor.....	0.75	1.0	1.0	0.5	0.8	0.8
	3.00	3.2	3.0	2.0	2.2	2.1
Averages.....	0.75	0.83		0.5	0.56	
	3.00	3.2		2.0	2.01	

¹ See comments.

COMMENTS BY COLLABORATORS.

C. L. Black: I found it necessary to use C. P. NaCl, and that even nicer oil layers are obtained by using half saturated sodium sulphate solution.

L. Patton and E. Bloomberg: Stated that the oily layer formed in the case of the dilute extracts did not cover the brine, and that, therefore, an exact reading was not possible; that, however, the amount of oil present is more than a mere trace.

C. B. Gnadinger: Without being advised as to the amount of oil actually present, the results appear to be excellent. If they are reasonably correct, I believe the method should be made official.

E. W. Thornton: The separation in each case was satisfactory without centrifuging a third time.

Analyses of spearmint, peppermint, and wintergreen extracts.

COLLABORATORS	OIL PRES- ENT	OIL FOUND				
		Spearmint		Peppermint		Wintergreen
E. H. Berry.....	0.75	0.8		0.8		0.8
	3.00	3.0		3.0		3.1
R. D. Cook.....	0.75	0.9		0.9		0.8
	3.00	3.0		3.1		2.7
C. B. Gnadinger.....	0.75	0.7	0.8	0.8	0.8	0.8 0.8
	3.00	2.8	2.9	3.0	2.9	3.0 3.0
J. S. McCune.....	0.75	0.7	0.8	0.8	0.8	0.7 0.7
	3.00	3.0	3.0	3.0	3.0	3.0 2.8
E. J. Munch.....	0.75	0.8	0.8	0.6	0.6	0.8 0.8
	3.00	2.8	2.9	3.0	3.0	3.0 3.0
K. J. Osterhout.....	0.75	0.7	0.8	0.8	0.8	0.8 0.8
	3.00	2.8	2.7	3.0	3.0	3.0 3.0
G. W. Trainor.....	0.75	0.9	1.0	0.9	0.9	0.9 0.9
	3.00	2.8	3.0	3.0	3.0	2.9 2.9
Averages.....	0.75	0.8		0.8		0.8
	3.00	2.9		3.0		2.9

COMMENTS BY COLLABORATORS.

E. H. Berry: This method worked very satisfactorily, as it did last year. It certainly should be made official.

C. B. Gnadinger: I think the method is an excellent one, and should be made official.

K. J. Osterhout: Thinks that there may be some volatilization of oil in the case of spearmint.

CONCLUSIONS.

The results throughout are quite satisfactory. The saponification method for wintergreen yielded results which are truly remarkable in accuracy and in concordance. As to the details for determining the amount of salicylic acid separated, there seems to be very little choice between the gravimetric and volumetric procedure. The gravimetric results appear to be a little closer, and it would seem preferable to retain the method in its original form.

In the brine method for anise and nutmegs, the results for the standard extracts are excellent. Even for the dilute preparations, the reports are very satisfactory.

The carbon bisulphid method for peppermint, spearmint, and wintergreen yielded most satisfactory results.

Referring particularly to wintergreen extract, it would seem desirable to adopt both the saponification method and the bisulphid method, for the reason that the former gives the percentage of actual methyl salicylate present, while the latter gives the percentage of oil. The presence of a foreign oil, therefore, will become apparent if both methods are used. It is suggested, however, that the saponification method be described as a method for methyl salicylate.

RECOMMENDATIONS.

In view of the above results, supplementing and corroborating the outcome of last year's study on flavoring extracts, the following recommendations are respectfully submitted:

(1) That the saponification method of Hortvet and West for methyl salicylate in wintergreen extract, as described in the Journal of Industrial and Engineering Chemistry, 1909, No. 1, and slightly modified in U. S. Bureau of Chemistry Bulletin 152, page 141, by the then associate referee, R. S. Hiltner, be adopted as provisional. The method follows:

Mix 10 cc. extract in a 100 cc. beaker with 10 cc. of potassium hydroxid solution (10%). Heat on a boiling water bath until volume is reduced about one-half. Add a distinct excess of dilute hydrochloric acid, cool, and extract with three portions of ether, 40 cc., 30 cc., and 20 cc., respectively. Filter the combined ether extracts through a dry filter into a weighed dish, wash with 10 cc. ether, and evaporate spontaneously. Dry over calcium chloride in a desiccator and weigh. The weight of salicylic acid thus obtained multiplied by 9.33 gives the percentage of oil of wintergreen by volume.

(2) That the following method, devised by Hortvet and West, and described in the Journal of Industrial and Engineering Chemistry, volume 1, No. 1, be made provisional for anise and nutmeg extracts:

To 10 cc. extract in a Babcock milk flask add 1 cc. of hydrochloric acid (1 : 1), then sufficient half-saturated salt solution previously heated to 60°C. to fill the flask nearly to the neck. Cork and let stand in water at 60°C. for about 15 minutes, occasionally giving the flask a twisting motion, and centrifuge for 10 minutes at about 800 revolutions per minute. Add brine till the oil rises into the neck of the bottle, and again centrifuge for ten minutes. If the separation is not satisfactory, or the liquid is not clear, cool to about 10°C. and centrifuge for an additional ten minutes. Multiply the reading by 2 to obtain the percentage of oil by volume.

(3) That the following slight modification of the Howard-Mitchell method, which has been studied during the last two years, be now provisionally adopted for peppermint and spearmint extracts and for the determination of oil in wintergreen extract. The method follows:

Pipette 10 cc. of the extract into a Babcock milk bottle, add 1 cc. of carbon disulphid, mix thoroughly, then add 25 cc. of cold water and 1 cc. concentrated hydrochloric acid. Close the mouth of the bottle with the thumb and shake vigorously, whirl the bottle in a centrifuge for six minutes, and remove all but 3 or 4 cc. of the supernatant liquid, which should be practically clear, by means of a glass tube of small bore, and aspiration.

Connect the stem of the bottle with a filter pump, immerse the bottle in water kept at approximately 70°C. for three minutes, removing from the bath every 15 seconds and shaking vigorously. Continue in the same manner for 45 seconds, using a boiling water bath. Remove from the bath and shake while cooling.

Disconnect from the suction and fill the bottle to the neck with saturated salt solution at room temperature, centrifuge for two minutes, and read the volume of the separated oil from the top of the meniscus. Multiply the reading by 2 to obtain the percentage of oil by volume.

In the case of wintergreen, use as floating medium a mixture of 1 volume of concentrated sulphuric acid and 3 volumes of saturated sodium solution.

Since submitting the above report on flavoring extracts, two further collaborators submitted their reports. The first was by A. G. Woodman, of Boston, whose results by the methods submitted are similar to those obtained by the other collaborators, but he makes a very interesting statement regarding a nephelometric method which he has tried on very dilute extracts, and he reported his results on the dilute samples of anise and nutmeg submitted. His results were:

	OIL PRESENT	OIL FOUND
	<i>per cent</i>	<i>per cent</i>
Anise.....	0.75	1.00
Nutmeg.....	0.50	0.50

While Mr. Woodman's details were not available, a few simple tests were made along the line suggested, and it is believed that the idea is promising and that when Mr. Woodman's paper is available it may be desirable to study the method for use on very dilute preparations.

The second paper received was by C. O. Dodge, of the Bureau of Chemistry in Washington. His results on the wintergreen extracts by the saponification method, slightly modified by himself, are highly accurate and are as follows:

	OIL PRESENT	OIL FOUND
	<i>per cent</i>	<i>per cent</i>
Wintergreen extracts:		
No. 1.....	0.75	0.75
No. 2.....	3.00	2.99

Mr. Dodge suggests that the details submitted may be improved by keeping down the bulk of the solution to be extracted and using a smaller amount of ether. However, the details suggested by him involve four extractions with a total of 60 cc. of the solvent, while the original method requires only three extractions with a total of 90 cc. of ether. Inasmuch as the results obtained by the collaborators were very satisfactory, there is doubt that the changes are necessary.

Mr. Dodge raises the further point that oil of wintergreen may contain only 98% methyl salicylate, and suggests that the calculation of the result should be based on this figure. It seems, however, that this objection is overcome by the recommendation that this determination be considered a determination of methyl salicylate rather than a determination of oil.

REPORT ON BAKING POWDERS.

BY H. E. PATTEN (Bureau of Chemistry, Washington, D. C.),
Associate Referee.

Tests of three gravimetric methods for the determination of lead in phosphate baking powders were made. Five analysts coöperated. The methods studied were the Seeker-Clayton, Remington modification of the Seeker-Clayton, and Exner methods. Directions for these methods were sent out as follows:

DESCRIPTION OF METHODS.

SEEKER-CLAYTON METHOD.

Place 20 grams of the baking powder in a large beaker and add about 25 cc. of water in small portions at a time to avoid excessive frothing. Add 20 cc. of concentrated hydrochloric acid, a little at a time for the same reason, and digest on a steam bath until the solution is perfectly clear and limpid or until a drop of the solution gives no reaction for starch with iodine and potassium iodide solution. Add sufficient solution of ammonium citrate¹ (lead-free) to correspond to 20 grams of citric acid and render slightly alkaline to litmus with ammonia, density 0.95. The latter is added a little at a time, with care to keep the solution cool, to avoid precipitation of calcium salts. Dilute to about 400–450 cc., add 10 cc. of 10% hydrochloric acid, cool to room temperature, saturate with hydrogen sulphide, and allow to stand overnight.

Filter through a moistened filter, using suction if necessary, and wash the precipitate² with hydrogen sulphide water and finally with a little water. Dissolve the precipitate by passing through the filter three 5 cc. portions of boiling 10% hydrochloric acid followed by three 5 cc. portions of boiling 25% nitric acid, and collect the filtrate in a 100–150 cc. beaker. Finally wash the filter with a little hot water, add 2 cc. of concentrated sulphuric acid to the filtrate and washings, and evaporate on a hot plate until fumes of sulphuric acid are copiously evolved. The solution should now be practically colorless, but if not so add a little nitric acid and again evaporate until fumes appear. Cool, add 10 cc. of water and 20 cc. of alcohol, and allow to stand overnight. Filter through a Gooch on asbestos and wash with alcohol.

¹ *Ammonium citrate solution.*—This reagent may be prepared by dissolving 100 grams of citric acid in 100 cc. of hot water, cooling, adding a little at a time sufficient ammonia to leave a slight excess, again cooling, and then saturating with hydrogen sulphide. Allow to stand overnight, or until the sulphides have settled out, filter, boil the filtrate to expel excess of hydrogen sulphide and ammonia, cool, and make up to 200 cc. Lead-free citric acid may be used instead of this solution, but it has the disadvantage of causing a considerable evolution of heat in the subsequent neutralization with ammonia, resulting in a precipitate of calcium citrate.

² In some cases, especially when a very large amount of calcium phosphate is present, a white precipitate of calcium citrate will settle out on standing overnight. In such a case, decant the supernatant liquid through a filter and dissolve the precipitate in a small amount of dilute hydrochloric acid, add an excess of ammonium citrate, cool, render slightly alkaline with ammonia, cool, saturate with hydrogen sulphide, and allow to stand from 6 to 12 hours. The precipitated sulphides are then filtered off and treated as in the regular method. (The material remaining on the filter from the liquid first decanted has in the meantime been washed with hydrogen sulphide water and dissolved in hot hydrochloric and nitric acid, the solution being finally combined with that from the second crop of lead sulphide.)

Place the Gooch in a small beaker and treat the contents with a few drops of concentrated ammonia. Then pour 10 cc. of 50% ammonium acetate into the crucible and allow it to stand for about 15 minutes. Remove the crucible from the beaker and carefully wash the bottom and sides with water, allowing the washings to run into the beaker. Now, by placing the lips over the top of the crucible, blow the solution still remaining in the crucible into the beaker. Wash the crucible with a little water, forcing the washings through the asbestos pad in the manner just described. Rinse the bottom of the crucible with a jet of water and fit it into a bell jar arranged for filtering by suction. Filter the contents of the beaker through the Gooch, collecting the filtrate in a second beaker placed under the bell jar, and wash thoroughly with hot water. Acidify the filtrate with acetic acid, heat nearly to boiling, add an excess of potassium bichromate, and allow to stand overnight. Then filter through a tared Gooch, dry for 20-30 minutes on a hot plate, cool, and weigh as PbCrO_4 .

REMINGTON MODIFICATION OF THE SEEKER-CLAYTON METHOD.

Weigh 100 grams of the baking powder into a large beaker (1.5-2 liters) and add 500 cc. of the cold citrate solution¹ slowly with stirring. If the powder contains albumin a troublesome frothing may result, which can be controlled by the use of a few cubic centimeters of ether. Stir thoroughly several times, and set aside for several hours.

Decant the supernatant liquid, and wash the starch twice by decantation with about 50 cc. of water, adding the washings to the original solution. (A centrifuge will be of great assistance in the decantation and washing.) Heat the solution on the steam bath until the albumin is coagulated, and pass in a slow stream of hydrogen sulphid for several hours, or overnight. (If a heavy white precipitate is formed, add ammonia cautiously until it is dissolved, taking care that the solution remains acid.) Filter off the precipitated sulphids and wash with water containing hydrogen sulphid.

To the residue of starch add 250 cc. of water and 25 cc. of hydrochloric acid, and heat on the steam bath until the starch is completely hydrolized. Neutralize with ammonia, and make just acid to litmus with hydrochloric acid. Pass in a slow stream of hydrogen sulphid for several hours, or overnight. Filter and wash with water containing hydrogen sulphid.

Place the filters containing both precipitates in a tall 100 cc. beaker, add 5 cc. of sulphuric acid and 10 cc. of nitric acid, and heat on the hot plate, with frequent additions of nitric acid until all organic matter is destroyed and the liquid is nearly colorless. Take down to fumes of sulphuric acid, cool, add 20 cc. of water and 40 cc. of alcohol, and allow to stand overnight.

From this point proceed as in the original Seeker-Clayton method, as follows:

Filter through a Gooch on asbestos, and wash with alcohol. Place the Gooch in the original beaker and moisten the contents with a few drops of concentrated ammonia. Then pour 10 cc. of 50% ammonium acetate solution (previously treated with H_2S and boiled) into the crucible and allow it to stand for about fifteen minutes. Remove the crucible from the beaker and carefully wash the bottom and sides with water, allowing the washings to run into the beaker. Now, by placing the lips over the top of the crucible, blow the solution still remaining in the crucible

¹Preparation of ammonium citrate reagent.—Dissolve 200 grams of ammonium citrate in a liter of water, add 10 cc. of concentrated hydrochloric acid, warm slightly, and pass in a slow stream of hydrogen sulphid for several hours. Filter and heat to boiling to expel excess of H_2S .

into the beaker. Wash the crucible with a little water, forcing the washings through the asbestos pad in the manner just described. Rinse the bottom of the crucible with a jet of water, and fit it into a bell glass fitted for filtering with suction. Pass the ammonium acetate solution through the Gooch, filtering twice if necessary to secure a perfectly clear filtrate, and wash thoroughly with a little hot water. Acidify the filtrate with acetic acid, heat nearly to boiling, add an excess of potassium dichromate, and allow to stand overnight. Filter on a small tared Gooch, wash the precipitated lead chromate with cold water, dry the crucible and contents by heating for 20-30 minutes on a hot plate, cool, and weigh as PbCrO_4 .

EXNER METHOD.

Weigh out 200 grams¹ of the sample and transfer this to a 3-liter Jena flask. Add 300 cc. concentrated nitric acid in portions, with thorough shaking after each addition. The mixture, which first forms a thick paste, is slowly heated on an asbestos gauze with repeated shaking. The mass becomes thinner, and by the time the oxidation of the starch begins it is quite fluid. As soon as brown fumes begin to appear at the mouth of the flask, heat is removed and a stemless funnel is inserted in the neck of the flask. The reaction soon becomes very vigorous, and the flask should be set in a hood with a good draft. The reaction, however, will not become so violent as to cause loss of material by foaming over. When the action has moderated, the flask is set back on the asbestos gauze over a moderate Bunsen flame. When the action becomes weak, as shown by the fumes in the flask becoming lighter, 90 cc. concentrated sulphuric acid is slowly added to the contents of the flask, and heating is again continued until the fumes fade. Then 25 cc. of concentrated nitric acid are added from time to time, with continued heating, until all the starch is completely oxidized. This can be told from the behavior of the last addition of nitric acid, which boils out with but little decomposition. Usually three or four additions of 25 cc. portions of nitric acid suffice. The nitric acid should finally be expelled as completely as possible without endangering the flask. Cool and add 400 cc. distilled water, shake, and allow to settle. The soluble sulphates of sodium, potassium, aluminum, iron, etc., go into solution, while calcium sulphate and most of the lead sulphate will be precipitated. Filter through an 18 cm. folded filter into a liter Erlenmeyer flask; rinse the 3-liter flask two or three times with small portions of water and pour the rinsings through the filter. There is no need of a thorough washing of the precipitate. The latter is transferred to a 2-liter Erlenmeyer flask. This is best done by opening the filter containing the precipitate over a 600 cc. beaker and rinsing thoroughly with water from a wash bottle. The contents of the beaker are then transferred to the Erlenmeyer, together with whatever precipitate remains in the 3-liter flask. The contents are then diluted so as to nearly fill the flask, stirred thoroughly to dissolve the calcium sulphate, 20 cc. strong acetic acid added, and the liquid thoroughly saturated with hydrogen sulphid. This may be done quickly by preparing a solution of sodium sulphid by saturating a 5% sodium hydroxid solution completely with hydrogen sulphid, drawing up this liquid into a pipette, which is then dipped into the solution in the Erlenmeyer and stirred while the sulphid runs out. Only a small part of the free acid need be neutralized to produce complete saturation with hydrogen sulphid. This procedure saves both time and gas. The flask is then corked and set aside until the precipitate has settled. The liquid is then siphoned off, saving much time in filtering. The lower end of the siphon tube which is introduced into the liquid has a short bend at right angles so

¹ If other weights have to be used, take reagents in proportion.

that it will not suck the precipitate from the bottom. When there is much calcium sulphate, one such treatment will not suffice to bring all into solution, but the flask is refilled with distilled water, is again acidified and saturated with hydrogen sulphid and allowed to settle till the calcium sulphate is practically all dissolved and the residue of sulphids is dark colored. When much calcium sulphate is present, solution may be hastened by the addition of lead-free sodium acetate to the water, 50-75 grams to each 2 liters of water. If the solution of the salt gives no darkening with hydrogen sulphid, it may be safely used.

The liquid containing the soluble sulphates is treated separately to recover the trace of lead which it may contain. The acidity is partially neutralized with ammonium hydroxid just short of the point of producing a permanent precipitate of aluminum phosphate. It is then saturated with hydrogen sulphid and the precipitate allowed to settle. Some iron sulphid will usually be precipitated also. It is important that the sulphid precipitations be made in very slightly acid solutions, otherwise lead sulphid will not be completely precipitated. When both precipitates have finally settled, the liquids may be siphoned off, and the precipitates transferred on to separate 11 cm. filters and washed with hydrogen sulphid water. The filters with their precipitates are placed into a 200 cc. Erlenmeyer flask, 10 cc. concentrated nitric acid and 5 cc. concentrated sulphuric acid are added, a stemless funnel inserted in the neck, and the flask heated to completely oxidize the material. When the nitric acid has all been expelled and the residue darkens, more nitric acid is added until no such darkening occurs. The residue is finally heated till fumes of sulphur trioxide are given off. Then cool and add 15 cc. of water. Filter through 7 cm. filter, rinse, and then wash the filter twice with small portions of dilute sulphuric acid, and finally with a little water. There is no need of removing all sulphuric acid. Place a clean 150 cc. beaker under the filter, dissolve the precipitate with 15-25 cc. ammonium acetate and wash thoroughly with water. The ammonium acetate is best prepared by mixing 1 part 99% acetic acid, 1 part water, and 1 part ammonium hydroxid sp. gr. 0.90, introducing a piece of litmus paper, and cautiously adding more ammonium hydroxid until neutral.

The solution of lead in ammonium acetate is acidified with acetic acid and precipitated with potassium bichromate. The mixture is heated on the steam bath and then allowed to cool and settle. It is filtered on to a tared Gooch having a thick felt, washed with water, dried at about 125°C., and weighed as lead chromate.

It is taken as a matter of course that the water used and the reagents must be free from lead. There seems to be no danger from lead in vessels when Jena glass is used.

SAMPLE SOLUTIONS.

In view of the lack of confidence expressed by many analysts with whom we consulted in the Seeker-Clayton method as originally given, it was decided to send out for a preliminary test samples containing the lead in a soluble form. A very slight amount of organic material was added to necessitate wet oxidation as recommended by the methods. Some of the collaborators very properly objected on the ground that these samples did not offer the same analytical difficulties met with in the analysis of phosphate baking powders. They have, nevertheless, worked faithfully upon the samples and have sent in data which, in the judgment of the associate referee, fully justify the style of sample used

for this preliminary test of the comparative merits of the methods. Further, to eliminate personal equations and any autosuggestion in the analysts, three sample solutions labeled A, B, and C were sent out, each containing exactly the same concentration of lead (as $\text{Pb}(\text{NO}_3)_2$), but without indicating to the analysts that these solutions were duplicates one of the other so far as the lead was concerned. The ingredients of the solutions are given below.

It will be noted that the diluted solutions made from these samples which were finally to be taken for the lead tests contained a quantity of lead the same as would be found in a baking powder containing 20 parts of lead per million. Thus, 200 grams of baking powder (Exner method) containing 20 parts per million of lead would have 4 mg. of lead. One hundred grams (Remington method) would contain 2 mg. of lead, and 20 grams (Seeker-Clayton method) would contain 0.4 mg. The solutions measured out for analysis contained these respective amounts.

INGREDIENTS OF SOLUTIONS.

Solution A contained 10 grams of sugar per 100 cc. and 40 mg. of lead per 100 cc. (as $\text{Pb}(\text{NO}_3)_2$), with a few drops of concentrated nitric acid to repress hydrolysis.

Solution B was an exact duplicate of solution A and was taken from the same mix.

Solution C was identical with solutions A and B except that 2.5 grams $\text{CaH}_4(\text{PO}_4)_2 \cdot \text{H}_2\text{O}$ (found to be lead free) was added per 100 cc.

DIRECTIONS ACCOMPANYING SAMPLE SOLUTIONS.

The following directions were sent out with the solutions:

Take 50 cc. of each solution at 25°C. and make up with distilled water to 500 cc. at 25°C. Then for use in the Exner method use 100 cc. of this diluted solution; for the Remington method use 50 cc.; and for the Seeker-Clayton method use 10 cc. The results should be reported in milligrams per liter of original solution.

REPORTS OF COLLABORATORS.

The collaborators reported results which are tabulated as milligrams per liter of original sample solution in Table 1. These results have been recalculated to a basis of parts per million of a phosphate baking powder, using the weights of baking powder recommended for analysis in each of the methods, respectively, and are given in Table 2.

One of the collaborators, Dr. T. J. Bryan, in addition to the requested work, added to the diluted solutions to be taken for analysis an amount of lead-free baking powder required by the different methods and then

TABLE 1.

Lead in milligrams per liter of original solution as reported by the collaborators.

[Solutions contained 400 mg. of lead per liter.]

COLLABORATOR AND ANALYST	LEAD IN MILLIGRAMS PER LITER OF SOLUTION								
	Solution A			Solution B			Solution C		
	Seeker-Clayton	Remington	Exner	Seeker-Clayton	Remington	Exner	Seeker-Clayton	Remington	Exner
Loomis (Wichman)...	128	333	333	128	307	358	128	422	416
	1220	640	870	640	500	370	380	460	420
Hortvet (Pettijohn)...	2750	990	700	380	450	210
				700					
Morey (Hoyt).....	260	550	359	380	360	436	260	396	378
	380	475	372	320	422	429	190	410	359
	384	371	454	833	358	211	1020	269	358
Bryan.....	897	294	371	512	179	237	1020	294	397
	513	436	442	513	410	410	448	576	481
Stallings (Clarke)....	448	422	416	448	422	384	513	512	512
	385	448	577
				422					

TABLE 2.

Recalculation of results given in Table 1 to a basis¹ of parts per million in a phosphate baking powder.

[Solutions contained 20 parts of lead per million.]

COLLABORATOR AND ANALYST	LEAD IN PARTS PER MILLION								
	Solution A			Solution B			Solution C		
	Seeker-Clayton	Remington	Exner	Seeker-Clayton	Remington	Exner	Seeker-Clayton	Remington	Exner
Loomis (Wichman)...	6	17	17	6	15	18	6	21	21
	61	32	44	32	25	19	19	23	21
Hortvet (Pettijohn)...	138	50	35	19	22	11
				35					
Morey (Hoyt).....	13	28	18	19	18	22	13	20	19
	19	24	19	16	21	21	10	21	18
	19	19	23	42	18	11	51	13	18
Bryan.....	45	15	19	26	9	12	51	15	19
	26	22	22	26	21	21	22	29	24
Stallings (Clarke)....	22	21	21	22	21	19	26	26	26
	19	22	22	29
				21					

¹ For synthesis of samples see page 213.

TABLE 3.

Lead found in solutions to which lead-free baking powder was added in the proportions required by the various methods.

[Solutions contained 400 mg. of lead per liter.]

METHOD	LEAD IN MILLIGRAMS PER LITER OF SOLUTION		
	Solution A	Solution B	Solution C
Seeker-Clayton.....	704	320	384
	448	192	192
Remington.....	410	179	102
	397	38	140
	134	121	313
Exner.....	281	294	

subjected the solutions to analysis. These results are shown in Table 3. Table 4 contains the results given in Table 3 recalculated to parts per million of lead in a phosphate baking powder in order to make them comparable with similar results given in Table 2.

TABLE 4.

Recalculation of results of Table 3 to a basis of parts per million in the phosphate baking powder added.

[Solutions contained 20 parts of lead per million.]

METHOD	LEAD IN PARTS PER MILLION		
	Solution A	Solution B	Solution C
Seeker-Clayton.....	35	16	19
	22	10	10
Remington.....	22	9	5
	20	2	7
Exner.....	7	6	16
	14	15	

DISCUSSION OF RESULTS.

Recurring now to the objections which have been made by some of the collaborators (H. M. Loomis, T. J. Bryan, C. B. Morey): They were of the opinion that these methods for the estimation of lead should be used upon lead-free baking-powder samples to which known quantities of lead had been added, or upon sample solutions containing the same quantity and ratio of ingredients that would occur upon solution of a phosphate baking powder in acid.

These methods have two objects: One, to determine the lead quantitatively, and the other to separate the lead from the mixture in which it is found. It seemed advisable to study first their respective merits in determining lead under conditions where there was practically no interference from baking-powder ingredients. If a method can not stand this test, manifestly there is no use of trying it out where there is added the further difficulty of separating the lead from a complex mixture of ingredients.

The reason that the associate referee sent out these very simple samples for analysis was to ascertain if these three methods could give comparable results in the hands of capable analysts under the most simple conditions. If this had proved to be the case, it was then the intention to carry out further work upon more complicated samples fully simulating the actual conditions found in phosphate baking powders. Owing to the lateness of the date at which the present associate referee took up this work, it has been possible to proceed only as far as the report indicates.

In the work done by Mr. Pettijohn, under Dr. Hortvet, the prescribed

methods were not followed exactly in all cases; and for this reason, together with the wide variation in results presented, it seems fair to judge the methods on the results reported by the other four analysts.

SEEKER-CLAYTON METHOD.

VARIATIONS REPORTED.

The Seeker-Clayton method might possibly be adapted for this work by substituting a colorimetric reading at the end instead of the present gravimetric determination, but in its present form it gives results varying from (Table 2) 6 parts per million to 51 parts per million, using one solution with four different analysts, and from 19 parts per million to 51 parts per million for one analyst on three solutions (Bryan).

WEIGHING LIMITATION ERROR.

The inherent difficulty with the Seeker-Clayton method is the small amount of sample taken. With the baking-powder standard for lead set at 20 parts per million upper limit, there will be 0.0004 gram of lead in a 20-gram sample containing this highest limit. Since the lead is weighed as PbCrO_4 , the actual mass weighed would be 0.000624 gram. Assuming that the balance weighs accurately 0.0001 gram, then any weight between 0.00055 gram and 0.00065 gram will be read as 0.0006 gram of PbCrO_4 . This weight calculated to lead gives 0.000385 gram lead, with a range from 0.000353 gram to 0.000417. Calculating to parts per million of lead, 0.0006 gram PbCrO_4 represents 19.25 parts per million, and the range is 3.2 parts per million (from 17.65 to 20.85). Hence we have a possible error of $\pm 8.3\%$ inherent in the limitations of the balance when estimating lead in baking powders containing approximately 20 parts per million, even when the balance weighs accurately to 0.0001 gram. The above standard referred to (20 parts per million of lead) is for baking-powder acid-ingredient chemicals. The standard for complete baking powders is one-half of this, or 10 parts per million. A baking powder containing this upper limit would yield only 0.0003 gram PbCrO_4 and, the weighing errors remaining the same, the percentage of error due to the limitations of the balance would be doubled.

WEIGHT STANDARDIZATION ERROR.

The above discussion of weighing errors assumes that the set of metric weights used with the balance is rigidly standardized, and that each correction of standardization is used throughout all operations. To illustrate how great such an error may be, test 11388, U. S. Bureau of Standards, of a new set of weights (Becker's Sons, Rotterdam) showed the following: One of the 2-gram weights was too heavy by 0.1 mg., each 100-mg. weight was 0.04 mg. too light, and the 50-mg. weight was 0.02

mg. too light. Thus, a weighing in which one of the 2-gram weights was replaced by the other would show 0.0001 gram more or less than the correct mass. Similarly, a weighing in which the two 100-mg. weights and the 50-mg. weights were used together would lack 0.0001 gram of the true mass. Thus the possible error through neglecting the standardization of weights would be as large as the above mentioned error due to the limitations of the balance. And even when weight standardization is taken care of, constant use and corrosion of weights may introduce error.

OPTIMUM LOAD.

In making such close estimates as required by the Seeker-Clayton method, the effect of the balance load on the sensibility must be taken into account. It would, of course, be advisable to determine the load of greatest sensibility, and then add weights to the pan containing the Gooch crucible until this optimum load is approximated; or a lighter crucible might be chosen in case the optimum load were lower.

DRYING ERROR.

The method of drying the PbCrO_4 precipitate on the hot plate according to the Seeker-Clayton directions has been found to be subject to an error of 0.0002 gram by Dr. T. J. Bryan in his report, although he does not specify exact reasons for this error. Mr. Pettijohn also has criticized this procedure, but without quantitative expression. This error may be partially due to absorption of moisture during the weighing or in the balance case.

RECOMMENDATIONS ON SEEKER-CLAYTON METHOD.

The Seeker-Clayton method, because of the small quantity of sample and reagents required, may well serve as a semiquantitative method in the hands of one skilled in its use; but taking into account all of the factors mentioned, it would seem inadvisable to recommend the Seeker-Clayton method for further study.

REMINGTON MODIFICATION.

VARIATIONS REPORTED.

The Remington modification method showed much closer checking than the Seeker-Clayton, the greatest variation being from 13 to 29 parts per million with one solution and four different analysts, and from 9 to 19 parts per million with three solutions for one analyst (Bryan).

The majority of determinations, however, came reasonably close to the true value of 20 parts per million. Out of a total of 23 determinations by four analysts, over half (13) were within 10% of the correct value. Fifteen were within 20%, and four showed an error greater than 30%.

WEIGHING ERRORS.

With this method the larger weight of sample taken (100 grams) reduces the error in weighing due to the limitations of the balance to ± 0.32 parts per million when a total of 20 parts per million is being estimated. Other weighing errors are correspondingly reduced. An error of 0.0001 gram in weighing the PbCrO_4 would produce an error of 0.64 parts per million with the 20 parts per million total.

RECOMMENDATIONS ON REMINGTON MODIFICATION.

From the results obtained with this method, it would appear that the Remington method does not possess quite the accuracy of the Exner method. Consequently it is recommended that no further study be made of the Remington modification method at present.

EXNER METHOD.

VARIATIONS REPORTED.

Still closer checking was found in the results with the Exner method. The greatest variation was from 11 to 22 parts per million with one solution and four analysts, and from 11 to 23 parts per million with one analyst (Bryan), using three solutions. Fifteen out of a total of 21 determinations were within 10% of 20 parts per million, and only three showed an error greater than 20%.

WEIGHING ERRORS.

With the Exner method weighing errors are still further reduced, the error due to weighing limitations of the balance being only ± 0.16 parts per million in estimating 20 parts per million of lead. An error of 0.0001 gram in weighing the lead chromate precipitate would give an error of 0.32 parts per million for this 20 parts per million total.

USE OF ALCOHOL IN PRECIPITATING LEAD SULPHATE.

Mr. Pettijohn recommends that alcohol be added in the Exner method, at the point where the PbSO_4 (recently formed from the sulphid by the action of nitric and sulphuric acids) is treated with water, in order to precipitate the lead sulphate completely. It appears to the associate referee that this would delay the determination and not add appreciably to the accuracy. The solubility of PbSO_4 in water is 0.0042 gram PbSO_4 per 100 grams distilled water at 19°C . (Rothmund: Löslichkeit und Löslichkeitsbeeinflussung). This means that when *saturated*, 15 cc. of water would hold only 0.0006 gram of PbSO_4 . The 15 cc. of water used in transferring the sulphate precipitate does not have time to reach saturation; hence the use of alcohol to recover this minute quantity of PbSO_4

is without material effect upon the accuracy of the method in general, if carried out according to Exner's directions. However, where it is desired to use the Exner method with fractional quantities of the 200-gram sample, the error introduced by the loss of the PbSO_4 in the transference water might need consideration.

RECOMMENDATIONS ON THE EXNER METHOD.

In view of the shorter time required and the greater accuracy both from a priori considerations and actual experimental data, the Exner method seems well worthy of further study, where a gravimetric method is to be used. In case the full quantity recommended (200 grams) is used, it is advisable to take the substance from the can, thoroughly mix, and secure each complete sample taken for analysis as a composite of many fractions lifted from different portions of the mix.

WICHMANN MODIFICATION OF SEEKER-CLAYTON METHOD.

Mr. H. J. Wichmann in his report presents the following method, which he recommends as dependable:

One hundred grams of baking powder are weighed out into a 1.3-liter beaker and 10% HCl added to excess in small portions. Excessive frothing may be kept down with ether. The mixture of acid and starch is heated until the starch is hydrolyzed and the solution is quite limpid. After cooling, 200 cc. 50% ammonium citrate solution are added. The solution is kept cold while an excess of ammonia is slowly and carefully added. If a precipitate forms, sufficient ammonium citrate is added to dissolve it. Fifteen cubic centimeters of saturated HgCl_2 solution are added and the solution diluted to about 1,200 cc. Hydrogen sulphid is then passed to saturation and the beaker set aside. The precipitate settles rapidly and can readily be filtered. A centrifuge can be used to advantage. There is no need of standing overnight, as the heavy HgS will drag down the PbS . If the solution stands overnight a precipitate of calcium citrate is apt to settle out.

The filter paper with the sulphids is placed in a small casserole, or the sulphids are washed into it from the cylinder, if a centrifuge is used. Ten cubic centimeters of HNO_3 and 2 cc. of concentrated H_2SO_4 are added. The nitric acid is evaporated and the sulphuric acid slowly fumed off. If a gas muffle furnace is available the casserole can be placed on the platform in front of the muffle until the H_2SO_4 has disappeared and then slowly heated inside to a light red heat. The mercury salts will vaporize, and any ferric sulphate present will be broken up to the oxid. After cooling, the lead sulphate is leached out from the residue with an ammoniacal 25% solution of ammonium acetate. Several leachings must be made. The filter paper is then washed with hot water. The filtrate is acidified with acetic acid and the lead precipitated as chromate with potassium dichromate. After standing overnight the lead chromate is collected on a Gooch, dried by heating to 125°C ., cooled, and weighed.

In answer to the objection that the mercury would tend to carry off some of the lead with it during volatilization, Mr. Wichmann assures

us that he has not found this error to interfere with the accuracy of the method. Mr. Clayton states that in his experience such a volatilization of mercury salts gave low results for the lead; but considering the very much smaller quantities of lead present in each test sample when following the Seeker-Clayton method, it seems likely that the large relative error obtained by Mr. Clayton would become insignificant when using 100-gram samples as recommended by Mr. Wichmann.

RECOMMENDATIONS.

The associate referee recommends that further study be carried out upon the Exner and Wichmann methods. The Exner method is recommended for adoption as a provisional method.

Special emphasis should be laid upon speed of operations, as well as upon clean separations, close measurement of wash water, and accurate weighing. The use of a stirrer during hydrogen sulphid precipitation, followed by centrifuging, should be studied with a view to eliminating the standing overnight. The Wichmann method presents the only gravimetric determination which can be carried out within approximately 24 hours, and is therefore especially commended for careful study. It is recommended that the determinations be accompanied by a time schedule, showing actual time involved in each operation. This is especially important in view of the large number of samples and the short time available in handling many carloads of material at the factories.

The associate referee wishes to thank the collaborators, T. J. Bryan, J. Hortvet, H. M. Loomis, C. B. Morey, and R. E. Stallings, and the analysts, J. O. Clarke, F. L. Hoyt, E. Pettijohn, and H. J. Wichmann, for their helpful spirit in carrying out the work. Acknowledgment is also made to Mr. G. H. Mains for assistance in discussing data and compiling this report.

THE RÔLE OF CALCIUM SULPHATE IN PHOSPHATE BAKING POWDERS.

(ABSTRACT.)

By H. E. PATTEN (Bureau of Chemistry, Washington, D. C.).

The present investigation was undertaken in view of the different opinions which have been expressed as to the function of calcium sulphate in phosphate baking powders. Some chemists hold that the calcium sulphate is merely a neutral inert body acting possibly as a drying agent in the baking powder, but contributing nothing to the chemical changes which alone render the powder of service in baking. Others maintain

that the calcium sulphate acts chemically in the dough, and finally is not found in its original form in the finished biscuit, but broken down chemically and recombined into a soluble product (sodium sulphate) and an insoluble product [calcium phosphate $(\text{CaO})_x(\text{P}_2\text{O}_5)_y$] of variable composition. The presence of calcium sulphate is also held to retard the evolution of gas by the baking powder and thus renders the leavening action more gradual.

There is also question in the minds of some as to whether the reaction between sodium bicarbonate and monocalcium phosphate yields dicalcium phosphate (CaHPO_4) or tricalcium phosphate $\text{Ca}_3(\text{PO}_4)_2$; and also question as to the effect of flour and starch (in the dough of the biscuit) upon the reaction. Indeed some claim that experiments made with sodium bicarbonate, calcium phosphate, calcium sulphate, and water cannot be taken as good evidence of what reactions will take place when these baking-powder ingredients are actually mixed with flour and water and baked into a finished biscuit.

A survey of the literature at the time this work was undertaken¹ showed that accurately controlled heterogeneous equilibria studies had been carried out in which carbonates were investigated as to their action with sulphate and chlorid, but not with phosphate; also that phosphates had been studied in connection with sulphate and chlorid, but not with carbonate. Consequently, there was a large field of reaction possibility still unexplored, even where the baking-powder ingredients alone were taken in aqueous solution, no flour being added.

From these equilibria experiments, and the diagrams graphically representing their data, tentative inferences were made as to the chemical changes which might be expected in the baking reaction, as follows (using symbols for brevity):

If $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$ or CaSO_4 anhydrous be added to NaHCO_3 in presence of excess of water—²

(1) CaCO_3 will be formed (since it is about 1,000 times less soluble than CaSO_4) along with Na_2SO_4 and H_2CO_3 .

(2) The H_2CO_3 will then decompose and CO_2 gas will be evolved.

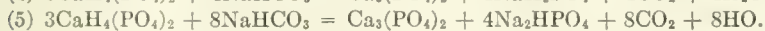
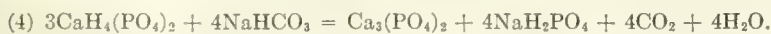
(3) The main effect of rise in temperature above 25°C . is to liberate CO_2 held in solution by CaCO_3 ,³ and to deposit the CaCO_3 (released by the CO_2) as a precipitate; also temperature rise changes any residual $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$ to the anhydrous form, CaSO_4 .

¹ This problem was referred to the author in 1911, some time in November, by Dr. W. D. Bigelow, at the instance of Dr. H. W. Wiley. A preliminary report was submitted to Dr. Bigelow on January 15, 1912.

² Compare U. S. Patent 316,863, April 28, 1885. CO_2 formed from $\text{CaSO}_4 +$ alkaline bicarbonates.

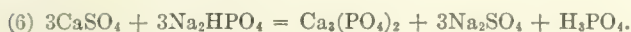
³ $\text{CaH}_2(\text{CO}_3)_2$ calcium bicarbonate has not been shown to exist, despite the frequent references to it in chemical writings.

If $\text{CaH}_4(\text{PO}_4)_2$ be added to NaHCO_3 in water the reaction varies with the quantity of NaHCO_3 used, thus:



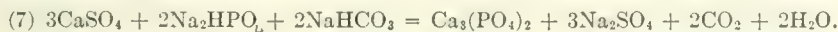
It is found that "tricalcium phosphate" is always formed and the variation of NaHCO_3 produces a variation in NaH_2PO_4 and Na_2HPO_4 and CO_2 and H_2O .

If we add CaSO_4 or $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$ to $\text{CaH}_4(\text{PO}_4)_2$ and NaHCO_3 in water, the reaction is modified in that an end-product, Na_2HPO_4 , is acted upon, thus:



The tricalcium phosphate (a solid solution $(\text{CaO})_x(\text{P}_2\text{O}_5)_y$ is about 1,000 times less soluble in water than CaSO_4 , hence this reaction proceeds till the solubility limit of the $\text{Ca}_3(\text{PO}_4)_2$ in the acid solution resulting is reached. This equation, of course, does not represent the reality, since the composition of the solid solution ($\text{Ca}_3(\text{PO}_4)_2$?) varies with the composition of the liquid phase in contact with it.

Reaction No. 6 above does not take account of the NaHCO_3 present. When this is done we have:



No hint is given in the literature examined of a compound, $\text{CaNa}_4(\text{PO}_4)_2$, either as a solid phase or in solution.

Without regard to exact equations, such as 4, 5, 6, and 7, it may be inferred from the published isotherms referred to in the literature that—

(8) With calcium sulphate present in excess as a solid phase, very little phosphorus pentoxid will be found in solution.

(9) If the content of CaSO_4 is high in the baking powder it is likely that CaSO_4 or $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$ will exist as a solid phase after the baking powder reaction is ended, and some CaCO_3 may be present. But if the CaSO_4 content is low in the baking powder, then there would be a solid solution left as residue after baking, containing CaO and P_2O_5 , but not necessarily in a simple molecular ratio to each other; here, too, some CaCO_3 might be left in the solid residue.

(10) Tricalcium phosphate is not permanent or of fixed chemical composition, at least when it is formed in the usual way by precipitation; the P_2O_5 may be washed from it with excess of water, leaving a basic compound, or, conversely, its lime may be extracted by acid solutions, leaving the solid solution relatively higher in P_2O_5 content than in CaO .

(11) The effect of adding salt (sodium chlorid, NaCl) in mixing dough probably is to decrease the P_2O_5 in solution. This effect is observed where chlorids act on calcium phosphates alone in aqueous solution.¹

¹ U. S. Dept. Agr., Bur. Soils Bul. 41, p. 34. KCl decomposes $\text{Ca}_3(\text{PO}_4)_2$ and decreases P_2O_5 in solution.

(12) Precipitation of calcium carbonate in the cold is retarded by the presence of phosphate salts.¹ This would tend, though perhaps only slightly, to hold more CO₂ in solution and give it up at higher temperature in the oven.

The double titration of phosphates, using methyl orange and phenolphthalein and a neutral salt such as calcium chlorid, is outlined in detail by H. N. Morse in his Analytical Chemistry, and has long been a matter of common knowledge. Wadman² in 1894 published results on testing phosphate baking powders, using essentially this method.

Although the foregoing review of the subject makes it appear that calcium sulphate is not inert in the baking-powder reaction, a few experiments have been carried out to ascertain the effect of the calcium sulphate, not only in quantitative experiments with the chemicals and water but also in baking tests.

The following table shows the amount of calcium sulphate not decomposed when varying amounts were used in the baking-powder reaction in flasks with given amounts of monocalcium phosphate and sodium bicarbonate.

Quantity of calcium sulphate left in solid residue after baking-powder reaction.¹

CALCIUM SULPHATE ADDED		CALCIUM SULPHATE LEFT IN SOLID RESIDUE
Grams	Per cent by weight of CaH ₄ (PO ₄) ₂ ·H ₂ O	Gram
0.378	15	0
0.756	30	0.054
1.009	40	0.080
1.512	60	0.175
2.018	80	0.545

¹ Reaction: 2.5215 grams 3CaH₄(PO₄)₂·H₂O + 2.2460 grams 8 NaHCO₃ + (Q.) CaSO₄ + 100 grams H₂O.

It appears that with the proportions of monocalcium phosphate and sodium bicarbonate used, the calcium sulphate was entirely decomposed when its amount as CaSO₄ was 15% of the weight of the phosphate (CaH₄(PO₄)₂·H₂O).

In regard to the effect of CaSO₄ upon the production of carbon dioxide, it may be said that since both calcium sulphate and monocalcium phosphate react with sodium bicarbonate, giving off carbon dioxide, when mixed they will still react, as shown in equations 4, 5, 6, and 7 above, which in general are borne out in our experiments, results subject, of course, to variations produced by change in concentrations and temperatures.

Further, a monocalcium phosphate which has been neutralized to phenolphthalein by addition of N/10 sodium hydroxid shows a marked

¹ Storer, Am. J. Sci., 1858, **35**: (2) 41; Dulong, Ann. Chim., 1812, **82**: 273; Spiller, J. Chem. Soc., 1858, **10**: 110; Kippenberger, Z. anorg. Chem., 1894, **6**: 177.

² J. Am. Chem. Soc., 1894, **16**: 333.

increase in acidity when calcium sulphate is added. This excess acidity may be titrated with standard alkali, or it may be determined quantitatively by the electrometric method, using the hydrogen electrode to evaluate the increase in the so-called "hydrogen ion concentration." Both methods were used in this investigation.

The electrometric method has the advantage that it does not disturb the equilibrium as does the addition of alkali in titrating.

Baking experiments conducted in coöperation with Miss Hannah Wessling confirmed the conclusions from the chemical studies. Addition of calcium sulphate appreciably raised the limit of alkali which could be used with a given amount of phosphate in the baking-powder mixture without leaving an alkaline taste in the biscuit.

The experiments reported at this time have settled only the question as to whether the calcium sulphate does take part in the reaction. The exact limits of the reaction are yet to be determined.

REPORT ON MEAT AND FISH.

By E. D. CLARK (Food Research Laboratory, Philadelphia, Pa.),
Associate Referee.

Studies this year are included in two groups. The first was to determine the accuracy and general application of the Price method for starch in meat products by the analysis of known samples through the aid of the collaborators. The second group of studies included methods of detecting incipient decomposition and changes in fish flesh by determining the various forms of nitrogen in the so-called "nitrogen distribution," the changes in fat constants, etc.

PRICE METHOD FOR STARCH IN MEAT PRODUCTS.

For the last two years it has been recommended by the association that Price's method be studied further, with a view to its adoption as provisional. It seemed desirable to make a final study of this method before recommending its adoption. Previous work reported on the method shows that the directions as given are easily carried out and that theoretical values are obtained for known weights of starch added to meat. Accordingly, the immediate problem seemed to be a study of the uniformity of results obtained on the same meat sample by different analysts, rather than any further investigation of theoretical considerations.

In collaborative work of this sort on starch in meat it is impossible to prepare uniform and exact samples because of the difficulty of thoroughly mixing dry starch or cereal with a wet and greasy material like sausage meat. Accordingly, two large samples of ground meat were prepared,

using borax as a preservative. To each of these samples potato starch was added and the whole thoroughly mixed by running through a meat chopper ten times in succession. Allowing for the moisture, etc., in the starch, it was possible to prepare fairly uniform samples of chopped meat containing an approximately known weight of starch. Portions of the large samples were analyzed for starch, and then 4-ounce samples were sent out to collaborators.

In the original Price method, reduced copper is determined by titration, as proposed by Low. However, in many laboratories it is customary to determine the precipitated cuprous oxid by weighing directly, according to Munson and Walker's procedure. Therefore, in order to get comparative figures, the collaborators were requested to determine the starch in the second sample by the use of both the volumetric and gravimetric methods for reduced copper.

Results of collaborative work on the Price method for starch in meat products.

ANALYST	AVERAGES OF TWO OR MORE DETERMINATIONS			
	Sample 1 (\pm 1.30% starch)		Sample 2 (\pm 1.65% starch)	
	Volumetric	Gravimetric	Volumetric	Gravimetric
	per cent	per cent	per cent	per cent
L. H. Almy, Philadelphia.....	1.25	1.23	1.67	1.69
E. D. Clark, Philadelphia.....	1.22	1.42	1.45
R. D. Cook, Chicago.....	1.24	1.44
M. O. Johnson, Washington.....	1.33
T. R. Le Compte, Washington.....	1.67	1.65
R. M. Mehurin, Washington.....	1.57	1.53
C. G. Sutton, St. Paul.....	1.11	1.11	1.42	1.47
A. S. Thatcher, Washington.....	1.36
G. W. Trainor, Chicago.....	1.28	1.74
Average.....	1.26	1.17	1.56	1.56

A study of the collaborators' figures shows that they are in satisfactory agreement with each other and with the approximately known weight of starch added in making up the sample. The Price method has an accuracy and an ease of manipulation that make it worthy of adoption as a provisional method. To accommodate those analysts who prefer to determine the copper gravimetrically as cuprous oxid, rather than by titrating according to Low's method, the Price method should be modified to permit the weighing of cuprous oxid as an optional procedure. From our experience, and that of our collaborators, it seems that this method should be adopted as provisional in place of that of Mayrhofer.

PRELIMINARY STUDIES ON CHEMICAL METHODS OF DETECTING
DETERIORATION IN FISH FLESH.

BY E. D. CLARK AND L. H. ALMY.

The quantitative study of changes in fish flesh upon standing or during cold storage is a comparatively new one. The researches of Richardson and Scherubel, Emmett and Grindley, and others, upon meat during storage opened the way in that field. Wiley, Pennington, and collaborators, have done the fundamental work on the chemistry of decomposition processes in the flesh of poultry, game, etc. The investigations of Smith¹ and of Perlzweig and Gies² on changes in fish during cold storage are the pioneer contributions to our knowledge of the behavior of fish flesh under refrigeration.

Owing to the fact that there were few generally accepted methods for studying chemical changes in fish flesh, and also that the perishable nature of fresh fish made it impossible to send samples to collaborators, it was decided to make the preliminary studies in our own laboratory. This work was done in connection with the investigations of the laboratory on the effects of cold storage upon fish. Therefore, this report is a result of the year's experience with various methods applied to the study of fish.

The following list shows the determinations made for the tabulation of results on changes in the fish during cold storage. It was intended to include every determination that offered any possibility of giving information about fish flesh and its changes. Some of these determinations are unnecessary and some may be of little value, but all of them will help to give definite information about the adequacy of the methods themselves as applied to this problem. Most of the determinations have been adopted from the work of early investigators upon meats and poultry. The determination of the acidity of the fish extract was suggested by Smith's publication. Determination of amino acids by both the Van Slyke and Sørensen methods was a new departure. Other new methods or modifications will be apparent.

Tabulation of complete analysis of fish.

(A) SAMPLE.

- | | |
|-------------------------------------|--------------------|
| 1. Description of fish and history. | 3. Edible portion. |
| 2. Total weight. | 4. Refuse. |

(B) GROSS ANALYSIS.

- | | |
|-------------------|---|
| 1. Moisture. | 5. Total nitrogen |
| 2. Solids. | 6. Nitrogen \times 6.25 (protein). |
| 3. Ash. | 7. Water-insoluble nitrogen. |
| 4. Ether extract. | 8. Ammonia and amine nitrogen by aëra-
tion. |

¹ Smith. Biochem. Bul., 1913, 3: 54.² Perlzweig and Gies. Biochem. Bul., 1913, 3: 69.

(c) ANALYSIS OF AQUEOUS EXTRACT.

- | | |
|--|--|
| 1. Acidity of extract in terms of N/10 NaOH solution with phenolphthalein.
2. Solids.
3. Ash.
4. Organic matter.
5. Nitrogen distribution in extract.
(a) Total nitrogen of extract, or soluble nitrogen.
(b) Coagulable nitrogen. | (c) Nitrogen precipitated by—
(I) Tannin-salt method.
(II) Zinc sulphate method.
(d) Nitrogen "bases," etc., by difference.
(e) "Amino-acid nitrogen" by—
(I) Van Slyke method.
(II) Sørensen titration method.
(f) Ammonia and amine nitrogen by aëration. |
|--|--|

(d) STUDY OF FAT EXTRACTED FROM FLESH.

- | | |
|---|---|
| 1. Specific gravity.
2. Index of refraction.
3. Iodin number. | 4. Saponification number.
5. Acid number.
6. Acetyl number. |
|---|---|

There is no need to discuss the ordinary determinations like total nitrogen, ash, fat constants, etc., but in the case of ammonia nitrogen there are certain complications that must be considered. In the determination of the so-called ammonia nitrogen we used a modification of the Folin aëration method. We followed Smith in using the Steel¹ modification of Folin's method because Perlzweig and Gies reported the presence of crystals of ammonium magnesium phosphate in fish flesh even early in the process of deterioration. The use of sodium hydroxid and sodium chlorid in place of the sodium carbonate, as suggested by Steel, seems to be necessary in cases like the analysis of fish where magnesium ammonium phosphate is known to be present. Benedict and Osterberg² have made a detailed study of the use of Steel's modification as well as the original Folin method upon urines, and they report that the former method apparently gives correct results for ammonia under nearly every condition. In fish flesh not only do we have the interference of this magnesium compound but also the added complication of amines, unknown as to nature and amount. Later we will show that certain classes of amines may be determined quantitatively by the aëration method, either when alone or with ammonium salts. Since this is true, and since we know that amines occur in fish, we found it desirable to use Steel's modification, and later to make a study of its efficiency in determining amines, either in the presence or absence of ammonium salts.

In order to study the interference, if any, of the amines with the determination of ammonia, we carried out the following experiments: Standard solutions of methyl amine, trimethyl amine, and ammonium chlorid were prepared. The exact amount of nitrogen contained in 5 cc. portions of these solutions was ascertained by duplicate Kjeldahl deter-

¹ Steel. J. Biol. Chem., 1910, 8: 365.

² Benedict and Osterberg. Biochem. Bul., 1913, 3: 47.

minations. Then the same amounts of these standard solutions were analyzed by the Folin aëration method, using Steel's modification, as already noted. These solutions were analyzed separately and mixed in stated proportions. The figures by the aëration method are the averages of four separate determinations.

Determination of ammonia and amine nitrogen by the aëration method.

SUBSTANCE USED	NITROGEN BY AÉRATION	NITROGEN BY KJELDAHL METHOD
	gram	gram
Methyl amine solution alone.....	0.0052	0.0050
Trimethyl amine solution alone.....	0.0072	0.0077
Ammonium chlorid solution alone.....	0.0189	0.0188
Trimethyl amine + NH_4Cl , solutions mixed.....	0.0148	0.0143
Trimethyl amine + CH_3NH_2 , solutions mixed.....	0.0124	0.0127

It is evident from these results that the primary or tertiary amines, either alone or with ammonium salts, are determined quantitatively by the modified Folin method for ammonium nitrogen. Therefore, we consider that determinations by this method give "ammonia and amine" nitrogen. Proteins do not interfere.

We already have seen that the presence of amines and magnesium compounds in fish is a disturbing factor in the determination of the so-called "ammonia" nitrogen by the aëration method. In the past it usually has been customary to study the amino-acid content of flesh extracts when inquiring into the effect of cold storage upon such material. In our own work we used the Sørensen¹ titration method for amino acids in making parallel determinations along with the gasometric method of Van Slyke.² In determining amino acids in fish flesh, account must be taken of the presence of both ammonia and amines, which are very closely related to the amino acids, and have many properties in common with them. For this reason a study was made of the effect of ammonia and amines, separately and in combination, upon the Sørensen titration.

Our study of the behavior of ammonia and amines in amino-acid titrations was carried out in much the same way as that upon the aëration method for ammonia described above. Standard solutions of leucin, of ammonia, and of different amines were prepared and analyzed for their "amino-acid nitrogen" according to the Sørensen method. Later, definite volumes of these same solutions were mixed and then analyzed for amino-acid nitrogen in the same way. It is obvious that if there were no interference between the ammonia, amines, and amino acids the results of the titrations upon the mixed solutions would be the same as those upon

¹ Sørensen. *Biochem. Z.*, 1907-8, **7**: 44.

² Van Slyke. *J. Biol. Chem.*, 1912, **12**: 275, and *idem.*, 1913, **16**: 121.

the solutions analyzed separately. Following are some results of this work:

Effect of ammonium chlorid and amines upon the determination of amino-acid nitrogen by the Sørensen method.

SUBSTANCES USED	NITROGEN DETERMINED SEPARATELY	NITROGEN DETERMINED IN MIXTURE
	gram	gram
Leucin solution alone.....	0.0059
Methyl amine solution alone.....	0.0072
Trimethyl amine solution alone.....	0.0017
Ammonium chlorid solution alone.....	0.0089
Leucin + NH_4Cl , solutions mixed.....	0.0148	0.0148
Leucin + CH_3NH_2 , solutions mixed.....	0.0131	0.0133
Leucin + $(\text{CH}_3)_3\text{N}$, solutions mixed.....	0.0076	0.0076
Leucin + CH_3NH_2 + NH_4Cl , solutions mixed.....	0.0219	0.0215
Leucin + $(\text{CH}_3)_3\text{N}$ + NH_4Cl , solutions mixed.....	0.0165	0.0159

An inspection of this table shows that the presence of ammonia in relatively small quantities does not seriously interfere with the amino-acid determination; neither does the presence of amines; but when amines and ammonia occur together the results obtained by the Sørensen titration are not reliable. Unfortunately, this is exactly the state of affairs existing in fish flesh. This work upon the interference of certain nitrogen compounds with the Sørensen amino-acid determination is a confirmation of the observations made by Henriques and Sørensen¹ upon the disturbing influence of ammonium salts and methyl amine. It is evident, then, that the determination of amino acids by the above method gives results that are inaccurate owing to the interference on the part of ammonia and amines. This interference, unfortunately, is not a definite one, and for that reason it is not allowable to correct the amino-acid results by subtracting the amount of nitrogen present as ammonia plus amines, determined by the aëration method. Therefore, in all the tables the percentage of amino acid as reported is subject to a varying and unknown correction, due to the presence of ammonia and amines in the fish flesh.

It is well known to organic chemists that various types of amines react in different and characteristic ways with nitrous acid; therefore, the Van Slyke gasometric method for amino acids is likewise subject to correction on account of the presence of various types of amines in fish flesh. If the time allowed for the reaction between the nitrous acid and the substance being analyzed is sufficiently long we know that the ammonia will be determined quantitatively by the Van Slyke method. However, it is not customary to allow the reaction to run more than three or five minutes. For that reason our results for amino acids determined by

¹ Henriques and Sørensen. Z. physiol. Chem., 1910, 64: 120.

the Van Slyke method are also subject to correction by an unknown amount, due to the presence in fish flesh of different types of amines and ammonia, and must not be viewed as giving accurate information on the percentage of amino acids alone.

To make the Sørensen and Van Slyke determinations of amino acids entirely accurate, it would, of course, have been possible to remove the ammonia and amines by distilling with calcium hydroxid according to Van Slyke's procedure. But this precaution seems hardly necessary in our work where the quantities of amino acids and ammonia are about 0.15% and 0.01%, respectively. Furthermore, the comparative value of our uncorrected amino-acid figures on extracts of fish flesh in studies of nitrogen distribution are not impaired, because the determinations are always made in exactly the same way on nearly identical material at definite periods during two years.

Many parallel determinations were made of nitrogen precipitated by the tannin-salt reagent and by zinc sulphate applied to the fish extracts. The results obtained by the latter method were constantly lower than those from the tannin-salt method. It is known that the zinc sulphate precipitates most sharply those substances commonly classed as proteoses, while the other method includes certain other nitrogenous substances as well as proteoses. However, in our work we always use both methods in order to have comparative figures running through the whole period of cold storage. We have found it very convenient and accurate to filter off the zinc sulphate precipitate on an asbestos mat in a Gooch crucible, wash mat and precipitate into a Kjeldahl flask and transfer for the nitrogen determinations.

In the space at our disposal it is impossible to indicate the details of analytical procedures used in our various determinations, as listed on pages 231-232. The strict precautions to be followed in preparing aqueous extracts of fish flesh, preserving them, taking aliquots, etc., may also be omitted at this time. This is also true of the details of extracting and analyzing the oils from the fish. In another year the results of all our work on frozen fish stored for two years will be completed and tabulated. At that time the comparative value of all the different determinations in showing changes in fish flesh will become evident and may be discussed more intelligently.

It is obvious that all of the methods so far applied in our work on fish aim to detect deterioration by changes in constituents *already present*, caused either by the action of bacteria or enzymes in the fish itself. For many purposes it seems very desirable to find methods that will show deterioration by the detection of decomposition products *not found* in perfectly fresh fish. We have in mind such things as indol, skatol, phenolic substances, and similar products of protein decomposition. The evidence

of the early stages of decomposition in fish is not easy to get, and no stone should be left unturned in a search for indications of the tainted condition in this class of foods.

RECOMMENDATIONS.

It is recommended—

(1) That Price's method (U. S. Bur. Chem. Circ. 108, p. 10) be made a provisional method for starch in meat products in place of Mayrhofer's method (U. S. Bur. Chem. Bul. 107 (rev.), p. 109, (b) (2)). With very little modification from Price's description, the official description should be:

In a 200 cc. beaker treat 10 grams of finely divided meat with 75 cc. of an 8% solution of potassium hydrate in 95% alcohol, and heat on the steam bath until all the meat is dissolved. This will require from 30 to 45 minutes. Add an equal volume of 95% alcohol, cool, and allow to stand at least one hour. Filter by suction through a thin layer of asbestos in a Gooch crucible. Wash twice with warm 4% potassium hydrate in 50% alcohol and then twice with warm 50% alcohol. Discard the washings. Endeavor to retain as much of the precipitate in the beaker as possible until the last washing. Place the crucible with contents in the original beaker, add 40 cc. of water and then 25 cc. of concentrated sulphuric acid. Stir during the addition of the acid and see that the acid comes in contact with all the precipitate. Allow to stand about 5 minutes, add 40 cc. of water, and heat just to boiling, stirring constantly. Transfer the solution to a 500 cc. graduated flask, add 2 cc. of a 20% aqueous solution of phosphotungstic acid, allow to cool to room temperature, and make up to mark with distilled water. Filter through a starch-free filter paper, and determine the dextrose present in a 50 cc. portion of the filtrate with Fehling's solution after neutralizing the acid, using Low's method, as given in U. S. Bureau of Chemistry Bulletin 107 (revised), page 241, for the determination of the copper in the cuprous oxide precipitate, or the latter may be weighed as directed on page 242. The amount of dextrose multiplied by 0.9 gives the equivalent in starch.

(2) That comparative studies be continued of the different determinations of the nitrogen distribution and the best manner of carrying them out as applied to extracts of fish flesh. Efforts should be made to standardize the method of determining coagulable proteins in such work. New methods proposed for the determination of proteoses, amino acids, amines, etc., in biological material should be investigated with a view to finding more sensitive and more accurate indices of deterioration in fish.

(3) That a study be made of the value of volatile distillation products like amines, indol, etc., in detecting incipient decomposition in fish.

REPORT ON FATS AND OILS.

By R. H. KERR (Bureau of Animal Industry, Washington, D. C.),
Associate Referee.

The attention of the associate referee on fats and oils has been given to a careful consideration of some of the new problems which have been raised by the rapid increase in the commercial practice of hardening oils by hydrogen. While the practice so far has been applied mainly to cottonseed oil, almost any edible oil may have its value so enhanced by this treatment as to make its hydrogenation a source of profit. It must be assumed, then, that the problems of those unlucky chemists who are responsible for detections of the adulteration of edible oils and fats are now to be increased and complicated by the addition to the list of possible adulterants of a long list of hydrogenated and partly hydrogenated fats. Aside from the evident fact that the would-be adulterator may now alter at will the iodine number, refractive index, and titer of any fat he may choose, the process of hydrogenation brings about other changes which are of far-reaching importance. The Halphen test of cottonseed oil disappears, the erucic acid of rape and mustard oils becomes behenic acid, the highly unsaturated acids of the fish oils are reduced to solid acids which no longer give the characteristic precipitate of insoluble bromids. It is to be seen at once that many of our tried and trusty tests will fail us completely before this new problem, and it is equally plain that we must devise new tests to meet the new condition or confess failure.

One of the most striking features of the problem is the absolute lack of definite information regarding the composition of fats. Although fats have been studied and worked over by chemists since the time of Chevreul, and the literature of the subject comprises a large number of bulky volumes, it remains true today that we do not know accurately the composition of a single fat. We know that cottonseed oil, for example, consists of the glycerids of palmitic, oleic, and linolic acids, but we do not know whether it consists of a simple mixture of the three triglycerids of these acids, or whether the whole array of possible mixed glycerids may not also be present. In fact, I am not aware that one single glycerid ever has been isolated from cottonseed oil and identified. Almost as much may be said of many other well-known fats. It was not known until two years ago that lard did not contain the glycerid tristearin despite all the thousands of analyses of lard made every year.

In dealing with the problem raised by the use of hydrogenated fats, it appears to me that the first requirement will be a much more extensive and intimate knowledge of the exact composition of fats. When we know of what glycerids a fat is composed we are then in a position to recognize

that fat wherever we find it, tedious and difficult as the process of identification may be. Accurate knowledge may, however, point the way to short and simple tests, just as Boemer's tedious fractional crystallization of lard finally led him to a method only a little less simple and speedy than the method already adopted by this association for the detection of the adulteration of lard. A modification of this method which I hope to present to this association at a later meeting will set the detection of beef fat, mutton fat, and all of the hydrogenated oils in lard upon a firm foundation and also will enable us to distinguish between natural lard and lard hardened by addition of hydrogen. The method, if presented, will be offered as a method for the detection of the glycerid tristearin in lard, not as a method for the detection of beef fat.

Our needs appear, then, to be (1) the collection of a large amount of wholly new information, and (2) new and less laborious methods whereby the needful knowledge may be acquired. It is along the second line that I have been working. The work I have been doing has not yet reached the stage of definite methods of analysis available for coöperative study, but some progress has been made along that line, and some improved methods probably will be available for study during the coming year. The ones suggested at present are:

- (1) A better method for the detection and identification of tristearin.
- (2) A better method for the separation of solid and liquid fatty acids.
- (3) A method for the separation of the liquid fatty acids from each other.
- (4) A method for the detection of cholesterol in mixture of animal and vegetable fats.

It is hoped that at least one, and possibly two, of these methods may be studied coöperatively and the results of the study presented at the next meeting of the association.

Mr. Hand at this point explained the operation of an apparatus for the extraction of fats which he exhibited.

REPORT ON DAIRY PRODUCTS (ADULTERATION).

By JULIUS HORTVET (State Dairy and Food Department, St. Paul, Minn.), *Associate Referee*.

The work of the past two years has magnified the importance of a special study of methods for the determination of fat in dairy products. It has, in fact, been realized more than heretofore that we are in urgent need of a thorough investigation of methods which have been recommended for study by this association; and, having in view the impor-

tance of advancing our work from year to year along the lines indicated by these recommendations, attention is first directed to the committee reports on this subject since 1906:

(1) At the meeting held in November, 1906, it was recommended, "That the Gottlieb method (Landw. Vers.-Sta., 1892, 40: 1) be made provisional for the determination of fat in milk. Referred to referee for 1907 for further recommendation. Adopted."¹

(2) At the meeting held in October, 1907, it was recommended that the study of methods of analysis of condensed milk be continued and, among other things, special attention be given to the "Roese-Gottlieb method for the determination of fat as described in the proceedings of the association for 1906. Adopted."²

(3) At the meeting held in November, 1908, it was recommended, "That the determination of fat in condensed milk be studied, special attention being given to solutions of less than 20% concentration."³

(4) At the meeting held in August, 1909, it was recommended, "That methods for determining fat in both sweetened and unsweetened condensed milk be studied, giving special attention to those modifications of the Babcock and extraction methods recently brought out in Bulletin 134 of the Indiana Agricultural Experiment Station. Adopted."⁴

(5) At the meeting held in November, 1910, it was recommended, "(1) That the Roese-Gottlieb method be adopted as provisional for the determination of fat in milks and condensed milk, both sweetened and unsweetened. Carried. (Recommended to the committee by G. E. Patriek as official.) (2) That the Roese-Gottlieb method be further studied for the analysis of ice cream, milk powders, malted milks, and milk chocolates. Adopted."⁵

(6) At the meeting held in November, 1911, it was recommended, "That Paul's method of extracting fat from dairy products be further studied, and that the fat obtained by this method be studied as to its chemical and physical constants. Adopted."⁶

During the meetings held in 1912, 1913, and 1914, the reports of the associate referee on dairy products were devoted chiefly to a continuation of the study of Paul's continuous-extraction method, together with the modifications of the method which have been proposed. Examination of the work on dairy products during the past eight years reveals the fact that the recommendations adopted during that period have not been consistently carried out. In fact, so far as the proceedings of the

¹ U. S. Bur. Chem. Bul. 105, p. 154.

² Ibid., 116, p. 117.

³ Ibid., 122, p. 188.

⁴ Ibid., 132, p. 188.

⁵ Ibid., 137, p. 169.

⁶ Ibid., 152, p. 188.

past eight years indicate, the Roesse-Gottlieb method has been adopted only as provisional. It appears to have been recommended for adoption as official, but, so far as can be learned, no further definite action along that line has been taken. Also, the further recommendations for study of the Roesse-Gottlieb method and other methods for the determination of fat in ice cream, milk powders, malted milk, etc., appear to have been overlooked. It therefore has been the aim of the associate referee during the past season to take up the study of methods for determining fat in dairy products along the lines recommended by this association between the years 1906 and 1911. The work on Paul's continuous-extraction method was practically disposed of at the meeting held in November, 1914, and no definite recommendation for the final adoption of that method was made, although it appeared that the method was valuable and serviceable in certain cases and for the special purpose for which it was originally designed. Taking up the work on dairy products at the stage indicated in the proceedings of 1911, the associate referee has accordingly enlisted the assistance of about twenty collaborators in a further study of the Roesse-Gottlieb method, with a view to placing this method in line for final adoption as official in 1916. Also, following out the instructions of this association as implied in the recommendations adopted at the meetings held in 1909 and 1910, the plan of work of the past season has included a comparative study of several modifications of the Babcock method which have been proposed during recent years. The following instructions were sent out to the collaborators:

INSTRUCTIONS TO COLLABORATORS.

Herewith I am sending you directions for collaborative work on dairy products for 1915. These directions include a description of various methods which have been proposed for the determination of fat in evaporated milk and ice cream. Also there is included a description of the Roesse-Gottlieb method, together with general instructions. You are requested to make a comparative study of all of these methods, and for that purpose to obtain in the local market two samples of each of the products named in the directions, viz., *whole milk, sweet cream, evaporated milk, and ice cream*. Instead of attempting to send out uniform samples to the collaborators, the above plan has been decided upon as preferable. Each collaborator will then carry out his work on samples obtained by himself, and, in addition, if time permits, may extend the work on as many additional samples as he may wish to investigate. You are requested to report your results in the plan shown in the tabular form herewith inclosed, and to write out in full your discussions and criticisms on the various methods.

During recent years a number of methods have been devised for testing butterfat in dairy products, and a considerable amount of uncertainty has arisen regarding the reliability of standard methods heretofore employed. It therefore has been deemed advisable to subject several of the most important of these tests to a careful critical study alongside of the well-tried-out Roesse-Gottlieb procedure adopted as provisional a few years ago. There is an urgent call for a reliable centrifugal

method for condensed milk and ice cream, and it is hoped that the collaborative work of the present season will bring about some agreement, at least on essential points, in connection with one or more of the methods which have been proposed.

Later in the season, as a result of correspondence with Dr. W. D. Bigelow, chief chemist of the National Canners' Association Laboratory, it was decided to supplement the work outlined in these instructions by determinations to be carried out on specially prepared samples of evaporated and condensed milk. These samples were put up under the careful supervision of Dr. Bigelow, at the factory of Borden's Condensed Milk Company, at Norwich, N. Y., and sent out direct to the collaborators. The samples included: (1) An unsweetened evaporated milk, (2) a sweetened condensed milk, (3) a sweetened condensed milk of composition different from sample 2. In connection with these samples, the following additional instructions were sent to the collaborators:

In addition to the work outlined in the instructions which I sent you some time ago, I will request that you make determinations of butterfat on these three samples by the Roese-Gottlieb method, and, in addition, by any other method which you believe may give correct results. You may select any one of the methods described in the outline sent you; or if you have another method which you have employed in your laboratory, I should be pleased to have you report results by that method. Results obtained on these samples are to be reported at the same time that you send me your statement covering the work included in my outline already sent you.

Carrying out the plan of work as originally outlined in the first letter of instructions, the collaborators were given the following description of methods:

DESCRIPTION OF METHODS.

BRINSMAID METHOD.

Place 2 cc. of glycerin in a Babcock milk test bottle and rotate so that the lower half surface is coated with the glycerin. In a small beaker mix 12 cc. of sulphuric acid (C. P., sp. gr. 1.84) with 18 cc. of 80% acetic acid. Weigh 9 grams of the well-mixed sample into the bottle and pour in the mixed acids so that the neck of the bottle is well washed down, then rotate the bottle so that the milk and acids are mixed and all white lumps dissolved. After thorough mixing, set the bottle aside for 15 minutes, the mixture in the meantime becoming black as in the ordinary Babcock test. Arrange a suitable rack or support so that the bottle may be placed standing in boiling water, allowing water of sufficient depth to cover the bottle above its contents. Leave the bottle in the water for 20 minutes, mixing the contents several times during this period, run in the centrifuge 5 minutes, add hot water so that the contents of the bottle rise to the bottom of the neck, then run 2 minutes, add hot water until the fat rises to the upper part of the neck, and complete the test by making a final run of 1 minute. Measure the fat column and express the result in the manner described under the heading, "Reading the fat column."

Notes on the method.—The preliminary coating of the inside of the bottle with glycerin to some extent prevents the milk from adhering to the sides. A good

method of introducing the glycerin into the bottle is to take an ordinary 2 cc. pipette, cut off the tapering point, draw the glycerin into the pipette, wipe off the outside around the end, place the pipette down into the bottle, and allow the glycerin to drain out. This procedure will prevent the glycerin from coming in contact with the neck of the bottle. Considerable heat is developed when the acids are mixed, but there is no danger of the mixture becoming too hot. A thorough mixing of the milk and acids is necessary for good results. This method appears to have been in use for several years in the laboratory of the Illinois Dairy and Food Commission. Mr. David Klein, Illinois State analyst, proposes that in the case of condensed milk a correction figure of 0.24 be subtracted from the result after multiplying the reading by 2.

GRIGSBY METHOD.

Weigh 9 grams of the thoroughly mixed sample into a Babcock milk test bottle and add 10 cc. of glacial acetic acid, washing down the neck of the bottle with the acid, then heat the mixture by putting the body of the bottle in hot water until the contents appear smooth, uniform, and free from lumps. While hot, add ordinary Babcock sulphuric acid, about 1 or 2 cc. at a time, mixing the contents after each addition, until the color of the mixture becomes dark chocolate brown, centrifuge 5 minutes, run in hot water to the bottom of the neck, centrifuge 2 minutes longer, then run in hot water until the fat column comes nearly to the top of the neck, and complete with a final run in the centrifuge of 1 minute. Measure the fat column and express the result as directed in the paragraph headed, "Reading the fat column."

Notes on the method.—Too long heating or overheating of the mixture of acetic acid and sample has practically no effect on the result, heating for 5 to 10 minutes in water slightly below boiling temperature being usually sufficient. The hotter the mixture the less sulphuric acid will ordinarily be required to produce the proper result. The fat column should be perfectly clear and free from curd or char. In case either curd or char appears, the test should be repeated, adding sulphuric acid so as to produce a darker color in the case of interfering curd or so as to produce a lighter color in the case of interfering char.

WENDLER METHOD¹.

Prepare a reagent having the following composition: 125 grams sodium hydroxid, 25 grams Rochelle salt, 25 grams sodium chlorid; dissolve in water, mix the solution, and make volume up to 500 cc. In testing whole milk, run 19 cc. of the reagent into a Babcock test bottle, add 17.6 cc. of the sample in such manner as to float over the reagent, then add 10 or 12 drops of isobutyl alcohol. In testing cream, weigh 18 grams of the sample into the test bottle, run in 19 cc. of the reagent so as to displace the cream, then add 10 or 12 drops of isobutyl alcohol. In testing evaporated milk, weigh 9 grams of the sample in the bottle, add 9 cc. of water and thoroughly mix, then run in 19 cc. of reagent so as to displace the sample, and add 10 or 12 drops of isobutyl alcohol. In testing ice cream proceed as in the case of evaporated milk. In each case, after introducing sample and reagents, shake the mixture for several minutes, place the test bottle in a water bath at 50°C. for 3 to 5 minutes in the case of milk and cream, and in the case of evaporated milk and ice cream heat the mixture at 60°C. 5 to 7 minutes. Shake thoroughly during the heating, then centrifuge at highest possible speed for 4 minutes. Add boiling hot water to bring the fat column up to the graduations in the

¹ Z. öffent. Chem., 1906, 12: 41-58.

neck, centrifuge an additional minute, measure the fat column, and express results as directed in paragraph headed, "Reading the fat column."

In addition to the methods to be subjected to comparative study described above, determine fat in each sample according to the Roesse-Gottlieb method. Also, in the case of milk and cream samples make the fat determination according to the Babcock method carried out according to procedure commonly recognized as correct.

For convenience in making comparative tests by the Roesse-Gottlieb method, the following description is added.

ROESSE-GOTTLIEB METHOD.

Weigh 40 grams of the properly prepared sample, preferably in a tared weighing dish used for sugar analysis, transfer by washing to a 100 cc. graduated sugar flask, and make up to the mark with water. Measure 10 cc. of this solution into a Röhrig tube or into a suitable size Werner-Schmidt extraction apparatus, using for the purpose not more than 10 cc. of water to transfer sample to the tube. To the material in the extraction tube add 1.25 cc. of concentrated ammonium hydroxid (2 cc. if the sample be sour) and mix thoroughly. Add 10 cc. of 95% alcohol and mix well, then add 25 cc. of washed ethyl ether, shake vigorously for half a minute, add 25 cc. of petroleum ether (redistilled slowly at a temperature below 60° preferably) and shake for half a minute. Let stand 20 minutes, or until the upper liquid is practically clear and its lower level constant. Draw off the ether-fat solution as much as possible (usually 0.5 to 0.9 will be left) into a weighed flask through a small quick-acting filter. Re-extract the liquid remaining in the tube, this time with only 15 cc. of each ether, shaking vigorously half a minute, and allow to settle. Draw off the clear solution through the small filter into the same flask as before and wash the tip of the outlet, the funnel, and filter with a few cc. of a mixture of the two ethers in equal parts. Extract again and wash in the manner just described. Evaporate the ether slowly on a steam bath, then dry the fat in a water-oven until loss of weight ceases.

Notes on the method.—Cream and evaporated milk are best sampled by taking 40 grams and dilute to 100 cc. A 10 cc. sample of this mixture is taken for analysis. In the case of ice cream or other products which are not smooth and which contain air bubbles, 4 grams of the thoroughly mixed sample are weighed and introduced into a Röhrig tube with the aid of 10 cc. of water, then ammonia, alcohol, and ether are added in the amounts called for by the method. In the case of whole milk it has been found convenient to weigh out 10 grams directly. If an emulsion occurs in the Röhrig tube the addition of a little alcohol followed by shaking will usually break it up. Also, the use of a tube of larger caliber will tend to overcome this difficulty. Or, use a large test tube provided with a blowing-off device, as in the Werner-Schmidt method. Care should be taken to weigh the residue after evaporation of ether under exactly the same conditions as those under which the dish was weighed. If the fat contains casein or foreign matter, re-extract with ether, evaporate, and weigh again. If all proper precautions are taken, results on a given sample should check within 0.02 to 0.05%. If the result is not satisfactory repeat the analysis.

GENERAL INSTRUCTIONS.

(1) *Condition of samples.*—All samples should be in the best condition possible. An evaporated milk should be perfectly smooth and contain no small lumps of fat or curded matter.

(2) *Sampling ice cream.*—Allow the sample to soften at room temperature, with frequent stirring with a spoon. A good method of obtaining a uniform sample is

to mix with an egg beater just before weighing. It has been found difficult to obtain a fair sample after the ice cream has been heated on the water bath, owing to the fact that the butterfat is melted and quickly rises to the surface.

(3) *Babcock test bottle*.—Use an 18-gram milk test bottle, reading to 8% and graduated to 0.1 of 1%.

(4) *Reading the fat column*.—Measure with a pair of dividers and read (a) from the extreme bottom to the extreme top, (b) from the extreme bottom to the lower line of the meniscus. Record and report both results. In cases where a 9-gram sample has been taken for the test, add the two readings (a) and (b) to obtain the percentage of fat.

(5) *Experience*.—In the case of all methods make preliminary trials. Repeat as many times as seem necessary in order to become adequately prepared for the regular determinations to be made on selected samples. The importance of ample experience cannot be overestimated, especially in connection with the Roese-Gottlieb method.

(6) *Plan of work*.—The following outline will serve to indicate the determinations which are to be made on the various samples named in the heading.

MILK	CREAM	EVAPORATED MILK	ICE CREAM
Roese-Gottlieb	Roese-Gottlieb	Roese-Gottlieb	Roese-Gottlieb
Wendler	Wendler	Wendler	Wendler
Babcock	Babcock	Brinsmaid	Grigsby

The results reported by the collaborators have been compiled and arranged for purposes of comparison as shown in the following tables.

The results shown in Tables 1 and 2 indicate chiefly a comparison of the Wendler and Brinsmaid methods with the Roese-Gottlieb as described in the instructions to the collaborators. In methods directing the use of a 9-gram sample of evaporated or condensed milk the collaborators were instructed to compute the fat percentages by three different formulas. A comparison of the results reported by these methods indicates that the sum of the readings designated (a) and (b) represents a fair basis on which to make the comparisons. The results obtained by multiplying reading (a) by 2 and deducting 0.24 were for the most part apparently too low. This method of expressing the fat measurement was, however, originally intended for tests applied to ice cream. With very few exceptions, the results obtained by multiplying reading (b) by 2 and adding 0.15 agreed substantially with the results obtained by adding fat-column readings (a) and (b). It therefore has been decided, for purposes of comparison of results by the various methods, to confine the Wendler and Brinsmaid figures to the sum of readings (a) and (b). Results by various miscellaneous methods are also shown in the tabulations.

Mr. J. T. Keister, of the dairy laboratory, U. S. Bureau of Chemistry, has made a special study of the methods which were described in the instructions to the collaborators. These methods have been subjected to a large number of comparative tests on samples of milk, cream, evaporated

TABLE 1.
Fat determinations in condensed milk.
 COLLABORATORS' SAMPLE No. 1—EVAPORATED MILK.

ANALYST	ROESE- GOTTLIEB	WENDLER	BRINSMAID	OTHER METHODS
H. B. Burnett.....	8 05	8 10	8 20
R. E. Stevenson	8 06	7 65	7 85
C. Bahlman.....	8 17	8 05	8 15
	7 98		
C. G. Sutton.....	7 96	7 95	8 15
	7 93	7 96	8 15
	7 94		
G. B. Taylor.....			7 60
	7 56		7 85
H. S. Bailey.....	8 06		
E. W. Thornton.....	8 01	8 10	7 90
	7 63		7 95
C. L. Black.....	7 66		8 10
	7 81	8 21	8 14	27.58
	7 81	8 12	8 13	27.40
C. N. Austin.....	7 83	8 16	8 12	27.62
	7 82	8 14		27.78
	7 81			27.54
				27.32
	7 98		
W. D. Strack.....	7 97			28.3
	7 98			28.3
J. H. Bornmann.....	7 95		8 00	28.05
	7 92		7 95	27.96
F. C. Broeman.....	7 76	7 85	7 95
	7 98		
	7 97		
F. F. Fitzgerald.....	7 96		
	7 96		
	7 96		
	7 98		
H. M. Miller.....	7 99		
	8 00		
	8 03		
	7 96		
P. J. Donk.....	7 97		
	7 97		
	7 94		
Indiana Condensed Milk Co...	7 98		
	7 98		
W. C. Geagley.....	7 97	6 8	8 0	28.0
	8 00	5 8	8 0	28.0
	7 97		
C. L. Munroe.....	7 94		
	7 86		
	7 90		
H. E. Otting.....			8 20
	8 10		8 10	28.60
H. A. Halvorson	7 94		
	7 89		
J. T. Keister.....	8 05			28 08
	8 06			27 96
O. L. Evenson.....				28 02
	8 01			28 05
Maximum.....	8 10		
Minimum.....	7 81		
Average.....	7 96		

¹ Not included in calculations.

² Griggsby method.

³ Beinling method.

⁴ Hunziker method.

⁵ Biesterfeld-Evenson method.

TABLE 1.—Continued.
COLLABORATORS' SAMPLE No. 2—SWEETENED CONDENSED MILK.

ANALYST	ROESE- GOTTLIEB	GRIGSBY	BRINSMAID	WENDLER	OTHER METHODS
L. B. Burnett.....	9.05	9.05			
R. E. Stevenson.....	9.13	9.30			
C. Bahlman.....	9.07		8.35	8.20	
C. G. Sutton.....	9.10				
	9.13				
G. B. Taylor.....	8.96				
H. S. Bailey.....	9.00				
E. W. Thornton.....	¹⁹ 9.38				
	¹⁹ 9.49				
C. L. Black.....	¹⁸ 8.57				
	9.14	8.80	9.20		
	9.10	8.92	9.11		
C. N. Austin.....	9.11	8.81	9.18		
		9.04	9.24		
	9.09				
W. D. Strack.....	9.08				²⁰ 9.2
	9.07				²⁰ 9.1
J. H. Bornmann.....	9.17				³⁸ 8.1
	9.12				³⁸ 8.4
E. C. Broeman.....	¹⁷ 7.95				
F. F. Fitzgerald.....	9.07				
	9.09				
	9.09				
H. M. Miller.....	9.09				
	9.08				
	9.07				
P. J. Donk.....	9.08				
	¹⁹ 9.58				
C. L. Monroe.....	¹⁸ 8.62				
	¹⁹ 9.24				⁴⁹ 9.20
J. T. Keister.....	¹⁹ 9.245				⁴⁹ 9.25
	9.12				⁴⁹ 9.12
O. L. Evenson.....	9.05				⁴⁹ 9.12
	9.08				
H. A. Halvorson.....	8.99				
Maximum.....	9.17				
Minimum.....	8.96				
Average.....	9.08				

¹ Not included in calculations.
² Beimling method.
³ Leach method.
⁴ Biesterfeld-Evenson method.

TABLE 1.—*Continued.*
COLLABORATORS' SAMPLE No. 53N3A—SWEETENED CONDENSED MILK.

ANALYST	ROESE- GOTTLIEB	GRIGSBY	BRINSMAID	OTHER METHODS
L. B. Burnett.....	18.95	9.20		
R. E. Stevenson.....	29.15	8.80		
C. G. Sutton.....	8.86			³ 8.85
	8.84			³ 8.92
G. B. Taylor.....	8.85			
H. S. Bailey.....	8.99			
E. W. Thornton.....	28.69			
C. L. Black.....	28.06			
	28.04			
			9.21	
C. N. Austin.....	8.90	8.52	9.12	
	8.90	8.59	9.14	
	8.91	8.81	9.24	
	8.83			
W. D. Strack.....	8.83			⁴ 9.10
	8.84			⁴ 9.00
J. H. Bornmann.....	8.89			⁵ 8.55
	8.86			⁵ 8.40
F. C. Broeman.....	27.38			
F. F. Fitzgerald.....	8.97			
	8.95			
	8.95			
H. M. Miller.....	8.96			
	8.93			
	8.97			
P. J. Donk.....	8.93			
	8.93			
C. L. Munroe.....	28.58			
	28.62			
J. T. Keister.....	29.105			³ 9.116
	29.13			³ 9.075
O. L. Evenson.....	8.95			³ 9.00
	8.98			³ 9.05
H. A. Halvorson.....	8.93			
	8.96			
	8.81		6.8	⁶ 6.2
	8.82		6.8	⁶ 5.9
W. C. Geagley.....	8.83			⁷ 7.2
				⁷ 7.2
Maximum.....	8.99			
Minimum.....	8.81			
Average.....	8.90			

¹ Average of the following determinations: 8.93, 8.97, 8.94, 8.96, 8.96, 8.94.

² Not included in calculations.

³ Biesterfeld-Evenson method.

⁴ Beilming method.

⁵ Leach method.

⁶ Wendler method.

⁷ Hunziker method.

TABLE 2.
Collaborative fat determinations in milk, cream, and ice cream.
MISCELLANEOUS MILKS.

ANALYST	ROESE- GOTTLIEB	WENDLER	BABCOCK	VARIATIONS FROM ROESE-GOTTLIEB	
		Average of (a) and (b) readings	Average of (a) and (b) readings	Wendler	Babcock
H. B. Burnett.....	3.70	3.69	3.52	-0.01	-0.18
	3.79	3.75	3.52	-0.04	-0.27
R. E. Stevenson.....	3.70	3.60	3.40	-0.10	-0.30
	3.62	3.65	3.45	+0.03	-0.17
C. Bahlman.....	3.56	3.68	3.60	+0.12	+0.04
	4.69	4.78	4.80	+0.09	+0.11
C. G. Sutton.....	¹³ 3.96	¹⁴ 4.09	¹⁴ 4.00	+0.13	+0.04
G. B. Taylor.....	3.78	3.65	¹³ 3.80	-0.13	+0.02
E. W. Thornton.....	4.58	4.58	4.53	0.00	-0.05
	¹³ 3.18	¹³ 3.05	¹³ 3.23	-0.13	+0.05
C. L. Black.....	¹⁴ 4.18	¹⁴ 4.04	¹⁴ 4.28	-0.14	+0.10
	¹³ 3.32	¹³ 3.33	3.25	+0.01	-0.08
J. H. Bornmann.....	¹³ 3.56	¹³ 3.28	3.55	-0.28	-0.01
	4.61	4.65	4.55	+0.04	-0.06
H. A. Halvorson.....	3.06	3.05	3.00	-0.01	-0.06
W. C. Geagley.....	3.45	3.40	3.43	-0.05	-0.02

MISCELLANEOUS CREAMS.

H. B. Burnett.....	18.25	18.00	17.60	-0.25	-0.65
	18.37	18.80	17.60	+0.43	-1.20
	18.05	18.40	17.48	+0.35	-0.57
R. E. Stevenson.....	18.45	19.20	18.40	+0.75	-0.05
	17.69	17.88	18.50	+0.19	+0.81
C. Bahlman.....	11.13	11.25	11.50	+0.12	+0.37
C. G. Sutton.....	¹² 22.15	¹² 22.90	¹² 23.00	+0.75	+0.85
G. B. Taylor.....	16.88	15.25	¹⁸ 15.50	-1.63	+1.62
E. W. Thornton.....	29.21	31.00	31.00	+0.79	+0.79
	¹¹ 16.80	¹¹ 16.35	¹¹ 16.88	-0.45	+0.08
C. L. Black.....	¹¹ 15.41	¹¹ 15.08	¹¹ 15.43	-0.33	+0.02
	¹¹ 16.44	¹¹ 18.00	+1.56
J. H. Bornmann.....	¹¹ 19.35	¹¹ 19.80	+0.45
W. C. Geagley.....	16.37	17.10	17.00	+0.73	+0.63

MISCELLANEOUS ICE CREAMS.

ANALYST	ROESE- GOTTLIEB	WENDLER	GRIGSBY	VARIATIONS FROM ROESE-GOTTLIEB	
		Sum of (a) and (b) readings	Sum of (a) and (b) readings	Wendler	Grigsby
H. B. Burnett.....	9.11	9.40	9.20	+0.29	+0.09
	6.04	5.60	5.80	-0.44	-0.24
R. E. Stevenson.....	9.00	8.84	9.15	-0.16	+0.15
	6.30	5.23	5.13	-1.07	-1.17
C. Bahlman.....	11.02	10.60	10.75	-0.42	-0.27
C. G. Sutton.....	¹¹ 11.63	¹¹ 11.45	¹¹ 11.75	-0.18	+0.12
G. B. Taylor.....	¹¹ 13.47	13.00	¹¹ 13.80	-0.47	+0.33
E. W. Thornton.....	14.77	14.80	14.66	+0.03	-0.11
	¹¹ 11.06	¹¹ 10.13	¹¹ 10.90	-0.93	-0.16
C. L. Black.....	¹¹ 11.03	⁹ 9.83	¹¹ 10.93	-1.20	-0.10
	⁹ 9.63	⁹ 9.73	+0.10
J. H. Bornmann.....	10.29	⁹ 9.98	-0.31
	¹¹ 10.46	10.70	10.30	+0.24	-0.16
H. A. Halvorson.....	¹¹ 12.28	12.40	12.60	+0.12	+0.32
W. C. Geagley.....	¹¹ 11.31	¹¹ 11.40	¹¹ 11.20	+0.09	-0.11

¹ Average of two or more results.

milk, and ice cream. The report of Mr. Keister as a collaborator during the past season shows a large amount of painstaking thorough work, and is so extensive and complete as to necessitate its incorporation here-with separately under Tables 3 and 4.

The following comments have been submitted by the collaborators:

COMMENTS BY COLLABORATORS.

C. L. Munroe, Miner Laboratories, Chicago, Ill.: Was unable to obtain any results by the Wendler method. In only one or two instances (while working with other samples of unsweetened condensed milk) was I able to obtain a fat column and those obtained were very unsatisfactory, as they were not clear and did not have a definite lower limit. In all other cases no fat column at all was obtained. I then tried several minor variations in the procedure outlined in the written directions, as follows: After mixing the sample, water, reagent, and isobutyl alcohol (this mixing was done very carefully and thoroughly in every case, shaking each bottle five minutes), I heated the bottles in a water bath at 55°C. for 20 minutes, with shaking. Then centrifuged at high speed for 5 minutes, added boiling water up to the neck, centrifuged 2 minutes at high speed, added boiling water to bring fat column up into neck, centrifuged 1 minute slowly and 1 minute at high speed. Obtained a fat column, but it was not clear, contained lumps of casein, and the lower limit was not sharply defined.

W. D. Bigelow, National Canners' Association, Washington, D. C.: When the samples were examined in this laboratory we had mislaid your letter, and therefore we examined them by the method that we ordinarily employ; that is, making only two extractions. It came to my attention afterwards that your letter directed three extractions. The results of Messrs. Fitzgerald, Miller, and Donk are, therefore, 0.02 or 0.03 lower than they would have been if your directions had been followed exactly.

F. C. Broeman, F. C. Broeman & Co., Cincinnati, O.: The Babcock method and its modifications gave such poor results on the sugared evaporated milk that I would not deem them worthy of consideration. Charring and indistinct separations made it impossible to get check readings on different determinations of same sample.

J. H. Bornmann, U. S. Food and Drug Inspection Laboratory, Chicago, Ill.: Brinsmaid method: While this method gives good results, it is objectionable because it requires the coating of the bottle with glycerol and heating in boiling water for 20 minutes.

Grigsby method: This method has the advantage of rapidity and simplicity over the Brinsmaid method. One disadvantage is the difficulty of knowing when just enough, and not too much, acid has been added.

Wendler method: Whereas this method gave satisfactory results on milk, it was a failure on all the other products. The fact that a comparatively harmless solution at moderately high temperature is used in place of sulphuric acid at a high temperature is one point in favor of this method. The fat column in the case of milk was very clear and sharply defined, which was not often the case to so marked a degree with the other methods employed.

The Roese-Gottlieb method requires a longer lapse of time before the results are obtained, but the total time spent on the manipulation is not so great when compared with the time spent on a centrifugal method that gives trouble. It is evident that the fat can be weighed more accurately than measured.

The Leach method (precipitation with copper sulphate and removal of the sugar solution in sweetened milk) seems to give low results.

H. A. Halvorson, State Dairy and Food Department, St. Paul, Minn.: After a large amount of preliminary work I found that I was able to obtain very satisfactory results with the Wendler method on whole milk, evaporated milk, and ice cream; with the Brinsmaid method on evaporated milk; and with the Grigsby method on ice cream. The importance of sufficient preliminary experience with all these methods, especially the Wendler method, cannot be overestimated. The following paragraphs show briefly the results that can be obtained. All the results are averages of closely agreeing duplicates.

Wendler method: On three samples of miscellaneous whole milks, results (the average of (a) and (b) readings) on two samples ran 0.04 and 0.09% higher and results on one sample ran 0.01% lower than the corresponding results by the Roesse-Gottlieb method. On four samples of miscellaneous evaporated milks, results (sum of (a) and (b) readings) on three samples ran 0.02, 0.03, and 0.01% higher and results on one sample ran 0.04% lower than the corresponding Roesse-Gottlieb results. On six samples of miscellaneous ice creams, results (sum of (a) and (b) readings) on five samples ran 0.21, 0.12, 0.30, 0.04, and 0.24% higher and results on one sample ran 0.14% lower than the corresponding Roesse-Gottlieb results.

Brinsmaid method: On four samples of miscellaneous evaporated milk, results (sum of (a) and (b) readings) ran 0.22 and 0.03% higher and results on the other two samples ran 0.09 and 0.09% lower than the corresponding Roesse-Gottlieb results.

Grigsby method: On five samples of miscellaneous ice creams, results (sum of (a) and (b) readings) on four samples were 0.21, 0.32, 0.35, 0.26% higher and results on one sample were 0.01% lower than the corresponding results by the Roesse-Gottlieb method.

Roesse-Gottlieb method: I consider this method superior to any other when great accuracy is required. An experienced analyst, with care, can obtain results that agree very closely, say, within less than 0.1%. In order that different analysts may obtain concordant results on the same sample, it is more desirable (due to errors made in measuring) to weigh the sample directly rather than to take 10 cc. of solution containing 40 grams in 100 cc. This is especially true in the case of ice cream, which usually contains a large amount of air even after being melted. I think that results nearer the truth would be obtained if the directions required that the sample be weighted out directly.

E. W. Thornton, Department of Agriculture, Raleigh, N. C.: I experienced some difficulty in the Wendler method on ice creams, due apparently to slight saponification of the butterfat. The results reported on the ice cream by this method are on a pure ice cream which contained no thickening agent.

My results on evaporated milk are on the sample No. 1 (unsweetened) which was sent to me. Both centrifugal methods on this product gave very clear and satisfactory fat readings, but with a uniform difference of 0.2% between readings (a) and (b). I was unable to obtain satisfactory results by either the Wendler or Brinsmaid methods on sweetened condensed milk.

C. N. Austin, Sears, Roebuck & Co., Chicago, Ill.: Roesse-Gottlieb method: In this work the fat flasks were dried to constant weight before the ether extracts were added. A counterpoise flask was used, which underwent the same heating as the flasks used in weighing the fat. It was found to be impossible to weigh a series of flasks with uniform rapidity. To compensate for the change in weight of the flasks, which takes place when they are removed from the desiccator, each one was allowed to stand on the balance pan until constant weight had been reached before a weighing was recorded. Another error that was difficult to avoid was caused by casein or water-soluble material passing through the filter with the ether extracts. To avoid this the extracts were allowed to stand about 30

minutes, and the filters were moistened with petroleum ether before adding the extracts. In making the extractions petroleum ether redistilled and coming over below 60° was used. Blanks were run to allow for the extractive matter in the reagents and nonvolatile residue in the ether. These caused a correction of 0.02 to 0.04 in the percentage. We were able to obtain results checking within 0.05%.

Babcock method: The modifications of the Babcock methods suggested were used in making determinations of the fat on the three samples submitted, as well as on a number of samples used for preliminary work. Bottles of the form specified could not be obtained in time for the work. The bottles used were of the ordinary form, reading to 10% and graduated to fifths of a per cent. The height from 0 to 8.0 on the stem of the 10% bottles was 1.5 cm. less than the height of the corresponding reading of the 8% bottles. A number of determinations were afterward made on other samples using the two kinds of bottles. This showed an average difference in the height of meniscus for the two kinds of bottles of only 0.01 on the scale. No correction was applied, however, since it was ascertained that the correction of -0.24, recommended for the Brinsmaid method, was originally determined from results obtained with bottles of the kind used by us. With all of the Babcock methods it was found difficult to obtain a sharply defined column of clear fat, due sometimes to a slight char, but more often to the presence of a slight amount of fine caseous material that prevented an accurate reading of the lower limit of the fat column. The fact that errors are multiplied by the necessity of using 9-gram samples would appear to be an added objection to the Babcock methods.

The Brinsmaid method was found to give results higher than the Roesse-Gottlieb results, even after the correction had been applied. However, the difference was not uniform for the three samples. The method gave results that check closely within themselves, and seemed to be as good for sweetened condensed milk as for evaporated milk.

The Grigsby method was found to be very unsatisfactory. It was very difficult to obtain fat columns free from curd or char, and even perfectly clear fat columns would sometimes give readings varying by as much as 0.2. The results were lower than those obtained by the other Babcock methods. With sweetened condensed milk this method was only applicable when the reagents were added to the drained curd. The presence of a small amount of water prevented the complete solution of the caseous material. When the results were tabulated the correction for the Brinsmaid method was applied to these results also as indicated in the instructions.

In the case of the Wendler method we substituted amyl alcohol for the isobutyl alcohol recommended, since the latter could not be obtained, and the former appeared to be effectual in preventing saponification of the fat. The method thus modified gave results with evaporated milk that accorded closely with those obtained by the Brinsmaid method, but with sweetened condensed milk no results could be obtained, since the precipitated curd did not dissolve completely.

H. S. Bailey, Bureau of Chemistry, Washington, D. C.: Just for my own satisfaction I tried to see how close duplicates could be obtained on a sweetened condensed milk, weighing up the original milk after dilution instead of merely using a 10 cc. pipette. Somewhat to my surprise the results were 8.996, 8.994, and 8.992. By using the weighing burette so that only one weighing more than the number of portions weighed out is required, very little more time is required than if we used a pipette, as there is no occasion for rinsing. To my mind the one drawback in the Roesse-Gottlieb method as outlined is the danger of using too much water in rinsing out the pipette. I find that if only 2 or 3 cc. are used there is no emulsion formed, while if one takes 10 cc. it is necessary to let the determination stand for a long time before it is possible to draw off the ether-fat column.

L. B. Burnett, Bureau of Chemistry, Washington, D. C.: Of the two centrifuge methods tried on whole milk, the Wendler method gave the best results, the fat column being perfectly clear, the lower meniscus horizontal. On evaporated milk the Wendler method did not work quite as well as on whole milk. More consistent results were obtained if the sample was not shaken during the heating, but rotated. With a little experience good results were obtained. On cream and ice cream the Wendler method seemed to give satisfactory results. The variations in the results on the second sample of cream and ice cream were not due to any fault of the method, as clear fat columns were obtained, but to the condition of the sample.

The Grigsby method was found to be satisfactory for sweetened condensed milk. The sample being diluted with one-third its weight of water and 12 instead of 9 grams being weighed out.

The Brinsmaid method did not appear to offer any special advantages, but satisfactory results were obtained with it on evaporated milk.

In the Wendler method in making up the fat column to the top of the graduations, it was found that if a dilute alcohol was employed instead of water the flocculent material was less apt to come to the top; however, this is not necessary if the sample is not shaken violently during the heating.

C. L. Black, U. S. Food and Drug Inspection Laboratory, Philadelphia, Pa.: I found the Wendler method most unsatisfactory in that in no case was I able to get all the curd dissolved by the amount of heating and shaking directed in the method, this being especially noticeable in the case of ice cream. The method would appear very unsatisfactory as a routine laboratory method in that, owing to the amount of vigorous shaking required, so few determinations can be made at one time, and these require so much more attention from the analyst than any of the ordinary methods used for similar products. In addition, it appears that the prolonged shaking seems to cause a decided loss of fat.

Geo. B. Taylor, State Board of Health, New Orleans, La.: Wendler method proved to be very unsatisfactory. In nearly every instance the clear fat would be underlain by curd. In a few instances the fat refused to rise.

It is my opinion that the Roesse-Gottlieb method as outlined (40 grams to 100 cc., 10 cc. = 4 grams taken) is unsatisfactory where materials heavy in fat are concerned. Small lumps of fat will invariably form when the solution is mixed, and remain in the pipette after the required amount is run into the Röhrig tube. When large amounts of fat are present it is practically impossible to get to constant weight by heating at the temperature of boiling water. An appreciable loss is almost always noted on heating for intervals of an hour.

During the last two years this laboratory has used, with good results, a slight modification of the Grigsby method for analyzing ice cream. We use 17.6 cc. of glacial acetic acid, mix well, but do not heat, adding commercial sulphuric acid to dark coffee brown color (about 10 cc.). This gives a clear reading. The results duplicate well.

On the other hand, it has been difficult to obtain good duplicates with the Roesse-Gottlieb method, especially with fruit ice creams. No method is satisfactory which requires the weighing out of 9 grams of sweetened condensed milk into an 8% Babcock milk bottle. This was done on triplicate samples, and worked by the Brinsmaid method, with no fat rising. A 40% solution was made up and 15 cc. equivalent to 6 grams were taken. No results were obtained. The Roesse-Gottlieb method is the only satisfactory method for sweetened condensed milk.

Clarence Bahlman, Board of Health, Cincinnati, Ohio.—When 18 grams of cream were treated with Wendler's reagent, there was a slight curd under the fat, when heated 5 minutes at 50°C. This was entirely avoided when the mixture of cream

and reagent was heated 10 minutes at 60°C. In the case of the evaporated milk, the same trouble was experienced when heating 5 minutes at 50°C., but was remedied by heating 10 minutes at 60°C. With the sweetened condensed milk, the Brinsmaid method gave considerable trouble. The fat layer showed a dark curd. Four tests were made; in only one were we able to obtain a satisfactory fat. Also, the Wendler method showed some curd when heating 5 minutes at 50°C., but gave clear separation when heated 10 minutes at 60°C.

J. T. Keister, Bureau of Chemistry, Washington, D. C.: In the Brinsmaid method very good readings were obtained in most cases and the results indicate that it gives figures varying only about 0.10 to 0.15 from the truth, when the readings are made to lower line of meniscus.

The Wendler method was a failure for evaporated milk on the 9-gram charge. Using a 6-gram charge was a considerable improvement, but as only two samples were tested by this modification no conclusion can be drawn.

The Grigsby method gave very satisfactory results on plain and nut ice cream and is about all that could be expected of a centrifugal method. In case of plain cream not a sufficient number of figures have been recorded to form any conclusion as to whether the correction -0.24 is necessary or not, as only two samples were examined, with some variation in results. In case of nut ice creams these results indicate clearly that the above correction is not necessary. The method worked successfully on the sample of pineapple cream, but in case of the other fruit creams (peach and strawberry) the method was not a success, the fat not being completely liberated and the fruit pulp also interfering with readings.

In case of the Wendler method, the results show that in its present form the method is not applicable to ice cream.

DISCUSSION OF RESULTS.

Reports from collaborators, most of whom reported determinations in duplicate, are on the whole a very satisfactory demonstration of the reliability of the Roese-Gottlieb method, especially when applied to such products as unsweetened and sweetened evaporated milk. With the exception of a few modifications, as suggested by some of the collaborators, the directions for carrying out the Roese-Gottlieb method will doubtless remain in the form already adopted as provisional. The preferred modification consists in weighing the sample. With this modification and one or two not very material changes in description, the method is in form for another season's study, anticipating its final adoption as official.

The associate referee selected, finally, for comparative study the methods devised by Wendler,¹ Brinsmaid,² and Grigsby.² The first of these, it is true, was described as applicable to the testing of milk, but preliminary trials of the method on other products led to the hope that good results might be forthcoming if the collaborators were directed to apply it to all samples submitted. The Brinsmaid method was devised for evaporated milk and the Grigsby method for ice cream. Unfortunately, in planning the work for the present year, the method described by Hunziker, and recommended for study at the meeting held in 1909, was overlooked.

¹ Z. öfent. Chem., 1906, **12**: 41-58; Analyst, **31**: 118.

² Correspondence from Mr. David Klein of the Illinois Food Commission.

There are, however, some results by this method reported by two of the collaborators, and these, together with information received from reliable sources, afford a basis for the belief that the method should be included in the coöperative study to be carried out next year. So far as can be judged from results reported by the collaborators, it appears that the methods devised by Brinsmaid, Grigsby, and Hunziker are worthy of further study, attention being given primarily to the products for which they were designed; and, in addition to these, there may probably be included the methods described by Beimling and Manchester. In the selection of methods it will, however, be borne in mind that a test like that described by Beimling has the serious defect arising from the probable solution of amyl alcohol in the fat column, thus causing too high results. A modified Roesch-Gottlieb procedure has for some time been under process of perfection in the dairy laboratory at the U. S. Bureau of Chemistry, but there is no assurance as yet that this is available for collaborative study with a view to adoption by this association. It may seem like a venture somewhat far from the aims of the association to put these modified centrifugal methods so much to the fore in our collaborative work in comparison with a method now well-nigh permanently established. There are, however, two purposes in view, either one of which seems to justify the past season's plan of work. In the first place, there is a persistent demand for a reliable quick method for factory control work or as a reliable sorting-out test for use in State and municipal food and dairy laboratories. In the second place, in the interest of sound policy and future stability in our work as analysts, this group of miscellaneous proposed so-called modified Babcock tests should be subjected to careful criticism and fair treatment. Then, any of these modifications that are not to be depended upon for the purposes for which they were devised should be consigned to oblivion. The supply of such methods in the last half dozen years has perhaps much exceeded the demand; most of them have gone into print without anything like adequate analytical backing; and the originators have been for the most part overconfident and somewhat lax in their descriptions. Some analysts, in fact, seem to imagine that one or two of these methods are applicable to all kinds of dairy products, from plain milk to ice cream and infant foods. The program which will be offered for the coming year will at any rate contemplate a weeding out among the score or more of these centrifugal methods with a view to determining which, if any, are really dependable for the products for which they were originally designed.

RECOMMENDATIONS.

It is recommended—

(1) That the Roese-Gottlieb method be adopted as official for the determination of fat in milk and condensed milk, both unsweetened and sweetened.

(2) That a further study be made of the Roese-Gottlieb method in the analysis of ice cream, milk powders, malted milks, and milk chocolates.

(3) That a special further study be made of modifications of the Babcock method as applied to condensed milk, both unsweetened and sweetened, and to ice cream.

TABLE 3.
Comparative study of fat methods for milk, cream, and evaporated milk.
MILK.

SAMPLE	WENDLER METHOD ¹				BABCOCK METHOD				ROESE-GOTTLIEB METHOD
	(a)		(b)		(a)		(b)		
	Reading	Per cent fat	Reading	Per cent fat	Reading	Per cent fat	Reading	Per cent fat	
No. 1.....	3.95	3.95	3.75	3.75	3.75	3.75	3.55	3.55	3.69 3.696
No. 2.....	3.40	3.40	3.20	3.20	3.20	3.20	3.00	3.00	3.225 3.218

¹ Reading O. K.

CREAM.

METHOD	READING	PER CENT FAT	READING	PER CENT FAT	READING	PER CENT FAT
	(a)		(b)		(c) with glymol	
Wendler: ¹						
No. 1.....	17.9	17.88	17.30	17.29	17.40	17.39
No. 2.....	16.4	16.12	15.8	15.58	15.7	15.48
Babcock:						
No. 1.....	17.9	17.74	17.3	17.15	17.4	17.25
No. 2.....	15.9	15.70	15.3	15.11	15.4	15.20

¹ Found it difficult to get uniform readings with cream. Method not considered as reliable as Babcock.

Roese-Gottlieb:

	Per cent fat
No. 1.....	17.32
No. 2.....	{ 15.16 15.23

TABLE 3.—Continued.
EVAPORATED MILK.

SAMPLE	WEIGHT OF SAMPLE	READINGS		PER CENT FAT				REMARKS
		(a)	(b)	¹ (a)×2	(b)×2	(a)×2 -0.24	(b)×2 +0.15	

BRINSMAID METHOD.								
	grams							
a {	9.0947	3.8	3.6	7.52	7.12	7.28	7.27	Readings O. K.
	9.0720	3.8	3.6	7.54	7.14	7.30	7.29	Slight undissolved curd.
b {	9.0622	3.8	3.6	7.54	7.15	7.30	7.30	Some charring.
	9.0600	3.9	3.7	7.74	7.35	7.50	7.50	Do.
c {	9.0747	3.9	3.7	7.73	7.33	7.49	7.48	Reading O. K.
	9.0550	3.9	3.7	7.75	7.35	7.51	7.50	Little brown "mud."
d {	9.0395	3.9	3.7	7.76	7.36	7.52	7.51	Cloudy fat but good reading.
	9.9780	3.9	3.7	7.73	7.33	7.49	7.48	Do.
e {	9.1914	4.4	4.2	8.61	8.22	8.37	7.91	Very good readings.
	8.9388	4.3	4.1	8.66	8.25	8.42	8.02	Do.
f {	9.0417	4.10	3.90	8.16	7.76	7.92	7.91	Readings O. K.
	9.0276	4.15	3.95	8.26	7.87	8.02	8.02	This one charred.
g {	9.0305	4.40	4.20	8.77	8.37	8.53	8.52	Slight char; not perfect.
	9.1270	4.45	4.25	8.776	8.38	8.54	8.53	Do.
h {	9.0245	4.3	4.1	8.57	8.17	8.33	8.32	Some char.
	9.1210	4.2	4.0	8.29	7.89	8.05	8.04	Reading O. K.

WENDLER METHOD.								
a {	9.0437	3.45	3.25	6.86	6.46	6.62	6.61	Undissolved casein, not a success.
	9.0722	3.30	3.10	6.54	6.15	6.30	6.30	
b {	9.1882	3.55	3.35	6.95	6.56	6.71	6.71	Some white curd.
	9.0677	3.50	3.30	6.94	6.55	6.70	6.70	Do.
c {	9.1330	3.20	3.00	6.37	5.98	6.13	6.13	Much white curd.
	9.0575	3.10	2.90	6.18	5.77	5.94	5.92	Do.
i {	6.0700	2.60	2.40	7.71	7.12	7.47	7.27	Clear reading; (a) × 3.
	6.2470	2.70	2.50	7.77	7.20	7.53	7.35	Do.
j {	6.0235	2.50	2.30	7.48	6.87	7.24	7.04	Reading O. K.; (a) × 3.
	6.0651	2.50	2.30	7.42	6.82	7.18	6.97	Slight brown "mud."

ROESE-GOTTLIEB METHOD.							
SAMPLE	PER CENT	SAMPLE	PER CENT	SAMPLE	PER CENT	SAMPLE	PER CENT
a.... {	7.172	c.... {	7.41	e.... {	8.172	h.... {	8.05
	7.144		7.41		8.197		8.06
b.... {	7.10	d.... {	7.478	f.... {	7.765	i.....	7.12
	7.15		7.519	g.....	7.779	j.....	7.457
					8.59		

¹ (a) is corrected to 9 grams.

TABLE 4.
Comparative study of fat methods with ice cream.

GRIGSBY METHOD.

SAMPLE	VARIETY	WEIGHT OF SAMPLE. (GRAMS)	READINGS		PER CENT FAT					REMARKS
			(a)	(b)	(a) + (b) ¹	(a) × 2	(b) × 2	(a) × 2 - 0.24	(b) × 2 + 0.15	
a	Vanilla.....	9.0915	4.95	4.78	9.62	9.80	9.40	9.56	9.55	Reading O. K.
		9.2712	5.10	4.90	9.70	9.90	9.51	9.66	9.66	Do.
b	Chocolate.....	9.0716	4.55	4.35	8.82	9.025	9.60	8.78	8.75	Reading perfect.
		9.1961	4.60	4.40	8.82	9.04	8.62	8.80	8.77	Few small lumps of chocolate in neck; not objectionable.
c	Vanilla.....	9.0130	4.60	4.40	8.97	9.18	8.78	8.94	8.93	Reading O. K.
		9.0232	4.70	4.50	9.16	9.37	8.97	9.13	9.12	Do.
d	Pineapple.....	9.0484	4.05	3.85	7.86	8.06	7.66	7.82	7.81	Fruit pulp was in neck of bottle, but separated from fat, and did not interfere with readings.
		9.0067	4.10	3.90	7.98	8.19	7.77	7.95	7.93	Do.
e	Peach.....	9.2447	3.50	3.30	6.61	6.81	6.42	6.57	6.57	Fat and pulp not in contact; no interference with reading.
		9.1150	3.50	3.30	6.71	6.91	6.50	6.67	6.65	Fairly good reading; pulp interfered somewhat.
f	Strawberry.....	9.0820	4.00	3.80	7.73	7.93	7.52	7.69	7.67	Only fair reading, pulp interfered.
		9.1520	4.20	4.00	8.06	8.26	7.86	8.02	8.01	Do.
g	Maple nut.....	9.5625	4.70	4.50	8.65	8.84	8.46	8.60	8.61	Reading O. K.; nuts in neck of bottle, but separated from fat.
		9.0225	4.40	4.20	8.58	8.78	8.33	8.54	8.53	Very good reading obtained by heating H ₂ O bath slightly above 60°, which caused separation of nuts from fat.
h	Peach.....	9.0210	3.90	3.70	7.53	7.78	7.38	7.54	7.53	Fruit pulp interfered with readings, some fat being entangled with pulp.
		9.2230	3.8	3.60	7.27	7.52	7.02	7.78	7.17	Do.

WENDLER METHOD.

a	Vanilla.....	9.1160	4.30	4.10	8.28	8.49	8.08	8.25	8.23	Only fair test some curd.
		9.0710	4.5	4.30	8.72	8.92	8.52	8.68	8.67	Too much curd to read accurately.
b	Chocolate.....	9.0855	4.45	4.25	8.61	8.81	8.42	8.57	8.57	White "mud" at bottom of fat column; not a success.
		9.1282	4.50	4.30	8.68	8.87	8.48	8.63	8.63	Do.
c	Vanilla.....	9.1370	4.60	4.40	8.86	9.06	8.66	8.82	8.81	White "mud" reading only approximate.
		9.0612	4.50	4.30	8.74	8.94	8.54	8.70	8.69	Do.
d	Pineapple.....	9.1090	4.00	3.80	7.70	7.90	7.66	White curd or "mud" at bottom of fat column.
		9.0730	4.00	3.80	7.74	7.93	7.69	Do.
e	Peach.....	9.0890	3.25	3.05	6.44	White curd or "mud;" not a success.
		9.1540	3.25	3.05	6.45	Do.

ROESE-GOTTLIEB METHOD.

SAMPLE	VARIETY	PER CENT FAT	SAMPLE	VARIETY	PER CENT FAT
a	Vanilla.....	9.58	e	Peach.....	7.63
b	Chocolate.....	9.57	f	Strawberry.....	7.66
		9.02			8.136
c	Vanilla.....	9.03	g	Maple nut.....	8.138
		9.216			8.75
d	Pineapple.....	9.25	h	Peach.....	8.776
		8.14			8.47
		8.16			8.508

¹ Corrected to 18 grams.

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REPORT

OF THE

COMMITTEE ON EDITING TENTATIVE
AND OFFICIAL

METHODS OF ANALYSIS

THE ASSOCIATION OF OFFICIAL AGRICULTURAL CHEMISTS

WILLIAMS & WILKINS COMPANY

BALTIMORE, U. S. A.

1916

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REPORT OF THE COMMITTEE ON EDITING METHODS OF ANALYSIS.

WASHINGTON, D. C., November 16, 1915.

TO THE PRESIDENT AND MEMBERS OF THE ASSOCIATION OF OFFICIAL AGRICULTURAL CHEMISTS.

Gentlemen:—Your committee on editing methods of analysis begs leave to report that it has completed the work assigned and herewith submits for your consideration the revised methods. The committee has included all authorized changes and additions, has eliminated obsolete methods in so far as possible, rewritten the text where parts appeared obscure, and made such consolidations of general methods and rearrangements as in its opinion would promote brevity and clearness.

In order that the members of the Association may have an opportunity to criticize the revised methods it is suggested by your committee that this report be published in the JOURNAL of the Association with a view to final adoption of the methods in 1916.

Respectfully submitted,

COMMITTEE ON EDITING METHODS OF ANALYSIS.

R. E. DOOLITTLE, *Chairman*, A. F. SEEKER,

W. A. WITHERS,

G. W. HOOVER,

J. P. STREET,

B. L. HARTWEIL.

Editorial Note:—The Board of Editors submits the following comment upon the report of the Committee on Editing Methods of Analysis:

Inasmuch as the methods of analysis as prescribed by this Association have been adopted, by regulation, for the enforcement of the Federal Food and Drugs Act and, by law, in many of the States for the enforcement of State laws, certain safeguards to protect their integrity are provided by the Association's Constitution and By-Laws.

The Constitution and By-Laws provide for Official and Provisional Methods only. At the last meeting of the Association it was suggested that the "Provisional" methods be designated as "Tentative" and the committee was instructed at the afternoon session, Wednesday, November 17, 1915, to make their report accordingly. The phraseology of the Constitution was not changed, however, but was referred to a committee to be reported upon at the 1916 meeting. Therefore, in the present report the word "Tentative" has been substituted for the word "Provisional", though the term "Provisional" must be formally retained until the Constitution shall have been changed.

Changes in methods cannot be made until an opportunity has been given members of the Association to test them. Before a method can be adopted as Official, it must have been tested through coöperative work and must have been recommended by the appropriate referee for at least two years. Official Methods, therefore, are those which have been thoroughly tested and which, in the opinion of the Association, yield accurate results in the hands of its members. In other words, they are believed to render as absolutely correct results as are possible in the existing state of knowledge concerning the determination in question. Provisional Methods are those which, while in the opinion of the Association yielding dependable results for comparison, have not as yet been tested so thoroughly as those that have been adopted officially. They are believed to be the best of the kind which have been tested by the Association but which for reasons stated have not the standing of Official Methods.

The methods in the form last promulgated by the Association, either in Bulletin 107 (Revised) or as published in their Proceedings of the last eight years, are as yet the only ones adopted by the Association.

It should be clearly understood that the revised methods as published herewith are simply the report of the Committee on Editing Methods of Analysis, including the recommendations adopted at the 1915 meeting, and at the present time have not been officially adopted. The methods are printed at this time to enable the members of the Association to study and criticize them so that they may be able to vote on them intelligently at the 1916 meeting at which time the matter will come up for final action.

RECOMMENDATIONS FOR OFFICIAL AND TENTATIVE
METHODS OF ANALYSIS, AS PRESENTED AT
THE ANNUAL MEETING OF THE ASSO-
CIATION OF OFFICIAL AGRICUL-
TURAL CHEMISTS, NOVEMBER
16 AND 17, 1915.

I. FERTILIZERS.

GENERAL METHODS.

1 MECHANICAL ANALYSIS OF BONE AND TANKAGE.—TENTATIVE.

Transfer 100 grams of the original material to a sieve having circular openings 1/50 inch (0.5 mm.) in diameter. Sift, breaking the lumps by means of a soft rubber pestle if the material has a tendency to cake. Weigh the coarse portion remaining on the sieve. Determine the fine portion by difference.

2 PREPARATION OF SAMPLE.—OFFICIAL.

Reduce the gross sample by quartering to an amount sufficient for analytical purposes. Transfer to a sieve having circular openings 1/25 inch (1 mm.) in diameter, sift, breaking the lumps with a soft rubber pestle. Grind in a mortar the part remaining on the sieve until the particles will pass through. Mix thoroughly and preserve in tightly stoppered bottles. Grind and sift as rapidly as possible to avoid loss or gain of moisture during the operation.

3 MOISTURE.—OFFICIAL.

Heat 2 grams of the sample prepared as in 2 for 5 hours in a water oven at the temperature of boiling water. In the case of potash salts, sodium nitrate, and ammonium sulphate heat at about 130°C. to constant weight. The loss in weight is considered as moisture.

TOTAL PHOSPHORIC ACID.

Gravimetric Method.—Official.

4 REAGENTS.

(a) *Molybdate solution.*—Dissolve 100 grams of molybdic acid in dilute ammonium hydroxid (144 cc. of ammonium hydroxid (sp. gr. 0.90) and 271 cc. of water); pour this solution slowly and with constant stirring into dilute nitric acid (489 cc. of nitric acid (sp. gr. 1.42) and 1148 cc. of water). Keep the mixture in a warm place for several days or until a portion heated to 40°C. deposits no yellow precipitate of ammonium phosphomolybdate. Decant the solution from any sediment and preserve in glass-stoppered vessels.

(b) *Ammonium nitrate solution.*—Dissolve 200 grams of commercial ammonium nitrate, phosphate free, in water and dilute to 2 liters.

(c) *Magnesia mixture.*—Dissolve 22 grams of recently ignited calcined magnesia in dilute hydrochloric acid, avoiding an excess of the latter. Add a little calcined magnesia in excess, and boil a few minutes to precipitate iron, aluminium, and phos-

phoric acid; filter; add 280 grams of ammonium chlorid, 261 cc. of ammonium hydroxid (sp. gr. 0.90) and dilute to 2 liters. Instead of the solution of 22 grams of calcined magnesia, 110 grams of crystallized magnesium chlorid ($\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$) dissolved in water may be used, then add 280 grams of ammonium chlorid and proceed as above.

(d) *Dilute ammonium hydroxid for washing.*—Dilute 100 cc. of ammonium hydroxid (sp. gr. 0.90) to 1 liter.

(e) *Magnesium nitrate solution.*—Dissolve 320 grams of calcined magnesia in nitric acid, avoiding an excess of the latter; then add a little calcined magnesia in excess, boil, filter from the excess of magnesia, ferric oxid, etc., and dilute to 2 liters.

5

PREPARATION OF SOLUTION.

Treat 2 or 2.5 grams of the sample by one of the methods given below:

(a) Ignite, and dissolve in hydrochloric acid.

(b) Evaporate with 5 cc. of magnesium nitrate, ignite, and dissolve in hydrochloric acid.

(c) Boil with 20–30 cc. of strong sulphuric acid in a Kjeldahl flask, adding 2–4 grams of sodium or potassium nitrate at the beginning of the digestion and a small quantity after the solution has become nearly colorless, or adding the nitrate in small portions from time to time. After the solution is colorless add 150 cc. of water, and boil for a few minutes.

(d) Digest in a Kjeldahl flask with strong sulphuric acid and such other reagents as are used in either the plain or modified Kjeldahl or Gunning method for estimating nitrogen. Do not add any potassium permanganate, but after the solution has become colorless add about 100 cc. of water and boil for a few minutes.

(e) Dissolve in 30 cc. of concentrated nitric acid and a small quantity of hydrochloric acid and boil until organic matter is destroyed.

(f) Add 30 cc. of concentrated hydrochloric acid, heat and add cautiously, in small quantities at a time, about 0.5 gram of finely pulverized potassium chlorate to destroy organic matter.

(g) Dissolve in 15–30 cc. of strong hydrochloric acid and 3–10 cc. of nitric acid. This method is recommended for fertilizers containing much iron or aluminium phosphate.

After solution, cool, dilute to 200 cc. or to 250 cc. if a 2.5 gram sample was used. Mix, and pour on a dry filter.

6

DETERMINATION.

Take an aliquot of the solution prepared as directed above, corresponding to 0.25 gram, 0.50 gram, or 1 gram, neutralize with ammonium hydroxid, and clear with a few drops of nitric acid. In case hydrochloric or sulphuric acid has been used as a solvent, add about 15 grams of dry ammonium nitrate or a solution containing that amount. To the hot solution add 60–80 cc. of the molybdate solution for every decigram of phosphoric acid (P_2O_5) that is present. Digest at about 65°C . for an hour, and determine if the phosphoric acid has been completely precipitated by the addition of more molybdate solution to the clear supernatant liquid. Filter and wash with cold water or, preferably, ammonium nitrate solution. Dissolve the precipitate on the filter with ammonium hydroxid and hot water and wash into a beaker to a bulk of not more than 100 cc. Nearly neutralize with hydrochloric acid, cool, and add magnesia mixture from a burette; add slowly (about 1 drop per second), stirring vigorously. After 15 minutes add 12 cc. of ammonium hydroxid (sp. gr. 0.90). Let stand till the supernatant liquid is clear (2 hours is usually enough) filter, wash with the dilute ammonium hydroxid until the washings are

practically free from chlorin, dry the filter and precipitate and transfer the latter to a weighed porcelain crucible. Ignite the filter separately and add its ash to the precipitate in the crucible. Ignite to whiteness or to a grayish white, weigh, and calculate to phosphoric acid (P_2O_5).

Volumetric Method.—Official.

7

REAGENTS.

(a) *Molybdate solution.*—To 100 cc. of molybdate solution prepared as directed in 4 (a), add 5 cc. of nitric acid (sp. gr. 1.42). This solution should be filtered immediately before using.

(b) *Standard sodium or potassium hydroxid solution.*—Dilute 323.81 cc. of N/1 alkali, free from carbonates, to 1 liter. 100 cc. of the solution should neutralize 32.38 cc. of N/1 acid; 1 cc. is equivalent to 1 mg. of P_2O_5 (1% of P_2O_5 on a basis of 0.1 gram of substance).

(c) *Standard acid solution.*—Prepare an acid solution corresponding to the, or to one-half of the, strength of (b), and standardize by titration against that solution, using phenolphthalein as indicator. Hydrochloric or nitric acid may be used.

(d) *Phenolphthalein solution.*—Dissolve 1 gram of phenolphthalein in 100 cc. of alcohol.

8

PREPARATION OF SOLUTION.

Dissolve according to 5 (b), (e), (f), or (g), preferably by (e), when these acids are a suitable solvent, and dilute to 200 cc. with water.

9

DETERMINATION.

(a) For percentages of 5 or below use an aliquot corresponding to 0.4 gram of substance, for percentages between 5 and 20 use an aliquot corresponding to 0.2 gram of substance, and for percentages above 20 use an aliquot corresponding to 0.1 gram of substance. Add 5–10 cc. of nitric acid, depending on the method of solution (or the equivalent in ammonium nitrate), nearly neutralize with ammonium hydroxid, dilute to 75–100 cc., heat in a water bath to 60°–65°C., and for percentages below 5 add 20–25 cc. of freshly filtered molybdate solution. For percentages between 5 and 20 add 30–35 cc. of molybdate solution. For percentages greater than 20 add sufficient molybdate solution to insure complete precipitation. Stir, let stand in the bath about 15 minutes, filter *at once*, wash once or twice with water by decantation, using 25–30 cc. each time, agitate the precipitate thoroughly and allow to settle; transfer to the filter and wash with cold water until the filtrate from 2 fillings of the filter yields a pink color upon the addition of phenolphthalein and 1 drop of the standard alkali. Transfer the precipitate and filter to a beaker or precipitating vessel, dissolve the precipitate in a small excess of the standard alkali, add a few drops of phenolphthalein solution, and titrate with the standard acid.

(b) Proceed as in (a) with this exception: Heat in a water bath at 45°–50°C., add the molybdate solution, and allow to remain in the bath with occasional stirring for 30 minutes.

(c) Proceed as in (a) to the point where the solution is ready to place in the water bath. Then cool the solution to room temperature, add molybdate solution at the rate of 75 cc. for each decigram of phosphoric acid present, place the stoppered flask containing the solution in a shaking apparatus and shake for 30 minutes at room temperature, filter at once, wash, and titrate as in (a).

WATER-SOLUBLE PHOSPHORIC ACID.

10

Gravimetric Method.—Official.

Place 2 grams of the sample on a 9 cm. filter, wash with successive small portions of water, allowing each portion to pass through before adding more, until the filtrate measures about 250 cc. If the filtrate is turbid, add a little nitric acid. Make up to any convenient volume, mix well, use an aliquot, and proceed as under 6.

11

Volumetric Method.—Official.

Treat the sample as directed under 10. To an aliquot of the solution corresponding to 0.2 or 0.4 gram, add 10 cc. of concentrated nitric acid and ammonium hydroxid until a slight permanent precipitate is formed, dilute to 60 cc., and proceed as directed under 9.

CITRATE-INSOLUBLE PHOSPHORIC ACID.—OFFICIAL.

12

REAGENTS.

In addition to the reagents given under 4 and 7 prepare ammonium citrate solution by one of the following methods:

Ammonium citrate solution.—(1) Dissolve 370 grams of commercial citric acid in 1500 cc. of water; nearly neutralize with commercial ammonium hydroxid; cool; add ammonium hydroxid until exactly neutral (testing with litmus or azolitmin paper), and dilute sufficiently to make the specific gravity 1.09 at 20°C. The volume will be about 2 liters; or,

(2) To 370 grams of commercial citric acid add commercial ammonium hydroxid until nearly neutral; reduce the specific gravity to slightly more than 1.09 at 20°C. and make exactly neutral, testing as follows: Prepare a solution of fused calcium chlorid, 200 grams to the liter, and add 4 volumes of strong alcohol. Make this solution exactly neutral, using a small amount of freshly prepared corallin solution as preliminary indicator, and test finally by withdrawing a portion, diluting with an equal volume of water, and testing with cochineal solution; 50 cc. of this solution will precipitate the citric acid from 10 cc. of the citrate solution. To 10 cc. of the nearly neutral citrate solution add 50 cc. of the alcoholic calcium chlorid solution, stir well, filter at once through a folded filter, dilute with an equal volume of water, and test the reaction with neutral solution of cochineal. If acid or alkaline, add ammonium hydroxid or citric acid, as the case may be, to the citrate solution, mix, and test again, as before. Repeat this process until a neutral reaction is obtained. Add sufficient water to make the specific gravity 1.09 at 20°C.

13

DETERMINATION.

(a) *Acidulated samples.*—Heat 100 cc. of strictly neutral ammonium citrate solution (sp. gr. 1.09) to 65°C. in a 250 cc. Erlenmeyer flask placed in a warm water bath, keeping the flask loosely stoppered to prevent evaporation. The level of the water in the bath should be above that of the citrate solution in the flask. When the citrate solution has reached 65°C., drop into it the filter containing the washed residue from the water-soluble phosphoric acid solution in 10, close tightly with a smooth rubber stopper, and shake violently until the filter paper is reduced to a pulp, relieving the pressure by momentarily removing the stopper. Place the flask in the bath and maintain its contents at exactly 65°C. Shake the flask every 5 minutes. At the expiration of exactly 30 minutes from the time the filter and the residue are introduced, remove the flask from the bath and immediately filter the contents as rapidly as possible through a quick-acting filter paper. Wash with

water at 65°C. until the volume of the filtrate is about 350 cc., allowing time for thorough draining before adding new portions of water. (1) Transfer the filter and its contents to a crucible, ignite until all organic matter is destroyed, add 10–15 cc. of strong hydrochloric acid, and digest until all phosphate is dissolved; or, (2) Return the filter with contents to the digestion flask, add 30–35 cc. of strong nitric acid, 5–10 cc. of strong hydrochloric acid, and boil until all phosphate is dissolved. Dilute the solution as prepared in (1) or (2) to 200 cc. If desired, the filter and its contents may be treated according to methods 5 (b), (c) or (d). Mix well, filter through a dry filter and proceed as directed under 6 or 9.

(b) *Non-acidulated samples.*—In case a determination of citrate-insoluble phosphoric acid is required in non-acidulated samples treat 2 grams of the phosphatic material without previous washing with water, precisely as in (a), except when the substance contains much animal matter (bone, fish, etc.), in which case dissolve the residue insoluble in ammonium citrate by any one of the processes described under 5 (b), (c) or (d) and determine phosphoric acid as directed in 6 or 9.

14

CITRATE-SOLUBLE PHOSPHORIC ACID.—OFFICIAL.

The sum of the water-soluble and citrate-insoluble subtracted from the total gives the citrate-soluble phosphoric acid.

15

DETECTION OF NITRATES.—OFFICIAL.

Mix 5 grams of the fertilizer with 25 cc. of hot water and filter. To a portion of this solution add 2 volumes of concentrated sulphuric acid, free from nitric acid and oxids of nitrogen, and allow the mixture to cool. Add cautiously a few drops of a concentrated solution of ferrous sulphate so that the fluids do not mix. If nitrates are present the junction shows at first a purple, afterwards a brown, color or if only a very minute quantity be present, a reddish color. To another portion of the solution add 1 cc. of a 1% solution of sodium nitrate and test as before to determine whether sufficient sulphuric acid were added in the first test.

ORGANIC AND AMMONIACAL NITROGEN ONLY.

Kjeldahl Method.—Official.

16

REAGENTS.

For ordinary work N/2 acid is recommended. For work in determining very small amounts of nitrogen N/10 acid is recommended. In titrating mineral acids against ammonium hydroxid solution use cochineal or methyl red as indicator.

(a) *Standard hydrochloric acid.*—Determine the absolute strength as follows: *Preliminary test.*—Place a measured portion of the acid to be standardized in an Erlenmeyer flask with excess of calcium carbonate, to neutralize free acid, and a few drops of potassium chromate as indicator. By titration with silver nitrate solution determine exactly the quantity required to precipitate the chlorin. *Final determination.*—To a measured portion of the acid to be standardized add from the burette 1 drop in excess of the required quantity of silver nitrate solution as determined by the preceding test. Heat to boiling, protect from the light, and allow to stand until the precipitate is granular. Filter on a tared Gooch crucible, previously heated to 140°–150°C., wash with hot water, testing the filtrate to prove excess of silver nitrate. Dry the silver chlorid at 140°–150°C., cool and weigh.

(b) *Standard sulphuric acid.*—Determine the absolute strength of the acid by precipitation with barium chlorid solution as follows: Dilute a measured quantity of the acid to be standardized to approximately 100 cc., heat to boiling and add drop

by drop a 10 % solution of barium chlorid until no further precipitation occurs. Continue the boiling for about 5 minutes, allow to stand for 5 hours or longer in a warm place, pour the supernatant liquid on a tared Gooch or on an ashless filter, treat the precipitate with 25-30 cc. of boiling water, transfer to the filter and wash with boiling water until the filtrate is free from chlorin. Dry, ignite over a Bunsen burner and weigh as barium sulphate.

(c) *Standard alkali solution*.—Accurately determine the strength of this solution by titration against the standard acid. N/10 solution is recommended.

(d) *Sulphuric acid*.—Sp. gr. 1.81 and free from nitrates and ammonium sulphate.

(e) *Metallic mercury, or mercuric oxid*.—Mercuric oxid should be prepared in the wet way, but not from mercuric nitrate.

(f) *Copper sulphate*.—Crystallized.

(g) *Potassium permanganate*.—Finely pulverized.

(h) *Granulated zinc or pumice stone*.—Added to the contents of the distillation flask if necessary to prevent bumping.

(i) *Potassium sulphid solution*.—Dissolve 40 grams of commercial potassium sulphid in 1 liter of water.

(j) *Sodium hydroxid solution*.—A saturated solution, free from nitrates.

(k) *Cochineal solution*.—Digest, with frequent agitation, 3 grams of pulverized cochineal in a mixture of 50 cc. of strong alcohol and 200 cc. of water for 1 or 2 days at ordinary temperature, and then filter.

(l) *Methyl red solution*.—Dissolve 1 gram of methyl red (dimethyl-amino-azobenzene-ortho-carbonic acid) in 100 cc. of 95% alcohol.

17

APPARATUS.

(a) *Kjeldahl flasks for both digestion and distillation*.—Total capacity of about 550 cc., made of hard, moderately thick, and well-annealed glass.

(b) *Distillation flasks*.—For distillation any suitable flask of about 550 cc. capacity may be used. It is fitted with a rubber stopper through which passes the lower end of a Kjeldahl connecting bulb to prevent sodium hydroxid being carried over mechanically during distillation. The bulb should be about 3 cm. in diameter, and the tubes should be of the same diameter as the condenser tube with which the upper end of the bulb tube is connected by means of rubber tubing.

18

DETERMINATION.

Place 0.7-3.5 grams, according to the nitrogen content, of the substance to be analyzed in a digestion flask with approximately 0.7 gram of mercuric oxid, or its equivalent in metallic mercury, and add 20-30 cc. of sulphuric acid (0.1-0.3 gram of crystallized copper sulphate may also be used in addition to the mercury, or in place of it). Place the flask in an inclined position and heat below the boiling point of the acid until frothing has ceased. (A small piece of paraffin may be added to prevent extreme foaming.) Then raise the heat until the acid boils briskly and digest for a time after the mixture is colorless or nearly so, or until oxidation is complete. Remove the flask from the flame, hold it upright, and while still hot add carefully potassium permanganate in small quantities at a time until, after shaking, the liquid remains green or purple.

After cooling dilute with about 200 cc. of water, add a few pieces of granulated zinc or pumice stone, if necessary to prevent bumping, and 25 cc. of potassium sulphid solution with shaking. Next add sufficient sodium hydroxid solution to make the reaction strongly alkaline, 50 cc. are usually enough, pouring it down the side of the flask so that it does not mix at once with the acid solution. Connect the flask

with the condenser, mix the contents by shaking, distil until all ammonia has passed over into a measured quantity of the standard acid and titrate with the standard alkali. The first 150 cc. of the distillate will generally contain all the ammonia.

The use of mercuric oxid in this operation greatly shortens the time necessary for digestion, which is rarely over an hour and a half in case of substances most difficult to oxidize, and is more commonly less than an hour. In most instances the use of potassium permanganate is quite unnecessary, but it is believed that in exceptional cases it is required for complete oxidation, and in view of the uncertainty it is always used. The potassium sulphid removes all the mercury from the solution, and so prevents the formation of mercurio-ammonium compounds which are not completely decomposed by the sodium hydroxid. The addition of zinc gives rise to an evolution of hydrogen and prevents violent bumping.

Previous to use the reagents should be tested by a blank experiment with sugar. The sugar partially reduces any nitrates present that might otherwise escape notice.

Gunning Method.—Official.

19

REAGENTS.

Potassium sulphate.—Pulverized.

The other reagents and standard solutions used are described under 16.

20

APPARATUS.

The apparatus used is described under 17.

21

DETERMINATION.

Place 0.7–3.5 grams, according to the nitrogen content, of the substance to be analyzed in a digestion flask. Add 10 grams of powdered potassium sulphate and 15–25 cc. (ordinarily about 20 cc.) of sulphuric acid (0.1–0.3 gram of crystallized copper sulphate may also be added). Conduct the digestion as in the Kjeldahl process, starting with a temperature below the boiling point and increasing the heat gradually until frothing ceases. Digest for a time after the mixture is colorless or nearly so, or until oxidation is complete. Do not add either potassium permanganate or potassium sulphid. Cool, dilute, neutralize, distil, and titrate with the standard alkali. In neutralizing before distilling it is convenient to add a few drops of phenolphthalein indicator or litmus paper. The pink color given by phenolphthalein indicating an alkaline reaction is destroyed by a considerable excess of strong fixed alkali.

Kjeldahl-Gunning-Arnold Method.—Official.

22

REAGENTS AND APPARATUS.

Described under 16, 17 and 19.

23

DETERMINATION.

Place 0.7–3.5 grams, according to the nitrogen content, of the substance to be analyzed in a digestion flask. Add 15–18 grams of potassium sulphate, 1 gram of copper sulphate, 1 gram of mercuric oxid, or its equivalent in metallic mercury, and 25 cc. of sulphuric acid. Heat gently until frothing ceases, then boil the mixture briskly, and continue the digestion for a time after the mixture is colorless or nearly so or until oxidation is complete. Cool, dilute with about 200 cc. of water, add 50 cc. of potassium sulphid solution, make strongly alkaline with sodium hydroxid solution and complete the determination as directed under 18.

TOTAL NITROGEN.

Kjeldahl Method Modified to include the Nitrogen of Nitrates.—Official.

24

REAGENTS.

(a) *Zinc dust*.—Impalpable powder. Granulated zinc or zinc filings will not answer.

(b) *Sodium thiosulphate*.

(c) *Commercial salicylic acid*.

The other reagents and standard solutions are described under 16.

25

APPARATUS.

The apparatus used is described under 17.

26

DETERMINATION.

Place 0.7–3.5 grams, according to the nitrogen content, of the substance to be analyzed in a Kjeldahl digestion flask. (1) Add 30 cc. of sulphuric acid containing 1 gram of salicylic acid, shake until thoroughly mixed, allow to stand for at least 30 minutes, and then add 5 grams of crystallized sodium thiosulphate and digest as directed below; or, (2) Add to the substance 30 cc. of sulphuric acid containing 2 grams of salicylic acid, allow to stand at least 30 minutes and then add gradually 2 grams of zinc dust, shaking the contents of the flask at the same time and digest as follows:

Place the flask on the stand for holding the digestion flasks and heat over a low flame until all danger from frothing has passed. Then raise the heat until the acid boils briskly and continue the boiling until white fumes no longer escape from the flask. This requires about 5–10 minutes. Add approximately 0.7 gram of mercuric oxid, or its equivalent in metallic mercury, and continue the boiling until the liquid in the flask is colorless, or nearly so. In case the contents of the flask are likely to become solid before this point is reached, add 10 cc. more of sulphuric acid. Complete the oxidation with a little potassium permanganate in the usual way and proceed as directed under 18. The reagents should be tested by blank experiments.

Gunning Method Modified to include the Nitrogen of Nitrates.—Official.

27

REAGENTS AND APPARATUS.

The reagents and standard solutions are described under 16, 17, 19 and 24.

28

DETERMINATION.

Place 0.7–3.5 grams, according to the nitrogen content, of the substance to be analyzed in a digestion flask. Add 30–35 cc. of salicylic acid mixture (30 cc. of sulphuric acid to 1 gram of salicylic acid); shake until thoroughly mixed, and allow to stand for at least 30 minutes with frequent shaking. Add 5 grams of sodium thiosulphate and heat the solution for 5 minutes; cool; add 10 grams of potassium sulphate and heat very gently until foaming ceases, then strongly until nearly colorless, and proceed as directed under 21.

Absolute or Cupric Oxid Method.—Official.

29

REAGENTS.

(a) *Coarse cupric oxid*.—Ignite and cool before using.

(b) *Fine cupric oxid*.—Grind (a).

(c) *Metallic copper*.—Granulated copper, or fine copper gauze, heated and cooled in a current of hydrogen or by dropping the heated copper into a test tube containing a few cc. of methyl alcohol.

(d) *Sodium bicarbonate*.—Free from organic matter.

(e) *Caustic potash solution*.—A supersaturated solution of caustic potash in hot water.

30

APPARATUS.

(a) *Combustion tube*.—Hard Bohemian glass, about 65 cm. long, 12.7 mm. internal diameter and sealed at one end.

(b) *Azotometer*.—Capacity 100 cc., accurately calibrated.

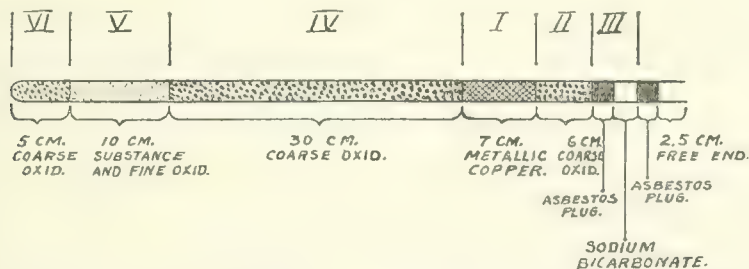
(c) *Sprengel mercury air pump*.

(d) *Small paper scoop*.—Made from stiff writing paper.

31

DETERMINATION.

Use 1-2 grams of ordinary commercial fertilizers. In the case of highly nitrogenized substances, the amount to be used is governed by the amount of nitrogen estimated to be present. Fill the tube (Fig. 1) as follows: (1) about 5 cm. of coarse



THE ROMAN NUMERALS REFER TO THE ORDER IN WHICH THE DIFFERENT PORTIONS ARE TO BE HEATED.

FIG. 1.

cupric oxid; (2) place on the small paper scoop a sufficient amount of the fine cupric oxid which, when mixed with the substance to be analyzed, will fill about 10 cm. of the tube; pour on this the substance, rinsing the watch glass with a little of the fine oxid, and mix thoroughly with a spatula, pour into the tube, rinsing the scoop with a little fine oxid; (3) about 30 cm. of coarse cupric oxid; (4) about 7 cm. of metallic copper; (5) about 6 cm. of coarse cupric oxid; (6) a small plug of asbestos; (7) 0.8-1 gram of sodium bicarbonate; (8) a large loose plug of asbestos.

After the tube is filled hold in a horizontal position and tap gently on the table in order that a canal may be formed in the upper portion of the fine cupric oxid. Place the tube in the combustion furnace, leave about 2.5 cm. of it projecting and connect with the pump by a rubber stopper smeared with glycerol, taking care to make the connection perfectly tight. In order to protect the latter from the heat, place an asbestos plate, having a circular opening in the center, over the projecting end of the tube.

Exhaust the air from the tube by means of the pump. When a vacuum has been obtained, allow the flow of mercury to continue; light the gas under that part of the

tube containing the metallic copper, the anterior layer of cupric oxid and the sodium bicarbonate. As soon as the vacuum is destroyed and the apparatus filled with carbon dioxid, shut off the flow of mercury and at once introduce the delivery tube of the pump into the receiving arm of the azotometer just below the surface of the mercury seal so that the escaping bubbles will pass into the air and not into the tube, to avoid the useless saturation of the caustic potash solution.

When the flow of carbon dioxid has very nearly or completely ceased, pass the delivery tube down into the receiving arm so that the bubbles will escape into the azotometer. Light the gas under the 30 cm. layer of oxid, heat gently for a few minutes, to drive out any moisture that may be present, and then bring to a red heat. Heat gradually the mixture of substance and oxid, lighting a jet at a time. Avoid a too rapid evolution of bubbles, which should be allowed to escape at the rate of about one per second or a little faster. When the burners under the mixture have all been turned on, light the gas under the layer of oxid at the end of the tube. When the evolution of bubbles has ceased, turn out all the burners except those under the metallic copper and anterior layer of oxid, and allow to cool for a few minutes. Exhaust with the pump and remove the azotometer before the flow of mercury has stopped. Break the connection of the tube with the pump, stop the flow of mercury, and extinguish the burners. Allow the azotometer to stand for at least an hour, or cool with a stream of water until the volume and temperature become constant.

Adjust accurately the level of the potassium hydroxid solution in the bulb to that in the azotometer; note the volume of the nitrogen, temperature, and height of barometer; calculate the weight of the nitrogen as usual.

AMMONIACAL NITROGEN.

32

Magnesium Oxid Method.—Official.

Place 0.7–3.5 grams, according to the ammonia content, of the substance to be analyzed in a distillation flask with about 200 cc. of water and 5 grams or more of magnesium oxid, free from carbonates. Then connect the flask with a condenser and distil 100 cc. of the liquid into a measured quantity of standard acid and titrate with standard alkali solution.

NITRIC AND AMMONIACAL NITROGEN.

33

Ulsch-Street Method.—Official.

Place 1 gram of the sample in a half-liter flask, add about 30 cc. of water and 2–3 grams of reduced iron, and, after standing sufficiently long to insure solution of the soluble nitrates and ammonium salts, add 10 cc. of a mixture of strong sulphuric acid with an equal volume of water; shake thoroughly, place a long-stemmed funnel in the neck of the flask to prevent mechanical loss, and allow to stand for a short time until the violence of the reaction has moderated. Heat the solution slowly, boil for 5 minutes, and cool. Add about 100 cc. of water, a little paraffin, and 7–10 grams of magnesium oxid, free or nearly free from carbonates. Connect with a condenser, such as is used in the Kjeldahl method, and boil the mixture for 40 minutes, nearly to dryness; collect the ammonia in a measured quantity of standard acid, and titrate with standard alkali solution in the usual manner. The nitrogen obtained represents the nitrates plus the ammonium salts contained in the sample.

In the analysis of nitrate salts proceed as above, except that 25 cc. of the nitrate solution, equivalent to 0.25 gram of the sample, are employed with 5 grams of reduced iron. After boiling add 75 cc. of water and an excess of sodium hydroxid solution and complete the determination as above.

34

Zinc-Iron Method.—Official.

Dissolve 10 grams of the sample in water and dilute to 500 cc. Place 25 cc. of this solution, corresponding to 0.5 gram of the substance, in a 400 cc. distillation flask, add 120 cc. of water, 5 grams of well-washed and dried zinc dust, and 5 grams of reduced iron. To the solution add 80 cc. of saturated sodium hydroxid solution, connect the flask with the condensing apparatus and conduct the distillation simultaneously with the reduction, collecting the ammonia in standard acid. Continue the distillation until 100 cc. have been distilled and titrate with standard alkali solution.

NITROGEN IN NITRATE SALTS.

35

Ferrous Sulphate-Zinc-Soda Method.—Tentative.

Place 0.5 gram of the nitrate salt in a 600–700 cc. flask, add 200 cc. of water, 5 grams of powdered zinc, 1–2 grams of ferrous sulphate, and 50 cc. of sodium hydroxid solution (36° Baumé). Connect with the distilling apparatus, distil, collect the distillate in the usual way in N/10 sulphuric acid and titrate with standard alkali solution.

ORGANIC NITROGEN SOLUBLE IN NEUTRAL PERMANGANATE.—OFFICIAL.

36

Preliminary Test (Determination of Water-Insoluble Organic Nitrogen).

Place 1 gram of the material on an 11 cm. filter paper and wash with water at room temperature until the filtrate measures 250 cc. Dry and determine nitrogen in the residue as in **18** or **21**, making a correction for the nitrogen of the filter, if necessary.

37

DETERMINATION.

Place a quantity of the fertilizer, equivalent to 50 mg. of water-insoluble organic nitrogen as determined in **36**, on a moistened 11 cm. filter paper and wash with water at room temperature until the filtrate measures 250 cc. Transfer the insoluble residue with 25 cc. of tepid water to a 300 cc. Griffin low-form beaker, add 1 gram of sodium carbonate, mix, and add 100 cc. of 2% permanganate. Cover with a watch glass and immerse for 30 minutes in a steam or hot water bath so that the level of the liquid in the beaker is below that of the water in the bath. Stir twice at intervals of 10 minutes. At the end of the digestion remove from the bath, add immediately 100 cc. of cold water, and filter through a heavy 15 cm. folded filter. Wash with small quantities of cold water until the filtrate measures about 400 cc. Determine nitrogen in the residue and filter, as in **18** or **21**, correcting for the nitrogen contained in the latter. The nitrogen thus obtained is the inactive water-insoluble organic nitrogen. Subtract this result from that obtained in **36** to obtain the percentage of organic nitrogen soluble in neutral permanganate.

ORGANIC NITROGEN SOLUBLE IN ALKALINE PERMANGANATE.—OFFICIAL.

(Not applicable to fertilizers containing cottonseed meal or castor pomace.)

38

PREPARATION OF SAMPLE.

(a) *Mixed fertilizers.*—Place an amount of material, equivalent to 50 mg. of water-insoluble organic nitrogen determined as directed under **36**, on a filter paper and wash with water at room temperature until the filtrate measures 250 cc.

(b) *Raw materials*.—Place an amount of material, equivalent to 50 mg. of water-insoluble organic nitrogen determined as directed under 36, in a small mortar, add about 2 grams of powdered rock phosphate, mix thoroughly, transfer to a filter paper, and wash with water at room temperature until the filtrate measures 250 cc. When much oil or fat is present, it is well to wash with ether before extracting with water.

39

DETERMINATION.

Dry the residue of 38 at a temperature not exceeding 80°C. and transfer from the filter to a 500–600 cc. Kjeldahl distillation flask. Add 20 cc. of water, 15–20 small glass beads, or fragments of pumice stone, a piece of paraffin the size of a pea, and 100 cc. of alkaline permanganate solution (25 grams of pure potassium permanganate and 150 grams of sodium hydroxid, separately dissolved in water, the solutions cooled, mixed, and made to a volume of 1 liter). Connect with an upright condenser to the lower end of which a receiver containing standard acid has been attached. Digest slowly, for at least 30 minutes, below distillation point, with a very low flame, using coarse wire gauze and asbestos paper between the flask and flame. Gradually raise the temperature and, after any danger from frothing has passed, distil until 95 cc. of the distillate are obtained, and titrate as usual. When a tendency to froth is noticed, lengthen the digestion period and no trouble will be experienced when the distillation is begun. During the digestion gently rotate the flask occasionally, particularly if the material shows a tendency to adhere to the sides. The nitrogen thus obtained is the active water-insoluble organic nitrogen.

POTASH.

Method I.

Lindo-Gladding Method.—Official.

40

REAGENTS.

(a) *Ammonium chlorid solution*.—Dissolve 100 grams of ammonium chlorid in 500 cc. of water, add 5–10 grams of pulverized potassium-platinic chlorid, and shake at intervals for 6–8 hours. Allow the mixture to settle overnight and filter. The residue may be used for the preparation of a fresh supply.

(b) *Platinum solution*.—A platinic chlorid solution containing the equivalent of 1 gram of metallic platinum (2.1 grams of H_2PtCl_6) in every 10 cc.

(c) *80% alcohol*.—Sp. gr. 0.8645 at $\frac{15^\circ\text{C.}}{15^\circ}$. Denatured alcohol, made up according to formula 1 (U. S. Internal Rev., Reg. No. 30, Revised, Aug. 22, 1911, p. 45) and diluted with water to make 80% alcohol by volume, may also be used.

41

PREPARATION OF SOLUTION.

(a) *Mixed fertilizers*.—Place 2.5 grams of the sample upon a 12.5 cm. filter paper and wash with boiling water until the filtrate amounts to about 200 cc. Add to the filtrate 2 cc. of concentrated hydrochloric acid, heat to boiling, transfer to a 250 cc. graduated flask and add to the hot solution a slight excess of ammonium hydroxid and sufficient ammonium oxalate to precipitate all the lime present, cool, dilute to 250 cc., mix, and pass through a dry filter.

(b) *Potash salts; muriate and sulphate of potash, sulphate of potash and magnesia, and kainit*.—Dissolve 2.5 grams and dilute to 250 cc. without the addition of ammonium hydroxid and ammonium oxalate.

(c) *Organic compounds*.—When it is desired to determine the total amount of potash in organic substances, such as cottonseed meal, tobacco stems, etc., saturate

10 grams of the sample with strong sulphuric acid and ignite in a muffle at a low red heat to destroy organic matter. Add a little strong hydrochloric acid, warm slightly in order to loosen the mass from the dish, transfer to a 250 cc. graduated flask, add ammonia and ammonium oxalate and proceed as in (a).

42

DETERMINATION.

(a) *Mixed fertilizers*.—Evaporate 50 cc. of the solution in 41 (a) nearly to dryness, add 1 cc. of dilute sulphuric acid (1 to 1), evaporate to dryness, and ignite to whiteness. Maintain a full red heat until the residue is perfectly white. Dissolve the residue in hot water, using at least 20 cc. for each decigram of potassium oxid present, add a few drops of hydrochloric acid, and platinum solution in excess. Evaporate on a water bath to a thick paste. Treat the residue with 80% alcohol, avoiding exposure to ammonia. Filter, wash the precipitate thoroughly with 80% alcohol both by decantation and on the filter, continuing the washing after the filtrate is colorless. Then wash with 10 cc. of the ammonium chlorid solution to remove impurities from the precipitate and repeat 5 or 6 times. Wash again thoroughly with 80% alcohol and dry the precipitate for 30 minutes at 100°C. Weigh and calculate to potassium oxid. The precipitate should be perfectly soluble in water.

(b) *Muriate of potash*.—Acidify 50 cc. of the solution prepared according to 41 (b) with a few drops of hydrochloric acid, add 10 cc. of platinum solution and evaporate to a thick paste. Treat the residue as under (a).

(c) *Sulphate of potash; sulphate of potash and magnesia; and kainit*.—Acidify 50 cc. of the solution prepared according to 41 (b) with a few drops of hydrochloric acid and add 15 cc. of platinum solution. Evaporate the mixture and proceed as directed under (a), except that 25 cc. portions of the ammonium chlorid solution should be used.

(d) *Water-soluble potash in wood ashes and cotton hull ashes*.—Prepare the solution according to 41 (a) and proceed as directed under (a), paying special attention to the last sentence.

Method II.—Official.

(The Lindo-Gladding method is preferable in the presence of soluble sulphates.)

43

REAGENTS.

Described under 40.

44

PREPARATION OF SOLUTION.

Prepare the solution as directed under 41, omitting in all cases the addition of ammonium hydroxid and ammonium oxalate.

45

DETERMINATION.

Dilute 25 cc. of the solution made as directed under 44 (50 cc. if less than 10% of potassium oxid be present) to 150 cc., heat to 100°C., and add, drop by drop, with constant stirring, a slight excess of barium chlorid solution. Without filtering, add in the same manner barium hydroxid solution in slight excess. Filter while hot and wash until the precipitate is free from chlorin. Add to the filtrate 1 cc. of strong ammonium hydroxid, and then a saturated solution of ammonium carbonate until the excess of barium is precipitated. Heat and add, in fine powder, 0.5 gram of pure oxalic acid or 0.75 gram of ammonium oxalate. Filter, wash free from chlorin, evaporate the filtrate to dryness in a platinum dish, and ignite carefully over the free

flame, below a red heat, until all volatile matter is driven off. Digest the residue with hot water, filter through a small filter and dilute the filtrate, if necessary, so that for each decigram of potassium oxid there will be at least 20 cc. of liquid. Acidify with a few drops of hydrochloric acid and add platinum solution in excess. Evaporate on a water bath to a thick paste and treat the residue with 80% alcohol, both by decantation and after collecting on a Gooch or other form of filter, dry for 30 minutes at 100°C. and weigh. If there is an appearance of foreign matter in the double salt, it should be washed as in 42 (a) with several portions of 10 cc. each of the ammonium chlorid solution.

THOMAS OR BASIC SLAG.—TENTATIVE.

46

MECHANICAL ANALYSIS.

Proceed as directed under 1, using 10 grams of material.

47

PREPARATION OF SAMPLE.

Prepare the sample as directed under 2.

TOTAL PHOSPHORIC ACID.

Gravimetric Method.

48

PREPARATION OF SOLUTION.

Prepare the solution for analysis as directed under 5 (e), or in strong hydrochloric acid alone. In the latter case after the portion for analysis is measured out, add nitric acid and heat for a few minutes.

49

DETERMINATION.

Dehydrate an aliquot (20 cc.) of 48 by evaporating to dryness on a steam or hot water bath; treat with 5 cc. of hydrochloric acid and 25 cc. of hot water; digest in order to complete solution and filter off silica. From this point proceed as directed under 6. Before precipitating with magnesia mixture, add 5 cc. of 5% sodium acetate.

50

Volumetric Method.

Prepare the solution as directed under 5 (e) and determine the phosphoric acid in an aliquot of this solution as directed under 9, standardizing the solutions against a standard phosphate material of approximately the same composition as the sample under examination.

CITRATE-SOLUBLE PHOSPHORIC ACID.

Gravimetric Method.—(Wagner's Method.)

51

PREPARATION OF SOLUTION.

Weigh 5 grams of the slag into a 500 cc. Wagner flask containing 5 cc. of 95% alcohol. (The flask should have a neck width of at least 22 mm. and should be marked at least 8 cm. below the mouth.) Make up to the mark with 2% citric acid solution of a temperature of 17.5°C. Fit the flask with a rubber stopper and place at once in a rotatory apparatus, shaking the flask for 30 minutes at the rate of 30–40 revolutions per minute, at the end of which time remove the flask, filter immediately on a dry filter and analyze the solution at once.

52

DETERMINATION.

To 50 cc. of the clear filtrate in a beaker add 100 cc. of molybdate solution prepared as directed under 4 (a). Place the beaker in a water bath, until the temperature of the beaker's contents reaches 65°C., remove from the bath and cool to room temperature. Filter and wash the yellow precipitate of ammonium phosphomolybdate 4 or 5 times with 1% nitric acid. Dissolve the precipitate in 100 cc. of cold 2% ammonium hydroxid, nearly neutralize with hydrochloric acid and add to the solution, drop by drop, with continuous stirring, 15 cc. of magnesia mixture prepared as directed under 4 (c) and proceed as under 6.

53

Volumetric Method.

In an aliquot of the clear solution prepared as in 51, determine the phosphoric acid as directed under 9.

II. SOILS.

1

DIRECTIONS FOR TAKING SAMPLES.—OFFICIAL.

Sampling should be done preferably when the soil is reasonably dry. Remove from the surface all vegetable material not incorporated with the soil. With a soil auger or tube, whichever may be better adapted to the soil conditions, take a sufficient number of sub-samples at properly distributed points to secure composite samples representative of the entire tract.

(a) *Surface soil*.—Take a composite sample representative of the entire tract to a depth of either (1) 6 inches, (2) the average depth of the plowed soil if this exceeds 6 inches, or (3) a maximum depth of 12 inches when there is no clear line of demarcation between the soil and sub-soil above this depth.

(b) *Sub-soil*.—Take a composite sample of each important and distinctly different soil stratum below the surface section already sampled to a total depth of 40 inches. If a soil auger is used, before taking sub-soil samples the hole should be enlarged and carefully cleaned out with the auger to prevent contamination of the several substrata when the sample is being withdrawn.

Mix each composite sample thoroughly and, after cutting down by quartering to about 2–4 pounds, air-dry in a cool, well-ventilated place.

It is recommended that the weight of a given volume of the soil as it lies in the field be taken for calculating the percentage results obtained by analysis to pounds per given area of the soil.

2

PREPARATION OF SAMPLE.—OFFICIAL.

After air-drying and weighing the sample, pulverize in a porcelain mortar, using a rubber-tipped pestle to avoid the reduction of rock fragments, and pass through a sieve with circular openings $\frac{1}{25}$ inch (1 mm.) in diameter. Discard the detritus and weigh. Thoroughly mix the sifted material and preserve in a suitable stoppered container.

For the quantitative determination of any of the constituents, prepare a very finely pulverized sub-sample of the sifted material, using an agate mortar.

3

MOISTURE.—OFFICIAL.

Dry 2 or more grams of the sample, as prepared under 2, in a tared platinum dish for 5 hours at the temperature of boiling water; cover the dish, cool in a desiccator, and weigh rapidly to prevent the absorption of moisture. Heat, cool, and weigh at intervals of 2 hours to constant weight. The loss of weight is reported as moisture.

4

VOLATILE MATTER.—OFFICIAL.

Heat the dish and dry soil from 3 to full redness, stirring occasionally, until all organic matter is destroyed. If the soil contains appreciable quantities of carbonates, cool and moisten with a few drops of saturated ammonium carbonate solution, dry and heat to dull redness to expel ammonium salts; cool in a desiccator and weigh.

ORGANIC CARBON.—OFFICIAL.

5

APPARATUS.

(a) *A calorimeter bomb.*—Use a type that permits the recovery and transfer of the entire solid residue of the exploded charge to a small vessel by means of a jet of water.

(b) *Parr's apparatus for determining carbon dioxid.*¹—Illustrated in Fig. 2.

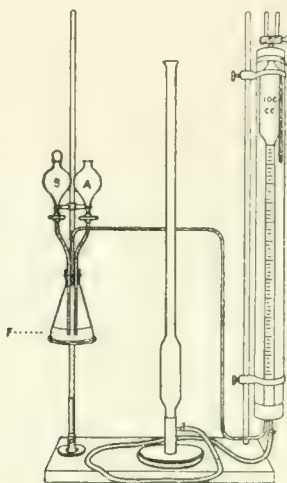


FIG. 2. PARR'S APPARATUS FOR THE DETERMINATION OF CARBON DIOXID.

This consists of a 150 cc. Erlenmeyer flask (*F*) fitted with a 3-holed stopper through 2 of which the stems of 2 dropping funnels (*S*) and (*A*) extend almost to the bottom of the flask. A capillary tube, passing through the third hole and flush with the bottom of the stopper, connects with the gas burette (*B*).

(c) *A simple Hempel gas pipette.*—Contains 30% potassium hydroxid solution.

6

DETERMINATION.

Introduce 2 grams of soil as prepared under 2 (1 gram if high in organic matter), 0.75 gram of magnesium powder, and 10 grams of sodium peroxid, into the closed dry calorimeter bomb, and mix thoroughly by shaking the bomb back and forth. Explode the charge by means of an electric spark or by dropping a red hot plug into the bomb through an automatic valve which closes immediately after the plug enters. Remove the residue from the bomb, using as little hot water as possible, heat to boiling, and transfer to the receiving funnel (*S*) of Parr's apparatus. From the acid funnel (*A*) run 50 cc. of sulphuric acid (1 to 2) into the flask (*F*). Connect the apparatus and slowly add the contents from the receiving funnel (*S*). The carbon dioxid generated passes through the capillary tube into the graduated burette (*B*). Heat the contents of the flask (*F*) to boiling and boil for 1 minute, then force the gases into the graduated burette (*B*) by introducing water into the flask (*F*) through the funnel (*S*). Read the burette, recording the temperature and pressure. Pass the gas into an ordinary absorption pipette containing 30% potassium hydroxid solution. Shake the gas with the solution until carbon dioxid is wholly absorbed. Return the

residual gas to the graduated burette (*B*), and again read the burette noting the temperature and pressure. The difference in readings calculated to standard conditions of temperature and pressure gives the number of cc. of carbon dioxide derived from the total carbon in the sample. Conduct a blank determination upon the reagents used. If an appreciable amount of carbon dioxide is obtained in the blank, the result expressed in terms of total carbon must be corrected accordingly.

Determine the inorganic carbon as directed under **9** and subtract it from the total carbon to obtain the organic carbon.

INORGANIC CARBON.

Modified Marr Method.²—Tentative.

7

REAGENTS.

(a) *N/10 hydrochloric acid.*

(b) *N/10 sodium hydroxid.*

(c) *Dilute hydrochloric acid.*—Dilute 25 cc. of concentrated hydrochloric acid (sp. gr. 1.19) to 250 cc. with carbon dioxide-free water.

(d) *Barium hydroxid solution.*—Prepare a saturated aqueous solution of barium hydroxid, filter through asbestos into a large container through which air free from carbon dioxide has been aspirated for some time, and provide an arrangement whereby the solution may be delivered by air pressure or gravity and kept from contact with carbon dioxide by means of soda lime tubes.

(e) *Carbon dioxide-free water.*—Use recently boiled and cooled water, or water from which carbon dioxide has been removed by aeration for a sufficient length of time with carbon dioxide-free air. Keep in a container provided with a similar attachment as in (d).

8

APPARATUS.

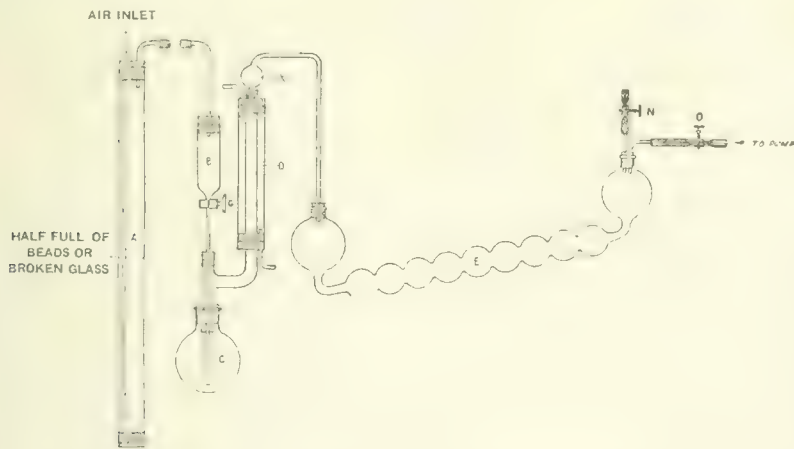


FIG. 3. MODIFIED MARR APPARATUS FOR DETERMINING CARBON DIOXID.

The apparatus required (Fig. 3), consists of a tube (*A*), 50–60 cm. long, partly filled with beads or broken glass and containing strong potassium hydroxid solution (1 to 2), a cylindrical open top separatory funnel (*B*), capacity 50 cc., marked at 20 and 40 cc., the stem of which extends almost to the bottom of the 250 cc. flask (*C*) into the

mouth of which is fitted, by means of a rubber stopper, a section of glass tubing 10 cm. in length and 1 cm. internal diameter, which in turn is furnished with a side tube extending through the condenser jacket (*D*), and connected by means of a rubber stopper to the small trap (*K*), which is attached to the Meyer absorption apparatus (*E*) as shown in the figure. The Meyer absorption apparatus is provided with 2 large bulbs, each of about 250 cc. capacity, and 10 smaller connecting bulbs, each of about 10 cc. capacity. The connections between the various bulbs should have an internal diameter of 8–10 mm. A perforated rubber stopper carrying 2 short pieces of capillary tubing each provided with rubber tubing and pinch-cocks (*N* and *O*) is fitted into the other opening of the Meyer absorption apparatus; (*N*) is for the addition of reagents, (*O*) is connected to the vacuum pump.

All parts of the apparatus must be capable of withstanding a vacuum of approximately 70 cm. and be perfectly air tight.

9

DETERMINATION.

Place 5–20 grams of soil as prepared under **2** (depending upon the carbonate content as indicated by qualitative examination) in the flask (*C*), and connect up the apparatus, but do not connect (*A*) to (*B*). Close the stop-cock (*G*) and the pinch-cock (*N*); open the pinch-cock (*O*) and exhaust the apparatus to a vacuum of approximately 70 cm. Close the pinch-cock (*O*). Connect the barium hydroxid container by means of the pinch-cock (*N*) and rubber tube; open the pinch-cock (*N*) and allow sufficient barium hydroxid solution to flow into the Meyer absorption apparatus to fill 3 or 4 of the small bulbs; close the pinch-cock (*N*); substitute the barium hydroxid container by the carbon dioxid-free water container; open the pinch-cock (*N*) and add sufficient carbon dioxid-free water to fill all the small bulbs and most of the lower large bulb. Through the separatory funnel (*B*), add 80 cc. of carbon dioxid-free water to the sample in the flask (*C*), avoiding the entrance of any air, open the pinch-cock (*O*) and heat to boiling, protecting the flask from the direct flame by a wire gauze with an asbestos center. Adjust the burner so that 2–3 minutes are required for the contents of the flask to reach boiling temperature and boil until bubbles no longer pass through the Meyer absorption apparatus. Then close the pinch-cock (*O*), and run into the flask (*C*), avoiding the entrance of any air, 20 cc. of the dilute hydrochloric acid through the separatory funnel which is then connected with the tube (*A*). This proportion of hydrochloric acid plus the 80 cc. of water previously added, gives an acid (2 to 100) for the decomposition of carbonates. If the nature of the soil is such that a greater strength of acid is considered necessary, an amount of acid (3 to 100) may be used for digesting the soil. Shake the Meyer apparatus (*E*) gently, so that the liquid in the lower large bulb is brought into contact with the gas therein, and open (*O*) carefully, but do not allow more than a few bubbles to escape before shaking again. Repeat the operation several times until bubbles no longer pass through the Meyer apparatus (*E*) when (*O*) is opened. Leave (*O*) open, and continue the boiling for about 25 minutes until carbon dioxid gas is no longer evolved from the sample in the flask (*C*). Maintain a constant flow of cold water through the condenser (*D*). Do not allow the boiling to become so violent that liquid is drawn up into the condenser tube. If foaming is troublesome, add a drop of non-volatile oil through the separatory funnel (*B*). When the evolution of carbon dioxid has ceased, close (*O*) and break the vacuum by cautiously opening the stop-cock (*G*) drawing in air through the tube (*A*).

Disconnect the Meyer apparatus (*E*) and filter by the Cain method³ as follows: Prepare a filter by covering a perforated porcelain plate within a carbon funnel with a layer of asbestos, and a layer of ground quartz, both of which have previously been

purified by digestion with hydrochloric acid and thorough washing with water. The filter tube passes through a 1-holed rubber stopper which fits into a side arm filtering flask. The side arm of this flask is connected to the suction pipe by a 2-way stop-cock. In the top of the filter tube is fitted a 2-holed rubber stopper carrying glass tubes bent at right angles. To these tubes are attached rubber tubing bearing pinch-cocks. One of these rubber tubes terminates in a tube containing soda lime. The other rubber tube is connected with one end of the Meyer apparatus. The other end of the Meyer apparatus is attached to a bottle containing carbon dioxid-free water by means of a well-washed rubber tube and a glass tube extending to the bottom of the bottle. This glass tube passes through a 2-holed rubber stopper. In the other hole of the stopper is placed a tube containing soda lime. A pinch-cock is placed on the tube joining the water bottle and the Meyer apparatus. This cock is kept closed until the precipitate on the filter is ready for washing. With the pinch-cock on the air outlet of the filter tube closed and the pinch-cock from the filter tube to the Meyer apparatus open, apply gentle suction to the filter flask until the contents of the Meyer apparatus have been transferred to the filter. When necessary, the pinch-cock to the air inlet of the filter tube is opened to admit air behind the liquid in the Meyer apparatus. Open the pinch-cock between the wash-water bottle and the Meyer apparatus and open the stop-cock leading from the filter flask so as to maintain a gentle suction. By manipulation of the Meyer apparatus the wash water comes in contact with all parts of the interior of the apparatus, after which the water is sucked through the filter. After this thorough washing admit air through the side opening of the stop-cock leading to the filter flask.

Disconnect the apparatus, remove the filter pad with the barium carbonate from the filter tube by means of a glass rod, place in a beaker and add a measured amount of N/10 hydrochloric acid in excess, first rinsing the Meyer bulbs with a small amount, carefully measured, of this acid and water. Titrate the excess of acid with N/10 sodium hydroxid, using methyl orange as indicator. Make a blank determination under the same conditions and apply the necessary correction. From the amount of N/10 hydrochloric acid required to neutralize the barium carbonate formed by the carbon dioxid in the sample, calculate the quantity of inorganic carbon. One cc. of N/10 acid corresponds to 0.0006 gram of carbon.

10

TOTAL NITROGEN.—OFFICIAL.

Place 7–14 grams of the soil, as prepared under 2, in a 300 cc. Kjeldahl digestion flask with 30 cc., or more if necessary, of concentrated sulphuric acid and 0.7 gram of mercuric oxid, or 0.65 gram of mercury. Mix immediately by shaking to prevent the soil from adhering to the sides of the flask. Heat over a low flame, increase the heat gradually, and rotate the flask frequently or shake if necessary to prevent the contents from sticking to the bottom of the flask. When all the organic matter is destroyed, continue the digestion for 1 hour. Oxidize the residue with potassium permanganate, carefully adding small portions at a time to the hot liquid until, after shaking, the liquid remains green or purple. After cooling, dilute the contents of the flask with 100 cc. of water and transfer to a 700 cc. copper flask, using about 150 cc. of water to wash out the digestion flask. Add an excess of strong alkali solution containing potassium sulphid, connect the flask with a distilling apparatus, mix the contents thoroughly and complete the determination as directed in I, 18.

STRONG ACID DIGESTION OF THE SOIL.

11

PREPARATION OF SOIL SOLUTION.—OFFICIAL.

Place an amount of soil, as prepared under **2** and equivalent to 10 grams on a moisture-free basis, in a 200–300 cc. non-soluble glass Erlenmeyer flask to which is fitted, by a ground joint or 1-holed rubber stopper, a reflux tube 20 inches or more in length. Add 100 cc. of hydrochloric acid of constant boiling point (approximate sp. gr. 1.115, 1350 cc. of acid (sp. gr. 1.19), and 1000 cc. of water), and digest continuously for 10 hours on a steam or water bath, shaking the flask every hour. Allow to settle, and avoiding more than very small quantities of the sediment, decant the solution into a porcelain dish or non-soluble glass beaker. Transfer by means of hot water the insoluble residue to a filter, wash until free from chlorin and add the washings to the original solution. Concentrate, oxidize the organic matter present in the solution with a few drops of nitric acid and evaporate to dryness on a water bath. Treat with hot water, add a few cc. of hydrochloric acid and again evaporate to complete dryness. When the final evaporation is complete and the dish cooled, moisten the residue with a few drops of strong hydrochloric acid. Add 10–20 cc. of water, warm on the bath to secure complete solution of the soluble salts, filter and wash until free from chlorin. Again evaporate the solution to dryness to render insoluble any silica that may remain in solution, and treat as above. The filtrate constitutes the acid extract freed of soluble silica, and is made up to a definite volume (250 or 500 cc.) and designated as *A*.

12

INSOLUBLE RESIDUE.—OFFICIAL.

Combine the filters and the main residue obtained in **11** in a small dish, dry, ignite over a Bunsen flame for an hour or more, carefully at first, then completely over a blast lamp to constant weight. Weigh and calculate as the insoluble residue. This residue may be analyzed by the usual methods applicable to silicates or it may be employed in the determination of total alkalies as described under **26**. If it is desired to determine the silica soluble in alkalies, treat a separate portion of the soil as directed in **11**, except that all filtrations must be made through the same *hardened* filter, then, without igniting, wash the insoluble residue into a platinum dish, dry at 100°C. and complete the determination as directed under **III, 4 (a)**.

13

IRON, ALUMINIUM AND PHOSPHORIC ACID, COLLECTIVELY.—OFFICIAL.

(1) To an aliquot (50 or 100 cc., according to the probable amount of iron present) of *A*, under **11**, add ammonium hydroxid, drop by drop, until the precipitate formed requires several seconds to dissolve, thus leaving the solution but faintly acid. Heat nearly to the boiling point, and add ammonium hydroxid to precipitate all of the iron, aluminium, etc. Boil in a covered beaker for about 1 minute, remove, and if no ammonia is given off (detected by smelling) continue the addition, drop by drop, until ammonia can be detected. Do not allow the precipitate to settle, but stir and pour on the filter. Wash immediately with hot water, using a fine jet which is played around the edge of the precipitate, thus cutting it free from the paper in order to produce rapid filtration. Wash the precipitate several times, return it to the original beaker, dissolve with a few drops of hydrochloric acid and warm. Reprecipitate the iron, aluminium and phosphoric acid with ammonium hydroxid as above and wash until free from chlorin. Designate the filtrate as *B*.

Dry the filter and precipitate, remove the latter from the filter, ignite the filter separately and add to its ash the precipitate. Then ignite to bright redness, cool

in a desiccator and weigh as ferric oxid (Fe_2O_3), aluminium oxid (Al_2O_3), and phosphorus pentoxid (P_2O_5). Transfer this residue to a flask, digest with several cc. of sulphuric acid (1 to 4), and heat to accelerate solution. When solution is complete reduce with zinc and determine the ferrous iron by titration with a standard permanganate solution, and calculate to ferric oxid; or, (2) In lieu of the above, evaporate 50 or 100 cc. of *A*, under **11**, with the addition of 10 cc. of sulphuric acid until all hydrochloric acid is expelled, dilute with water, reduce with zinc and determine the ferrous iron by titration with a standard permanganate solution and calculate to ferric oxid.

The weight of ferric oxid, plus that of the phosphorus pentoxid, determined under **17** or **19**, subtracted from the collective weight of ferric oxid, aluminium oxid, and phosphorus pentoxid, gives the weight of the aluminium oxid.

14**MANGANESE.—OFFICIAL.**

Concentrate *B*, under **13**, to about 50 cc., cool, add bromin water until the solution is colored, make alkaline with ammonium hydroxid, and heat to boiling in a covered beaker; cool, and repeat the addition of bromin water, of ammonium hydroxid and boil again. If a precipitate is obtained, slightly acidify the solution with acetic acid, filter immediately, and wash with hot water. Dry the precipitate, ignite and weigh as manganomanganic oxid (Mn_3O_4). Designate the filtrate, or if there is no precipitate, the original solution, as *C*.

15**CALCIUM.—OFFICIAL.**

Concentrate *C*, under **14**, to about 50 cc., make slightly alkaline with ammonium hydroxid, and add, while still hot, ammonium oxalate solution, drop by drop, slightly in excess of complete precipitation, to convert the magnesium also into oxalate. Heat to boiling, allow the precipitate to settle completely, decant the clear solution on a filter, pour 15–20 cc. of hot water on the precipitate, and again decant the clear solution on the filter. Dissolve the precipitate in the beaker with a few drops of hydrochloric acid, add a little water, repeat the precipitation as above, and filter through the same filter; transfer the precipitate to the filter and wash free from chlorin with hot water; dry, ignite the precipitate over the blast lamp to constant weight, and weigh as calcium oxid. Designate the filtrate and washings as *D*.

16**MAGNESIUM.—OFFICIAL.**

Evaporate *D*, under **15**, to dryness on the water bath and heat carefully to expel ammonium salts. Treat the residue with 20–25 cc. of hot water and about 5 cc. of hydrochloric acid, filter and wash. Concentrate to about 50 cc., cool, and add sufficient disodium hydrogen phosphate solution to precipitate the magnesium; then add gradually ammonium hydroxid, with constant stirring, until the solution is distinctly alkaline. Determine if the precipitation is complete by the addition of more of the disodium hydrogen phosphate solution. After 30 minutes, add gradually 10 cc. of strong ammonium hydroxid, cover to prevent the escape of ammonia, and let stand in the cold. Filter after 12 hours, wash the precipitate free from chlorin, using dilute ammonium hydroxid [**I**, **4** (**d**)], dry the filter and precipitate and transfer the latter to a weighed porcelain crucible. Ignite the filter separately and add its ash to the precipitate in the crucible. Burn at first at a moderate heat, then ignite to whiteness or to a grayish white, weigh as magnesium pyrophosphate ($\text{Mg}_2\text{P}_2\text{O}_7$) and calculate to magnesium oxid (MgO).

PHOSPHORIC ACID.

17

Gravimetric Method.—Official.

Concentrate 100–200 cc. of *A*, under **11**, to about 25–30 cc., neutralize with ammonium hydroxid and add about 10 cc. additional. Dissolve the precipitate by the slow addition of dilute nitric acid, stirring constantly and avoiding a large excess, add gradually about 20 cc. of molybdate solution [**I, 4 (a)**], and allow to stand for 1 or 2 hours in a water bath at a temperature of 40°C. After an hour determine if the precipitation is complete, as follows: Pipette about 5 cc. of the clear liquid into 5 cc. of warm molybdate solution. If any precipitate is produced, return the test liquid to the main portion, add more molybdate solution, and repeat the operation until all the phosphoric acid is precipitated. Then allow to stand for several hours at room temperature, preferably overnight. Filter off the ammonium phosphomolybdate, wash the precipitate thoroughly with cold water, dissolve with ammonium hydroxid, and determine as magnesium pyrophosphate, as directed under **I, 6** and calculate to phosphorus pentoxid (P_2O_5).

Volumetric Method.—Tentative.

18

REAGENTS.

(a) *Standard sodium or potassium hydroxid solution.*—Strength such that 1 cc. of this solution is equivalent to 0.0005 gram of phosphorus pentoxid (P_2O_5).

(b) *Standard nitric acid solution.*—Strength same as the standard alkali solution described under (a) as determined by titration, using phenolphthalein as indicator.

19

DETERMINATION.

Proceed as in **17** until all the phosphoric acid is precipitated and then complete the determination in the following manner:

Allow the solution containing the yellow precipitate to stand for at least 3 hours at a temperature not above 40°C., filter on a small filter paper or on a Gooch crucible and wash with cold water until the filtrate from 2 fillings of the filter yields a slight pink color on the addition of phenolphthalein and 1 drop of the standard alkali. Return the filter and precipitate to the same beaker used for precipitating the phosphomolybdate, dissolve the yellow precipitate in the standard sodium or potassium hydroxid solution, add a few drops of phenolphthalein and titrate the excess of alkali with the standard acid. Calculate to phosphorus pentoxid (P_2O_5).

20

SULPHURIC ACID.—OFFICIAL.

Evaporate 100–200 cc. of *A*, under **11**, nearly to dryness on a water bath to expel the excess of acid, add 50 cc. of water, heat to boiling and add, drop by drop, 10% barium chlorid solution until no further precipitation occurs. Continue the boiling for about 5 minutes and allow to stand for 5 hours or longer in a warm place. Decant the liquid on an ashless filter or tared Gooch, previously heated, treat the precipitate with 15–20 cc. of boiling water, transfer to the filter and wash free from chlorin with boiling water. Dry the precipitate and filter, ignite, weigh as barium sulphate and calculate to sulphur trioxid (SO_3).

POTASSIUM AND SODIUM.

21

Method I.—Official.

(1) Treat the filtrate from **20** with ammonium hydroxid exactly as in **13**.

Evaporate the filtrate and washings to dryness, heat below redness until ammonium salts are expelled, dissolve in hot water, add 5 cc. of barium hydroxid solution,

and heat to boiling; let settle for a few minutes, and determine if the precipitation is complete by the addition of barium hydroxid solution to a little of the clear liquid. When no further precipitate is produced, filter and wash thoroughly with hot water. Heat the filtrate to boiling, add ammonium hydroxid and ammonium carbonate to complete the precipitation of the barium, calcium, etc., let stand a short time on the water bath, filter, and wash the precipitate thoroughly with hot water; evaporate the filtrate and washings to dryness, expel ammonium salts by heating below redness, treat with a little hot water, add a few drops of ammonium hydroxid, 1 or 2 drops of ammonium carbonate, and a few drops of ammonium oxalate; let stand a few minutes on the water bath, set aside for a few hours, filter, evaporate to complete dryness on the water bath, and heat to dull redness until all ammonium salts are expelled and the residue is nearly or quite white. Dissolve in a minimum amount of water, filter into a tared platinum dish, add a few drops of hydrochloric acid, evaporate to dryness on the water bath, heat to dull redness, cool in a desiccator, and weigh as potassium and sodium chlorids. Repeat the heating until constant weight is obtained. Dissolve in a small amount of water; if any residue remains, the separation must be repeated until the residue of potassium and sodium chlorids is entirely soluble. Dissolve the residue with water, add an excess of platinic chlorid solution [I, 40 (b)], proceed as directed under I, 45 and calculate to potassium oxid (K_2O); or, (2) Instead of the foregoing, evaporate to dryness a fresh aliquot of A, under 11, redissolve in water, treat directly with barium hydroxid solution, and from this point proceed as directed above in (1).

22

Method II.—Tentative.

Proceed as in 21 through "let stand a short time on the water bath" (the point at which the barium, calcium, etc., have been precipitated with ammonium hydroxid and ammonium carbonate) and then proceed as follows:

Filter into a beaker, add 1 or 2 drops of hydrochloric acid and 1 cc. of ammonium sulphate (75 grams to 1 liter), digest several hours on a water bath, and filter into a tared platinum dish. Evaporate to dryness, heat to full redness, add 1 gram of powdered ammonium carbonate; heat to expel excess of ammonium carbonate, cool, and weigh the sulphates of sodium and potassium. Determine potassium as directed under I, 42 (a) and calculate to potassium oxid (K_2O).

TOTAL PHOSPHORUS.

23

Magnesium Nitrate Method.—Official.

Place 5 grams of soil, as prepared under 2, in a porcelain dish. Moisten with 5–7 cc. of magnesium nitrate solution [I, 4 (e)]. Dry on the water bath and burn off the organic matter at low redness. Cool, moisten slightly with water, add 10 cc. of concentrated hydrochloric acid, and digest 2 hours on the water bath, keeping the dish covered with a watch glass and stirring 2 or 3 times during the digestion. Transfer to a 250 cc. graduated flask, cool, fill to the mark, mix well, and pass through a dry folded filter, pouring back on the filter until the filtrate becomes clear. Pipette an aliquot corresponding to 2 or 4 grams of the soil, depending upon the amount of phosphorus present, into a porcelain dish, evaporate to dryness, treat with hydrochloric acid and water, filter, and wash; the combined volume should not exceed 40 cc. Make alkaline with ammonium hydroxid, and dissolve the precipitate by the slow addition of concentrated nitric acid, using a slight excess. Add gradually, while shaking, 5–15 cc. of molybdate solution [I, 7 (a)]. Keep the solution at 40°–50°C. for an hour, let stand overnight at room temperature, filter, and wash well with cold water. Return the filter and precipitate to the same flask and determine phosphorus volumetrically, as directed under 19.

24

Sodium Peroxid Method.—Official.

Place 10 grams of sodium peroxid in an iron or porcelain crucible and thoroughly mix with 5 grams of the soil as prepared under 2. If the soil has very little organic matter, add a little starch to hasten the action. Heat the mixture carefully by applying the flame of a Bunsen burner directly upon the surface of the charge and the sides of the crucible until the action starts. Cover the crucible until the reaction is over and keep at a low red heat for 15 minutes. Do not allow fusion to take place. By means of a large funnel and a stream of hot water, wash the charge into a beaker, acidify with hydrochloric acid and boil. Transfer to a 500 cc. graduated flask, cool and fill to the mark. If the action has taken place properly there should be no undecomposed soil in the bottom of the flask. Allow the silica to settle and draw off 200 cc. of the clear solution.

Precipitate the iron, aluminium, and phosphorus with ammonium hydroxid; filter, wash several times with hot water, wash the precipitate back into the beaker with a stream of hot water, and dissolve the precipitate in hot hydrochloric acid, pouring the acid upon the filter to dissolve any precipitate adhering to it. Evaporate the solution and washings to dryness on a water bath. Treat with dilute hydrochloric acid, heating if necessary, and remove the silica by filtration. Concentrate the filtrate and washings to about 10 cc., add 2 cc. of strong nitric acid, and make alkaline with ammonium hydroxid. Add nitric acid very slowly and with constant stirring until the solution is clear, avoiding an excess. Heat at 40°–50°C. on a water bath, add 15 cc. of molybdate solution, [I, 7 (a)], and maintain this temperature for 1–2 hours. Let stand overnight, filter, and wash free from acid with 0.1% solution of ammonium nitrate, and, finally, once or twice with cold water. Transfer the filter and precipitate to the same beaker and determine phosphorus volumetrically as directed under 19.

25

TOTAL POTASSIUM.—OFFICIAL.

Decompose the soil by the J. L. Smith method⁴ as follows: Triturate gently 0.5 or 1 gram of the finely ground soil with 1 gram of dry ammonium chlorid in a smooth mortar, then add 8 parts of calcium carbonate and mix intimately. Transfer the mixture to a platinum crucible, rinsing the mortar with a little calcium carbonate. Heat the crucible gradually until fumes of ammonium salts no longer appear, and continue until the lower *three-fourths only* of the crucible are brought to a red heat. Maintain this temperature 40–60 minutes. The temperature should be sufficient to keep the calcium chlorid formed by the reaction of ammonium chlorid with calcium carbonate in a state of fusion. The mass, however, does not become liquid since the fused calcium chlorid is absorbed by the large quantity of calcium carbonate present. If the silicate is fused by the application of too strong heat, disintegration of the mass at the end of the operation with water cannot be effected. Moreover, too high a temperature causes volatilization of alkali chlorids. The mass contracts in volume during the ignition, and is usually easily detached from the crucible. Transfer the fused mass to a porcelain dish, thoroughly slake with hot water, and grind thoroughly with an agate pestle. After washing 5 times by decantation with hot water, transfer to a filter and wash well, 300 cc. of wash water being sufficient. To the filtrate add 10 cc. of concentrated hydrochloric acid, and evaporate nearly to dryness in a porcelain dish. Treat with hot water and 2 cc. of hydrochloric acid and filter by means of suction through a small filter into a 150 cc. Jena beaker. Concentrate the solution to 30 cc., add 1.5 cc. of platinic chlorid solution [I, 40 (b)], evaporate to a sirupy consistency, and add 15 cc. of 2.25 N/1 acidulated alcohol (prepared by passing hydrochloric acid gas into a mix-

ture of 2000 cc. of 95% alcohol and 152 cc. of hydrochloric acid, sp. gr. 1.20). Filter by means of suction through a small filter, wash with 80% alcohol, then with ammonium chlorid solution [I, 40 (a)], and finally with 80% alcohol. Dry the precipitate on the filter and wash the precipitate with hot water into a weighed platinum dish, using suction. Evaporate to dryness, heat in a drying oven for an hour at 120°C., cool in a desiccator, weigh and calculate to potassium oxid (K_2O).

TOTAL ALKALIES.

26

*J. Lawrence Smith Method.*⁴—Official.

(1) Proceed as directed under 25 to the point indicated by the phrase "300 cc. of wash water being sufficient" (the point at which the ignited mass has been disintegrated and thoroughly washed with water). The filtrate contains the silicate alkalies in the form of chlorids together with calcium chlorid and hydroxid. Precipitate the calcium at once with ammonium carbonate solution; allow to settle, decant the supernatant liquid into a porcelain (or platinum) dish, concentrate and finally transfer the precipitate to the dish. When the volume is reduced to about 30 cc., add a little ammonium carbonate solution and ammonium hydroxid, heat and filter into a porcelain (or platinum) dish, evaporate the filtrate to dryness on a water bath and expel ammonium salts by ignition. Dissolve the residual alkali chlorids in 3-5 cc. of water; a little black or dark brown flocculent matter usually remains undissolved, while the solution may also contain traces of calcium. Add 2-3 drops of ammonium carbonate and ammonium hydroxid, warm gently, and filter through a very small filter into a weighed platinum vessel. Evaporate to dryness on a water bath, heat the alkali chlorids to incipient fusion, cool, and weigh as sodium and potassium chlorids; or,

(2) Determine, by the above method, the quantity of alkalies in the insoluble residue, 12, and add that obtained under 21 or 22.

27

PHOSPHORUS SOLUBLE IN N/5 NITRIC ACID.—TENTATIVE.

Digest 10 grams of air-dried soil in a stoppered flask, with 100 cc. of N/5 nitric acid, for exactly 5 hours in a water bath kept at a temperature of 40°C. Filter the solution through a dry paper, cool to room temperature, and titrate 20 cc. of the filtrate with standard potassium hydroxid solution (carbonate-free), using phenolphthalein as indicator. From the data thus secured calculate the number of cc. of N/1 acid and of water to make exactly 1 or 2 liters of acid of N/5 strength after allowing for the quantity neutralized by the amount of soil to be used in the following procedure:

Place 200 grams of the air-dried soil in a large, dry, glass-stoppered bottle and add exactly 2000 cc. of N/5 nitric acid corrected for neutralization as above described. With soils rich in available phosphoric acid, 100 grams of soil and 1000 cc. of acid will be sufficient. Digest in a large water bath at a temperature of 40°C. for exactly 5 hours, shaking thoroughly each half hour. At the end of the digestion shake the contents of the bottle well and pour quickly upon a large, dry, ribbed filter of 2 thicknesses of paper and of sufficient size to receive the entire contents of the bottle. Collect the filtrate in a dry vessel, pouring back on the filter until the filtrate becomes clear. Evaporate 1000 or 500 cc. of the filtrate, according to the quantity of soil used, to dryness in a porcelain dish; add a few cc. of nitric acid to oxidize organic matter, etc., moisten the residue with hydrochloric acid, digest with water, and filter into a 500 cc. flask. Add a solution containing 15 grams of ammonium nitrate; then strong ammonium hydroxid until a permanent precipitate forms, and then concentrated nitric acid slowly until the precipitate dissolves.

Dilute to about 100 cc., if less than that volume, place a thermometer in the flask, and heat to 85°C. Add 75 cc. of recently prepared molybdate solution [I, 4 (a)], digest in a water bath at 80°C. for 15 minutes, with occasional shaking, remove from the bath and allow to stand at least 10 minutes before filtering. Continue the determination as directed under I, 6 and calculate to phosphorus pentoxid (P_2O_5).

28

CALCIUM CARBONATE REQUIRED.—TENTATIVE.

Place 100 grams of soil, as prepared under 2, in a 400 cc. wide-mouthed bottle, add 250 cc. of N/1 potassium nitrate, stopper, and shake continuously for 3 hours in a shaking machine, or every 5 minutes by hand. Let stand overnight. Draw off 125 cc. of the clear, supernatant liquid, boil 10 minutes to expel carbon dioxide, cool, and titrate with standard sodium hydroxid solution, 1 cc. of which is equivalent to 4 mg. of calcium carbonate (0.001% on basis of the weight of soil used), using phenolphthalein as indicator.

29

STATEMENT OF RESULTS.—OFFICIAL.

Calculate all results of soil analysis as per cent of the soil dried to constant weight as under 3 and state in the following order:

Insoluble residue.....
Soluble silica.....
Manganomanganic oxid (Mn_2O_3).....
Potassium oxid (K_2O).....
Sodium oxid (Na_2O).....
Calcium oxid (CaO).....
Magnesium oxid (MgO).....
Ferric oxid (Fe_2O_3).....
Aluminium oxid (Al_2O_3).....
Phosphorus pentoxid (P_2O_5).....
Sulphur trioxid (SO_3).....
Organic carbon.....
Inorganic carbon.....
Volatile matter.....
Total nitrogen.....
Total phosphorus.....
Total potassium.....
Phosphorus soluble in N, 5 acid.....
Calcium carbonate required.....
Total.....

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III. PLANT CONSTITUENTS.

1 PREPARATION OF SAMPLE.—OFFICIAL.

Thoroughly cleanse the material from all foreign matter, especially from adhering soil, air-dry, grind, and preserve the sample in tightly stoppered bottles.

2 PREPARATION OF ASH.—OFFICIAL.

Ignite 10-20 grams of the substance, in a flat-bottomed platinum dish in a muffle, at a comparatively low temperature. Do not employ a full red heat because of the danger of volatilizing alkali chlorids, etc. If rich in silica and alkalies, char the material, treat with water to dissolve soluble salts, filter through an ashless filter, dry the filter and paper and incinerate, add the filtrate to the incinerated residue, evaporate to dryness and ignite at a low red heat. If rich in phosphates, e.g., seeds and animal substances, char the material, dissolve soluble salts in dilute acetic acid, filter through an ashless filter, wash with water, dry and incinerate the filter and residue, add the filtrates to the incinerated residue, evaporate to dryness, and ignite gently. While still warm, pulverize the whole of the ash as obtained above, mix intimately and preserve in a tightly stoppered, dry bottle. If after incineration the ash has absorbed moisture, dry thoroughly at low redness before bottling.

3 CARBON DIOXID.—OFFICIAL.

Determine carbon dioxid in a weighed portion of the ash prepared under 2. Liberate the carbon dioxid by treatment with dilute hydrochloric acid in any of the usual forms of apparatus, and determine the increase in weight of the potash bulbs. The efficiency of the apparatus should be tested by blank determinations conducted upon weighed portions of pure calcite.

4 CARBON, SAND AND SILICA.—OFFICIAL.

Transfer the residue from the carbon dioxid determination to a beaker or evaporating dish; evaporate to dryness; pulverize and dry thoroughly to render the silica insoluble. Moisten the dry residue with 5-10 cc. of hydrochloric acid, add about 50 cc. of water, allow to stand on the water bath for a few minutes, filter through a hardened filter and wash thoroughly. Dilute the solution and washings to 250 cc. or other convenient volume. Designate as A.

Wash the residue from the filter into a platinum dish and boil for about 5 minutes with approximately 20 cc. of a saturated solution of pure sodium carbonate, add a few drops of pure sodium hydroxid solution, allow the solution to settle and decant through a tared Gooch filter. Boil the residue in the dish with sodium carbonate solution and decant as before. Repeat the process again, then transfer the residue to the Gooch filter, wash thoroughly, first with hot water, then with a little dilute hydrochloric acid, and finally with hot water until free from chlorin. Dry the filter and contents to constant weight at 110°C. to determine the combined weight of carbonaceous material and sand. Incinerate; the loss in weight represents the carbonaceous material; the residue is sand. Confirm by microscopic examination. Determine the soluble silica as follows: (1) Combine the alkaline filtrate and washings, acidify with hydrochloric acid, evaporate to dryness and determine the silica

in the usual way; or, (2) Treat a weighed portion of the ash, as prepared under **2**, with dilute hydrochloric acid. Evaporate to dryness; pulverize and dry thoroughly to render the silica insoluble. Moisten the dry residue with 5–10 cc. of hydrochloric acid, add about 50 cc. of water, allow to stand on the water bath for a few minutes, filter on an ashless filter, wash, dry, ignite and weigh to determine the combined weight of the silica and sand. Deduct the weight of the sand found above to obtain that of the silica. The soluble silica cannot be separated from the residue after ignition.

5**CARBON-FREE ASH.—OFFICIAL.**

Subtract the weights of the carbon found in **4** and the carbon dioxide found in **3** from that of the total ash used in **3**.

6**FERRIC AND ALUMINIUM OXIDS.—OFFICIAL.**

(Applicable for plant materials other than seeds.)

Pipette an aliquot of *A*, under **4**, corresponding to 0.5 gram of ash, into a 250 cc. beaker. If ferrous iron is present, oxidize it by boiling with a few cc. of hydrogen peroxid or of concentrated nitric acid. Cool, add ammonium hydroxid until a precipitate begins to form, then nitric acid until just clear, and finally add 2–3 cc. of concentrated nitric acid in excess. Add 25 cc. of 50% ammonium nitrate solution, phosphate free, heat to 40°C., and add slowly, with constant shaking, a moderate excess of molybdate solution [**I, 4 (a)**], and allow to stand for 1 or 2 hours at a temperature not exceeding 40°C. After standing for an hour pipette 5 cc. of the clear solution into an equal volume of warm molybdate solution. If a precipitate forms in the test portion return it to the original solution and add more molybdate solution. Allow to stand at room temperature for several hours, preferably overnight. Filter, wash with about 75 cc. of ammonium nitrate solution (2.5%, phosphate free, and slightly acidified with nitric acid) and combine the filtrate and washings. Designate as *B*. Reserve the precipitate for the determination of phosphoric acid as described under **11**.

Without concentrating solution *B*, cautiously neutralize with ammonium hydroxid, add a very slight excess of the alkali, avoiding a temperature higher than 40°C., and allow to stand at this temperature until the precipitate completely settles. Filter the clear supernatant liquid, wash the precipitate a few times by decantation with hot water before transferring to the filter, then wash 4 or 5 times on the filter. Dissolve the precipitate on the filter with hot nitric acid (1 to 5), wash and reprecipitate as before. The combined filtrates and washings from the first and second precipitations should not exceed 500 cc. and should not be concentrated by evaporation. Designate as *C* and reserve for the determination of calcium and magnesium as described under **8**. The same filter may be used for the second filtration, and the volume of the solution for the reprecipitation need not exceed 100 cc. Before the second filtration is made, a small quantity of ashless filter paper pulp should be added in order to facilitate the washing and leave the precipitate finely divided after the ignition, so that it can be easily fused with potassium hydrogen sulphate for the iron determination. Dry and ignite the precipitate and weigh as ferric and aluminium oxids.

The iron oxid is determined in the following manner: Fuse, in a platinum crucible, the ignited precipitate with about 4 grams of fused potassium hydrogen sulphate. This fusion takes but a few minutes and must not be continued beyond the time actually needed. When completed the crucible is set aside and allowed to cool. Add 5 cc. of concentrated sulphuric acid and heat until copious fumes of sulphuric

acid are given off. Cool, transfer to a flask, add water, and digest till the solution is clear. Reduce with zinc, cool, titrate with N/50 potassium permanganate and calculate to ferric oxid.

If it is desired to use a larger amount of the sample for the iron determination, evaporate a suitable aliquot of *A*, under **4**, with sulphuric acid, reduce with zinc, and titrate as above.

MANGANESE, CALCIUM AND MAGNESIUM.

(Applicable for plant materials other than seeds.)

7

Method I.—Official.

To an aliquot of *A*, under **4**, corresponding to 0.5–2 grams of ash, add a quantity of pure ferric chlorid solution, more than sufficient to combine with the phosphoric acid which may be present, and neutralize with ammonium hydroxid. Dissolve the precipitate in a very slight excess of hydrochloric acid and add 1–2 grams of sodium acetate. Boil for 1–2 minutes, filter at once, and wash with boiling water. Dissolve the precipitate in hydrochloric acid and reprecipitate as above. Concentrate the combined filtrates and washings to about 50 cc. and determine manganese, calcium and magnesium as directed under **II**, **14**, **15**, and **16**.

8

Method II.—Tentative.

Calcium.—Make alkaline with ammonium hydroxid the combined filtrates and washings, *C*, under **6**, and proceed as directed under **II**, **15**. If the ignited calcium oxid has a brown coloration, due to manganese, dissolve in dilute hydrochloric acid and determine the manganese as directed below. Deduct the weight of manganomanganic oxid thus obtained from the weight of the impure calcium oxid.

Manganese.—Acidify the combined filtrates and washings from the calcium determination and evaporate to dryness in a porcelain casserole. Expel the ammonium salts by carefully heating the casserole from above, treat with a few cc. of hydrochloric acid and water, filter off molybdic acid, and wash the precipitate until it is free from chlorin. Bring the filtrate to a volume of 100 cc., add 1–2 drops of bromin, make alkaline with ammonium hydroxid and let stand for several minutes without agitation. Filter off the precipitated manganese, wash, dry, ignite, and weigh the precipitate as manganomanganic oxid. To this add the weight of the manganomanganic oxid found as an impurity in the calcium determination.

Magnesium.—Concentrate the alkaline filtrate from the manganese determination to 75 cc. and determine magnesium as directed under **II**, **16**.

PHOSPHORIC ACID.

9

Method I.—Official.

Determine phosphoric acid in an aliquot of *A*, under **4**, corresponding to 0.2–1 gram of ash, as directed under **I**, **6** or **9**.

10

Method II.—Official.

Determine phosphoric acid in the plant substance as directed under **I**, **6**, using sufficient material to give 0.2–1 gram of ash in the aliquot of the solution employed.

11

Method III.—Official.

The phosphomolybdate precipitate obtained in **6** is used for the determination of phosphoric acid as directed under **I**, **6**, beginning with "Dissolve the precipitate

on the filter with ammonium hydroxid, etc." or as under **I, 9 (a)**, beginning with "Wash with cold water until the filtrate from 2 fillings of the filter yields a pink color, etc."

12 **SULPHURIC ACID, SODIUM, AND POTASSIUM.—OFFICIAL.**

Boil an aliquot of **A**, under **4**, corresponding to 0.5–1 gram of ash, add barium chlorid solution in small quantities until no further precipitate is formed, and proceed as directed under **II, 20** and **21**.

13 **CHLORIN.—OFFICIAL.**

Determine as silver chlorid, either gravimetrically or by one of the standard volumetric procedures (as the Volhard method given below), in a nitric acid or aqueous solution of the ash.

*Volhard Method.*¹

14

REAGENTS.

(a) *N/10 silver nitrate.*

(b) *N/10 ammonium or potassium sulphocyanate.*

(c) *Ferric indicator.*—Saturated solution of ferric alum.

(d) *Nitric acid.*—Free from lower oxids of nitrogen, secured by diluting the usual pure acid with about 1/4 part of water, and boiling till perfectly colorless.

15

DETERMINATION.

Dissolve a weighed portion of the ash, prepared under **2**, in nitric acid (1 to 10), filter and wash with water. Add a known volume of the *N/10* silver nitrate in slight excess to the combined filtrate and washings. Stir well, filter, and wash the silver chlorid precipitate thoroughly. To the filtrate and washings add 5 cc. of the ferric indicator and a few cc. of the nitric acid. Titrate the excess of silver with the *N/10* sulphocyanate until a permanent light brown color appears. Calculate the amount of chlorin.

16

POTASSIUM IN PLANTS.—OFFICIAL.

Determine potassium as directed under **I, 42**, using sufficient plant material to yield 0.5–1 gram of ash in the aliquot of the solution used for the potassium determination.

SULPHUR IN PLANTS.

17

Peroxid Method.—Official.

Place 1.5–2.5 grams of material in a nickel crucible of about 100 cc. capacity and add 5 grams of pure anhydrous sodium carbonate. Mix thoroughly, using a nickel or platinum rod, and moisten with approximately 2 cc. of water. Add pure sodium peroxid, approximately 0.5 gram at a time, thoroughly mixing the charge after each addition. Continue adding the peroxid until the mixture becomes nearly dry and quite granular, requiring usually about 5 grams of peroxid. Place the crucible over a low alcohol or other sulphur-free flame and heat carefully with occasional stirring until the contents are fused. (Should the material ignite, the determination is worthless.) After fusion remove the crucible, allow to cool somewhat, and cover the hardened mass with peroxid to a depth of about 0.5 cm. Heat gradually, and finally with full flame until fusion again takes place, rotating the crucible from time to time in order to bring any particles adhering to the sides into contact with the oxidizing material. Continue the heating for 10 minutes after fusion is complete.

Cool somewhat, place the warm crucible and contents in a 600 cc. beaker and carefully add about 100 cc. of water. After the initial violent action has ceased, wash the material out of the crucible, make slightly acid with hydrochloric acid (adding small portions at a time), transfer to a 500 cc. flask, cool, and make to volume. Filter, and determine sulphates in 200 cc. of the filtrate as directed under **II, 20**.

18**CHLORIN IN PLANTS.—TENTATIVE.**

Moisten 5 grams of the substance in a platinum dish with 20 cc. of a 5% solution of sodium carbonate, evaporate to dryness, and ignite as thoroughly as possible at a temperature not exceeding dull redness. Extract with hot water, filter and wash. Return the residue to the platinum dish and ignite to an ash; dissolve in nitric acid, add this solution to the water extract and determine chlorin as directed under **15**.

BIBLIOGRAPHY.

¹ Sutton. Volumetric Analysis. 10th ed., 1911, p. 145.

IV. WATERS.

POTABLE WATER.

TURBIDITY.—TENTATIVE.

1

REAGENTS.

(a) *Standard turbidity solution*.—Weigh out 1 gram of elutriated fuller's earth previously dried and sifted through a 200 mesh sieve. Make up to a liter. If the fuller's earth is of good quality and the proper degree of fineness, this stock solution has a turbidity of 1000. Check the stock solution with a Jackson turbidimeter.

(b) *Turbidity standards*.—Prepared by dilution of (a).

2

DETERMINATION.

Determine the turbidity of the sample with a Jackson turbidimeter equipped with either candle or electric light. If the turbidity is less than 100, which prohibits the use of the turbidimeter, determine by direct comparison with turbidity standards contained in bottles of clear white glass.

COLOR.—TENTATIVE.

3

REAGENTS.

(a) *Standard color solution*.—Dissolve 1.246 grams of potassium platonic chlorid ($\text{PtCl}_4\cdot 2\text{KCl}$) and 1 gram of crystallized cobalt chlorid ($\text{CoCl}_2\cdot 6\text{H}_2\text{O}$) in a small quantity of water, add 100 cc. of concentrated hydrochloric acid and make up to 1 liter with water. This stock solution has a color of 500.

(b) *Color standards*.—Prepared by dilution of (a).

4

DETERMINATION.

Compare the color of the sample, freed from suspended matter, with color standards in tubes of clear white glass.

5

ODOR.—TENTATIVE.

Shake the vessel containing the sample and note the odor. Heat a portion of the sample just to boiling and note the odor.

6

TOTAL SOLIDS.—OFFICIAL.

Thoroughly shake the vessel containing the sample and pipette 100 cc. of the unfiltered water into a weighed platinum dish. If the sample contains much suspended matter, shake, pour rapidly into a 100 cc. measuring cylinder, and transfer without delay to a weighed platinum dish; evaporate to dryness and heat to constant weight at 105°C. In the case of highly mineralized waters it is advisable to weigh again after drying at 180°C.

7

SOLIDS IN SOLUTION.—OFFICIAL.

Allow the sample to stand until all sediment has settled, and filter if necessary to secure a perfectly clear liquid. Occasionally a clear filtrate can only be obtained by the use of alumina cream but this should be avoided if possible. Evaporate 100—

250 cc. to dryness in a weighed platinum dish. Heat to constant weight at 105°C. In the case of highly mineralized waters it is advisable to weigh again after drying at 180°C.

8**SUSPENDED MATTER.—OFFICIAL.**

(1) The difference between the values for total solids and dissolved solids represents the suspended matter; or, (2) Determine the suspended matter directly by filtering a suitable quantity of the water through a tared Gooch crucible, suitably prepared, and weighing after drying at 105°C.

9**IGNITED RESIDUE.—OFFICIAL.**

Ignite the residue from **6** at a low red heat until the ash is white or nearly so. Note any odor or change in color produced during ignition. Record the weight of the ignited residue and calculate the loss on ignition.

FREE AND ALBUMINOID AMMONIA.—OFFICIAL.**10****REAGENTS.**

(a) *Saturated solution of sodium carbonate.*

(b) *Ammonia-free water.*

(c) *Standard ammonium chlorid solution.*—One cc. is equivalent to 0.01 mg. of nitrogen in the form of ammonia (NH_3).

(d) *Nessler reagent.*—Dissolve 50 grams of potassium iodid in a minimum quantity of cold water. Add a saturated solution of mercuric chlorid until a slight permanent precipitate is formed. Add 400 cc. of 50% solution of potassium hydroxid (or an equivalent quantity of sodium hydroxid), dilute to 1 liter, allow to settle, and decant.

(e) *Alkaline potassium permanganate solution.*—Dissolve 200 grams of potassium hydroxid and 8 grams of potassium permanganate in water and dilute to 1 liter.

11**DETERMINATION.**

Connect a flask of about 1500 cc. capacity with an upright bulb condenser by means of a rather large glass tube and a soft rubber stopper or a recently extracted cork stopper. Place in the flask 5 cc. of the saturated solution of sodium carbonate and 500 cc. of ammonia-free water. Distil into 50 cc. Nessler tubes until no further traces of ammonia are indicated on the addition of 2 cc. of the Nessler reagent to 50 cc. of the distillate. Continue the distillation until the volume of the solution in the flask has been reduced to about 200 cc. Cool slightly, add 500 cc. of the water under examination, and distil, at the rate of 1 tubeful in 15 minutes, into 50 cc. Nessler tubes until ammonia ceases to be given off (4 or 5 tubes are usually sufficient). Add 2 cc. of the Nessler reagent to each tube and let stand 10 minutes. Freshly prepare in a similar manner other tubes containing known amounts of the standard ammonium chlorid, made up to 50 cc. with ammonia-free water, and compare the nesslerized distillates with these. Report as milligrams per liter of nitrogen in the form of free ammonia (NH_3). Cool the flask and add 50 cc. of the alkaline permanganate recently boiled. Distil, at the rate of 1 tubeful in 15 minutes, into 50 cc. Nessler tubes until ammonia ceases to come off. Nesslerize and compare as in the determination of free ammonia. Report as milligrams per liter of nitrogen in the form of albuminoid ammonia (NH_3).

NITROGEN IN THE FORM OF NITRITE.—OFFICIAL.

12

REAGENTS.

- (a) *Concentrated hydrochloric acid*.—Sp. gr. 1.2.
- (b) *Sulphanilic acid solution*.—Dissolve 1 gram of sulphanilic acid in 100 cc. of hot water.
- (c) *Alpha-naphthylamin hydrochlorid solution*.—Boil 0.5 gram of the salt with 100 cc. of water for 10 minutes at constant volume.
- (d) *Standard nitrite solution*.—Dissolve 1.1 gram of silver nitrite in nitrite-free water, precipitate the silver with sodium chlorid solution and dilute to 1 liter, mix and allow to settle. Dilute 100 cc. to 1 liter and then 10 cc. of this solution to 1 liter, using in each case nitrite-free water. Each cc. of the last solution is equivalent to 0.0001 mg. of nitrogen as nitrite.

13

DETERMINATION.

Place 100 cc. of the water in a 100 cc. Nessler tube and treat with 1 or 2 drops of concentrated hydrochloric acid. Add 1 cc. of the sulphanilic acid, 1 cc. of the alpha-naphthylamin hydrochlorid, and thoroughly mix. Set aside for 30 minutes with other Nessler tubes containing known amounts of the standard nitrite made up to 100 cc. with nitrite-free water, and treated with hydrochloric acid, sulphanilic acid, and alpha-naphthylamin hydrochlorid in the manner just described. Determine the amount of nitrite by comparing the depth of pink color in the known and unknown solutions. Record as nitrogen in the form of nitrite.

NITROGEN IN THE FORM OF NITRATE.

Phenoldisulphonic Acid Method.—Official.

(For water of low chlorin content.)

14

REAGENTS.

- (a) *Phenoldisulphonic acid solution*.—Dissolve 25 grams of pure white phenol in 150 cc. of concentrated sulphuric acid, add 75 cc. of fuming sulphuric acid (13–15% SO_3) and heat at 100°C. for 2 hours.
- (b) *Standard nitrate solution*.—Dissolve 0.722 gram of pure potassium nitrate in 1 liter of nitrate-free water. Evaporate 50 cc. of this solution to dryness in a porcelain dish; treat with 2 cc. of the phenoldisulphonic acid solution, rubbing with a glass rod to insure intimate contact. Dilute to 500 cc.; 1 cc. is equivalent to 0.01 mg. of nitrogen as nitrate. This solution is permanent. Standards for comparison are prepared by adding ammonium hydroxid to measured volumes of it in 100 cc. Nessler tubes.
- (c) *Standard silver sulphate solution*.—Dissolve 4.3969 grams of silver sulphate, free from nitrate, in 1 liter of water; 1 cc. is equivalent to 1 mg. of chlorin.
- (d) *Ammonium hydroxid*.—Sp. gr. 0.90.

15

DETERMINATION.

Take 100 cc. of the sample, or an amount that will contain 0.05 mg. or less of nitrogen as nitrate, and add standard silver sulphate, precipitating all but about 0.5 mg. of the chlorin. Heat to boiling, allow to settle, or add a little alumina cream, filter, and wash with small amounts of hot water. Evaporate the filtrate to dryness in a porcelain dish on the water bath; when cool, treat with 2 cc. of the phenoldisulphonic acid solution as in 14 (b). Dilute with water and add slowly ammonium hy-

dioxid until the maximum color is developed. Transfer to a colorimetric cylinder, filter if necessary, and compare with the standards in the usual manner. Record as nitrogen in the form of nitrate.

Reduction Method.—Official.

(For water of high chlorin content.)

16

REAGENTS.

(a) *Sodium or potassium hydroxid solution.*—Dissolve 250 grams of the purest hydroxid obtainable in 1250 cc. of water and boil down to 1 liter.

(b) *Aluminium foil.*—Use strips about 10 cm. long, weighing about 0.5 gram.

17

DETERMINATION.

Place 10 cc. of the sample in a 100 cc. test tube and dilute to exactly 50 cc.; add 5 cc. of the sodium hydroxid and a strip of the aluminium foil. Close the mouth of the test tube with a rubber stopper carrying a U-shaped glass tube connected with a second test tube containing about 50 cc. of ammonia-free water acidified with hydrochloric acid, which serves as a trap to catch any ammonia which might escape. Allow to stand at room temperature for 12 hours or more until reduction is complete. Transfer the contents of the first tube to a Kjeldahl flask and distil. Cool the distillates and nesslerize as directed under 11; also nesslerize the solution in the trap. Record as nitrogen in the form of nitrate.

CHLORIN.—OFFICIAL.

18

REAGENTS.

(a) *N/20 sulphuric acid.*

(b) *N/20 sodium carbonate.*

(c) *Potassium chromate indicator.*—Dissolve 5 grams of potassium chromate in water, add a solution of silver nitrate until a slight permanent red precipitate is produced, filter, and dilute to 100 cc.

(d) *Standard silver nitrate solution.*—Dissolve 4.791 grams of silver nitrate in water and dilute to 1 liter; 1 cc. is equivalent to 1 mg. of chlorin. Check by titration against a standardized solution of sodium chlorid.

19

DETERMINATION.

To 100 cc. of the water add a few drops of phenolphthalein. If a red color appears, titrate the carbonates thus indicated to bicarbonates with N/10 or N/20 sulphuric acid. If the water is acid to methyl orange, add N/20 sodium carbonate to neutralize the acidity. Add 1 cc. of the potassium chromate and titrate with the standard silver nitrate. Correct for the amount of silver nitrate necessary to give in 100 cc. of chlorin-free water with 1 cc. of the chromate, the shade obtained at the end of the titration of the sample. Iodids and bromids are not usually found in interfering quantities in potable water. However, if they are present make the equivalent correction.

If chlorids are present in very small quantities concentrate 500 or 1000 cc. in a porcelain dish to 100 cc., rub down the sides of the dish carefully, add 1 cc. of the indicator and titrate as described above. If sufficient chlorids are present in 100 cc. of the water to consume more than 25 cc. of the standard silver nitrate, determine by precipitation in nitric acid solution and weigh the silver chlorid.

OXYGEN REQUIRED.

Method I.—Official.

20

REAGENTS.

(a) *Standard potassium permanganate solution.*—Dissolve 0.3952 gram of potassium permanganate in 1 liter of water; each cc. has 0.1 mg. of oxygen available for oxidation.

(b) *Standard oxalic acid solution.*—Dissolve 0.7875 gram of crystallized oxalic acid in 1 liter of water.

Determine the value of the oxalic acid in terms of the permanganate by boiling 10 cc. of the oxalic acid and 200 cc. of redistilled water (prepared by treating distilled water with alkaline permanganate and distilling) with 10 cc. of sulphuric acid (1 to 3) and titrating, while still boiling, with the standard permanganate to the appearance of a pink color.

21

DETERMINATION.

Add 10 cc. of sulphuric acid (1 to 3) to 200 cc. of the water in a porcelain dish and heat to boiling. Add from a burette the standard permanganate until the water is distinctly red and boil for 10 minutes, adding more of the standard permanganate from time to time to maintain the red color. Add 10 cc. of the standard oxalic acid and titrate back with the standard permanganate to a pink color. From the total number of cc. used of the permanganate subtract the number of cc. equivalent to 10 cc. of the oxalic acid. The result gives the number of cc. of the permanganate required for 200 cc. of the water. Correct for sulphids, nitrites and ferrous salts, if present, by subtracting the number of cc. of the standard permanganate absorbed by another 200 cc. portion of the sample when treated as above, digesting at room temperature for 3 minutes.

Method II. (Schulze-Trommsdorf Method.¹)—Tentative.

(To be used when the chlorine content of the sample is high.)

22

REAGENTS.

(a) *50% sodium hydroxid solution.*

Other reagents and standard solutions are described under 20.

23

DETERMINATION.

Introduce 100 cc. of the water to be examined in a 300 cc. flask, add 0.5 cc. of the sodium hydroxid and 10 cc. of the permanganate, boil for 10 minutes, allow to cool to 50°–60°C. and add 5 cc. of the dilute sulphuric acid and 10 cc. of the standard oxalic acid. As soon as the liquid has become perfectly colorless, and while constantly agitating, cautiously add from a burette, drop by drop, the standard permanganate, until the liquid acquires a faint permanent redness. The permanganate required to effect this is the quantity required for the decomposition of the organic matter in the 100 cc. of water.

If 100 cc. of the water require more than 4 cc. of the permanganate for the oxidation of organic matter, a second determination must be made using more of the permanganate and a correspondingly larger quantity of the sodium hydroxid, as undecomposed permanganate remaining after boiling must be at least twice as great as the quantity decomposed.

DISSOLVED OXYGEN.

Method I. (Winkler Method as Modified by Drown and Hazen.²)—Tentative.

(When less than 0.1 mg. of nitrite nitrogen per liter is present.)

24

REAGENTS.

(a) *Manganous sulphate solution.*—Dissolve 48 grams of manganous sulphate in 100 cc. of water.

(b) *Sodium hydroxid-potassium iodid solution.*—Dissolve 360 grams of sodium hydroxid and 100 grams of potassium iodid in 1 liter of water.

(c) *Sulphuric acid.*—(Sp. gr. 1.4). Mix equal weights of concentrated sulphuric acid and water.

(d) *Standard sodium thiosulphate solution.*—Dissolve 6.2 grams of recrystallized sodium thiosulphate in 1 liter of water. This gives a N/40 solution, each cc. of which is equivalent to 0.2 mg. of oxygen or 0.1395 cc. of oxygen at 0°C. and 760 mm. pressure. This solution should be standardized occasionally against N/40 potassium dichromate.

(e) *Starch indicator.*—Mix about 2 grams of clean starch with cold water to a thin paste; pour into about 200 cc. of boiling water. Boil for a few minutes. This solution should be freshly prepared.

25

COLLECTION OF SAMPLE.

Collect the sample in a carefully calibrated glass stoppered bottle, approximately 250 cc. capacity, by means of an apparatus designed to avoid the entrainment or absorption of any oxygen from the atmosphere. Note the temperature.

26

DETERMINATION.

Add approximately 2 cc. of the manganous sulphate and 2 cc. of the sodium hydroxid-potassium iodid, delivering both of these solutions beneath the surface of the liquid by means of a pipette. Insert the stopper and mix the contents of the bottle by shaking. Allow the precipitate to settle. Remove the stopper; add about 2 cc. of sulphuric acid and mix thoroughly. Rinse the contents of the bottle into a flask; titrate with N/40 sodium thiosulphate, using a few cc. of the starch indicator toward the end of the titration. Do not add the starch until the color has become a faint yellow; titrate until the blue color disappears. Express the results in milligrams per liter and in percentage of saturation.³ This latter determination is the ratio of the amount of gas present to the maximum amount capable of being dissolved by distilled water at the same temperature and pressure.

Method II. (Winkler Method as Modified by Rideal and Stewart.⁴)—Tentative.

(When more than 0.1 mg. of nitrite nitrogen per liter is present.)

27

REAGENTS.

(a) *N/10 potassium permanganate.*

(b) *2% potassium oxalate solution.*

Other reagents are described under 24.

28

COLLECTION OF SAMPLE.

Proceed as directed under 25.

29

DETERMINATION.

Preliminary test.—Determine the amount of the permanganate required to oxidize the nitrite to nitrate by acidifying a preliminary sample of 50 cc. with 1 cc. of the sulphuric acid and adding the permanganate until a slight pink color remains after standing 10 minutes. Calculate the amount of the permanganate required for a sample collected as described under 25.

To the sample add 1 cc. of the sulphuric acid and about 0.1 cc. of the permanganate in excess of the calculated amount required to oxidize the nitrite to nitrate. If more than 10 cc. of the permanganate are required add an additional 1 cc. of the sulphuric acid. Rotate the bottle and allow to stand for 10 minutes, after which destroy any excess of the permanganate by adding from a pipette 0.5–1 cc. of the oxalate. Insert the stopper and rotate as before. The color quickly disappears, and when decolorized add approximately 2 cc. of the manganous sulphate and 2 cc. of the sodium hydroxid-potassium iodid and proceed as in 26. Express the results in milligrams per liter and percentage saturation.³

MINERAL WATER.

30

SPECIFIC GRAVITY.—TENTATIVE.

Determine specific gravity at $\frac{20^{\circ}\text{C.}}{20^{\circ}}$ by means of a pycnometer.

31

SOLIDS IN SOLUTION.—OFFICIAL.

Determine as directed under 7.

32

IGNITED RESIDUE.—OFFICIAL.

Determine as directed under 9.

33

FREE AND ALBUMINOID AMMONIA.—OFFICIAL.

Determine as directed under 11.

34

NITROGEN IN THE FORM OF NITRITE.—OFFICIAL.

Determine as directed under 13.

35

NITROGEN IN THE FORM OF NITRATE.—OFFICIAL.

Determine as directed under 15 or 17.

36

CHLORIN.—OFFICIAL.

Determine as directed under 19.

37

HYDROGEN SULPHID.—TENTATIVE.

Place 0.5–2 cc. of N/100 iodine in a 500 cc. flask and add the water until the color of the iodine disappears. Add 5 cc. of the starch indicator and then N/100 iodine until a blue color appears. Fill the flask to the mark with water, noting the amount added. Subtract the quantity of water, iodine solution, and the starch indicator added, to determine the quantity of the water titrated. An excess of iodine is required to produce a blue color. A correction is obtained by adding 5 cc. of the starch indicator to 500 cc. of water and then adding N/100 iodine until the color matches that of the sample under examination. Correct the original titration by the amount of iodine used in the blank.

38

FREE CARBON DIOXID.—TENTATIVE.

If the water reacts acid to phenolphthalein and alkaline to methyl orange, titrate 100 cc. with N/20 sodium carbonate (free from bicarbonate) until the solution is neutral to phenolphthalein. The number of cc. used multiplied by 1.1 gives the milligrams of free carbon dioxide in 100 cc. Express results in milligrams per liter.

39

CARBONIC AND BICARBONIC ACIDS.—OFFICIAL.

To 100 cc. of the water add a few drops of phenolphthalein and, if a pink color is produced, titrate with N/20 crystallized potassium hydrogen sulphate or sulphuric acid, adding a drop every 2 or 3 seconds, until the red color disappears. Multiply the burette reading by the factor 3 which gives the milligrams of the carbonic acid ion in 100 cc. To the colorless solution from this titration, or to the original solution if no color is produced with phenolphthalein, add 1 or 2 drops of methyl orange; continue the titration without refilling the burette and note the total reading. If carbonic acid is absent, multiply the total burette reading by the factor 3.05, which gives the value of the bicarbonic acid ion in milligrams per 100 cc. If carbonic acid is present, multiply the reading with phenolphthalein by 2 and subtract from the total reading of the burette. Multiply the difference by 3.05, which gives the bicarbonic acid ion in milligrams per 100 cc. Express results in milligrams per liter.

SILICA, IRON, ALUMINIUM, CALCIUM, STRONTIUM AND MAGNESIUM.

40

SILICA.—OFFICIAL.

Make a preliminary examination, using 100–250 cc. of water to determine the approximate quantity of calcium and magnesium present, in order to ascertain the quantity of water to be evaporated for the final analysis.

Evaporate a quantity, usually 1–5 liters, of the water sufficient to yield 0.1–0.6 gram of calcium oxide or 0.1–1 gram of magnesium pyrophosphate. Acidify the water with hydrochloric acid and evaporate on the water bath to dryness in a platinum dish; continue the drying for about an hour. Thoroughly moisten the residue with 5–15 cc. of hydrochloric acid (1 to 1). Allow to stand 10–15 minutes and add sufficient water to bring the soluble salts into solution. Heat on the steam bath until solution of the salts is effected. Filter to remove most of the silica and wash thoroughly with hot water. Evaporate the filtrate to dryness; treat with 5–10 cc. of the hydrochloric acid and sufficient water as above. Heat, filter, and wash thoroughly with hot water. Designate the filtrate as *A*. Transfer the 2 residues to a platinum crucible, ignite, heat over a blast lamp and weigh. Moisten the contents of the crucible with a few drops of water. Add a few drops of concentrated sulphuric acid and a few cc. of hydrofluoric acid and evaporate on the water bath under a good hood. Repeat the treatment if all the silica is not volatilized. Dry carefully on a hot plate, ignite, heat over a blast lamp, and weigh. The difference between the two weights is the weight of the silica. The residue in the crucible consists of aluminium and iron oxides. The weight of this residue is added to that of the total aluminium and iron oxides obtained in 41. (If the above residue weighs more than 0.5 mg., barium sulphate may be found here when barium is present in the water. If so, make the necessary correction and add to the weight of the total iron and aluminium oxides in 41.)

41

IRON AND ALUMINIUM.—OFFICIAL.

Concentrate *A*, under 40, to about 200 cc.; while still hot, add ammonium hydroxide slowly with constant stirring until alkaline to methyl orange. Boil, filter, and wash 2 or 3 times with hot water. Dissolve the precipitate in hot hydrochloric acid.

Dilute to approximately 25 cc., boil, and again precipitate with ammonium hydroxid; filter, wash thoroughly with hot water, dry, ignite, and weigh as iron and aluminium oxids. (In the presence of phosphoric acid, the weight of this residue must be corrected for the phosphorus pentoxid equivalent to the phosphoric acid found in 51, making due allowance for the difference in the volumes of the water used for these determinations.) Designate the filtrate as *B*.

IRON.

42

Colorimetric Method.

(If the amount of iron is less than 1 mg.)

Fuse in a platinum crucible the ignited precipitate of iron and aluminium oxids with fused potassium hydrogen sulphate, dissolve in water, and precipitate the iron and aluminium with ammonium hydroxid. Dissolve the precipitate on the filter paper in hydrochloric and nitric acids, dilute the solution, add ammonium sulphocyanate solution (1 to 20) and compare the color developed with that of calibrated color disks, or standards containing known amounts of iron.

43

Volumetric Method.

Fuse in a platinum crucible the residue of iron and aluminium oxids with fused potassium hydrogen sulphate. This fusion takes but a few minutes and must not be continued beyond the time actually needed. When completed, the crucible is set aside and allowed to cool. Add dilute sulphuric acid and heat the crucible until the fused mass is dissolved. Evaporate on the water bath as far as possible; then heat gradually until copious fumes of sulphuric acid are given off. Dissolve in water and allow to stand on the water bath. Cool, transfer to an Erlenmeyer flask, and make up to such a volume that the solution does not contain more than 2.5% of free sulphuric acid. Pass hydrogen sulphid through the solution to reduce the iron and precipitate any platinum contaminating the residue from the fusion. (Zinc may be used instead of hydrogen sulphid for reducing the iron.) Filter, wash and again pass hydrogen sulphid through the solution to be certain that all the iron is reduced. Expel the hydrogen sulphid by boiling, at the same time passing a current of carbon dioxid through the solution; test the escaping gas with lead acetate paper to ascertain the complete removal of hydrogen sulphid. When hydrogen sulphid has been removed discontinue boiling and let the flask cool somewhat without discontinuing the current of carbon dioxid. Titrate the reduced iron with a standard permanganate solution (1 cc. equivalent to 1 mg. of Fe) and calculate as iron.

44

ALUMINIUM.—OFFICIAL.

In the absence of phosphates, subtract from the weight of iron and aluminium oxids, under 41, the iron, under 42 or 43, calculated to oxid, to obtain the weight of aluminium oxid. Calculate to aluminium.

45

CALCIUM.—OFFICIAL.

Concentrate *B*, under 41, to 150–200 cc. and to this solution, containing not more than 0.6 gram of calcium, calculated as calcium oxid, or 1 gram of magnesium, calculated as magnesium pyrophosphate, add 1–2 grams of oxalic acid and sufficient hydrochloric acid to clear the solution. Heat to boiling and neutralize with ammonium hydroxid, stirring constantly. Add ammonium hydroxid in slight excess and allow to stand 3 hours in a warm place. Filter off the supernatant liquid and wash

the precipitate once or twice by decantation with 1% ammonium oxalate solution. Dissolve the precipitate in hydrochloric acid, dilute to 100–200 cc., add a little oxalic acid, and precipitate as above. After standing 3 hours, filter, wash with the ammonium oxalate solution as above, dry, ignite, heat over a blast lamp, and weigh as calcium and strontium oxids. Subtract from this weight, the weight of strontium oxid equivalent to the strontium under 46. The difference is the weight of calcium oxid. Calculate to calcium. Designate the filtrate and washings as *C*.

As a check on the calcium oxid, evaporate to dryness the filtrate from the strontium nitrate under 46, beginning with "Filter, wash with ether-alcohol mixture, etc.," dissolve the calcium nitrate in water, precipitate as oxalate, filter, wash, ignite, and weigh as calcium oxid.

46**STRONTIUM.—TENTATIVE.**

Dissolve the oxids under 45 in dilute nitric acid and test with the spectroscope for strontium. If strontium is present, transfer the nitric acid solution to a small Erlenmeyer flask. Evaporate nearly to dryness over a low flame and heat in an air bath at 150°–160°C. for 1 or 2 hours after the water is evaporated. Break up the dried material with a stirring rod, add 10–15 cc. of a mixture of equal parts of absolute alcohol and ether to dissolve the calcium nitrate. Cork the flask and allow to stand with frequent shaking for 2 hours or longer. Decant the solution through a 5.5 cm. filter, preserving the filtrate. Wash the residue several times by decantation with small portions of ether-alcohol solution. Dry the residue and the filter paper and wash the filter paper repeatedly with small portions of hot water, collecting the filtrate in the flask containing the main portion of the strontium nitrate residue. Add 1 or 2 drops of dilute nitric acid, evaporate, dry, pulverize, and treat with 10–15 cc. of ether-alcohol mixture as above. Cork the flask and let stand about 12 hours with occasional shaking. Filter, wash with ether-alcohol mixture until a few drops of the filtrate evaporated on a watch glass leave practically no residue. Dry the paper and precipitate. Dissolve the strontium nitrate in a few cc. of hot water. Add a few drops of sulphuric acid, then a volume of alcohol equal to the volume of the solution and allow to stand 12 hours. Filter, ignite, weigh as strontium sulphate and calculate to strontium. Test spectroscopically for absence of calcium.

47**MAGNESIUM.—OFFICIAL.**

Concentrate *C*, under 45, to about 200 cc.; add 2–3 grams of diammonium hydrogen phosphate and sufficient hydrochloric acid to clear the solution when the ammonium phosphate is all dissolved; disodium hydrogen phosphate or sodium ammonium hydrogen phosphate may be used instead of the diammonium hydrogen phosphate. When cold, make slightly alkaline with ammonium hydroxid, stirring constantly. Add 1–2 cc. excess of ammonium hydroxid and allow to stand about 12 hours. Filter off the supernatant liquid and wash 3 or 4 times by decantation with a solution of 2.5% ammonium hydroxid. Dissolve the precipitate in hydrochloric acid, dilute to about 150 cc., add a little diammonium hydrogen phosphate and precipitate with ammonium hydroxid as before. Allow to stand 6–12 hours, filter, wash free from chlorin, ignite, heat over a blast lamp, and weigh as magnesium pyrophosphate. (Cf. II, 16). Calculate to magnesium.

SULPHURIC ACID, SODIUM, POTASSIUM AND LITHIUM.**48****SULPHURIC ACID.—OFFICIAL.**

Make a preliminary examination, using 100–250 cc. of the water to determine the approximate quantity of sulphates. The alkali salts present can be approximated

by calculating the amount of sodium necessary to combine with the excess of acids (hydrochloric, sulphuric, and bicarbonic) over the calcium and magnesium.

Take a quantity, usually 1-5 liters, of the water sufficient to yield not more than 1 gram of barium sulphate and not more than 0.5 gram of mixed chlorids. Acidify with hydrochloric acid, evaporate to dryness in a platinum dish and remove silica by 2 evaporations as under 40, using not more than 2 cc. of hydrochloric acid (1 to 1) for the final solution. Combine the filtrate and washings from the silica determinations, and concentrate to about 150-200 cc. Heat to boiling and precipitate with slight excess of 10% barium chlorid solution, added very slowly and with constant stirring. Cover and allow to stand on the steam bath about 12 hours. Filter, wash thoroughly the precipitate of barium sulphate with hot water, dry, ignite over a Bunsen burner, and weigh.

If the content of sulphate in the sample is unusually large proceed as far as the concentration of the silica filtrates as directed above. Add 50 cc. of concentrated hydrochloric acid, heat to boiling and precipitate with barium chlorid solution as before. Evaporate to dryness, wash the precipitate repeatedly by decantation and filter. Complete the washing of the precipitate; ignite and weigh. Calculate to the sulphuric acid ion. Designate the filtrate as *E*.

49

SODIUM, POTASSIUM AND LITHIUM.—OFFICIAL.

Evaporate to dryness *E*, under 48, in a platinum dish and ignite the residue to faint redness to remove all traces of ammonium salts. Dissolve the residue in the dish in about 200 cc. of water and precipitate with milk of lime or a solution of barium hydroxid. Boil, allow to stand 30 minutes, and filter off the insoluble magnesium hydroxid. Thoroughly wash the precipitate with hot water and combine the filtrate and washings. If the precipitate of magnesium is large, it is advisable to dissolve in a small amount of hydrochloric acid, evaporate to dryness, take up with water, and precipitate as before. Concentrate the 2 filtrates and washings to 200-250 cc. Add ammonium hydroxid and sufficient ammonium carbonate solution to precipitate the calcium and barium. Allow to stand on a steam bath 1-2 hours. Filter off the supernatant liquid, dissolve the precipitate in hydrochloric acid, reprecipitate as above, and wash thoroughly with hot water. Evaporate the combined filtrates and washings to dryness and drive off the ammonium salts by gentle heat. Treat the residue with water; filter through a small filter, using as little wash water as possible; evaporate to a small volume and again precipitate with 1 or 2 drops of ammonium hydroxid and 2 or 3 drops of ammonium carbonate and oxalate. If any precipitate appears (which is usually not the case) filter and repeat the process. Evaporate the filtrate to dryness and drive off all ammonium salts by heating in platinum to faint redness. Treat the residue with a little water; filter into a small platinum dish; add a few drops of hydrochloric acid and evaporate to dryness. Dry in an oven, heat to faint redness, cool in a desiccator, and weigh the combined chlorids of potassium, sodium, and lithium. Repeat the heating to constant weight, (*x*). Dissolve the mixed chlorids in hot water; filter, and wash. Return the filter paper and residue to the dish, dry, ignite, and weigh, (*y*). The difference between (*x*) and (*y*) is the weight of the mixed chlorids.

The determination of lithium is then made according to the method of Gooch.⁵

Transfer the combined chlorids to a 50-100 cc. Erlenmeyer flask and evaporate the solution nearly, but not quite, to dryness. Add about 30 cc. of redistilled amyl alcohol. Connect the flask, the stopper of which carries a thermometer, using a condenser if desired, to avoid the escape of the irritating vapor of the amyl alcohol, and boil until the temperature rises approximately to the boiling point of amyl alco-

hol (130°C.) to remove the water. Cool slightly and add a drop of hydrochloric acid to convert small amounts of lithium hydroxid to lithium chlorid. Connect with the condenser and repeat the boiling until the temperature reaches the boiling point of amyl alcohol to again drive off the water. The content of the flask at this time is usually 15–20 cc. Filter through a small paper or a Gooch crucible into a graduated cylinder and note the exact quantity of the filtrate, which determines the subsequent correction. Wash the precipitate with small quantities of amyl alcohol. Evaporate the filtrates and washings in a small platinum dish to dryness on the steam bath, dissolve the residue in water, and add a few drops of sulphuric acid. Evaporate on a steam bath and expel the excess of sulphuric acid by heating gently over a Bunsen burner until the carbonaceous matter is completely burned off, repeating the addition of a few drops of sulphuric acid if necessary. Cool and weigh the dish and contents, (x). Dissolve in a small quantity of hot water, filter through a small filter, wash and return filter to dish; ignite and weigh, (y). The difference between (x) and (y) is the weight of impure lithium sulphate.

The purity of the lithium sulphate should be tested by adding small amounts of ammonium phosphate solution and ammonium hydroxid, which will precipitate any magnesium previously present in the lithium sulphate. Any precipitate appearing after standing overnight should be collected on a small filter, ignited, weighed as magnesium pyrophosphate, calculated to sulphate and subtracted from the weight of the impure lithium sulphate.

From this weight subtract 0.00113 gram of sodium and potassium sulphates for every 10 cc. of amyl alcohol filtrates, exclusive of the amyl alcohol used in washing the residue, on account of the solubility of sodium and potassium chlorids in amyl alcohol. Calculate to lithium from the corrected weight of lithium sulphate.

Dissolve the mixed chlorids from the flask and filter with hot water, evaporate to dryness, ignite gently to remove amyl alcohol, filter, and thoroughly wash; concentrate the filtrates and washings to 25–50 cc. Transfer to a porcelain dish, add sufficient platinic chlorid solution [I, 40 (b)] to convert sodium and potassium to their respective double chlorids and evaporate to dryness. Treat the residue with 80% alcohol, filter, and wash until the excess of platinic chlorid and sodium platinic chlorid has been removed. Dry the filter and precipitate, dissolve the residue in hot water, and transfer to a weighed platinum dish. Evaporate on the steam bath, dry for 30 minutes in the oven at 100°C. and weigh as potassium platinic chlorid; calculate to potassium chlorid. To the weight of potassium chlorid add 0.00051 gram for every 10 cc. of amyl alcohol used in the extraction of the lithium chlorid, which corrects for the solubility of the potassium chlorid in amyl alcohol. Calculate to potassium.

The weight of sodium chlorid is found by subtracting the combined corrected weights of lithium chlorid and potassium chlorid from the total weight of the 3 chlorids. Calculate the sodium chlorid to sodium.

PHOSPHORIC ACID.—OFFICIAL.

50

REAGENTS.

The reagents used are described under I, 7.

51

DETERMINATION.

Treat 500 cc. of the water, or a larger amount if necessary, with about 10 cc. of concentrated nitric acid and evaporate in a porcelain dish nearly to dryness to drive off hydrochloric acid. Treat the residue with water and filter, if necessary. Add ammonium hydroxid to alkalinity and then just enough nitric acid to restore acidity.

Add some solid ammonium nitrate and heat in the water bath at a temperature of 45°–50°C. Add the molybdate solution and keep at the above temperature for 30 minutes. The yellow precipitate formed at this point appears generally only in traces; if more than traces are present, filter and wash with cold water until entirely free from nitric and molybdic acids. Transfer the precipitate and filter to a beaker, add a little water, and beat the paper and contents to a pulp. Dissolve the yellow precipitate in a small amount of the standard potassium hydroxid; add phenolphthalein and titrate with the standard acid. From the data so obtained calculate the phosphoric acid ions in the water to milligrams per liter.

52 MANGANESE, IODIN, BROMIN, ARSENIC AND BORIC ACID.

Evaporate large quantities of water to dryness, after the addition of small amounts of solid sodium carbonate. Boil the residue thus obtained with water, transfer to a filter, and wash thoroughly with hot water. Make the alkaline filtrate up to a definite volume.

MANGANESE.—OFFICIAL.

53

REAGENTS.

- (a) *Dilute nitric acid (1 to 1).*
- (b) *0.2% silver nitrate solution.*
- (c) *Ammonium persulphate.*
- (d) *Standard manganous sulphate solution.*—Dissolve 0.2877 gram of pure potassium permanganate in a small amount of water, add an excess of sulphuric acid, reduce carefully with oxalic acid and make up to 1 liter. One cc. of this solution is equivalent to 0.1 mg. of manganese.

54

DETERMINATION.

Dissolve the insoluble residue under 52 in an excess of the dilute nitric acid, evaporate to dryness, treat with water, add about 1 cc. of strong nitric acid and a little of the silver nitrate. If a precipitate of silver chlorid appears, add more of the silver nitrate until all the chlorin is precipitated. Add an excess of about 10 cc. of the silver nitrate for each mg. of manganese present in the sample. Filter, add 1 gram of ammonium persulphate to the filtrate, and place the beaker or flask containing the solution on the steam bath until a pink color develops (usually about 20 minutes). Compare the color developed with standards similarly prepared by treating solutions containing known amounts of the standard manganous sulphate with nitric acid, silver nitrate, and ammonium persulphate.

IODIN AND BROMIN.—TENTATIVE.

55

REAGENTS.

- (a) *10% sodium hydroxid solution.*
- (b) *Sulphuric acid (1 to 5).*
- (c) *2% potassium or sodium nitrite solution.*
- (d) *Carbon disulphid.*—Freshly purified by distillation.
- (e) *Chlorin water.*—Saturated and freshly prepared.

56

DETERMINATION.

Evaporate to dryness an aliquot of the alkaline filtrate under 52, add 2–3 cc. of water to dissolve the residue and enough 95% alcohol to make the percentage of alcohol about 90. This precipitates the chlorids. Heat to boiling, filter and repeat the preceding solution and precipitation once or twice. Add 2 or 3 drops of the

sodium hydroxid to the combined alcoholic filtrates and evaporate to dryness. Dissolve this last residue in 2-3 cc. of water and repeat as above described the precipitation with alcohol, heating, and filtering. Add a drop of the sodium hydroxid to this alcoholic filtrate and evaporate to dryness. Dissolve this residue in a little water, acidify with the sulphuric acid, using 3 or 4 drops in excess, and transfer to a small flask. Add 4 drops of the potassium nitrite and about 5 cc. of the carbon disulphid. Shake until all the iodine is extracted, filter off the acid solution from the carbon disulphid, retaining the latter in the flask. Wash the flask, filter and contents with cold water and transfer the carbon disulphid (containing the iodine in solution) to a Nessler tube, using approximately 5 cc. of the carbon disulphid. In washing the filter make the contents of the tube up to definite volume, usually 12-15 cc., and compare the color with that of other tubes containing known amounts of iodine dissolved in carbon disulphid. Prepare these standard tubes by treating measured quantities of a solution of known potassium iodid content as described above. Transfer the sample and standards, from which the iodine has been removed, severally to small flasks. To the standards add definite measured quantities of a bromid solution of known strength, and to each of the flasks containing sample and standards add 5 cc. of purified carbon disulphid. Add the saturated chlorin water, 1 cc. at a time, shaking after each addition until all the bromine is set free. (Avoid a large excess of chlorin, since a bromo-chlorid may be formed which spoils the color reaction.) Filter off the water solution from the carbon disulphid through a moistened filter, wash the contents of the filter 2 or 3 times with water, and then transfer to a Nessler tube by means of about 1 cc. of carbon disulphid. Repeat this extraction of the filtrate twice, using 3 cc. of carbon disulphid each time. The combined carbon disulphid extracts usually amount to 11.5-12 cc. Add enough carbon disulphid to the tubes to bring them to a definite volume, usually 12-15 cc., and compare the sample with the standards. In some cases when using this method near its upper limit the amounts of carbon disulphid recommended do not extract all the bromine. In these cases, make 1 or 2 extra extractions with carbon disulphid, transfer the extracts to another tube, and compare the color with some of the lower standards and add the readings thus obtained to the others.

Results closely approximating the true values for iodine and bromine can be obtained on most samples by omitting the extractions with alcohol given above and by comparing the color of the carbon disulphid solutions directly in the extraction flasks, thus shortening the method.

ARSENIC.—OFFICIAL.

57

REAGENTS.

(a) *Zinc, arsenic-free.*

(b) *Sulphuric acid (1 to 5), arsenic-free.*

(c) *Standard arsenious oxid solution.*—Dissolve 0.0132 gram of pure arsenious oxid in 100 cc. of water containing about 50 mg. of sodium carbonate. One cc. of this solution is equivalent to 0.1 mg. of As.

58

DETERMINATION.

Evaporate to dryness an aliquot of the alkaline filtrate under 52. Acidify with the sulphuric acid and subject to the action of the zinc and the sulphuric acid in a Marsh-Berzelius apparatus. Compare the mirror obtained with a mirror prepared from an arsenious oxid solution of known strength. Calculate to the arsenic acid ion.

BORIC ACID.—OFFICIAL.

(Glassware containing boron must not be used in this determination.)

59

DETERMINATION.

Qualitative test.—Evaporate to dryness a part of the alkaline filtrate under 52, treat with 1-2 cc. of water, and slightly acidify with dilute hydrochloric acid (1 to 1). Add about 25 cc. of 95% alcohol, boil, filter, and repeat the extraction of the residue. Make the filtrate slightly alkaline with sodium hydroxid solution and evaporate to dryness. Add a little water, slightly acidify with dilute hydrochloric acid, and place a strip of turmeric paper in the liquid. Evaporate to dryness on the steam bath and continue the heating until the turmeric paper is dry. If boric acid is present the turmeric paper takes on a cherry-red color. As a confirmatory test, apply a drop of dilute ammonium hydroxid to the reddened paper, and a dark olive color will be due to boric acid.

Quantitative test.—It is not usually necessary to determine boric acid quantitatively. However, if it is necessary, the Gooch method⁶ is used.

60

METHOD OF REPORTING RESULTS.—TENTATIVE.

Report the bases and acids as positive and negative ions in milligrams per liter, except in the case of silica, which report as such without considering how much is present as the silicic acid ion and how much as free silica. Report iron and aluminium together when present in unimportant quantities, and in calculations consider it as iron. When iron and aluminium are present in larger quantities make the separation and report each separately.

In calculating the hypothetical combinations of acid and basic ions join sodium to nitrous, nitric, metaboric and arsenic acids; potassium to iodine and bromine; calcium to phosphoric acid. Assign the residual basic ions in the following order; ammonium, lithium, potassium, sodium, magnesium, calcium, strontium, manganese, iron and aluminium—to the residual acid ions in the following order: Chlorine, sulphuric acid ion, carbonic acid ion, and bicarbonic acid ion. In case the bicarbonic acid ion is not present in a sufficient quantity to join with all the calcium, the residual calcium is joined to silica to form calcium silicate, and manganese, iron, and aluminium are calculated to the oxides Mn_2O_4 , Fe_2O_3 , and Al_2O_3 , respectively.

INDUSTRIAL WATER.

61

SOLIDS IN SOLUTION.—OFFICIAL.

Determine as directed under 7.

62

CHLORIN.—OFFICIAL.

Determine as directed under 19.

63

COMBINED CARBONIC AND BICARBONIC ACIDS.—OFFICIAL.

Determine as directed under 39.

64

NITRATES.—OFFICIAL.

Determine as directed under 15 or 17.

65

SILICA.—OFFICIAL.

Determine as directed under 40. Generally one evaporation with hydrochloric acid for removal of silica is sufficient.

66

IRON AND ALUMINIUM.—OFFICIAL.

Determine as directed under 41.

67

CALCIUM.—OFFICIAL.

If no phosphoric acid is present, concentrate the filtrate from the determination of iron and precipitate with ammonium hydroxid and oxalate as directed under 45. Usually one precipitation is sufficient.

68

MAGNESIUM.—OFFICIAL.

Determine as directed under 47.

69

SULPHURIC ACID AND ALKALIES.—OFFICIAL.

Follow the methods described under 43 and 49. Generally, however, for technical purposes it is sufficiently accurate to determine the acids and the bases, except sodium and potassium, and then to calculate the excess of acid over basic ions to the sodium salt, and state the alkali thus found as sodium and potassium by difference.

70

TEMPORARY HARDNESS.⁷—TENTATIVE.

The difference between the alkalinity after boiling, 74, and the alkalinity before boiling, 72, is the temporary hardness in parts per million of calcium carbonate.

ALKALINITY—Before Boiling.

71

REAGENTS.

(a) *N/50 sulphuric acid.*

(b) *Erythrosin indicator.*—Dissolve 0.1 gram of the sodium salt in 1 liter of water.

(c) *Chloroform.*—Neutral to erythrosin.

72

DETERMINATION.

Measure 100 cc. of the water into a 250 cc. white, glass-stoppered bottle, add 2.5 cc. of the erythrosin and 5 cc. of the chloroform, add *N/50* sulphuric acid in small quantities, shaking the bottle vigorously after each addition of the acid. The rose color gradually disappears and is finally discharged by 1 or 2 drops of the acid. A white paper held back of the bottle facilitates the detection of the end point. Multiply the number of cc. of *N/50* sulphuric acid used by 10 to obtain the number of parts per million of alkalinity in terms of calcium carbonate.

ALKALINITY—After Boiling.

73

REAGENTS.

Described under 71.

74

DETERMINATION.

Boil 100 cc. of the water in a porcelain dish gently for 30 minutes. Cool, transfer to a 100 cc. volumetric flask and fill to the mark with recently boiled and cooled water. Filter through a dry paper and determine the alkalinity of the filtrate as directed under 72, making the proper calculation for the aliquot employed and calculating in terms of calcium carbonate the parts per million of alkalinity after boiling.

TOTAL HARDNESS.⁸—TENTATIVE.

75

REAGENTS.

(a) *Soda reagent*.—Prepare a N/10 alkali solution, using equal parts of sodium hydroxid and sodium carbonate. Standardize the solution by titration against N/20 sulphuric acid, using erythrosin as indicator.

76

DETERMINATION.

Add sufficient N/20 sulphuric acid to 200 cc. of the sample to neutralize the alkalinity, the amount required for this purpose being calculated from the results obtained as directed under 72. Concentrate to 100 cc., add 25 cc. of the soda reagent, and again boil down to 100 cc., using a porcelain, silver, or platinum dish. Cool, rinse into a 200 cc. volumetric flask and dilute to 200 cc. with freshly boiled and cooled water. Filter through a dry paper, reject the first 50 cc. of the filtrate, and titrate 100 cc. of the filtrate, using N/20 sulphuric acid and chloroform with erythrosin as indicator, as directed under 72. Calculate the total hardness by the following formula: $H = 12.5 (S - 2N)$ in which

H = total hardness expressed as parts per million of calcium carbonate.

S = number of cc. of N/20 sulphuric acid equivalent to the 25 cc. of the soda reagent used.

N = number of cc. of N/20 sulphuric acid used in titrating back the excess of the soda reagent.

77

PERMANENT OR NON-CARBONATE HARDNESS.—TENTATIVE.

The difference between the alkalinity before boiling 72 and the total hardness 76 is the permanent or non-carbonate hardness expressed as parts per million of calcium carbonate.

IRRIGATING WATER.

78

GENERAL METHODS.—OFFICIAL.

Determine the solids in solution, chlorin, carbonic and bicarbonic acids, sulphuric acid, calcium and magnesium as directed under 7, 19, 39, 43, 45, and 47 respectively. To make the hypothetical combination, calculate calcium and magnesium to the acid ions in the following order: bicarbonic, sulphuric and chlorin. Then calculate the remaining acid ions, including carbonic, to the corresponding salts of sodium.

BLACK ALKALI.—OFFICIAL.

79

REAGENTS.

(a) *N/50 sodium carbonate*.—One cc. of this solution is equivalent to 0.00106 gram of sodium carbonate.

(b) *N/50 sulphuric acid*.—One cc. of this solution is equivalent to 0.0010 gram of calcium carbonate or 0.00136 gram of calcium sulphate.

(c) *Erythrosin indicator*.—Dissolve 0.25 gram of the sodium salt in 1 liter of water.

(d) *Chloroform*.—Neutral to erythrosin.

80

DETERMINATION.

Transfer 200 cc. of the water to a platinum or silver dish, add 50–100 cc. of N/50 sodium carbonate according to the amount of soluble salts of calcium and magnesium present, and evaporate to dryness. Rub up the residue with carbon dioxide-free water.

For this purpose distilled water should be vigorously boiled until approximately one third of the original volume is evaporated, then cooled and stoppered. An ordinary laboratory wash bottle should not be used to transfer the residue, as the carbon dioxide from the breath of the operator is sufficient to vitiate the results.

Transfer to a 100 cc. graduated flask, make up to the mark, shake thoroughly, and allow to stand until clear (12-15 hours). Remove 50 cc. of the clear, supernatant liquid, equivalent to one half of the original quantity of water and sodium carbonate added, and transfer to a stoppered titrating bottle, of 250 cc. capacity, of clear glass without any tinge of pink. Add 5 cc. of the chloroform and 1 cc. of the erythrosin and titrate with the standard acid until the color disappears. Shake the solution vigorously after each addition of the acid; the chloroform produces a milky appearance which makes the reading of the end point sharp and certain.

(1) If less sulphuric acid is required than is equivalent to one half of the sodium carbonate added, due to some of the sodium carbonate reacting with soluble salts of calcium and magnesium, the solution originally contained no black alkali in excess but rather an excess of the so-called permanent or non-carbonate hardness. It is customary to express the hardness in terms of calcium carbonate or calcium sulphate. With irrigating waters the latter form is to be preferred. Therefore, the difference between the number of cc. of the sulphuric acid required and one half of the number of cc. of the sodium carbonate added multiplied by the factor 0.00136 gives the equivalent of calcium sulphate in 100 cc. of the water.

(2) If more sulphuric acid is required than that equivalent to one half of the sodium carbonate added, black alkali was originally present in the solution and the difference in cc. multiplied by the factor 0.00106 gives the black alkali in terms of sodium carbonate in 100 cc. of water.

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V. TANNING MATERIALS.—TENTATIVE.

EXTRACTS.

1

PREPARATION OF SOLUTION.

(a) *Solid extracts*.—Grind solid extracts in a large porcelain mortar, so that the material will pass through a 10 mesh sieve, mix thoroughly and weigh out a quantity containing 3.75–4.25 grams of tannin. This should be done as rapidly as possible to avoid change in moisture content. Pour into 100 cc. of water at 85°C., place on a steam bath and stir until a homogeneous solution is obtained. Transfer to a 1 liter flask with 800 cc. of water at 85°C. Cool rapidly to 20°C. and make up to 1 liter.

(b) *Fluid extracts*.—Allow fluid extracts to come to room temperature and mix thoroughly. Weigh out rapidly a quantity containing 3.75–4.25 grams of tannin. Dissolve by washing into a 1 liter flask with 900 cc. of water at 85°C. Cool rapidly to 20°C. and make up to 1 liter at 20°C.

After the preparation of the solutions, proceed at once with the analysis.

2

TOTAL SOLIDS.

Thoroughly mix the prepared solution, pipette at once 100 cc. into a tared flat-bottomed glass dish, 2½–3 inches in diameter, and (1) evaporate and dry for 16 hours in a combined evaporator and dryer¹ at 98°–100°C.; or, (2) after evaporating on the steam bath, dry for 12 hours on the bottom of a water oven at 98°–100°C. Remove immediately to desiccators containing sulphuric acid (place no more than 2 dishes in 1 desiccator) and weigh rapidly when cooled. Calculate the percentage of total solids.

SOLUBLE SOLIDS.

3

PREPARATION OF FILTER.

The kaolin used should be neutral to phenolphthalein and should not yield more than 1 mg. of soluble solids per 100 cc. of filtrate of a 1% suspension after an hour's digestion at 20°C. Dry on a water bath and preserve in a tightly stoppered bottle.

Add about 75 cc. of the solution, as prepared under 1, to 1 gram of the kaolin in a beaker. Stir and pour immediately into a single, 15 cm. No. 590, S. & S. folded filter. Return the filtrate to the paper when approximately 25 cc. have run through, repeat the operation for an hour, thus transferring all the kaolin to the paper. At the end of an hour, discard the solution on the filter by siphoning it off, disturbing the kaolin as little as possible.

4

DETERMINATION.

Bring about 150 cc. of the original solution, as prepared under 1, to exactly 20°C. Fill the filter, prepared as under 3, with this solution and discard the filtrate until it runs through clear. Keep the filter full, the temperature of the filtering solution at 20°–25°C., and the funnel and receiving vessel covered. Pipette at once 100 cc. of the clear filtrate into a tared dish, evaporate and dry as directed under 2. Calculate the percentage of soluble solids.

5

INSOLUBLE SOLIDS.

The difference between the percentage of the total solids and the percentage of soluble solids is the percentage of the substance insoluble in water at 20°–25°C.

NONTANNINS.

6

REAGENTS.

Hide powder.—This should be of woolly texture, well delimited, and 10 grams of the water-free powder should require 12–13 cc. of N/10 sodium hydroxid to neutralize it.

Calculate the amount of air-dry hide powder which will be required for the number of determinations to be made, on a basis of 13 grams of air-dry hide powder for each determination. Increase this calculated amount by 35 grams of dry hide powder to provide a sufficient amount for all the determinations.

Thoroughly digest the total amount of hide powder with 10 times its weight of water. Then for each gram of the hide powder, so digested, add 1 cc. of 3% chrome alum solution; and *either* agitate frequently for several hours and let stand overnight *or* agitate in some form of mechanical shaker for an hour. Transfer to a strong linen filter and squeeze thoroughly. Remove from the filter and digest for 15 minutes with a quantity of water equivalent to 15 times the weight of the dry hide powder employed. Filter and squeeze to approximately 73% of water, using a press if necessary. Very strong pressure is required to reduce the water content below 70%. Repeat the digestion and filtration 3 times. Determine moisture in 20 grams of the squeezed hide powder as directed under 2.

7

DETERMINATION.

Place 46 grams of the wet hide powder in a suitable container of about 300 cc. capacity, add 200 cc. of the tanning solution, as prepared under 1, and shake for 10 minutes in a mechanical shaker. Squeeze immediately through linen, add 2 grams of kaolin, as used under 3, to the filtrate which contains the nontannins, stir, and filter through a single, folded 18.5 cm. filter paper (No. 1F. Swedish, preferred), refiltering until the filtrate is clear. The filtrate should give no precipitate with a gelatin-salt solution (1% gelatin and 10% salt). Pipette 100 cc. of the filtrate into a tared dish and evaporate as directed under 2. Correct the weight of the nontannin residue for the dilution caused by the water retained in the wet hide powder. Calculate the percentage of nontannins.

8

TANNIN.

The difference between the percentage of the soluble solids and the percentage of nontannins is the percentage of tannin.

DETECTION OF SULPHITE-CELLULOSE.

9

REAGENTS.

Sulphite-cellulose solution.—Dissolve 0.5 gram of the total solids, derived from sulphite-cellulose, in 1 liter of water and add sufficient tanning material, free from sulphite-cellulose, to give a concentration of 3.75–4.25 grams of tannin per liter.

10

DETERMINATION.

Place 5 cc. of the tanning solution, prepared as under 1, in a test tube; add 0.5 cc. of anilin and shake well; then add 2 cc. of concentrated hydrochloric acid and mix again. Compare the precipitate formed with that produced when the above sul-

phite-cellulose solution is similarly treated. Sulphite-cellulose is held to be present, in the predetermined absence of the synthetic tanning material, Neradol-D, if the precipitates are approximately equivalent in amount.

LIQUORS.

11

PREPARATION OF SOLUTION.

Dilute the liquor with water at room temperature to contain approximately 0.7 gram of solids in 100 cc. of solution. If the liquor does not give a proper solution with water at room temperature, it may be diluted with water at 80°C., and then cooled rapidly to 20°C.

12

TOTAL SOLIDS.

Proceed as directed under 2.

13

SOLUBLE SOLIDS.

Proceed as directed under 4.

14

NONTANNINS.

Proceed as directed under 7, using the amount of wet chromed hide powder which will give the ratio between the tannin and hide powder shown in the following table:

TANNIN RANGE PER 100 CC.	DRY HIDE POWDER PER 200 CC.
<i>gram</i>	<i>grams</i>
0.35—0.45	9.0—11.0
0.25—0.35	6.5—9.0
0.15—0.25	4.0—6.5
0.00—0.15	0.0—4.0

TOTAL ACIDITY.

15

REAGENTS.

(a) *Hematin solution*.—Digest 0.5 gram of hematin in 100 cc. of cold neutral 95% alcohol.

(b) *Gelatin solution*.—Dissolve 10 grams of gelatin in hot water, cool, add 25 cc. of 95% alcohol and dilute. If the gelatin solution is acid or alkaline, neutralize with N/10 sodium hydroxid or N/10 acetic acid, respectively, using hematin solution as indicator and make up to 1 liter.

(c) *Kaolin*.—Digest with dilute hydrochloric acid; wash and dry as under 3.

(d) *N/10 sodium hydroxid*.

16

DETERMINATION.

Add 25 cc. of the gelatin solution to 25 cc. of the tanning liquor in a stoppered cylinder, dilute with water to 250 cc., add 15 grams of the kaolin and shake vigorously. Allow to settle for at least 15 minutes, remove 30 cc. of the supernatant liquid, dilute with 50 cc. of water and titrate with N/10 sodium hydroxid, using the hematin solution as indicator. Each cc. of N/10 sodium hydroxid is equivalent to 0.2% acid, calculated as acetic, in the liquor.

RAW AND SPENT MATERIALS.

(Under raw materials are included woods, barks, leaves, etc.)

17**MOISTURE IN SAMPLE AS RECEIVED.**

Cut or break up large pieces and mix the sample rapidly to avoid change in moisture content. Dry as directed under **2**, a suitable weighed quantity, dependent upon the physical condition and moisture content of the sample.

18**PREPARATION OF SAMPLE.**

Dry the remainder of the sample at a temperature not above 60°C. and grind to pass through a 20 mesh sieve.

19**MOISTURE IN PREPARED SAMPLE.**

Take 10 grams of the sample prepared in **18**, dry as directed under **2**, and calculate all results to an "as received", "air dry", or "moisture free" basis as desired.

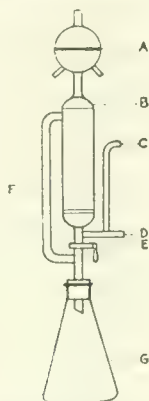
20**EXTRACTION.**

FIG. 4. METAL EXTRACTOR USED FOR EXTRACTING TANNING MATERIALS.

(For spent materials approximate the following quantities as closely as possible.)

Place a quantity of the dried sample, containing 3.75–4.25 grams of tannin, in a beaker and wet thoroughly with hot water. Place a perforated porcelain plate in a tin-lined copper extractor of the general form shown in Fig. 4, and on the plate place a layer of cotton and wet thoroughly with water. Connect the extractor with an 800 cc. Erlenmeyer flask (G), open the stop-cock (E) and close the outlets (C) and (D). Pour into the extractor the material to be extracted, washing it into the extractor with hot water. Return the percolate through the extractor until it is practically clear. Place a layer of cotton on top of the material. Close the stop-cock (E), connect with an 800 cc. Erlenmeyer flask containing about 650 cc. of water, connect (D) by a delivery tube with a liter graduated collecting flask, return the total percolate to the extractor and connect by means of the metal cap (B) with a block tin condenser (A) in such a way that the condensate will drip upon the layer of cotton. Boil the water in the flask, and collect 400–500 cc. of percolate from the side tube (D). Open the stop-cock (E) and close the side tube (D), add water to the flask (G), if necessary, until it contains about 200 cc. Continue the extraction with water at steam heat, allowing the percolate to run back into the boiling flask. Re-

peat with 2 successive portions (150–250 cc. each) of water for a total of 14 hours, heating at such a rate that approximately 330 cc. of water will be condensed per hour. Combine all the extracts in the graduated liter flask in which the first percolate was received. Heat to 80°C., cool, and make up to the mark.

21**ANALYSIS OF THE EXTRACT.**

Proceed as directed under **2–8**, inclusive. If more dilute solutions than the directions specify are employed in the determination of nontannins, the amount of hide powder used is reduced, as directed under **14**.

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VI. LEATHERS.—TENTATIVE.

VEGETABLE TANNED LEATHER.

1

PREPARATION OF SAMPLE.

Grind the sample, without undue heating, and pass through a 10 mesh sieve. The ground sample must not contain hard lumps. Plane heavily greased leathers (containing more than 20 % fat) into very thin shavings. Spread out the prepared sample and allow it to return to atmospheric moisture condition; mix thoroughly, and place in tightly covered containers.

2

MOISTURE.

Place 10 grams of the sample, as prepared under 1, in a tared, wide, shallow, weighing bottle (or a similar dish which can be covered tightly), and dry in a water oven for 15 hours at 98°–100°C. Cover the weighing bottle, cool in a desiccator containing sulphuric acid, and weigh. The moisture present in the leather as received may be determined by cutting it quickly into small pieces and drying without grinding as directed above.

3

TOTAL ASH.

Incinerate slowly 5 grams of the sample, as prepared under 1, at a dull red heat. If difficulty is experienced in burning off the carbon, leach the residue with hot water, filter on an ashless filter, dry and ignite the filter and residue, add the filtrate, evaporate to dryness and ignite. Cool in a desiccator containing sulphuric acid and weigh.

The ash may be examined for acids and bases by any suitable method. Aluminium, magnesium, sodium, barium, calcium and lead are the bases, and hydrochloric and sulphuric acids are the acids which it may be necessary to determine.

4

INSOLUBLE ASH.

Incinerate slowly the residue from the extraction of water-soluble material, obtained in 6 or 7, until all the carbon is burned off, cool in a desiccator containing sulphuric acid and weigh.

5

FATS.

Place, without packing, 15 grams of the leather, as prepared under 1, in a Soxhlet or Johnson extractor with a layer of fat-free cotton above and below the sample. Extract 8–10 hours with petroleum ether distilling between 50° and 80°C. Heavily greased leathers (containing 15% or more fat) will require the maximum time. Remove the receiving flask, evaporate the petroleum ether on the steam bath and dry the fat residue for 3 hours in a water oven at 98°–100°C., cool in a desiccator and weigh. Repeat the drying in the water oven for periods of 1–1½ hours, cooling and weighing as before, until no further loss in weight occurs. Retain the leather residue from the fat extraction for the extraction of water-soluble material in 6 or 7.

EXTRACTION OF WATER-SOLUBLE MATERIAL.

6

Method I.

Evaporate the petroleum ether from the fat-free leather, obtained under 5, and moisten thoroughly with from 100–150 cc. of water. Place a layer of cotton in the

bottom of a Soxhlet extractor designed for making extractions at temperatures below 100°C.

An extractor of this kind is furnished with a water jacket surrounding that portion of the apparatus containing the sample but does not enclose the side tube which carries the hot vapors to the condenser.

Transfer the moistened fat-free leather to the extractor, and cover this with another layer of cotton to avoid siphoning off solid particles. Maintain the temperature of the jacket surrounding the Soxhlet at 50°C. (1) Pour 200 cc. of water (including that used in moistening the leather) into the Soxhlet and allow it to siphon into the flask below, then heat and extract for an hour. Remove the flame and transfer the extract to a liter graduated flask. Then add water and continue the extraction as directed below, removing and transferring the extract to the liter flask before each fresh addition of water.

(2) Add 175 cc. of water and extract for 2 hours.

(3) Add 175 cc. of water and extract for 3 hours.

(4) Add 175 cc. of water and extract for 4 hours.

(5) Add 175 cc. of water and extract for 4 hours.

Transfer the last portion of the extract to the graduated flask. This gives 14 hours' extraction and an extract which does not exceed 1 liter in volume. Dilute to 1 liter at room temperature and mix thoroughly.

7

Method II.

(This method is the same in principle as the official method of the American Leather Chemists Association.¹)

Digest overnight 30 grams of the fat-free leather, obtained under 5, in approximately 200 cc. of water. Transfer the leather and extract to a percolator. Continue the extraction by percolating with water at 50°C. Collect 2 liters of percolate, regulating the flow of water at such a rate that 2 liters will be collected in 3 hours. Dilute to volume at room temperature and mix thoroughly.

To the extract, prepared according to 6 or 7, add a few drops of toluol to prevent fermentation of sugars, and reserve for the determination of glucose, total solids, soluble solids, and nontannins.

GLUCOSE.

8

PREPARATION OF SOLUTION.

To 200 cc. of the leather extract, as prepared under 6 or 7, add 25 cc. of a saturated solution of normal lead acetate, mix thoroughly, and filter at once through a dry, plaited paper, returning the first portions of the filtrate to the filter until the filtrate becomes clear. Keep the containers and the funnel covered during these operations. Without waiting for the entire filtrate to run through add 10-12 grams of solid potassium oxalate, shake frequently during 15-20 minutes and filter through a dry, plaited paper returning the first runnings to the filter until the filtrate runs clear. Pipette 150 cc. of the last filtrate into a 600 cc. Erlenmeyer flask, add 5 cc. of concentrated hydrochloric acid and boil under a reflux condenser for 2 hours. Cool, neutralize with solid sodium carbonate, using a little phenolphthalein as indicator, transfer to a 200 cc. volumetric flask and complete to volume with water. Filter through a double filter, and return the first runnings until the filtrate becomes perfectly clear. Determine the dextrose in the filtrate immediately.

9 DETERMINATION.

Determine dextrose in 50 cc. of the solution, as prepared under **8**, equivalent to 0.5 gram of leather, according to **VIII, 25** and express the result as glucose.

10 TOTAL SOLIDS.

Determine as directed under **V, 2**.

11 SOLUBLE SOLIDS.

Determine as directed under **V, 4**.

12 NONTANNINS.

Determine as directed under **V, 7**.

13 SOLUBLE TANNIN.

The difference between the percentage of the soluble solids and the corrected nontannins is the percentage of tannin.

14 NITROGEN.

Determine as directed under **I, 21**.

15 HIDE SUBSTANCE.

Multiply the percentage of nitrogen by 5.62. The result will be the percentage of hide substance present.

16 COMBINED TANNIN.

Deduct the sum of the percentages of moisture, under **2**, insoluble ash, under **4**, soluble solids, under **11**, and hide substance, under **15**, from 100. The result will be the percentage of combined tannin.

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VII. INSECTICIDES AND FUNGICIDES.

GENERAL METHOD.

1

PREPARATION OF SAMPLE.—TENTATIVE.

Mix thoroughly all samples before analysis. Make water-soluble arsenic determinations on samples as received without further pulverization or drying. In the case of lye, sodium cyanid or potassium cyanid, weigh large quantities in weighing bottles and analyze aliquots of the aqueous solutions.

PARIS GREEN.

2

MOISTURE.—TENTATIVE.

Dry 2 grams at 105°–110°C. for 5 hours and express the loss in weight as moisture.

TOTAL ARSENIC.¹—OFFICIAL.

(Arsenic, present as arsenate, is titrated as arsenious oxid.)

3

REAGENTS.

(a) *Starch indicator*.—Mix about 0.5 gram of finely powdered potato starch with cold water to a thin paste; pour into about 100 cc. of boiling water.

(b) *Standard arsenious oxid solution*.—Dissolve 2 grams of pure arsenious oxid in a beaker by boiling with about 150–200 cc. of water containing 10 cc. of concentrated sulphuric acid, cool, transfer to a 500 cc. graduated flask and dilute to the mark.

(c) *Standard iodine solution*.—Prepare an approximately N/20 solution as follows: Mix intimately 6.35 grams of pure iodine with twice its weight of pure potassium iodid. Dissolve in a small amount of water, filter and dilute the filtrate to 1 liter in a liter graduated flask. Standardize against (b) as follows: Pipette 50 cc. of the arsenious oxid into an Erlenmeyer flask, dilute to about 400 cc., neutralize with sodium bicarbonate, add 4–5 grams in excess, and add the standard iodine solution from a burette, shaking the flask continuously, until the yellow color disappears slowly from the solution, then add 5 cc. of the starch indicator and continue adding the iodine solution, drop by drop, until a permanent blue color is obtained. Calculate the value of the standard iodine solution in terms of *arsenious oxid* (As_2O_3) and *arsenic oxid* (As_2O_5). Occasionally restandardize the iodine against freshly prepared arsenious oxid solution.

4

APPARATUS.

The apparatus used is shown in Fig. 5. The distillation flask rests on a metal gauze which fits over a circular hole in a heavy sheet of asbestos board. The first 2 Erlenmeyer flasks are of 500 and 1000 cc. capacity and contain about 40 and 100 cc. of water, respectively. Both of these flasks should be placed in a pan and kept surrounded with cracked ice and water. The third flask, containing a small amount of water, is used as a trap.

5

DETERMINATION.

Weigh an amount of the sample equal to the arsenious oxid equivalent of 250 cc. of the standard iodine solution, and wash into the distillation flask by means of 100 cc. of concentrated hydrochloric acid (sp. gr. 1.19). Add 5 grams of cuprous chlorid (Cu_2Cl_2) and distil.

When the volume in the distillation flask is reduced to about 40 cc., add 50 cc. of concentrated hydrochloric acid by means of the dropping funnel and continue the distillation until 200 cc. of the acid distillate have passed over. Then wash down the condenser and all the connecting tubes carefully, transfer these washings and the contents of the 3 Erlenmeyer flasks to a liter graduated flask and dilute to the mark. Mix thoroughly, pipette 400 cc. into an Erlenmeyer flask and nearly neutralize with a saturated solution of sodium or potassium hydroxid, using a few drops of phenolphthalein as an indicator, keeping the solution well cooled.

Continue as directed under 3 C) beginning with "neutralize with sodium bicarbonate." The number of cc. of iodine used in this titration represents directly the total per cent of arsenic in the sample expressed as arsenious oxid (As_2O_3).

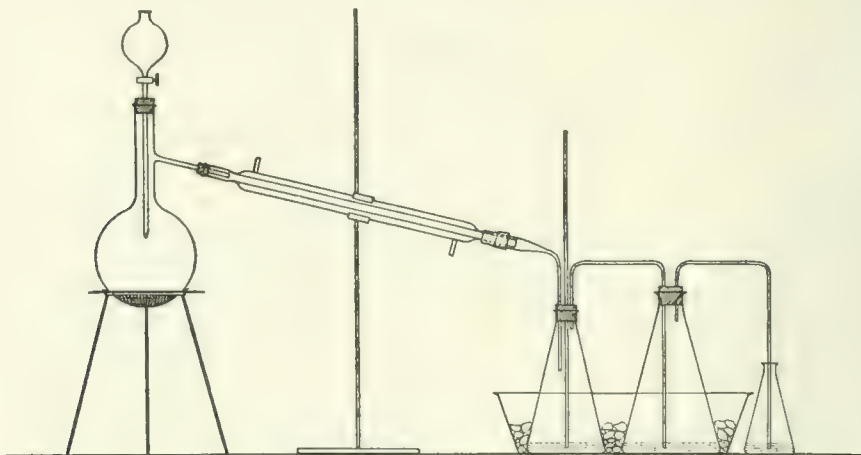


FIG. 5. APPARATUS FOR DISTILLATION OF ARSENIC CHLORID.

TOTAL ARSENIOS OXID.

(The following methods determine arsenic, and antimony if present, as the -ous oxids, As_2O_3 and Sb_2O_3 , respectively. Ferrous and cuprous salts vitiate the results.)

Method I.

C. C. Hedges Method,² Modified.³—Tentative.

6

REAGENTS.

The reagents and solutions used are described under 3.

7

DETERMINATION.

Weigh an amount of the sample equal to the arsenious oxid equivalent of 100 cc. of the standard iodine solution, wash into an Erlenmeyer flask with 10–15 cc. of dilute hydrochloric acid (1 to 1), followed by about 100 cc. of water, and heat on the steam-bath to complete solution, at a temperature not exceeding 60°C. Cool, neutralize

with sodium bicarbonate, add 4–5 grams in excess, and then sufficient 25% ammonium chlorid solution to dissolve the precipitated copper. Dilute somewhat and titrate as directed under 3 (C). A correction must be applied for the amount of iodine solution necessary to produce a blue color with starch in the presence of copper (using an equivalent weight of copper sulphate). The corrected number of cc. of the standard iodine solution used represents directly the per cent of arsenious oxide (As_2O_3) in the sample.

Method II.

- 8 *C. M. Smith Method,³ Modified.—Tentative.*
Proceed as directed in 7, using dilute sulphuric acid (1 to 4) instead of dilute hydrochloric. The solution in this case may be heated to boiling.

SODIUM ACETATE-SOLUBLE ARSENIOS OXID.—TENTATIVE.

- 9 **REAGENTS.**
(a) *Sodium acetate solution.*—Prepare a solution containing 12.5 grams of the crystallized salt ($\text{CH}_3\text{COONa} \cdot 3\text{H}_2\text{O}$) in each 25 cc.
The other reagents are described under 3.

- 10 **DETERMINATION.**
Place 1 gram of the sample in a 100 cc. flask and boil for 5 minutes with 25 cc. of the sodium acetate. Dilute to the mark, shake, and pass through a dry filter paper. Titrate an aliquot of this filtrate as directed under 3 (C). Calculate the amount of arsenious oxide (As_2O_3) present and express the result as per cent of sodium acetate-soluble arsenious oxide.

WATER-SOLUBLE ARSENIOS OXID.—TENTATIVE.

- 11 **REAGENTS.**
Described under 3.

- 12 **DETERMINATION.**
To 1 gram of the sample in a liter Florence flask add 1 liter of recently boiled water which has been cooled to exactly 32°C. Stopper the flask and place in a water bath kept at 32°C. by means of a thermostat. Digest for 24 hours, shaking hourly for 8 hours during this period. Filter through a dry filter and titrate 250 cc. of the filtrate as directed under 3 (C). Correct for the amount of the standard iodine necessary to produce the same color, using the same reagents and volume. Calculate the amount of arsenious oxide (As_2O_3) present and express the result as per cent of water-soluble arsenious oxide.

TOTAL COPPER OXID.

- 13 *Electrolytic Method.—Official.*
Treat 2 grams of the sample in a beaker with 100 cc. of water and about 2 grams of sodium hydroxide and boil thoroughly until all the copper is precipitated as cuprous oxide. Filter, wash well with hot water, dissolve the precipitate in hot dilute nitric acid, cool, transfer to a 250 cc. graduated flask and dilute to the mark. (1) Use 50–100 cc. of this solution for the electrolytic determination of copper as directed under VIII, 33 and calculate to per cent cupric oxide; or, (2) Electrolyze the aliquot in a weighed 150 cc. platinum dish, using a rotating spiral anode and a current of about

3 amperes. After all the copper is deposited (requiring about 30 minutes), wash the deposit with water by siphoning, then rinse with alcohol, dry for a few minutes in an oven, weigh and calculate to per cent cupric oxid.

14

*Thiosulphate Method.*⁵—Official.

Determine copper in another aliquot of the nitric acid solution of copper oxid, under 13, by titrating with N/20 thiosulphate solution, as directed under VIII, 29, and calculate to per cent cupric oxid.

LONDON PURPLE.

15

MOISTURE.—TENTATIVE.

Determined as directed under 2.

TOTAL ARSENIOUS OXID.⁶—Official.

16

REAGENTS.

Described under 3.

17

DETERMINATION.

Dissolve 2 grams of the sample in a mixture of about 80 cc. of water and 20 cc. of concentrated hydrochloric acid at a temperature of 60°–70°C.; filter and wash until the combined filtrate and washings measure 250 cc. Treat 100 cc. of this solution with sodium bicarbonate in excess, transfer to a 500 cc. volumetric flask and make up to the mark, adding a few drops of ether to destroy the bubbles. Mix thoroughly and pass through a dry filter. Titrate 250 cc. of the filtrate as directed under 3 (C) and calculate the per cent of arsenious oxid.

TOTAL ARSENIC OXID.⁷—Official.

18

REAGENTS.

The reagents and solutions used are described under 3.

19

DETERMINATION.

Boil, on a hot plate or over a low flame, 2 grams of the sample with 5 cc. of concentrated nitric acid and 20 cc. of concentrated sulphuric acid in a Kjeldahl digestion flask or a covered casserole. After 10–15 minutes add fuming nitric acid or powdered sodium nitrate, in small quantities at a time, until all organic matter is destroyed and the solution is practically colorless. Cool, add about 50 cc. of water (to decompose any nitro-sulphuric acid formed) and heat again until all nitric acid fumes are expelled. Cool, transfer to a 250 cc. volumetric flask, make up to the mark with water, mix thoroughly, and filter through a dry filter.

Transfer 50 cc. of this filtrate to a 400 cc. Erlenmeyer flask, dilute with water to 100 cc., add 1 gram of potassium iodid,⁸ heat to boiling and evaporate to about 40 cc. (not less). Cool, dilute to 150–200 cc., and remove the excess of iodine with N/20 sodium thiosulphate. In case the solution is slightly colored from organic matter or from any cause other than free iodine, add the thiosulphate until it is nearly colorless, then a few drops of the starch indicator, and continue adding the thiosulphate slowly until the blue color just disappears. Continue at once as directed under 3 (C) beginning with "neutralize with sodium bicarbonate." Subtract from this reading the number of cc. of the standard iodine solution corresponding to the arsenious oxid obtained in 17. Calculate the per cent of arsenic oxid in the sample.

20

WATER-SOLUBLE ARSENIOS OXID.—TENTATIVE.

Proceed as directed under 12, slightly acidifying the aliquot employed with hydrochloric acid before adding the excess of sodium bicarbonate.

WATER-SOLUBLE ARSENIC OXID.—TENTATIVE.

21

REAGENTS.

The solutions and reagents used are described under 3.

22

DETERMINATION.

Transfer an aliquot, 250 cc., of the water extract, from 20, to a casserole, add 5 cc. of concentrated sulphuric acid, evaporate to a small volume and heat on a hot plate till white fumes of sulphuric acid appear. Cover the casserole and add 1-2 cc. of fuming nitric acid and again heat till the appearance of white fumes. Cool, add a little water and, in order to expel the last traces of nitric acid, once more evaporate till white fumes appear. Cool, dilute to about 100 cc. with water, add 1 gram of potassium iodid³ and sufficient sulphuric acid to make the total amount present about 5 cc. Boil until the volume is reduced to about 40 cc. Cool, dilute to about 200 cc., remove the excess iodine with N/20 sodium thiosulphate and proceed as directed under 3 (c) beginning with "neutralize with sodium bicarbonate." Correct for the amount of the standard iodine solution necessary to produce the same color, using the same reagents and volume. Subtract from the corrected titration reading the number of cc. of the standard iodine solution corresponding to the arsenious oxid, obtained in 20. Calculate the per cent of arsenic oxid present.

LEAD ARSENATE.

23

MOISTURE.—TENTATIVE.

(a) *Powder*.—Dry 2 grams to constant weight at 105°–110° C. and report the loss in weight as moisture.

(b) *Paste*.—Proceed as under (a), using 50 grams.

Grind the dry sample to a fine powder, mix well, transfer a small portion to a sample bottle and again dry for 1-2 hours at 105°–110°C., and use this anhydrous material for the determination of total lead oxid and total arsenic.

TOTAL LEAD OXID.

24

Method I.⁹—Official.

Heat, on a hot plate, 0.6906 gram of the dry powdered sample with about 25 cc. of dilute nitric acid (1 to 4) in a 600 cc. beaker. If necessary, remove any insoluble residue by filtration. Dilute to at least 400 cc., heat nearly to boiling, add ammonium hydroxid to incipient precipitation, then dilute nitric acid (1 to 10) to redissolve the precipitate, adding 1-2 cc. in excess. Pipette into this solution, kept almost boiling, 50 cc. of a hot 10% potassium chromate solution, stirring constantly. Decant while hot through a weighed Gooch, previously heated at 140°–150°C., wash several times by decantation and then on the filter with boiling water until the washings are colorless. Dry the lead chromate at 140°–150°C. to constant weight. The weight of lead chromate multiplied by 100 gives the per cent of lead monoxid (PbO) in the dried sample.

The lead chromate precipitate may contain a small amount of lead arsenate which causes slightly high results. This error rarely amounts to more than 0.1-0.2%.

Method II.¹⁰—Tentative.

(Not applicable in the presence of calcium.)

25

REAGENT.

Acidified alcohol.—Mix water 100 parts; 95% alcohol 200 parts; and concentrated sulphuric acid 3 parts by volume.

26

DETERMINATION.

Heat, on a hot plate, 0.7360 gram of the dry powdered sample with about 25 cc. of dilute nitric acid (1 to 4) in a porcelain evaporating dish or casserole. Remove any insoluble residue by filtration. Add 3 cc. of concentrated sulphuric acid and evaporate on the hot plate to the appearance of white fumes. It is important that all nitric acid be expelled. Cool, add 50 cc. of water and about 100 cc. of 95% alcohol, let stand several hours (preferably over-night) and filter through a weighed Gooch crucible, previously washed with water, the acidified alcohol and 95% alcohol, and dried at 200°C. Wash the precipitate of lead sulphate in the crucible about 10 times with the acidified alcohol and then with 95% alcohol until free from sulphuric acid. Dry at 200°C. to constant weight, keeping the crucible covered to prevent loss by spattering. The weight of the lead sulphate multiplied by 100 gives the per cent of lead monoxid (PbO) in the dried sample.

TOTAL ARSENIC.

27

Method I.¹—Official.

Proceed as directed under 5, using an amount of the sample equal to the arsenic oxid equivalent of 500 cc. of the standard iodine solution and titrating a 200 cc. aliquot of the distillate. The number of cc. used of the standard iodine solution represents directly the total per cent of arsenic in the sample expressed as arsenic oxid (As_2O_5).

Method II.¹¹—Official.

(Not applicable in the presence of antimony.)

28

REAGENTS.

The reagents and solutions used are described under 3.

29

DETERMINATION.

Dissolve an amount of the powdered sample equal to the arsenic oxid equivalent of 400 cc. of the standard iodine solution, in dilute nitric acid in a porcelain casserole or evaporating dish. Add 5 cc. of concentrated sulphuric acid and heat on the hot plate to copious evolution of white fumes. Wash into a 200 cc. graduated flask with water, cool, make up to the mark and filter through a dry filter. Transfer 100 cc. of the filtrate to an Erlenmeyer flask and proceed as directed under 22, beginning with "add 1 gram of potassium iodide," to "Subtract from the corrected titration reading." The number of cc. of the standard iodine solution used, divided by 2, represents directly the per cent of total arsenic in the sample expressed as arsenic oxid (As_2O_5).

WATER-SOLUBLE ARSENIC OXID.—TENTATIVE.

30

REAGENTS.

The reagents and solutions used are described under 3.

31

DETERMINATION.

Treat 2 grams of the original sample, if in the form of a powder, or 4 grams, if a paste, as directed under 12 through "Filter through a dry filter."

Place 250-500 cc. of the *clear* filtrate in an Erlenmeyer flask, add 3 cc. of concentrated sulphuric acid and evaporate on a hot plate. When the volume is reduced to about 100 cc., proceed as directed under 22 to "Subtract from the corrected titration reading." Calculate and report as per cent of water-soluble arsenic oxid (As_2O_5).

CALCIUM ARSENATE.

32

TOTAL ARSENIC.¹—OFFICIAL.

Proceed as directed under 5, using an amount of the powdered sample equal to the arsenic oxid equivalent of 250 cc. of the standard iodine solution.

The number of cc. of the standard iodine solution used represents directly the total per cent of arsenic in the sample expressed as arsenic oxid (As_2O_5).

ZINC ARSENITE.

33

TOTAL ARSENIC.¹—OFFICIAL.

Proceed as directed under 5, using an amount of the powdered sample equal to the arsenious oxid equivalent of 500 cc. of the standard iodine solution and titrating a 200 cc. aliquot of the distillate. The number of cc. of the standard iodine solution used represents directly the per cent of total arsenic in the sample expressed as arsenious oxid (As_2O_3).

34

TOTAL ARSENIOUS OXID.—TENTATIVE.

Proceed as directed under 7 or 8.

COPPER CARBONATE.

35

COPPER OXID.—OFFICIAL.

Dissolve a weighed quantity of the substance in dilute nitric acid and determine copper as directed under 13 or 14.

BORDEAUX MIXTURE.

36

MOISTURE.—OFFICIAL.

(a) *Powder*.—Dry 2 grams to constant weight at 105°-110°C. and express the loss in weight as moisture.

(b) *Paste*.—Heat about 100 grams in an oven at 90-100°C. until dry enough to powder readily, and note the loss in weight. Powder this partially dried sample, and determine the remaining moisture in 2 grams as under (a). Determine carbon dioxid, as directed under 38, both in the original paste and in this partially dried sample. Calculate the total moisture by the following formula:

$M = a + (100-a)(b+c) - d$ in which

M = per cent total moisture in original paste;

a = per cent loss in weight of original paste during first drying;

b = per cent loss in weight of partially dried paste during second drying;

c = per cent carbon dioxid remaining in partially dried paste after first drying;

d = per cent total carbon dioxid in original paste.

CARBON DIOXID.¹²—OFFICIAL.

37

APPARATUS.

This consists of a 200 cc. Erlenmeyer flask closed with a 2-holed stopper; one of these holes is fitted with a dropping funnel the stem of which extends almost to the bottom of the flask; the outlet of a condenser, which is inclined upward at an angle of 30° from the horizontal, passes downward through the other hole. The upper end of the condenser is connected with a calcium chlorid tube which in turn is connected with a double U-tube filled in the middle with pumice fragments, previously saturated with copper sulphate solution and subsequently dehydrated, and with calcium chlorid at either end. Then follow 2 weighed U-tubes for absorbing the carbon dioxide, the first filled with porous soda-lime, and the second, one third with soda-lime and two thirds with calcium chlorid, the latter reagent being placed at the exit end of the train. A Geissler bulb, partly filled with sulphuric acid, is attached to the last U-tube to show the rate of gas flow. An aspirator is connected with the Geissler bulb to draw air through the apparatus. An absorption tower filled with soda-lime is connected with the mouth of the dropping funnel to remove carbon dioxide from the air entering the apparatus.

38

DETERMINATION.

Weigh 2 grams of the powder or 10 grams of the paste into the Erlenmeyer flask, add about 20 cc. of water, attach the flask to the apparatus omitting the 2 weighed U-tubes, and draw carbon dioxide-free air through the apparatus until the original air is displaced. Then attach the weighed U-tubes in the position as described in 37, close the stop-cock of the dropping funnel, fill half full with dilute hydrochloric acid (1 to 1), reconnect with the soda-lime tower, and allow the acid to flow into the Erlenmeyer flask, slowly if there is much carbon dioxide, rapidly if there is little. When effervescence diminishes, place a low Bunsen flame under the flask and start a flow of water through the condenser, a slow current of air being allowed to flow through the apparatus at the same time. Maintain a steady but quiet ebullition, and a slow air current through the apparatus. Boil for a few minutes after the water has begun to condense in the condenser, then remove the flame and continue the aspiration of air at the rate of about 2 bubbles per second until the apparatus is cool. Disconnect the tared absorption tubes, cool in the balance case and weigh. The increase in weight is due to carbon dioxide.

COPPER.

39

Electrolytic Method.—Official.

Dissolve 2 grams of the dry powdered sample in 20 cc. of water and 5 cc. of concentrated nitric acid, dilute to 100 cc., wash into a weighed 150 cc. platinum dish, and electrolyze, using a rotating spiral anode and a current of about 3 amperes. After all the copper is deposited (requiring about 30 minutes), wash the deposit with water by siphoning, then rinse with alcohol, dry for a few minutes in an oven, and weigh. Calculate the per cent of copper in the sample.

40

Thiosulphate Method.—Official.

Dissolve 2 grams of the dry powdered sample in about 50 cc. of 10% nitric acid, add ammonium hydroxid solution in excess and heat; then, without removing the precipitate which is formed, boil off the excess of ammonia, add 5-10 cc. of acetic acid, cool, add 10 cc. of 30% potassium iodid solution, and titrate as directed under VIII, 29.

BORDEAUX MIXTURE WITH PARIS GREEN.

41 MOISTURE.—OFFICIAL.

Proceed as directed under 36.

42 CARBON DIOXID.—OFFICIAL.

Proceed as directed under 38.

COPPER.

43 *Method I.—Tentative.*

Dissolve 2 grams of the dry powdered sample in a few cc. of strong nitric acid, add 25 cc. of a 3% solution of hydrogen peroxid and warm for 5–10 minutes. Make slightly alkaline with ammonium hydroxid and then slightly acid again with dilute nitric acid. Transfer to a weighed 150 cc. platinum dish, add 15–20 cc. of hydrogen peroxid, dilute to 100 cc. and electrolyze, using a rotating spiral anode and a current not exceeding 2 amperes. After the electrolysis has proceeded for about 20 minutes, add to the electrolyte 0.5 gram of ferric sulphate dissolved in a few cc. of water together with a drop or two of nitric acid. After all the copper is deposited, wash the deposit with water by siphoning, then rinse with alcohol, dry for a few minutes in an oven, weigh and calculate the per cent of copper. (Do not pass the current for more than 5–10 minutes after all the copper has been deposited without adding more ferric sulphate solution.)

44 *Method II.—Tentative.*

Treat 1 gram of the dry powdered sample with 20 cc. of water and 5–6 cc. of concentrated nitric acid, heat to boiling, cool, and add a slight excess of concentrated ammonium hydroxid. Wash the solution and precipitate into a weighed platinum dish of about 150 cc. capacity, and electrolyze, using a rotating anode and a current of about 4 amperes and 3–4 volts for about 90 minutes (or until all the copper is deposited). Wash the deposit by siphoning until the deposit is clean, being careful not to use too much wash water. Dissolve the copper in 5 cc. of concentrated nitric acid, dilute to 100 cc. and electrolyze as before, except that all the copper will be deposited in 30 minutes. Wash the deposit with water by siphoning, then rinse with alcohol, dry for a minute or so in an oven, weigh and calculate the per cent of copper.

45 TOTAL ARSENIC.¹—OFFICIAL.

Proceed as directed under 5, using an amount of the dry powdered sample equal to the arsenious oxid equivalent of 500 cc. of the standard iodine solution. The number of cc. of the standard iodine solution used, divided by 2, represents directly the per cent of total arsenic in the sample expressed as arsenious oxid (As_2O_3).

TOTAL ARSENIUS OXID.

46 *Method I.—Tentative.*

Proceed as directed under 7, using an amount of the dry, powdered sample equal to the arsenious oxid equivalent of 200 cc. of the standard iodine solution. Before titrating, all the copper must be in solution. The corrected number of cc. of the standard iodine solution used, divided by 2, represents directly the per cent of total arsenious oxid (As_2O_3) in the sample.

47 *Method II.—Tentative.*

Proceed as directed under 8.

48 WATER-SOLUBLE ARSENIOUS OXID.—TENTATIVE.

Proceed as directed under 20, using 2 grams of the sample.

BORDEAUX MIXTURE WITH LEAD ARSENATE.

49 MOISTURE.—OFFICIAL.

Proceed as directed under 36.

50 CARBON DIOXID.—OFFICIAL.

Proceed as directed under 38.

51 COPPER.—TENTATIVE.

Proceed as directed under 44.

52 LEAD OXID.³—TENTATIVE.

Dissolve the lead peroxid (which will contain a little arsenic) from the anodes used in the copper electrolysis, under 51, by means of dilute nitric acid and a little hydrogen peroxid, and add to this solution the washings from both electrolyses of copper. Add ammonium chlorid to dissolve any lead sulphate which may have precipitated out and make the solution up to 1 liter. Concentrate a 500 cc. aliquot of this solution to about 300 cc. (all hydrogen peroxid must be expelled from the solution), transfer to a 400 cc. beaker and precipitate the lead as lead chromate as directed under 24.

53 TOTAL ARSENIC.¹—OFFICIAL.

Proceed as directed under 5, using an amount of the dry, powdered sample equal to the arsenic oxid equivalent of 500 cc. of the standard iodine solution. The number of cc. of the standard iodine solution used, divided by 2, represents directly the per cent of total arsenic in the sample expressed as arsenic oxid (As_2O_5).

54 WATER-SOLUBLE ARSENIC OXID.—TENTATIVE.

Proceed as directed under 31.

SODIUM AND POTASSIUM CYANIDS.

55 CYANOGEN.¹²—OFFICIAL.

Weigh about 10 grams of the sample in a weighing bottle, dissolve in water, and make up to volume in a liter graduated flask. To a 50 cc. aliquot add N/20 silver nitrate, drop by drop, stirring constantly, until 1 drop produces a permanent turbidity. In calculating the results, 1 equivalent of silver is equal to 2 equivalents of cyanogen, according to the following equation:



Reserve the titrated solution for the determination of chlorin under 56.

56

CHLORIN.¹⁴—OFFICIAL.

After completion of the titration for cyanogen, as directed under 55, add a few cc. of 10% potassium chromate solution as indicator and titrate with N/20 silver nitrate until the appearance of the red-brown color of silver chromate.

The first titration with silver nitrate represents the cyanogen present according to the equation above. The second titration represents the cyanogen and chlorin according to the following equation: $\text{NaCNAgCN} + \text{NaCl} + 2\text{AgNO}_3 = 2\text{NaNO}_3 + 2\text{AgCN} + \text{AgCl}$. Therefore the second minus the first reading represents the chlorin present in terms of silver nitrate.

SOAP.

MOISTURE.

57

*Modified Method of Benedickt and Lewkowitsch.*¹⁵—Tentative.

Weigh about 5 grams of the sample in a tared, 100 cc. beaker, in which is previously placed a $\frac{1}{2}$ inch layer of recently ignited, dry sand, and a small glass rod; if the soap is hard, cut off the soap in very thin strips. Add 25 cc. of alcohol, or more if necessary, and dissolve on the water bath, stirring constantly. Evaporate the alcohol, heat in an oven at 110°C. until the soap is nearly dry, and weigh, then dry again for 30 minutes and weigh. Continue this alternate drying and weighing until the weight changes only a few milligrams during the course of 30 minutes' drying.

58

POTASSIUM AND SODIUM.¹⁶—TENTATIVE.

Dissolve about 5 grams of the soap in water; decompose with hydrochloric acid, filter off the water and wash the fat with cold water. Determine both potassium and sodium in the filtrate as directed under II, 21.

SODA LYE.

59

CARBONATE AND HYDROXID.¹⁷—OFFICIAL.

Weigh about 10 grams of the sample from the weighing bottle, dissolve in carbon dioxid-free water and make up to a definite volume. Titrate an aliquot of this solution with N/2 hydrochloric acid, using methyl orange as an indicator, and note the total alkalinity thus found. Transfer an equal aliquot to a graduated flask and add enough barium chlorid solution to precipitate all the carbonate, avoiding any unnecessary excess. Dilute to the mark with carbon dioxid-free water, stopper, shake, and set aside. When the liquid becomes clear, pipette off one half and titrate with N/2 hydrochloric acid, using phenolphthalein as an indicator. The number of cc. of N/2 acid, required for this titration, multiplied by 2 gives the number of cc. of N/2 acid required to neutralize the sodium hydroxid present in the original aliquot. The difference between this figure and the number of cc. of N/2 hydrochloric acid required for the total alkalinity represents the number of cc. of N/2 acid required to neutralize the sodium carbonate present in the aliquot. Calculate the percentages of sodium carbonate and hydroxid present in the sample.

TOBACCO AND TOBACCO EXTRACT.

NICOTIN.

Kissling Method.—Official.

60

REAGENTS.

(a) *Alcoholic sodium hydroxid solution.*—Dissolve 6 grams of sodium hydroxid in 40 cc. of water and 60 cc. of 90% alcohol.

- (b) 0.4% sodium hydroxid solution.
- (c) N/10 sulphuric acid.—One cc. is equivalent to 16.22 mg. of nicotin.
- (d) Phenacetolin solution.—Prepare a 0.5% alcoholic solution.
- (e) Cochineal solution.—Prepare as directed under I, 16 (k).

61

DETERMINATION.

Weigh 5–6 grams of tobacco extract, or 20 grams of finely powdered tobacco which has been previously dried at 60°C. if necessary, into a small beaker. Add 10 cc. of the alcoholic sodium hydroxid and follow, in the case of tobacco extract, with enough pure powdered calcium carbonate to form a moist but not lumpy mass. Mix thoroughly, transfer to a Soxhlet extractor and exhaust for about 5 hours with ether. Evaporate the ether at a low temperature, and take up the residue with 50 cc. of the 0.4% sodium hydroxid solution. Transfer this residue by means of water to a 500 cc. Kjeldahl flask, and distil with steam, passing the distillate through a condenser cooled by a rapidly flowing current of water. Use a 3-bend outflow tube, and, to prevent bumping and frothing, add a few pieces of pumice, and a small piece of paraffin. Distil till all the nicotin has passed over, the distillate usually varying from 400–500 cc. When completed, only about 15 cc. of the liquid should remain in the flask. Titrate the distillate with N/10 sulphuric acid, using the phenacetolin or cochineal solution as indicator.

*Silicotungstic Acid Method.*¹⁸—Official.

62

REAGENTS.

- (a) *Silicotungstic acid solution*.—Prepare a 12% solution of the silicotungstic acid having the following formula: $4\text{H}_2\text{O} \cdot \text{SiO}_2 \cdot 12\text{WO}_3 \cdot 22\text{H}_2\text{O}$.
- (b) *Sodium or potassium hydroxid solution* (1 to 2).
- (c) *Dilute hydrochloric acid* (1 to 4).

63

DETERMINATION.

Weigh such an amount of the preparation as will contain preferably between 0.1 and 1.0 gram of nicotin (if the sample contains very little nicotin, about 0.1%, do not increase the amount to the point where it interferes with the distillation); wash with water into a 500 cc. round-bottomed distillation flask; add a little paraffin to prevent frothing, a few small pieces of pumice and a slight excess of the sodium or potassium hydroxid, using phenolphthalein as an indicator. Distil rapidly in a current of steam through a well-cooled condenser, connected by means of an adapter with a suitable flask containing 10 cc. of the dilute hydrochloric acid. When distillation is well under way, heat the distillation flask to reduce the volume of the liquid as far as practicable without bumping or undue separation of insoluble matter. Distil until a few cc. of the distillate show no cloud or opalescence when treated with a drop of the silicotungstic acid and a drop of the dilute hydrochloric acid. Confirm the alkalinity of the residue in the distillation flask with phenolphthalein solution. Make up the distillate, which may amount to 1000–1500 cc., to a convenient volume (the solution may be concentrated on the steam bath without loss of nicotin), mix well and pass through a large dry filter if not clear. Test a portion with methyl orange to assure its acidity. Pipette an aliquot, containing about 0.1 gram of nicotin, into a beaker (if the samples contain very small amounts of nicotin, an aliquot containing as little as 0.01 gram of nicotin may be used), add to each 100 cc. of liquid 3 cc. of the dilute hydrochloric acid, or more if the necessity is indicated by the test with methyl orange, and add 1 cc. of the silicotungstic acid for each 0.01 gram of nico-

tin supposed to be present. Stir thoroughly and let stand overnight. Before filtering, stir the precipitate to see that it settles quickly and is in crystalline form; then filter on an ashless filter paper, and wash with cold dilute hydrochloric acid (1 to 1000). Transfer the paper and precipitate to a weighed platinum crucible, dry carefully, and ignite until all carbon is destroyed. Finally heat over a Teclu or Meker burner for not more than 10 minutes. The weight of the residue multiplied by 0.114 gives the weight of nicotine present in the aliquot.

FORMALDEHYDE SOLUTIONS.

FORMALDEHYDE.

*Hydrogen Peroxid Method.*¹⁹—Official.

64

REAGENTS.

- (a) *N/1 sulphuric acid.*
- (b) *N/1 sodium hydroxid.*—One cc. is equivalent to 30.02 mg. of formaldehyde.
- (c) *Hydrogen peroxid.*—An approximately 3% solution. If the hydrogen peroxid solution is acid, neutralize with (b), using litmus solution as indicator.
- (d) *Litmus solution.*—A solution of purified litmus.

65

DETERMINATION.

Measure 50 cc. of N/1 sodium hydroxid into a 500 cc. Erlenmeyer flask and add 50 cc. of the hydrogen peroxid. Then add 3 grams of the formaldehyde solution under examination, allowing the point of the pipette to reach nearly to the liquid in the flask. Place a funnel in the neck of the flask and heat on the steam bath for 5 minutes, shaking occasionally. Remove from the steam bath, wash the funnel with water, cool the flask to about room temperature, and titrate with N/1 acid, using the litmus solution as indicator. It is necessary to cool the flask before titration with the acid to get a sharp end point with the litmus. Calculate the per cent of formaldehyde.

*Cyanid Method.*²⁰—Official.

66

REAGENTS.

- (a) *N/10 silver nitrate.*
- (b) *N/10 ammonium sulphocyanate.*
- (c) *Potassium cyanid solution.*—Dissolve 3.1 grams of potassium cyanid in 500 cc. of water.
- (d) *50% nitric acid.*

67

DETERMINATION.

Treat 15 cc. of the N/10 silver nitrate with 6 drops of the 50% nitric acid in a 50 cc. volumetric flask; add 10 cc. of the potassium cyanid solution, dilute to the mark, shake well, filter through a dry filter and titrate 25 cc. of the filtrate with N/10 ammonium sulphocyanate as directed under **III, 15**. Acidify another 15 cc. portion of the N/10 silver nitrate with 6 drops of the 50% nitric acid and treat with 10 cc. of the potassium cyanid solution to which has been added a measured quantity (the weight of which must be calculated from the specific gravity) of the formaldehyde solution containing not over 2.5 grams of a 1% solution or the equivalent. Make up to 50 cc., filter and titrate a 25 cc. aliquot with the N/10 ammonium sulphocyanate for the excess of silver as before. The difference between the number of cc. of N/10 ammonium sulphocyanate used in these 2 titrations, multiplied

by 2, gives the number of cc. of N/10 ammonium sulphocyanate corresponding to the potassium cyanid used by the formaldehyde. Calculate the per cent of formaldehyde present 1 cc. of N/10 ammonium sulphocyanate is equivalent to 3 mg. of formaldehyde (HCHO)).

LIME-SULPHUR SOLUTIONS.²¹

TOTAL SULPHUR.—OFFICIAL.

68

PREPARATION OF SOLUTION.

Weigh 10 grams of the solution and dilute to the mark in a 250 cc. graduated flask with recently boiled and cooled water.

69

DETERMINATION.

Transfer a 10 cc. aliquot to a 400 cc. beaker, add about 3 grams of sodium peroxid, cover immediately with a watch glass and warm on the steam bath, with frequent shaking, until all the sulphur is oxidized to sulphate, adding more sodium peroxid if necessary. Dilute, acidify with hydrochloric acid, evaporate to dryness, treat with water acidified with hydrochloric acid, boil, and filter to remove silica, if present. Dilute the filtrate to 300 cc., add 50 cc. of concentrated hydrochloric acid,²² heat to boiling, and precipitate with 10% barium chlorid solution slowly and stirring constantly. (The rate is best regulated by attaching a suitable capillary tip to the burette containing the barium chlorid solution.) Evaporate to dryness on the steam bath, take up with hot water, filter through a quantitative filter paper, wash until free from chlorin, ignite and heat to constant weight over a Bunsen burner. Calculate the sulphur from the weight of barium sulphate. Previous to use test the reagents for sulphur and, if present, make corrections accordingly.

SULPHID SULPHUR.—OFFICIAL.

70

REAGENT.

Ammoniacal zinc solution.—Dissolve 50 grams of pure zinc chlorid in water, add ammonium hydroxid in sufficient quantity to redissolve the precipitate first formed, then add 50 grams of ammonium chlorid²³ and dilute to 1 liter.

71

DETERMINATION.

Dilute 10 cc. of the solution, prepared as directed under 68, to about 100 cc. and add the ammoniacal zinc solution until the sulphid is all precipitated, indicated by the addition of a drop of the clear solution to a few drops of nickel sulphate solution. Filter immediately, wash the precipitate thoroughly with cold water and transfer it and the filter paper to a beaker. Cover with water, disintegrate with a glass rod and add about 3 grams of sodium peroxid, keeping the beaker well covered with a watch glass. Warm on the steam bath with frequent shaking until all the sulphur is oxidized to sulphate, adding more sodium peroxid if necessary. Make slightly acid with hydrochloric acid, filter to remove shreds of filter paper, wash thoroughly with hot water, and determine the sulphur in the filtrate exactly as under 69.

72

THIOSULPHATE SULPHUR.—OFFICIAL.

Dilute 50 cc. of the solution, prepared as under 68, to about 100 cc. in a 200 cc. graduated flask. Add a slight excess of the ammoniacal zinc chlorid and dilute to the mark. Shake thoroughly and filter through a dry filter. To 100 cc. of the filtrate add a few drops of methyl orange and exactly neutralize with N/10 hydrochloric

acid. Titrate this neutral solution with approximately N/20 iodine, **3 (C)**, using a few drops of starch solution as indicator. From the number of cc. of iodine solution used, calculate the thiosulphate sulphur present.

73**SULPHATE SULPHUR.—OFFICIAL.**

To the solution from the determination in **72**, add 2 or 3 drops of hydrochloric acid, precipitate in the cold with 10% barium chlorid solution, allow to stand overnight, filter, calculate the sulphur from the weight of barium sulphate and report as sulphate sulphur.

74**TOTAL LIME.—OFFICIAL.**

To 25 cc. of the solution, prepared as under **68**, add 10 cc. of concentrated hydrochloric acid, evaporate to dryness on the steam bath, treat with water and a little hydrochloric acid, warm until all the calcium chlorid is dissolved, and filter from sulphur and any silica that may be present. Oxidize the filtrate by boiling with a little concentrated nitric acid, make ammoniacal, filter from iron and aluminium if present, heat to boiling and precipitate the calcium with ammonium oxalate solution. Filter, wash and ignite over a blast lamp to constant weight; weigh the residue as calcium oxid.

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VIII. FOODS AND FEEDING STUFFS.

1 PREPARATION OF SAMPLE.—OFFICIAL.

Grind the sample so that it will pass through a sieve having circular openings $\frac{1}{16}$ inch (1 mm.) in diameter. If the sample can not be ground, reduce it to as fine a state as possible.

MOISTURE.

2 *Direct Drying.—Official.*

Dry a quantity of the substance, representing about 2 grams of dry material, in a current of dry hydrogen or in vacuo at the temperature of boiling water to constant weight (approximately 5 hours). If the substance be held in a glass vessel, the latter should not be in contact with the boiling water.

3 *Drying in Vacuo without Heat.—Tentative.*

Mix the sample thoroughly and weigh by difference 2-5 gram portions from a stoppered weighing bottle into tared, covered crucibles. Where subsequent fat determinations are to be made, fat extraction cones may be used. Substances that dry down to horn-like material should be mixed with fat-free cotton or other suitable material (previously tared with the container). Place 200 cc. of fresh concentrated sulphuric acid in a strong, tight 6 inch vacuum desiccator. Put triplicate samples in separate desiccators, and exhaust by means of a vacuum pump. If a pump is not available, place 10 cc. of ether in a small beaker in the desiccator, and exhaust with a water filter pump.

Between the pump and the desiccator interpose an empty bottle, next to the desiccator, and a bottle of water. Draw the air from the desiccator through the water and turn the desiccator stop-cock at just the instant when the water begins to rise in the tube leading from the empty bottle.

Gently rotate the desiccator 4 or 5 times during the first 12 hours to mix the sulphuric acid with the water which has collected as an upper layer. At the end of 24 hours open the desiccator, forcing the incoming air to bubble through concentrated sulphuric acid, and make the first weighing. After weighing place in a desiccator containing fresh concentrated sulphuric acid and exhaust as before. Rotate the desiccator several times during the interval and weigh again after a suitable period of drying. Repeat this process of drying in vacuo over sulphuric acid until the weight is constant.

4 ASH.—OFFICIAL.

Char a quantity of the substance, representing about 2 grams of the dry material, and burn until free from carbon at a low heat, not to exceed dull redness. If a carbon-free ash can not be obtained in this manner, exhaust the charred mass with hot water, collect the insoluble residue on a filter, burn till the ash is white or nearly so, and then add the filtrate to the ash and evaporate to dryness. Heat to low redness till the ash is white or grayish white and weigh.

5

CRUDE PROTEIN.—OFFICIAL.

Determine nitrogen as directed under **I**, **18**, **21**, or **23**, and multiply the result by 6.25.

ALBUMINOID NITROGEN.—OFFICIAL.

6

REAGENT.

Stutzer's reagent.—Prepare cupric hydroxid as follows: Dissolve 100 grams of pure copper sulphate in 5 liters of water, add 2.5 cc. of glycerol, and then dilute sodium hydroxid solution until the liquid is just alkaline; filter, rub the precipitate up with water containing 5 cc. of glycerol per liter, and wash by decantation or filtration until the washings are no longer alkaline. Rub the precipitate up again in a mortar with water containing 10% of glycerol, thus preparing a uniform gelatinous mass that can be measured with a pipette. Determine the quantity of copper hydroxid per cc. of this mixture.

7

DETERMINATION.

Place 0.7 gram of the substance in a beaker, add 100 cc. of water, and heat to boiling; or, in case of substances rich in starch, heat on the water bath for 10 minutes; add a quantity of the Stutzer's reagent containing about 0.5 gram of the hydroxid; stir thoroughly, filter when cold, wash with cold water, and, without removing the precipitate from the filter, determine the nitrogen according to **I**, **18**, **21** or **23**, adding sufficient potassium sulphid solution to completely precipitate all of the copper and mercury. The filter paper used must be practically free from nitrogen. If the material (such as seeds, seed residue, or oil cake) is rich in alkaline phosphates, add, to decompose the alkaline phosphates, 1-2 cc. of a concentrated potash or soda alum solution, free from ammonia, then the copper hydroxid, and mix well by stirring. If this is not done, copper phosphate and free alkali may be formed, and the protein-copper precipitate partially dissolved in the alkaline liquid.

8

AMIDO NITROGEN.—OFFICIAL.

Subtract the amount of albuminoid nitrogen from the amount of total nitrogen to obtain the amido nitrogen.

CRUDE FAT OR ETHER EXTRACT.

Direct Method.—Official.

9

REAGENT.

Anhydrous ether.—Wash any of the commercial brands of ether with 2 or 3 successive portions of water, add solid sodium or potassium hydroxid, and let stand until most of the water has been abstracted from the ether. Decant into a dry bottle, add small pieces of carefully cleaned metallic sodium, and let stand until there is no further evolution of hydrogen gas. Keep the ether, thus dehydrated, over metallic sodium in lightly stoppered bottles.

10

DETERMINATION.

Large quantities of soluble carbohydrates may interfere with the complete extraction of the fat. In such cases extract with water before proceeding with the determination. Extract about 2 grams of material, dried as under **2** or **3**, with the anhydrous ether for 16 hours. Dry the extract at the temperature of boiling water for 30 minutes, cool in a desiccator, and weigh; continue, at 30 minutes intervals, this alternate drying and weighing to constant weight. For most feeds a period of 1-1½ hours is required.

11

Indirect Method.—Official.

Determine the moisture, as directed in 2 or 3, then extract the dried substance for 16 hours as directed under 10, dry again and regard the loss of weight as ether extract.

SUCROSE.

OPTICAL METHODS.

12

GENERAL DIRECTIONS FOR RAW SUGARS.—TENTATIVE.

(Rules¹ of the International Commission for Unifying Methods of Sugar Analysis.)

"In general all polarizations are to be made at 20°C."

"The verification of the saccharimeter must also be made at 20°C. For instruments using the Ventzke scale 26 grams of pure dry sucrose, weighed in air with brass weights, dissolved in 100 metric cc. at 20°C. and polarized in a room, the temperature of which is also 20°C., must give a saccharimeter reading of exactly 100.00. The temperature of the sugar solution during polarization must be kept constant at 20°C."

"For countries where the mean temperature is higher than 20°C., saccharimeters may be adjusted at 30°C. or any other suitable temperature, under the conditions specified above, provided that the sugar solution be made up to volume and polarized at this same temperature."

"In effecting the polarization of substances containing sugar employ only half-shade instruments." The saccharimeter used can be either single or double wedge and should be a half-shadow instrument with either double or triple field.

"During the observation keep the apparatus in a fixed position and so far removed from the source of light that the polarizing Nicol is not warmed."

"As sources of light employ lamps which give a strong illumination such as triple gas burner with metallic cylinder, lens and reflector; gas lamps with Auer (Welsbach) burner; electric lamp; petroleum duplex lamp; sodium light." Whenever there is any irregularity in the sources of light such as that due to the convolutions of the filament in the case of electric light or to the meshes of the gauze in the case of the Welsbach light, place a thin ground-glass plate between the source of light and the polariscope so as to render the illumination uniform.

"Before and after each set of observations the chemist must satisfy himself of the correct adjustment of his saccharimeter by means of standardized quartz plates. He must also previously satisfy himself of the accuracy of his weights, polarization flasks, observation tubes and cover-glasses. (Scratched cover-glasses must not be used.) Make several readings and take the mean thereof, but no one reading may be neglected." Such plates are standardized to read to the second decimal point and by their use a quick and at the same time accurate test can be made. In using such plates for testing saccharimeters, it is necessary that the instrument, as well as the plate, be at 20°C. before making a reading. Different points of the scale, preferably 20°, 50°, 80°, and 100°, (sugar scale) should be tested against the plates.

"In making a polarization use the whole normal weight for 100 cc. or a multiple thereof for any corresponding volume."

"As clarifying and decolorizing agents use either basic acetate of lead, alumina cream, or concentrated solution of alum. Boneblack and decolorizing powders are to be excluded." Whenever reducing sugars are determined in the solution for polarizing, use only neutral lead acetate for clarification as basic lead acetate causes precipitation of some of the reducing sugars. In addition to these clarifying agents,

neutral lead acetate and basic lead nitrate (Herles' solution) have been made official by the Association.

"After bringing the solution exactly to the mark at the proper temperature, and after wiping out the neck of the flask with filter paper, pour all of the well-shaken clarified sugar solution on a rapidly acting filter. Reject the first portions of the filtrate, and use the rest, which must be perfectly clear, for polarization." It is advisable to reject the first 20 cc. that run through, then cover the funnel with a watch glass and use the remainder for polarization. In no case should the whole solution or any part be returned to the filter. If cloudy after the 20 cc. have been rejected, begin a new determination.

"Whenever white light is used in polarimetric determinations, the same must be filtered through a solution of potassium dichromate of such a concentration that the percentage content of the solution multiplied by the length of the column of the solution in centimeters is equal to nine." This concentration must be doubled in reading carbohydrate materials of high rotation dispersion, such as commercial glucose, etc.

13

PREPARATION AND USE OF CLARIFYING REAGENTS.—TENTATIVE.

(a) *Basic lead acetate solution*.—Boil 430 grams of neutral lead acetate, 130 grams of litharge, and 1 liter of water for 30 minutes. Allow the mixture to cool and settle and dilute the supernatant liquid to a specific gravity of 1.25 with recently boiled water. Solid basic lead acetate may be substituted for the normal salt and litharge in the preparation of the solution.

(b) *Alumina cream*.—Prepare a cold saturated solution of alum in water. Add ammonium hydroxid with constant stirring until the solution is alkaline to litmus, allow the precipitate to settle and wash by decantation with water until the wash water gives only a slight test for sulphates with barium chlorid solution. Pour off the excess of water and store the residual cream in a stoppered bottle.

(c) *Dry basic lead acetate (Horne method)*.—This clarifying agent is obtained as a dry powdered salt and should contain 72.8% of lead, which corresponds to a composition of $3\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2 \cdot 2\text{PbO}$. Dissolve the normal or half-normal weight of the sugar solution in a flask with water and complete the volume. Add a small quantity of the dry salt and shake, then add more and shake again, repeating until completely precipitated but avoiding any excess. Of this salt 0.1346 gram is equivalent to 1 cc. of the basic lead acetate solution, described under (a). When molasses or any other substance producing a heavy precipitate is being clarified, some dry, coarse sand should be added to break up the balls of basic lead acetate and the precipitate. (This method is to have equal weight with the use of a solution of basic lead acetate in clarifying cane, sorghum, and beet products.)

(d) *Neutral lead acetate*.—Prepare a saturated solution of neutral lead acetate and add it to the sugar solution before completing to volume. Its use is imperative when determining the reducing sugars in the solution used for polarization.

(e) *Basic lead nitrate (Herles' solution)*.—(1) Dissolve 250 grams of lead nitrate in water and make up to 500 cc. (2) Dissolve 25 grams of sodium hydroxid in water and make up to 500 cc.

Add equal amounts of (1) and (2) to the sugar solution, shake, and add more if complete precipitation has not occurred, but avoid any excess. Then complete the volume with water. When this solution is used for clarification, the factor in the Clerget determination becomes 143.5 instead of 142.66.

DETERMINATION OF SUCROSE IN THE ABSENCE OF RAFFINOSE.

(In the presence of much levulose, as in honeys and fruits products, the optical method for sucrose gives too high a result.)

14 *By Polarization Before and After Inversion with Hydrochloric Acid.—Official.*

Dissolve the normal weight (26 grams) of the substance in water, add basic lead acetate carefully, avoiding any excess, then 1-2 cc. of alumina cream, shake, and dilute to 100 metric cc., filter, rejecting the first 20 cc. of the filtrate, cover the filter with a watch glass and, when sufficient filtrate is collected, polarize in a 200 mm. tube. The reading so obtained is the direct reading (P of formula given below) or polarization before inversion. For the invert reading, remove the lead from the solution either (1) by adding anhydrous potassium oxalate, a little at a time, to the remaining solution, avoiding an excess and removing the precipitated lead by filtration; or, (2) by adding anhydrous sodium carbonate under the same conditions. Introduce 50 cc. of the lead-free filtrate into a 100 cc. flask (if sodium carbonate was used for removing the lead, neutralize carefully the excess of sodium carbonate with a few drops of dilute hydrochloric acid) and add 25 cc. of water. Then add, little by little, while rotating the flask, 5 cc. of hydrochloric acid, (sp.gr. 1.20). Heat the flask after mixing, in a water bath kept at 70°C. The temperature of the solution in the flask should reach 67°-69°C. in 2½-3 minutes. Maintain a temperature of as nearly 69°C. as possible for 7-7½ minutes, making the total time of heating 10 minutes. Remove the flask and cool the contents rapidly to 20°C. and dilute to 100 cc. Polarize this solution in a tube provided with a lateral branch and a water jacket, maintaining a temperature of 20°C. This reading must be multiplied by 2 to obtain the invert reading. If it is necessary to work at a temperature other than 20°C., which is allowable within narrow limits, the volumes must be completed and both direct and invert polarizations must be made at exactly the same temperature.

The inversion may also be accomplished as follows: (1) To 50 cc. of the clarified solution, freed from lead, add 5 cc. of hydrochloric acid (sp. gr. 1.20) and set aside for 24 hours at a temperature not below 20°C.; or, (2) If the temperature be above 25°C. set aside for 10 hours. Make up to 100 cc. at 20°C. and polarize as directed above.

Calculate sucrose by one of the following formulas:

For substances in which the invert solution contains more than 12 grams of invert sugar per 100 cc.—The following formula is to be used when substances like raw sugars are polarized:

$$S = \frac{100 (P - I)}{142.66 - \frac{T}{2}} \text{ in which}$$

S = per cent of sucrose;

P = direct reading normal solution;

I = invert reading normal solution;

T = temperature at which readings are made.

For substances in which the concentration of the invert solution is less than 12 grams per 100 cc.—The following formula, which takes into account the concentration of the sugar in solution, should be used in all other cases.

$$S = \frac{100 (P - I)}{142.66 - \frac{T}{2} - 0.0065 \left[142.66 - \frac{T}{2} - (P - I) \right]} \text{ in which}$$

S = per cent of sucrose;

P = direct reading normal solution;

I = invert reading normal solution;

T = temperature.

By Polarization Before and After Inversion with Invertase.—Tentative.

15

REAGENT.

Invertase solution (Hudson Method.²)—Mix 1 kilo of pressed baker's or brewer's yeast with 1 liter of tap water and 50 cc. of toluene and keep at room temperature 2-3 days to allow autolysis to proceed to the stage of maximum inverting activity. Then add neutral lead acetate in slight excess, filter, precipitate the lead in the filtrate with hydrogen sulphid, filter again and then dialyze the filtrate thoroughly in a collodion sac. Preserve in an ice box the dialyzed solution with the addition of a little toluene to prevent the growth of micro-organisms. Note the optical activity of the invertase solution and correct the invert reading according to the amount of the solution used.

16

DETERMINATION.

Dissolve the normal weight (26 grams) of the substance in water, clarify, make up to volume, and take the direct polarization (P) as directed under 14. If lead has been used as a clarifying agent, remove the excess of lead from the filtrate, with anhydrous sodium carbonate or potassium oxalate, and filter. To 50 cc. of the filtrate in a 100 cc. flask add acetic acid, drop by drop, until the reaction is acid to litmus, add 10 cc. of the invertase solution, fill the flask with water nearly to 100 cc. and let stand in a warm place (about 40°C.) overnight. Cool and make up to 100 cc. at 20°C. Polarize at 20°C. in a 200 mm. tube. Allow the solution to remain in the tube for an hour and repeat the polarization. If there is no change from the previous reading, the inversion is complete, whereupon the reading and temperature of the solution are carefully noted. Correct the reading for the optical activity of the invertase solution and then multiply by 2. Calculate the percentage of sucrose by the following formula:

$$S = \frac{100 (P - I)}{142 - \frac{T}{2} - 0.0065 \left[142 - \frac{T}{2} - (P - I) \right]} \text{ in which}$$

S = per cent of sucrose;

P = direct reading;

I = invert reading;

T = temperature at which invert reading is made.

17

DETERMINATION OF SUCROSE AND RAFFINOSE.—OFFICIAL.

(Of value chiefly in the analysis of beet products.)

If the direct reading is more than 1° higher than the per cent of sucrose as calculated by the formula given under 14, raffinose is probably present. Calculate sucrose and raffinose by the following formula of Herzfeld:

$$S = \frac{0.5124 P - I}{0.839}; \quad R = \frac{P - S}{1.852} \text{ in which}$$

P = direct reading normal solution;

I = invert reading normal solution;

S = per cent of sucrose;

R = per cent of anhydrous raffinose.

The above formula assumes that the polarizations are made at exactly 20°C. If the temperature (T) is other than 20°C., the following formula should be used:

$$S = \frac{P(0.4724 + 0.002 T) - I}{0.899 - 0.003 T}$$

Having calculated S, then $R = \frac{P - S}{1.852}$

CHEMICAL METHODS.

18 DETERMINATION OF SUCROSE FROM REDUCING SUGARS BEFORE AND AFTER INVERSION.—TENTATIVE.

Determine the reducing sugars (clarification having been effected with *neutral* lead acetate, never with basic lead acetate), as directed under 25, and calculate to invert sugar from 27. Invert the solution as directed under 14 or 16, exactly neutralize the acid, and again determine the reducing sugars, but calculate them to invert sugar from the same table as referred to above, using the invert sugar column alone. Deduct the percentage of invert sugar obtained before inversion from that obtained after inversion, and multiply the difference by 0.95, the result being the per cent of sucrose. The solutions should be diluted in both determinations so that not more than 245 mg. of invert sugar are present in the amount taken for reduction. It is important that all lead be removed from the solution with potassium oxalate before reduction.

REDUCING SUGARS.

INVERT SUGAR.

Approximate Volumetric Method for Rapid Work.—Tentative.

19

REAGENT.

Soxhlet's Modification of Fehling's Solution.—Prepare by mixing, immediately before use, equal volumes of (a) and (b).

(a) *Copper sulphate solution.*—Dissolve 34.639 grams of copper sulphate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) in water, dilute to 500 cc. and filter through prepared asbestos.

(b) *Alkaline tartrate solution.*—Dissolve 173 grams of Rochelle salts and 50 grams of sodium hydroxid in water, dilute to 500 cc., allow to stand for 2 days and filter through prepared asbestos.

20

STANDARDIZATION OF COPPER SOLUTION.

Since the factor of calculation varies with the minute details of manipulation, every operator must determine a factor for himself, using a known solution of the pure sugar that he desires to determine, and keeping the conditions the same as those used for the determination.

Standardize the solution for invert sugar in the following manner:

Dissolve 4.75 grams of pure sucrose in 75 cc. of water, add 5 cc. of hydrochloric acid (sp. gr. 1.20) and invert as directed under 14. Neutralize the acid with sodium hydroxid solution and dilute to 1 liter. Ten cc. of this solution contain 0.050 gram of invert sugar, which should reduce 10 cc. of the reagent. The strength of the copper solution should never be taken as a constant, but should be checked against the sugar.

21

DETERMINATION.

Place 10 cc. of the reagent in a large test tube and add 10 cc. of water. Heat to boiling, and add gradually small portions of the solution of the material to be tested until the copper has been completely reduced, boiling after each addition to complete the reaction. Two minutes' boiling is required for complete reduction when the full amount of sugar solution has been added in one portion. When the end is nearly reached and the amount of sugar solution to be added can no longer be judged by the color of the solution, remove a small portion of the liquid and filter rapidly into a small porcelain crucible or on a test plate; acidify with dilute acetic acid, and test for copper with dilute potassium ferrocyanid solution. The sugar solution should be of such strength as will give a burette reading of 15-20 cc., and the number of successive additions should be as small as possible.

Soxhlet Volumetric Method.—Tentative.

22

REAGENT.

The reagent used is described under 19.

23

DETERMINATION.

Make a preliminary titration to determine the approximate percentage of reducing sugar in the material under examination. Prepare a solution which contains approximately 1% of reducing sugar. Place in a beaker 100 cc. of the reagent and approximately the amount of the sugar solution for its complete reduction. Boil for 2 minutes. Filter through a folded filter and test a portion of the filtrate for copper by use of dilute acetic acid and dilute potassium ferrocyanid solution. Repeat, varying the volume of sugar solution, until 2 successive amounts are found which differ by 0.1 cc., one giving complete reduction and the other leaving a small amount of copper in solution. The mean of these 2 readings is taken as the volume of the solution required for the complete precipitation of 100 cc. of the reagent.

Under these conditions 100 cc. of the reagent require 0.494 gram of invert sugar for complete reduction. Calculate the percentage by the following formula:

V = the volume of the sugar solution required for the complete reduction of 100 cc. of the reagent;

W = the weight of the sample in 1 cc. of the sugar solution;

$$\frac{100 \times 0.494}{VW} = \text{per cent of invert sugar.}$$

GRAVIMETRIC METHODS

Munson and Walker General Method.³—Tentative.

24

REAGENTS.

(a) *Asbestos*.—Digest the asbestos, which should be the amphibole variety, with dilute hydrochloric acid (1 to 3) for 2-3 days. Wash free from acid, digest for a

similar period with 10% sodium hydroxid solution, and then treat for a few hours with hot alkaline tartrate solution (old alkaline tartrate solutions that have stood for some time may be used for this purpose) of the strength employed in sugar determinations. Then wash the asbestos free from alkali, digest for several hours with dilute nitric acid (1 to 3) and, after washing free from acid, shake with water into a fine pulp. In preparing the Gooch crucible, make a film of asbestos $\frac{1}{4}$ inch thick and wash thoroughly with water to remove fine particles of asbestos. If the precipitated cuprous oxid is to be weighed as such, wash the crucible with 10 cc. of alcohol, then with 10 cc. of ether, dry for 30 minutes at 100°C., cool in a desiccator and weigh.

(b) The solution used is described under 19.

25

PRECIPITATION OF CUPROUS OXID.

Transfer 25 cc. each of the copper sulphate and alkaline tartrate solutions to a 400 cc. beaker of alkali-resisting glass and add 50 cc. of reducing sugar solution, or, if a smaller volume of sugar solution is used, add water to make the final volume 100 cc. Heat the beaker upon an asbestos gauze over a Bunsen burner, regulate the flame so that boiling begins in 4 minutes, and continue the boiling for exactly 2 minutes. (It is important that these directions be strictly observed and, in order to regulate the burner for this purpose, it is advisable to make preliminary tests, using 50 cc. of the reagent and 50 cc. of water before proceeding with the actual determination.) Keep the beaker covered with a watch glass during the heating. Filter the cuprous oxid at once on an asbestos mat in a porcelain Gooch crucible, using suction. Wash the cuprous oxid thoroughly with water at a temperature of about 60°C., and either weigh directly as cuprous oxid as in 26, or, determine the amount of reduced copper by one of the methods under 29-34, respectively. Conduct a blank determination, using 50 cc. of the reagent and 50 cc. of water, and, if the weight of cuprous oxid obtained exceeds 0.5 mg., correct the result of the reducing sugar determination accordingly. The alkaline tartrate solution deteriorates on standing and the amount of cuprous oxid obtained in the blank increases.

DETERMINATION OF REDUCED COPPER.

26

I. *Direct Weighing of Cuprous Oxid.—Tentative.*

Prepare a Gooch as directed under 24 (a).

Collect the precipitated cuprous oxid on the mat, as directed under 25, wash thoroughly with hot water, then with 10 cc. of alcohol, and finally with 10 cc. of ether. Dry the precipitate for 30 minutes in a water oven at the temperature of boiling water; cool and weigh. Calculate the weight of metallic copper. Obtain from 27 the weight of invert sugar equivalent to the weight of copper found.

This method should be used only for determinations in pure sugar solutions. In all other products the copper of the cuprous oxid should be determined by one of the following methods, since the cuprous oxid is very apt to be contaminated with foreign matter.

The number of milligrams of copper reduced by a given amount of reducing sugar differs when sucrose is present and when it is absent. In the tables the absence of sucrose is assumed except in the two columns under invert sugar, where one for mixtures of invert sugar and sucrose containing 0.4 gram of total sugar in 50 cc. of solution, and one for invert sugar and sucrose when the 50 cc. of solution contains 2 grams of total sugar are given, in addition to the column for invert sugar alone.

27

TABLE 1.—MUNSON AND WALKER'S TABLE.

For calculating dextrose, invert sugar alone, invert sugar in the presence of sucrose (0.4 gram and 2 grams total sugar), lactose (two forms), and maltose (anhydrous and crystallized).

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTROSE (D-GLUCOSE)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram total sugar	2 grams total sugar	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	
10	8.9	4.0	4.5	1.6	3.8	4.0	5.9	6.2	10
11	9.8	4.5	5.0	2.1	4.5	4.7	6.7	7.0	11
12	10.7	4.9	5.4	2.5	5.1	5.4	7.5	7.9	12
13	11.5	5.3	5.8	3.0	5.8	6.1	8.3	8.7	13
14	12.4	5.7	6.3	3.4	6.4	6.8	9.1	9.5	14
15	13.3	6.2	6.7	3.9	7.1	7.5	9.9	10.4	15
16	14.2	6.6	7.2	4.3	7.8	8.2	10.6	11.2	16
17	15.1	7.0	7.6	4.8	8.4	8.9	11.4	12.0	17
18	16.0	7.5	8.1	5.2	9.1	9.5	12.2	12.9	18
19	16.9	7.9	8.5	5.7	9.7	10.2	13.0	13.7	19
20	17.8	8.3	8.9	6.1	10.4	10.9	13.8	14.6	20
21	18.7	8.7	9.4	6.6	11.0	11.6	14.6	15.4	21
22	19.5	9.2	9.8	7.0	11.7	12.3	15.4	16.2	22
23	20.4	9.6	10.3	7.5	12.3	13.0	16.2	17.1	23
24	21.3	10.0	10.7	7.9	13.0	13.7	17.0	17.9	24
25	22.2	10.5	11.2	8.4	13.7	14.4	17.8	18.7	25
26	23.1	10.9	11.6	8.8	14.3	15.1	18.6	19.6	26
27	24.0	11.3	12.0	9.3	15.0	15.8	19.4	20.4	27
28	24.9	11.8	12.5	9.7	15.6	16.5	20.2	21.2	28
29	25.8	12.2	12.9	10.2	16.3	17.1	21.0	22.1	29
30	26.6	12.6	13.4	10.7	4.3	16.9	17.8	21.8	22.9	30
31	27.5	13.1	13.8	11.1	4.7	17.6	18.5	22.6	23.7	31
32	28.4	13.5	14.3	11.6	5.2	18.3	19.2	23.3	24.6	32
33	29.3	13.9	14.7	12.0	5.6	18.9	19.9	24.1	25.4	33
34	30.2	14.3	15.2	12.5	6.1	19.6	20.6	24.9	26.2	34
35	31.1	14.8	15.6	12.9	6.5	20.2	21.3	25.7	27.1	35
36	32.0	15.2	16.1	13.4	7.0	20.9	22.0	26.5	27.9	36
37	32.9	15.6	16.5	13.8	7.4	21.5	22.7	27.3	28.7	37
38	33.8	16.1	16.9	14.3	7.9	22.2	23.4	28.1	29.6	38
39	34.6	16.5	17.4	14.7	8.4	22.8	24.1	28.9	30.4	39
40	35.5	16.9	17.8	15.2	8.8	23.5	24.8	29.7	31.3	40
41	36.4	17.4	18.3	15.6	9.3	24.2	25.4	30.5	32.1	41
42	37.3	17.8	18.7	16.1	9.7	24.8	26.1	31.3	32.9	42
43	38.2	18.2	19.2	16.6	10.2	25.5	26.8	32.1	33.8	43
44	39.1	18.7	19.6	17.0	10.7	26.1	27.5	32.9	34.6	44
45	40.0	19.1	20.1	17.5	11.1	26.8	28.2	33.7	35.4	45
46	40.9	19.6	20.5	17.9	11.6	27.4	28.9	34.4	36.3	46
47	41.7	20.0	21.0	18.4	12.0	28.1	29.6	35.2	37.1	47
48	42.6	20.4	21.4	18.8	12.5	28.7	30.3	36.0	37.9	48
49	43.5	20.9	21.9	19.3	12.9	29.4	31.0	36.8	38.8	49
50	44.4	21.3	22.3	19.7	13.4	30.1	31.7	37.6	39.6	50
51	45.3	21.7	22.8	20.2	13.9	30.7	32.4	38.4	40.4	51
52	46.2	22.2	23.2	20.7	14.3	31.4	33.0	39.2	41.3	52
53	47.1	22.6	23.7	21.1	14.8	32.1	33.7	40.0	42.1	53
54	48.0	23.0	24.1	21.6	15.2	32.7	34.4	40.8	42.9	54
55	48.9	23.5	24.6	22.0	15.7	33.4	35.1	41.6	43.8	55
56	49.7	23.9	25.0	22.5	16.2	34.0	35.8	42.4	44.6	56
57	50.6	24.3	25.5	22.9	16.6	34.7	36.5	43.2	45.4	57
58	51.5	24.8	25.9	23.4	17.1	35.4	37.2	44.0	46.3	58
59	52.4	25.2	26.4	23.9	17.5	36.0	37.9	44.8	47.1	59
60	53.3	25.6	26.8	24.3	18.0	36.7	38.6	45.6	48.0	60
61	54.2	26.1	27.3	24.8	18.5	37.3	39.3	46.3	48.8	61
62	55.1	26.5	27.7	25.2	18.9	38.0	40.0	47.1	49.6	62
63	56.0	27.0	28.2	25.7	19.4	38.6	40.7	47.9	50.5	63
64	56.8	27.4	28.6	26.2	19.8	39.3	41.4	48.7	51.3	64

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTRINE (d-glucose)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram total sugar	2 grams total sugar	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	
65	57.7	27.8	29.1	26.6	20.3	40.0	42.1	49.5	52.1	65
66	53.6	28.3	29.5	27.1	20.8	40.6	42.8	50.3	53.0	66
67	59.5	28.7	30.0	27.5	21.2	41.3	43.5	51.1	53.8	67
68	60.4	29.2	30.4	28.0	21.7	41.9	44.2	51.9	54.6	68
69	61.3	29.6	30.9	28.5	22.2	42.6	44.8	52.7	55.5	69
70	62.2	30.0	31.3	28.9	22.6	43.3	45.5	53.5	56.3	70
71	63.1	30.5	31.8	29.4	23.1	43.9	46.2	54.3	57.1	71
72	64.0	30.9	32.3	29.8	23.5	44.6	46.9	55.1	58.0	72
73	64.8	31.4	32.7	30.3	24.0	45.2	47.6	55.9	58.8	73
74	65.7	31.8	33.2	30.8	24.5	45.9	48.3	56.7	59.6	74
75	66.6	32.2	33.6	31.2	24.9	46.6	49.0	57.5	60.5	75
76	67.5	32.7	34.1	31.7	25.4	47.2	49.7	58.2	61.3	76
77	68.4	33.1	34.5	32.1	25.9	47.9	50.4	59.0	62.1	77
78	69.3	33.6	35.0	32.6	26.3	48.5	51.1	59.8	63.0	78
79	70.2	34.0	35.4	33.1	26.8	49.2	51.8	60.6	63.8	79
80	71.1	34.4	35.9	33.5	27.3	49.9	52.5	61.4	64.6	80
81	71.9	34.9	36.3	34.0	27.7	50.5	53.2	62.2	65.5	81
82	72.8	35.3	36.8	34.5	28.2	51.2	53.9	63.0	66.3	82
83	73.7	35.8	37.3	34.9	28.6	51.8	54.6	63.8	67.1	83
84	74.6	36.2	37.7	35.4	29.1	52.5	55.3	64.6	68.0	84
85	75.5	36.7	38.2	35.8	29.6	53.1	56.0	65.4	68.8	85
86	76.4	37.1	38.6	36.3	30.0	53.8	56.6	66.2	69.7	86
87	77.3	37.5	39.1	36.8	30.5	54.5	57.3	67.0	70.5	87
88	78.2	38.0	39.5	37.2	31.0	55.1	58.0	67.8	71.3	88
89	79.1	38.4	40.0	37.7	31.4	55.8	58.7	68.5	72.2	89
90	79.9	38.9	40.4	38.2	31.9	56.4	59.4	69.3	73.0	90
91	80.8	39.3	40.9	38.6	32.4	57.1	60.1	70.1	73.8	91
92	81.7	39.8	41.4	39.1	32.8	57.8	60.8	70.9	74.7	92
93	82.6	40.2	41.8	39.6	33.3	58.4	61.5	71.7	75.5	93
94	83.5	40.6	42.3	40.0	33.8	59.1	62.2	72.5	76.3	94
95	84.4	41.1	42.7	40.5	34.2	59.7	62.9	73.3	77.2	95
96	85.3	41.5	43.2	41.0	34.7	60.4	63.6	74.1	78.0	96
97	86.2	42.0	43.7	41.4	35.2	61.1	64.3	74.9	78.8	97
98	87.1	42.4	44.1	41.9	35.6	61.7	65.0	75.7	79.7	98
99	87.9	42.9	44.6	42.4	36.1	62.4	65.7	76.5	80.5	99
100	88.8	43.3	45.0	42.8	36.6	63.0	66.4	77.3	81.3	100
101	89.7	43.8	45.5	43.3	37.0	63.7	67.1	78.1	82.2	101
102	90.6	44.2	46.0	43.8	37.5	64.4	67.8	78.8	83.0	102
103	91.5	44.7	46.4	44.2	38.0	65.0	68.5	79.6	83.8	103
104	92.4	45.1	46.9	44.7	38.5	65.7	69.1	80.4	84.7	104
105	93.3	45.5	47.3	45.2	38.9	66.4	69.8	81.2	85.5	105
106	94.2	46.0	47.8	45.6	39.4	67.0	70.5	82.0	86.3	106
107	95.0	46.4	48.3	46.1	39.9	67.7	71.2	82.8	87.2	107
108	95.9	46.9	48.7	46.6	40.3	68.3	71.9	83.6	88.0	108
109	96.8	47.3	49.2	47.0	40.8	69.0	72.6	84.4	88.8	109
110	97.7	47.8	49.6	47.5	41.3	69.7	73.3	85.2	89.7	110
111	98.6	48.2	50.1	48.0	41.7	70.3	74.0	86.0	90.5	111
112	99.5	48.7	50.6	48.4	42.2	71.0	74.7	86.8	91.3	112
113	100.4	49.1	51.0	48.9	42.7	71.6	75.4	87.6	92.2	113
114	101.3	49.6	51.5	49.4	43.2	72.3	76.1	88.4	93.0	114
115	102.2	50.0	51.9	49.8	43.6	73.0	76.8	89.2	93.9	115
116	103.0	50.5	52.4	50.3	44.1	73.6	77.5	90.0	94.7	116
117	103.9	50.9	52.9	50.8	44.6	74.3	78.2	90.7	95.5	117
118	104.8	51.4	53.3	51.2	45.0	75.0	78.9	91.5	96.4	118
119	105.7	51.8	53.8	51.7	45.5	75.6	79.6	92.3	97.2	119

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTRIOSE (d-GLUCOSE)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram total sugar	2 grams total sugar	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	
120	106.6	52.3	54.3	52.2	46.0	76.3	80.3	93.1	98.0	120
121	107.5	52.7	54.7	52.7	46.5	76.9	81.0	93.9	98.9	121
122	108.4	53.2	55.2	53.1	46.9	77.6	81.7	94.7	99.7	122
123	109.3	53.6	55.7	53.6	47.4	78.3	82.4	95.5	100.5	123
124	110.1	54.1	56.1	54.1	47.9	78.9	83.1	96.3	101.4	124
125	111.0	54.5	56.6	54.5	48.3	79.6	83.8	97.1	102.2	125
126	111.9	55.0	57.0	55.0	48.8	80.3	84.5	97.9	103.0	126
127	112.8	55.4	57.5	55.5	49.3	80.9	85.2	98.7	103.9	127
128	113.7	55.9	58.0	55.9	49.8	81.6	85.9	99.4	104.7	128
129	114.6	56.3	58.4	56.4	50.2	82.2	86.6	100.2	105.5	129
130	115.5	56.8	58.9	56.9	50.7	82.9	87.3	101.0	106.4	130
131	116.4	57.2	59.4	57.4	51.2	83.6	88.0	101.8	107.2	131
132	117.3	57.7	59.8	57.8	51.7	84.2	88.7	102.6	108.0	132
133	118.1	58.1	60.3	58.3	52.1	84.9	89.4	103.4	108.9	133
134	119.0	58.6	60.8	58.8	52.6	85.5	90.1	104.2	109.7	134
135	119.9	59.0	61.2	59.3	53.1	86.2	90.8	105.0	110.5	135
136	120.8	59.5	61.7	59.7	53.6	86.9	91.5	105.8	111.4	136
137	121.7	60.0	62.2	60.2	54.0	87.5	92.1	106.6	112.2	137
138	122.6	60.4	62.6	60.7	54.5	88.2	92.8	107.4	113.0	138
139	123.5	60.9	63.1	61.2	55.0	88.9	93.5	108.2	113.9	139
140	124.4	61.3	63.6	61.6	55.5	89.5	94.2	109.0	114.7	140
141	125.2	61.8	64.0	62.1	55.9	90.2	94.9	109.8	115.5	141
142	126.1	62.2	64.5	62.6	56.4	90.8	95.6	110.5	116.4	142
143	127.0	62.7	65.0	63.1	56.9	91.5	96.3	111.3	117.2	143
144	127.9	63.1	65.4	63.5	57.4	92.2	97.0	112.1	118.0	144
145	128.8	63.6	65.9	64.0	57.8	92.8	97.7	112.9	118.9	145
146	129.7	64.0	66.4	64.5	58.3	93.5	98.4	113.7	119.7	146
147	130.6	64.5	66.9	65.0	58.8	94.2	99.1	114.5	120.5	147
148	131.5	65.0	67.3	65.4	59.3	94.8	99.8	115.3	121.4	148
149	132.4	65.4	67.8	65.9	59.7	95.5	100.5	116.1	122.2	149
150	133.2	65.9	68.3	66.4	60.2	96.1	101.2	116.9	123.0	150
151	134.1	66.3	68.7	66.9	60.7	96.8	101.9	117.7	123.9	151
152	135.0	66.8	69.2	67.3	61.2	97.5	102.6	118.5	124.7	152
153	135.9	67.2	69.7	67.8	61.7	98.1	103.3	119.3	125.5	153
154	136.8	67.7	70.1	68.3	62.1	98.8	104.0	120.0	126.4	154
155	137.7	68.2	70.6	68.8	62.6	99.5	104.7	120.8	127.2	155
156	138.6	68.6	71.1	69.2	63.1	100.1	105.4	121.6	128.0	156
157	139.5	69.1	71.6	69.7	63.6	100.8	106.1	122.4	128.9	157
158	140.3	69.5	72.0	70.2	64.1	101.5	106.8	123.2	129.7	158
159	141.2	70.0	72.5	70.7	64.5	102.1	107.5	124.0	130.5	159
160	142.1	70.4	73.0	71.2	65.0	102.8	108.2	124.8	131.4	160
161	143.0	70.9	73.4	71.6	65.5	103.4	108.9	125.6	132.2	161
162	143.9	71.4	73.9	72.1	66.0	104.1	109.6	126.4	133.0	162
163	144.8	71.8	74.4	72.6	66.5	104.8	110.3	127.2	133.9	163
164	145.7	72.3	74.9	73.1	66.9	105.4	111.0	128.0	134.7	164
165	146.6	72.8	75.3	73.6	67.4	106.1	111.7	128.8	135.5	165
166	147.5	73.2	75.8	74.0	67.9	106.8	112.4	129.6	136.4	166
167	148.3	73.7	76.3	74.5	68.4	107.4	113.1	130.3	137.2	167
168	149.2	74.1	76.8	75.0	68.9	108.1	113.8	131.1	138.0	168
169	150.1	74.6	77.2	75.5	69.3	108.8	114.5	131.9	138.9	169
170	151.0	75.1	77.7	76.0	69.8	109.4	115.2	132.7	139.7	170
171	151.9	75.5	78.2	76.4	70.3	110.1	115.9	133.5	140.5	171
172	152.8	76.0	78.7	76.9	70.8	110.8	116.6	134.3	141.4	172
173	153.7	76.4	79.1	77.4	71.3	111.4	117.3	135.1	142.2	173
174	154.6	76.9	79.6	77.9	71.7	112.1	118.0	135.9	143.0	174

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTROSE (d-GLUCOSE)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram sugar total	2 grams sugar total	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ ·H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ ·H ₂ O	
175	155.5	77.4	80.1	78.4	72.13	112.8	118.7	136.7	143.9	175
176	156.3	77.8	80.6	78.8	72.7	113.4	119.4	137.5	144.7	176
177	157.2	78.3	81.0	79.3	73.2	114.1	120.1	138.3	145.5	177
178	158.1	78.8	81.5	79.8	73.7	114.8	120.8	139.1	146.4	178
179	159.0	79.2	82.0	80.3	74.2	115.4	121.5	139.8	147.2	179
180	159.9	79.7	82.5	80.8	74.6	116.1	122.2	140.6	148.0	180
181	160.8	80.1	82.9	81.3	75.1	116.7	122.9	141.4	148.9	181
182	161.7	80.6	83.4	81.7	75.6	117.4	123.6	142.2	149.7	182
183	162.6	81.1	83.9	82.2	76.1	118.1	124.3	143.0	150.5	183
184	163.4	81.5	84.4	82.7	76.6	118.7	125.0	143.8	151.4	184
185	164.3	82.0	84.9	83.2	77.1	119.4	125.7	144.6	152.2	185
186	165.2	82.5	85.3	83.7	77.6	120.1	126.4	145.4	153.0	186
187	166.1	82.9	85.8	84.2	78.0	120.7	127.1	146.2	153.9	187
188	167.0	83.4	86.3	84.6	78.5	121.4	127.8	147.0	154.7	188
189	167.9	83.9	86.8	85.1	79.0	122.1	128.5	147.8	155.5	189
190	168.8	84.3	87.2	85.6	79.5	122.7	129.2	148.6	156.4	190
191	169.7	84.8	87.7	86.1	80.0	123.4	129.9	149.3	157.2	191
192	170.5	85.3	88.2	86.6	80.5	124.1	130.6	150.1	158.0	192
193	171.4	85.7	88.7	87.1	81.0	124.7	131.3	150.9	158.9	193
194	172.3	86.2	89.2	87.6	81.4	125.4	132.0	151.7	159.7	194
195	173.2	86.7	89.6	88.0	81.9	126.1	132.7	152.5	160.5	195
196	174.1	87.1	90.1	88.5	82.4	126.7	133.4	153.3	161.4	196
197	175.0	87.6	90.6	89.0	82.9	127.4	134.1	154.1	162.2	197
198	175.9	88.1	91.1	89.5	83.4	128.1	134.8	154.9	163.0	198
199	176.8	88.5	91.6	90.0	83.9	128.7	135.5	155.7	163.9	199
200	177.7	89.0	92.0	90.5	84.4	129.4	136.2	156.5	164.7	200
201	178.5	89.5	92.5	91.0	84.8	130.0	136.9	157.3	165.5	201
202	179.4	89.9	93.0	91.4	85.3	130.7	137.6	158.1	166.4	202
203	180.3	90.4	93.5	91.9	85.8	131.4	138.3	158.8	167.2	203
204	181.2	90.9	94.0	92.4	86.3	132.0	139.0	159.6	168.0	204
205	182.1	91.4	94.5	92.9	86.8	132.7	139.7	160.4	168.9	205
206	183.0	91.8	94.9	93.4	87.3	133.4	140.4	161.2	169.7	206
207	183.9	92.3	95.4	93.9	87.8	134.0	141.1	162.0	170.5	207
208	184.8	92.8	95.9	94.4	88.3	134.7	141.8	162.8	171.4	208
209	185.6	93.2	96.4	94.9	88.8	135.4	142.5	163.6	172.2	209
210	186.5	93.7	96.9	95.4	89.2	136.0	143.2	164.4	173.0	210
211	187.4	94.2	97.4	95.8	89.7	136.7	143.9	165.2	173.8	211
212	188.3	94.6	97.8	96.3	90.2	137.4	144.6	166.0	174.7	212
213	189.2	95.1	98.3	96.8	90.7	138.0	145.3	166.8	175.5	213
214	190.1	95.6	98.8	97.3	91.2	138.7	146.0	167.5	176.4	214
215	191.0	96.1	99.3	97.8	91.7	139.4	146.7	168.3	177.2	215
216	191.9	96.5	99.8	98.3	92.2	140.0	147.4	169.1	178.0	216
217	192.8	97.0	100.3	98.8	92.7	140.7	148.1	169.9	178.9	217
218	193.6	97.5	100.8	99.3	93.2	141.4	148.8	170.7	179.7	218
219	194.5	98.0	101.2	99.8	93.7	142.0	149.5	171.5	180.5	219
220	195.4	98.4	101.7	100.3	94.2	142.7	150.2	172.3	181.4	220
221	196.3	98.9	102.2	100.8	94.7	143.4	150.9	173.1	182.2	221
222	197.2	99.4	102.7	101.2	95.1	144.0	151.6	173.9	183.0	222
223	198.1	99.9	103.2	101.7	95.6	144.7	152.3	174.7	183.9	223
224	199.0	100.3	103.7	102.2	96.1	145.4	153.0	175.5	184.7	224
225	199.9	100.8	104.2	102.7	96.6	146.0	153.7	176.2	185.5	225
226	200.7	101.3	104.6	103.2	97.1	146.7	154.4	177.0	186.4	226
227	201.6	101.8	105.1	103.7	97.6	147.4	155.1	177.8	187.2	227
228	202.5	102.3	105.6	104.2	98.1	148.0	155.8	178.6	188.0	228
229	203.4	102.7	106.1	104.7	98.6	148.7	156.5	179.4	188.8	229

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTROSE (d-GLUCOSE)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram total sugar	2 grams total sugar	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ ·H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ ·H ₂ O	
230	204.3	103.2	106.6	105.2	99.1	149.4	157.2	180.2	189.7	230
231	205.2	103.7	107.1	105.7	99.6	150.0	157.9	181.0	190.5	231
232	206.1	104.1	107.6	106.2	100.1	150.7	158.6	181.8	191.3	232
233	207.0	104.6	108.1	106.7	100.6	151.4	159.3	182.6	192.2	233
234	207.9	105.1	108.6	107.2	101.1	152.0	160.0	183.4	193.0	234
235	208.7	105.6	109.1	107.7	101.6	152.7	160.7	184.2	193.8	235
236	209.6	106.0	109.5	108.2	102.1	153.4	161.4	184.9	194.7	236
237	210.5	106.5	110.0	108.7	102.6	154.0	162.1	185.7	195.5	237
238	211.4	107.0	110.5	109.2	103.1	154.7	162.8	186.5	196.3	238
239	212.3	107.5	111.0	109.6	103.5	155.4	163.5	187.3	197.2	239
240	213.2	108.0	111.5	110.1	104.0	156.1	164.3	188.1	198.0	240
241	214.1	108.4	112.0	110.6	104.5	156.7	165.0	188.9	198.8	241
242	215.0	108.9	112.5	111.1	105.0	157.4	165.7	189.7	199.7	242
243	215.8	109.4	113.0	111.6	105.5	158.1	166.4	190.5	200.5	243
244	216.7	109.9	113.5	112.1	106.0	158.7	167.1	191.3	201.3	244
245	217.6	110.4	114.0	112.6	106.5	159.4	167.8	192.1	202.2	245
246	218.5	110.8	114.5	113.1	107.0	160.1	168.5	192.9	203.0	246
247	219.4	111.3	115.0	113.6	107.5	160.7	169.2	193.6	203.8	247
248	220.3	111.8	115.4	114.1	108.0	161.4	169.9	194.4	204.7	248
249	221.2	112.3	115.9	114.6	108.5	162.1	170.6	195.2	205.5	249
250	222.1	112.8	116.4	115.1	109.0	162.7	171.3	196.0	206.3	250
251	223.0	113.2	116.9	115.6	109.5	163.4	172.0	196.8	207.2	251
252	223.8	113.7	117.4	116.1	110.0	164.1	172.7	197.6	208.0	252
253	224.7	114.2	117.9	116.6	110.5	164.7	173.4	198.4	208.8	253
254	225.6	114.7	118.4	117.1	111.0	165.4	174.1	199.2	209.7	254
255	226.5	115.2	118.9	117.6	111.5	166.1	174.8	200.0	210.5	255
256	227.4	115.7	119.4	118.1	112.0	166.8	175.5	200.8	211.3	256
257	228.3	116.1	119.9	118.6	112.5	167.4	176.2	201.6	212.2	257
258	229.2	116.6	120.4	119.1	113.0	168.1	176.9	202.3	213.0	258
259	230.1	117.1	120.9	119.6	113.5	168.8	177.6	203.1	213.8	259
260	231.0	117.6	121.4	120.1	114.0	169.4	178.3	203.9	214.7	260
261	231.8	118.1	121.9	120.6	114.5	170.1	179.0	204.7	215.5	261
262	232.7	118.6	122.4	121.1	115.0	170.8	179.8	205.5	216.3	262
263	233.6	119.0	122.9	121.6	115.5	171.4	180.5	206.3	217.2	263
264	234.5	119.5	123.4	122.1	116.0	172.1	181.2	207.1	218.0	264
265	235.4	120.0	123.9	122.6	116.5	172.8	181.9	207.9	218.8	265
266	236.3	120.5	124.4	123.1	117.0	173.5	182.6	208.7	219.7	266
267	237.2	121.0	124.9	123.6	117.5	174.1	183.3	209.5	220.5	267
268	238.1	121.5	125.4	124.1	118.0	174.8	184.0	210.3	221.3	268
269	238.9	122.0	125.9	124.6	118.5	175.5	184.7	211.0	222.1	269
270	239.8	122.5	126.4	125.1	119.0	176.1	185.4	211.8	223.0	270
271	240.7	122.9	126.9	125.6	119.5	176.8	186.1	212.6	223.8	271
272	241.6	123.4	127.4	126.2	120.0	177.5	186.8	213.4	224.6	272
273	242.5	123.9	127.9	126.7	120.6	178.1	187.5	214.2	225.5	273
274	243.4	124.4	128.4	127.2	121.1	178.8	188.2	215.0	226.3	274
275	244.3	124.9	128.9	127.7	121.6	179.5	188.9	215.8	227.1	275
276	245.2	125.4	129.4	128.2	122.1	180.2	189.6	216.6	228.0	276
277	246.1	125.9	129.9	128.7	122.6	180.8	190.3	217.4	228.8	277
278	246.9	126.4	130.4	129.2	123.1	181.5	191.0	218.2	229.6	278
279	247.8	126.9	130.9	129.7	123.6	182.2	191.7	218.9	230.5	279
280	248.7	127.3	131.4	130.2	124.1	182.8	192.4	219.7	231.3	280
281	249.6	127.8	131.9	130.7	124.6	183.5	193.1	220.5	232.1	281
282	250.5	128.3	132.4	131.2	125.1	184.2	193.9	221.3	233.0	282
283	251.4	128.8	132.9	131.7	125.6	184.8	194.6	222.1	233.8	283
284	252.3	129.3	133.4	132.2	126.1	185.5	195.3	222.9	234.6	284

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTROSE (d-GLUCOSE)	INVERT SUGAR	INVERT SUGAR AND SUCR SE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram total sugar	2 grams total sugar	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	
285	253.2	129.8	133.9	132.7	126.6	186.2	196.0	223.7	235.5	285
286	254.0	130.3	134.4	133.2	127.1	186.9	196.7	224.5	236.3	286
287	254.9	130.8	134.9	133.7	127.6	187.5	197.4	225.3	237.1	287
288	255.8	131.3	135.4	134.3	128.1	188.2	198.1	226.1	238.0	288
289	256.7	131.8	135.9	134.8	128.6	188.9	198.8	226.9	238.8	289
290	257.6	132.3	136.4	135.3	129.2	189.5	199.5	227.6	239.6	290
291	258.5	132.7	136.9	135.8	129.7	190.2	200.2	228.4	240.5	291
292	259.4	133.2	137.4	136.3	130.2	190.9	200.9	229.2	241.3	292
293	260.3	133.7	137.9	136.8	130.7	191.5	201.6	230.0	242.1	293
294	261.2	134.2	138.4	137.3	131.2	192.2	202.3	230.8	242.9	294
295	262.0	134.7	138.9	137.8	131.7	192.9	203.0	231.6	243.8	295
296	262.9	135.2	139.4	138.3	132.2	193.6	203.7	232.4	244.6	296
297	263.8	135.7	140.0	138.8	132.7	194.2	204.4	233.2	245.4	297
298	264.7	136.2	140.5	139.4	133.2	194.9	205.1	234.0	246.3	298
299	265.6	136.7	141.0	139.9	133.7	195.6	205.8	234.8	247.1	299
300	266.5	137.2	141.5	140.4	134.2	196.2	206.6	235.5	247.9	300
301	267.4	137.7	142.0	140.9	134.8	196.9	207.3	236.3	248.8	301
302	268.3	138.2	142.5	141.4	135.3	197.6	208.0	237.1	249.6	302
303	269.1	138.7	143.0	141.9	135.8	198.3	208.7	237.9	250.4	303
304	270.0	139.2	143.5	142.4	136.3	198.9	209.4	238.7	251.3	304
305	270.9	139.7	144.0	142.9	136.8	199.6	210.1	239.5	252.1	305
306	271.8	140.2	144.5	143.4	137.3	200.3	210.8	240.3	252.9	306
307	272.7	140.7	145.0	144.0	137.8	201.0	211.5	241.1	253.8	307
308	273.6	141.2	145.5	144.5	138.3	201.6	212.2	241.9	254.6	308
309	274.5	141.7	146.1	145.0	138.8	202.3	212.9	242.7	255.4	309
310	275.4	142.2	146.6	145.5	139.4	203.0	213.7	243.5	256.3	310
311	276.3	142.7	147.1	146.0	139.9	203.6	214.4	244.2	257.1	311
312	277.1	143.2	147.6	146.5	140.4	204.3	215.1	245.0	257.9	312
313	278.0	143.7	148.1	147.0	140.9	205.0	215.8	245.8	258.8	313
314	278.9	144.2	148.6	147.6	141.4	205.7	216.5	246.6	259.6	314
315	279.8	144.7	149.1	148.1	141.9	206.3	217.2	247.4	260.4	315
316	280.7	145.2	149.6	148.6	142.4	207.0	217.9	248.2	261.2	316
317	281.6	145.7	150.1	149.1	143.0	207.7	218.6	249.0	262.1	317
318	282.5	146.2	150.7	149.6	143.5	208.4	219.3	249.8	262.9	318
319	283.4	146.7	151.2	150.1	144.0	209.0	220.0	250.6	263.7	319
320	284.2	147.2	151.7	150.7	144.5	209.7	220.7	251.3	264.6	320
321	285.1	147.7	152.2	151.2	145.0	210.4	221.4	252.1	265.4	321
322	286.0	148.2	152.7	151.7	145.5	211.0	222.2	252.9	266.2	322
323	286.9	148.7	153.2	152.2	146.0	211.7	222.9	253.7	267.1	323
324	287.8	149.2	153.7	152.7	146.6	212.4	223.6	254.5	267.9	324
325	288.7	149.7	154.3	153.2	147.1	213.1	224.3	255.3	268.7	325
326	289.6	150.2	154.8	153.8	147.6	213.7	225.0	256.1	269.6	326
327	290.5	150.7	155.3	154.3	148.1	214.4	225.7	256.9	270.4	327
328	291.4	151.2	155.8	154.8	148.6	215.1	226.4	257.7	271.2	328
329	292.2	151.7	156.3	155.3	149.1	215.8	227.1	258.5	272.1	329
330	293.1	152.2	156.8	155.8	149.7	216.4	227.8	259.3	272.9	330
331	294.0	152.7	157.3	156.4	150.2	217.1	228.5	260.0	273.7	331
332	294.9	153.2	157.9	156.9	150.7	217.8	229.2	260.8	274.6	332
333	295.8	153.7	158.4	157.4	151.2	218.4	230.0	261.6	275.4	333
334	296.7	154.2	158.9	157.9	151.7	219.1	230.7	262.4	276.2	334
335	297.6	154.7	159.4	158.4	152.3	219.8	231.4	263.2	277.0	335
336	298.5	155.2	159.9	159.0	152.8	220.5	232.1	264.0	277.9	336
337	299.3	155.8	160.5	159.5	153.3	221.1	232.8	264.8	278.7	337
338	300.2	156.3	161.0	160.0	153.8	221.8	233.5	265.6	279.5	338
339	301.1	156.8	161.5	160.5	154.3	222.5	234.2	266.4	280.4	339

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTROSE (d-glucose)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram sugar	2 grams sugar	C ₁₂ H ₂₂ O ₁₁ ·	C ₁₂ H ₂₂ O ₁₁ H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ H ₂ O	
340	302.0	157.3	162.0	161.0	154.8	223.2	234.9	267.1	281.2	340
341	302.9	157.8	162.5	161.6	155.4	223.8	235.6	267.9	282.0	341
342	303.8	158.3	163.1	162.1	155.9	224.5	236.3	268.7	282.9	342
343	304.7	158.8	163.6	162.6	156.4	225.2	237.0	269.5	283.7	343
344	305.6	159.3	164.1	163.1	156.9	225.9	237.8	270.3	284.5	344
345	306.5	159.8	164.6	163.7	157.5	226.5	238.5	271.1	285.4	345
346	307.3	160.3	165.1	164.2	158.0	227.2	239.2	271.9	286.2	346
347	308.2	160.8	165.7	164.7	158.5	227.9	239.9	272.7	287.0	347
348	309.1	161.4	166.2	165.2	159.0	228.5	240.6	273.5	287.9	348
349	310.0	161.9	166.7	165.7	159.5	229.2	241.3	274.3	288.7	349
350	310.9	162.4	167.2	166.3	160.1	229.9	242.0	275.0	289.5	350
351	311.8	162.9	167.7	166.8	160.6	230.6	242.7	275.8	290.4	351
352	312.7	163.4	168.3	167.3	161.1	231.2	243.4	276.6	291.2	352
353	313.6	163.9	168.8	167.8	161.6	231.9	244.1	277.4	292.0	353
354	314.4	164.4	169.3	168.4	162.2	232.6	244.8	278.2	292.8	354
355	315.3	164.9	169.8	168.9	162.7	233.3	245.6	279.0	293.7	355
356	316.2	165.4	170.4	169.4	163.2	233.9	246.3	279.8	294.5	356
357	317.1	166.0	170.9	170.0	163.7	234.6	247.0	280.6	295.3	357
358	318.0	166.5	171.4	170.5	164.3	235.3	247.7	281.4	296.2	358
359	318.9	167.0	171.9	171.0	164.8	236.0	248.4	282.2	297.0	359
360	319.8	167.5	172.5	171.5	165.3	236.7	249.1	282.9	297.8	360
361	320.7	168.0	173.0	172.1	165.8	237.3	249.8	283.7	298.7	361
362	321.6	168.5	173.5	172.6	166.4	238.0	250.5	284.5	299.5	362
363	322.4	169.0	174.0	173.1	166.9	238.7	251.2	285.3	300.3	363
364	323.3	169.6	174.6	173.7	167.4	239.4	252.0	286.1	301.2	364
365	324.2	170.1	175.1	174.2	167.9	240.0	252.7	286.9	302.0	365
366	325.1	170.6	175.6	174.7	168.5	240.7	253.4	287.7	302.8	366
367	326.0	171.1	176.1	175.2	169.0	241.4	254.1	288.5	303.6	367
368	326.9	171.6	176.7	175.8	169.5	242.1	254.8	289.3	304.5	368
369	327.8	172.1	177.2	176.3	170.0	242.7	255.5	290.0	305.3	369
370	328.7	172.7	177.7	176.8	170.6	243.4	256.2	290.8	306.1	370
371	329.5	173.2	178.3	177.4	171.1	244.1	256.9	291.6	307.0	371
372	330.4	173.7	178.8	177.9	171.6	244.8	257.7	292.4	307.8	372
373	331.3	174.2	179.3	178.4	172.2	245.4	258.4	293.2	308.6	373
374	332.2	174.7	179.8	179.0	172.7	246.1	259.1	294.0	309.5	374
375	333.1	175.3	180.4	179.5	173.2	246.8	259.8	294.8	310.3	375
376	334.0	175.8	180.9	180.0	173.7	247.5	260.5	295.6	311.1	376
377	334.9	176.3	181.4	180.6	174.3	248.1	261.2	296.4	312.0	377
378	335.8	176.8	182.0	181.1	174.8	248.8	261.9	297.2	312.8	378
379	336.7	177.3	182.5	181.6	175.3	249.5	262.6	297.9	313.6	379
380	337.5	177.9	183.0	182.1	175.9	250.2	263.4	298.7	314.5	380
381	338.4	178.4	183.6	182.7	176.4	250.8	264.1	299.5	315.3	381
382	339.3	178.9	184.1	183.2	176.9	251.5	264.8	300.3	316.1	382
383	340.2	179.4	184.6	183.8	177.5	252.2	265.5	301.1	316.9	383
384	341.1	180.0	185.2	184.3	178.0	252.9	266.2	301.9	317.8	384
385	342.0	180.5	185.7	184.8	178.5	253.6	266.9	302.7	318.6	385
386	342.9	181.0	186.2	185.4	179.1	254.2	267.6	303.5	319.4	386
387	343.8	181.5	186.8	185.9	179.6	254.9	268.3	304.2	320.3	387
388	344.6	182.0	187.3	186.4	180.1	255.6	269.0	305.0	321.1	388
389	345.5	182.6	187.8	187.0	180.6	256.3	269.8	305.8	321.9	389
390	346.4	183.1	188.4	187.5	181.2	256.9	270.5	306.6	322.8	390
391	347.3	183.6	188.9	188.0	181.7	257.6	271.2	307.4	323.6	391
392	348.2	184.1	189.4	188.6	182.3	258.3	271.9	308.2	324.4	392
393	349.1	184.7	190.0	189.1	182.8	259.0	272.6	309.0	325.2	393
394	350.0	185.2	190.5	189.7	183.3	259.6	273.3	309.8	326.1	394

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

[Expressed in milligrams.]

CUPROUS OXID (Cu ₂ O)	COPPER (Cu)	DEXTROROSE (d-GLUCOSE)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu ₂ O)
				0.4 gram sugar	2 grams sugar	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ ·H ₂ O	C ₁₂ H ₂₂ O ₁₁	C ₁₂ H ₂₂ O ₁₁ ·H ₂ O	
395	350.9	185.7	191.0	190.2	183.9	260.3	274.0	310.6	326.9	395
396	351.8	186.2	191.6	190.7	184.4	261.0	274.7	311.4	327.7	396
397	352.6	186.8	192.1	191.3	184.9	261.7	275.5	312.1	328.6	397
398	353.5	187.3	192.7	191.8	185.5	262.3	276.2	312.9	329.4	398
399	354.4	187.8	193.2	192.3	186.0	263.0	276.9	313.7	330.2	399
400	355.3	188.4	193.7	192.9	186.5	263.7	277.6	314.5	331.1	400
401	356.2	188.9	194.3	193.4	187.1	264.4	278.3	315.3	331.9	401
402	357.1	189.4	194.8	194.0	187.6	265.0	279.0	316.1	332.7	402
403	358.0	189.9	195.4	194.5	188.1	265.7	279.7	316.9	333.6	403
404	358.9	190.5	195.9	195.0	188.7	266.4	280.4	317.7	334.4	404
405	359.7	191.0	196.4	195.6	189.2	267.1	281.1	318.5	335.2	405
406	360.6	191.5	197.0	196.1	189.8	267.8	281.9	319.2	336.0	406
407	361.5	192.1	197.5	196.7	190.3	268.4	282.6	320.0	336.9	407
408	362.4	192.6	198.1	197.2	190.8	269.1	283.3	320.8	337.7	408
409	363.3	193.1	198.6	197.7	191.4	269.8	284.0	321.6	338.5	409
410	364.2	193.7	199.1	198.3	191.9	270.5	284.7	322.4	339.4	410
411	365.1	194.2	199.7	198.8	192.5	271.2	285.4	323.2	340.2	411
412	366.0	194.7	200.2	199.4	193.0	271.8	286.2	324.0	341.0	412
413	366.9	195.2	200.8	199.9	193.5	272.5	286.9	324.8	341.9	413
414	367.7	195.8	201.3	200.5	194.1	273.2	287.6	325.6	342.7	414
415	368.6	196.3	201.8	201.0	194.6	273.9	288.3	326.3	343.5	415
416	369.5	196.8	202.4	201.6	195.2	274.6	289.0	327.1	344.4	416
417	370.4	197.4	202.9	202.1	195.7	275.2	289.7	327.9	345.2	417
418	371.3	197.9	203.5	202.6	196.2	275.9	290.4	328.7	346.0	418
419	372.2	198.4	204.0	203.2	196.8	276.6	291.2	329.5	346.8	419
420	373.1	199.0	204.6	203.7	197.3	277.3	291.9	330.3	347.7	420
421	374.0	199.5	205.1	204.3	197.9	277.9	292.6	331.1	348.5	421
422	374.8	200.1	205.7	204.8	198.4	278.6	293.3	331.9	349.3	422
423	375.7	200.6	206.2	205.4	198.9	279.3	294.0	332.7	350.2	423
424	376.6	201.1	206.7	205.9	199.5	280.0	294.7	333.4	351.0	424
425	377.5	201.7	207.3	206.5	200.0	280.7	295.4	334.2	351.8	425
426	378.4	202.2	207.8	207.0	200.6	281.3	296.2	335.0	352.7	426
427	379.3	202.8	208.4	207.6	201.1	282.0	296.9	335.8	353.5	427
428	380.2	203.3	208.9	208.1	201.7	282.7	297.6	336.6	354.3	428
429	381.1	203.8	209.5	208.7	202.2	283.4	298.3	337.4	355.1	429
430	382.0	204.4	210.0	209.2	202.7	284.1	299.0	338.2	356.0	430
431	382.8	204.9	210.6	209.8	203.3	284.7	299.7	339.0	356.8	431
432	383.7	205.5	211.1	210.3	203.8	285.4	300.5	339.7	357.6	432
433	384.6	206.0	211.7	210.9	204.4	286.1	301.2	340.5	358.5	433
434	385.5	206.5	212.2	211.4	204.9	286.8	301.9	341.3	359.3	434
435	386.4	207.1	212.8	212.0	205.5	287.5	302.6	342.1	360.1	435
436	387.3	207.6	213.3	212.5	206.0	288.1	303.3	342.9	361.0	436
437	388.2	208.2	213.9	213.1	206.6	288.8	304.0	343.7	361.8	437
438	389.1	208.7	214.4	213.6	207.1	289.5	304.7	344.5	362.6	438
439	390.0	209.2	215.0	214.2	207.7	290.2	305.5	345.3	363.4	439
440	390.8	209.8	215.5	214.7	208.2	290.9	306.2	346.1	364.3	440
441	391.7	210.3	216.1	215.3	208.8	291.5	306.9	346.8	365.1	441
442	392.6	210.9	216.6	215.8	209.3	292.2	307.6	347.6	365.9	442
443	393.5	211.4	217.2	216.4	209.9	292.9	308.3	348.4	366.8	443
444	394.4	212.0	217.8	216.9	210.4	293.6	309.0	349.2	367.6	444
445	395.3	212.5	218.3	217.5	211.0	294.2	309.7	350.0	368.4	445
446	396.2	213.1	218.9	218.0	211.5	294.9	310.5	350.8	369.3	446
447	397.1	213.6	219.4	218.6	212.1	295.6	311.2	351.6	370.1	447
448	397.9	214.1	220.0	219.1	212.6	296.3	311.9	352.4	370.9	448
449	398.8	214.7	220.5	219.7	213.2	297.0	312.6	353.2	371.7	449

TABLE 1.—MUNSON AND WALKER'S TABLE.—Continued.

(Expressed in milligrams.)

CUPROUS OXID (Cu_2O)	COPPER (Cu)	DEXTROSE (d-glucose)	INVERT SUGAR	INVERT SUGAR AND SUCROSE		LACTOSE		MALTOSE		CUPROUS OXID (Cu_2O)
				0.4 gram sugar total	2 grams sugar total	$\text{C}_{12}\text{H}_{22}\text{O}_{11}$	$\text{C}_{12}\text{H}_{22}\text{O}_{11}\cdot\text{H}_2\text{O}$	$\text{C}_{12}\text{H}_{22}\text{O}_{11}$	$\text{C}_{12}\text{H}_{22}\text{O}_{11}\cdot\text{H}_2\text{O}$	
450	399.7	215.2	221.1	220.2	213.7	297.6	313.3	353.9	372.6	450
451	400.6	215.8	221.6	220.8	214.3	298.3	314.0	354.7	373.4	451
452	401.5	216.3	222.2	221.4	214.8	299.0	314.7	355.5	374.2	452
453	402.4	216.9	222.8	221.9	215.4	299.7	315.5	356.3	375.1	453
454	403.3	217.4	223.3	222.5	215.9	300.4	316.2	357.1	375.9	454
455	404.2	218.0	223.9	223.0	216.5	301.1	316.9	357.9	376.7	455
456	405.1	218.5	224.4	223.6	217.0	301.7	317.6	358.7	377.6	456
457	405.9	219.1	225.0	224.1	217.6	302.4	318.3	359.5	378.4	457
458	406.8	219.6	225.5	224.7	218.1	303.1	319.0	360.3	379.2	458
459	407.7	220.2	226.1	225.3	218.7	303.8	319.8	361.0	380.0	459
460	408.6	220.7	226.7	225.8	219.2	304.5	320.5	361.8	380.9	460
461	409.5	221.3	227.2	226.4	219.8	305.1	321.2	362.6	381.7	461
462	410.4	221.8	227.8	226.9	220.3	305.8	321.9	363.4	382.5	462
463	411.3	222.4	228.3	227.5	220.9	306.5	322.6	364.2	383.4	463
464	412.2	222.9	228.9	228.1	221.4	307.2	323.4	365.0	384.2	464
465	413.0	223.5	229.5	228.6	222.0	307.9	324.1	365.8	385.0	465
466	413.9	224.0	230.0	229.2	222.5	308.6	324.8	366.6	385.9	466
467	414.8	224.6	230.6	229.7	223.1	309.2	325.5	367.3	386.7	467
468	415.7	225.1	231.2	230.3	223.7	309.9	326.2	368.1	387.5	468
469	416.6	225.7	231.7	230.9	224.2	310.6	326.9	368.9	388.3	469
470	417.5	226.2	232.3	231.4	224.8	311.3	327.7	369.7	389.2	470
471	418.4	226.8	232.8	232.0	225.3	312.0	328.4	370.5	390.0	471
472	419.3	227.4	233.4	232.5	225.9	312.6	329.1	371.3	390.8	472
473	420.2	227.9	234.0	233.1	226.4	313.3	329.8	372.1	391.7	473
474	421.0	228.5	234.5	233.7	227.0	314.0	330.5	372.9	392.5	474
475	421.9	229.0	235.1	234.2	227.6	314.7	331.3	373.7	393.3	475
476	422.8	229.6	235.7	234.8	228.1	315.4	332.0	374.4	394.2	476
477	423.7	230.1	236.2	235.4	228.7	316.1	332.7	375.2	395.0	477
478	424.6	230.7	236.8	235.9	229.2	316.7	333.4	376.0	395.8	478
479	425.5	231.3	237.4	236.5	229.8	317.4	334.1	376.8	396.6	479
480	426.4	231.8	237.9	237.1	230.3	318.1	334.8	377.6	397.5	480
481	427.3	232.4	238.5	237.6	230.9	318.8	335.6	378.4	398.3	481
482	428.1	232.9	239.1	238.2	231.5	319.5	336.3	379.2	399.1	482
483	429.0	233.5	239.6	238.8	232.0	320.1	337.0	380.0	400.0	483
484	429.9	234.1	240.2	239.3	232.6	320.8	337.7	380.7	400.8	484
485	430.8	234.6	240.8	239.9	233.2	321.5	338.4	381.5	401.6	485
486	431.7	235.2	241.4	240.5	233.7	322.2	339.1	382.3	402.4	486
487	432.6	235.7	241.9	241.0	234.3	322.9	339.9	383.1	403.3	487
488	433.5	236.3	242.5	241.6	234.8	323.6	340.6	383.9	404.1	488
489	434.4	236.9	243.1	242.2	235.4	324.2	341.3	384.7	404.9	489
490	435.3	237.4	243.6	242.7	236.0	324.9	342.0	385.5	405.8	490

II. A. H. Low Volumetric Method, Modified.⁴—Tentative.

REAGENT.

Standard thiosulphate solution.—Prepare a solution of sodium thiosulphate containing 19 grams of pure crystals in 1 liter. Weigh accurately about 0.2 gram of pure copper foil and place in a flask of 250 cc. capacity. Dissolve by warming with 5 cc. of a mixture of equal volumes of strong nitric acid and water. Dilute to 50 cc., boil to expel the red fumes, add 5 cc. of strong bromine water, and boil

until the bromin is completely driven off. Remove from the heat and add a slight excess of strong ammonium hydroxid (about 7 cc. is required). Again boil until the excess of ammonia is expelled, as shown by a change of color of the liquid, and a partial precipitation. Then add a slight excess of strong acetic acid (3 or 4 cc. of 80% acid) and boil for a minute. Cool to room temperature and add 10 cc. of 30% potassium iodid solution. Titrate at once with the thiosulphate solution until the brown tinge has become weak, then add sufficient starch indicator [VII, 3 (a)] to produce a marked blue coloration. Continue the titration cautiously until the color due to free iodin has entirely vanished. The blue color changes toward the end to a faint lilac. If at this point the thiosulphate be added drop by drop and a little time allowed for complete reaction after each addition, there is no difficulty in determining the end point within a single drop. One cc. of the thiosulphate solution will be found to correspond to about 0.005 gram of copper.

29

DETERMINATION.

After washing the precipitated cuprous oxid, cover the Gooch with a watch glass and dissolve the oxid by means of 5 cc. of warm nitric acid (1 to 1) poured under the watch glass with a pipette. Catch the filtrate in a 250 cc. flask, wash the watch glass and Gooch free of copper, using about 50 cc. of water. Boil to expel red fumes, add 5 cc. of bromin water, boil off the bromin, and proceed exactly as in 28.

30

III. Volumetric Permanganate Method.—Tentative.

Filter and wash the cuprous oxid as directed under 25. Transfer the asbestos film to the beaker, add about 30 cc. of hot water, and beat the precipitate and asbestos thoroughly. Rinse the crucible with 50 cc. of a hot saturated solution of ferric sulphate in 20% sulphuric acid, receiving the rinsings in the beaker containing the precipitate. After the cuprous oxid is dissolved, wash the solution into a large Erlenmeyer flask and immediately titrate with a standard solution of potassium permanganate, 1 cc. of which should be equivalent to 0.010 gram of copper. Standardize this solution by making 6 or more determinations with the same sugar solution, titrating one half of the precipitates obtained, and determining the copper in the others by electrolysis. The average weight of copper obtained by electrolysis, divided by the average number of cc. of permanganate solution required for the titrations, gives the weight of copper equivalent to 1 cc. of the standard permanganate solution. A solution standardized with iron or oxalic acid will give too low a result.

31 *IV. Electrolytic Deposition from Sulphuric Acid Solution.—Tentative.*

Filter the cuprous oxid in a Gooch, wash the beaker and the precipitate thoroughly with hot water without transferring the precipitate to the filter. Wash the asbestos film and the adhering cuprous oxid into the beaker by means of hot dilute nitric acid. After the copper is all in solution, refilter through a thin film of asbestos in a Gooch and wash thoroughly with hot water. Add 10 cc. of sulphuric acid (1 to 4), and evaporate the filtrate on the steam bath until the copper salt has largely crystallized. Heat carefully on a hot plate or over asbestos until the evolution of white fumes shows that the excess of nitric acid is removed. Add 8–10 drops of nitric acid (sp. gr. 1.42) and rinse into a 100–125 cc. platinum dish. Deposit the copper by

electrolysis. Wash thoroughly with water, then break the current, wash with alcohol and ether successively, dry at about 50°C., and weigh. If preferred, the electrolysis can be conducted in a beaker, the copper being deposited upon a weighed platinum electrode.

32 *V. Electrolytic Deposition from Sulphuric and Nitric Acid Solution.—Tentative.*

Filter and wash as directed under **31**. Transfer the asbestos film from the crucible to the beaker by means of a glass rod and rinse the crucible with about 30 cc. of a boiling mixture of dilute sulphuric and nitric acids, containing 65 cc. of sulphuric acid (sp. gr. 1.84) and 50 cc. of nitric acid (sp. gr. 1.42) per liter. Heat and agitate until solution is complete; filter and electrolyze as under **31**.

33 *VI. Electrolytic Deposition from Nitric Acid Solution.—Tentative.*

Filter and wash as directed under **31**. Transfer the asbestos film and adhering oxid to the beaker. Dissolve the oxid still remaining in the crucible by means of 2 cc. of nitric acid (sp. gr. 1.42), adding it with a pipette and receiving the solution in the beaker containing the asbestos film. Rinse the crucible with a jet of water, allowing the rinsings to flow into the beaker. Heat the contents of the beaker until the copper is all in solution, filter, dilute the filtrate to a volume of 100 cc. or more, and electrolyze. When a nitrate solution is electrolyzed, the first washing of the deposit should be made with water acidulated with sulphuric acid, in order to remove all the nitric acid before the current is interrupted.

34 *VII. Reduction in Hydrogen.—Tentative.*

Deposit an asbestos film on a perforated platinum disc or cone contained in a hard glass filtering tube, wash free from loose fibers, dry and weigh. Through this tube, previously moistened, filter the cuprous oxid immediately, using suction. Transfer the cuprous oxid to the tube through a removable funnel, and wash thoroughly with hot water, alcohol and ether successively. After drying, connect the tube with a supply of dry hydrogen, heat gently until the cuprous oxid is completely reduced to metallic copper, cool in the current of hydrogen, and weigh. If preferred, a Gooch crucible may be used for the filtration.

Herzfeld Gravimetric Method.—Tentative.

Method I.

(For materials containing 1.5% or less of invert sugar and 98.5% or more of sucrose.)

35 REAGENTS.

The reagents and solutions used are described under **24**.

36 DETERMINATION.

Prepare the solution of the material to be examined so as to contain 20 grams in 100 cc., free from suspended impurities by filtration and from soluble impurities by neutral lead acetate, removing the excess of lead by means of sodium carbonate. Place 50 cc. of the reagent and 50 cc. of the sugar solution in a 250 cc. beaker. Heat this mixture at such a rate that approximately 4 minutes are required to bring it to the boiling point, and boil for exactly 2 minutes. Add 100 cc. of cold, recently boiled, water. Filter immediately through asbestos, and determine the copper by one of the methods under **26, 29-34**, respectively. Obtain the corresponding percentage of invert sugar from **37**.

37

TABLE 2.—HERZFELD'S TABLE.⁵

For the determination of invert sugar in materials containing 1.5%, or less, of invert sugar and 98.5%, or more, of sucrose.

COPPER REDUCED BY 10 GRAMS OF MATERIAL	INVERT SUGAR	COPPER REDUCED BY 10 GRAMS OF MATERIAL	INVERT SUGAR	COPPER REDUCED BY 10 GRAMS OF MATERIAL	INVERT SUGAR
mg.	per cent	mg.	per cent	mg.	per cent
50	0.05	140	0.51	230	1.02
55	0.07	145	0.53	235	1.05
60	0.09	150	0.56	240	1.07
65	0.11	155	0.59	245	1.10
70	0.14	160	0.62	250	1.13
75	0.16	165	0.65	255	1.16
80	0.19	170	0.68	260	1.18
85	0.21	175	0.71	265	1.21
90	0.24	180	0.74	270	1.24
95	0.27	185	0.76	275	1.27
100	0.30	190	0.79	280	1.30
105	0.32	195	0.82	285	1.33
110	0.35	200	0.85	290	1.36
115	0.38	205	0.88	295	1.38
120	0.40	210	0.90	300	1.41
125	0.43	215	0.93	305	1.44
130	0.45	220	0.96	310	1.47
135	0.48	225	0.99	315	1.50

Method II.

(For materials containing 1.5% or more of invert sugar and 98.5% or less of sucrose.)

38

REAGENTS.

Same as described under 24.

39

DETERMINATION.

Prepare a solution of the material to be examined in such a manner that it contains 20 grams in 100 cc. after clarification and removal of the excess of lead. Prepare a series of solutions in large test tubes by adding 1, 2, 3, 4, and 5 cc. of this solution to each tube successively. Add 5 cc. of the reagent to each, heat to boiling, boil 2 minutes, and filter. Note the volume of sugar solution which gives the filtrate lightest in tint, but still distinctly blue. Place 20 times this volume of the sugar solution in a 100 cc. flask, dilute to the mark, and mix well. Use 50 cc. of the solution for the determination, which is conducted as described under 36. For the calculation of the result use the following formulas and table of factors of Meissl and Hiller:

Let Cu = the weight of copper obtained;

P = the polarization of the sample;

W = the weight of the sample in the 50 cc. of the solution used for the determination;

F = the factor obtained from the table for the conversion of copper to invert sugar;

Then $\frac{\text{Cu}}{2} = Z$, approximate weight of invert sugar;

$Z \times \frac{100}{W} = Y$, approximate per cent of invert sugar;

$$\frac{100 P}{P + Y} = R, \text{ approximate per cent of sucrose in mixture of sugars;}$$

$$100 - R = I, \text{ approximate per cent of invert sugar;}$$

$$\frac{CuF}{W} = \text{per cent of invert sugar.}$$

The factor F for calculating copper to invert sugar is then found from 40.

40

TABLE 3.

Meissl and Hiller's^s factors for determinations in materials in which, of the total sugars present, 1.5%, or more, is invert sugar, and 98.5%, or less, is sucrose.

RATIO OF SUCROSE TO INVERT SUGAR = R: I.	APPROXIMATE ABSOLUTE WEIGHT OF INVERT SUGAR (Z)						
	200	175	150	125	100	75	50
	milligrams	milligrams	milligrams	milligrams	milligrams	milligrams	milligrams
	per cent	per cent	per cent	per cent	per cent	per cent	per cent
0: 100	56.4	55.4	54.5	53.8	53.2	53.0	53.0
10: 90	56.3	55.3	54.4	53.8	53.2	52.9	52.9
20: 80	56.2	55.2	54.3	53.7	53.2	52.7	52.7
30: 70	56.1	55.1	54.2	53.7	53.2	52.6	52.6
40: 60	55.9	55.0	54.1	53.6	53.1	52.5	52.4
50: 50	55.7	54.9	54.0	53.5	53.1	52.3	52.2
60: 40	55.6	54.7	53.8	53.2	52.8	52.1	51.9
70: 30	55.5	54.5	53.5	52.9	52.5	51.9	51.6
80: 20	55.4	54.3	53.3	52.7	52.2	51.7	51.3
90: 10	54.6	53.6	53.1	52.6	52.1	51.6	51.2
91: 9	54.1	53.6	52.6	52.1	51.6	51.2	50.7
92: 8	53.6	53.1	52.1	51.6	51.2	50.7	50.3
93: 7	53.6	53.1	52.1	51.2	50.7	50.3	49.8
94: 6	53.1	52.6	51.6	50.7	50.3	49.8	48.9
95: 5	52.6	52.1	51.2	50.3	49.4	48.9	48.5
96: 4	52.1	51.2	50.7	49.8	48.9	47.7	46.9
97: 3	50.7	50.3	49.8	48.9	47.7	46.2	45.1
98: 2	49.9	48.9	48.5	47.3	45.8	43.3	40.0
99: 1	47.7	47.3	46.5	45.1	43.3	41.2	38.1

Example: The polarization of a sugar is 86.4, and 50 cc. of solution containing 3.256 grams of sample gave 0.290 gram of copper.

$$\frac{Cu}{2} = \frac{0.290}{2} = 0.145 = Z$$

$$\frac{Z \times 100}{W} = 0.145 \times \frac{100}{3.256} = 4.45 = Y$$

$$100 - R = 100 - 95.1 = I = 4.9$$

$$R: I = 95.1: 4.9$$

By consulting the table it will be seen that the vertical column headed 150 is nearest to Z, 145, and the horizontal column headed 95: 5 is nearest to the ratio of R to I, 95.1: 4.9. Where these columns meet, we find the factor 51.2 which enters into the final calculation:

$$\frac{CuF}{W} = \frac{0.290 \times 51.2}{3.256} = 4.56 \text{ per cent of invert sugar.}$$

In case there is no sucrose present, the following table may be used instead of the factors given in 40.

TABLE 4.—MEISSL'S TABLE.⁷*For the determination of invert sugar alone.*

[According to Wein.]

[Expressed in milligrams.]

COPPER	INVERT SUGAR	COPPER	INVERT SUGAR	COPPER	INVERT SUGAR	COPPER	INVERT SUGAR
90	46.9	135	70.8	180	95.2	225	120.4
91	47.4	136	71.3	181	95.7	226	120.9
92	47.9	137	71.9	182	96.2	227	121.5
93	48.4	138	72.4	183	96.8	228	122.1
94	48.9	139	72.9	184	97.3	229	122.6
95	49.5	140	73.5	185	97.8	230	123.2
96	50.0	141	74.0	186	98.4	231	123.8
97	50.5	142	74.5	187	99.0	232	124.3
98	51.1	143	75.1	188	99.5	233	124.9
99	51.6	144	75.6	189	100.1	234	125.5
100	52.1	145	76.1	190	100.6	235	126.0
101	52.7	146	76.7	191	101.2	236	126.6
102	53.2	147	77.2	192	101.7	237	127.2
103	53.7	148	77.8	193	102.3	238	127.8
104	54.3	149	78.3	194	102.9	239	128.3
105	54.8	150	78.9	195	103.4	240	128.9
106	55.3	151	79.4	196	104.0	241	129.5
107	55.9	152	80.0	197	104.6	242	130.0
108	56.4	153	80.5	198	105.1	243	130.6
109	56.9	154	81.0	199	105.7	244	131.2
110	57.5	155	81.6	200	106.3	245	131.8
111	58.0	156	82.1	201	106.8	246	132.3
112	58.5	157	82.7	202	107.4	247	132.9
113	59.1	158	83.2	203	107.9	248	133.5
114	59.6	159	83.8	204	108.5	249	134.1
115	60.1	160	84.3	205	109.1	250	134.6
116	60.7	161	84.8	206	109.6	251	135.2
117	61.2	162	85.4	207	110.2	252	135.8
118	61.7	163	85.9	208	110.8	253	136.3
119	62.3	164	86.5	209	111.3	254	136.9
120	62.8	165	87.0	210	111.9	255	137.5
121	63.3	166	87.6	211	112.5	256	138.1
122	63.9	167	88.1	212	113.0	257	138.6
123	64.4	168	88.6	213	113.6	258	139.2
124	64.9	169	89.2	214	114.2	259	139.8
125	65.5	170	89.7	215	114.7	260	140.4
126	66.0	171	90.3	216	115.3	261	140.9
127	66.5	172	90.8	217	115.8	262	141.5
128	67.1	173	91.4	218	116.4	263	142.1
129	67.6	174	91.9	219	117.0	264	142.7
130	68.1	175	92.4	220	117.5	265	143.2
131	68.7	176	93.0	221	118.1	266	143.8
132	69.2	177	93.5	222	118.7	267	144.4
133	69.7	178	94.1	223	119.2	268	144.9
134	70.3	179	94.6	224	119.8	269	145.5

41

TABLE 4.—MEISSL'S TABLE.—Continued.

[Expressed in milligrams.]

COPPER	INVERT SUGAR	COPPER	INVERT SUGAR	COPPER	INVERT SUGAR	COPPER	INVERT SUGAR
270	146.1	310	169.7	350	193.8	390	218.7
271	146.7	311	170.3	351	194.4	391	219.3
272	147.2	312	170.9	352	195.0	392	219.9
273	147.8	313	171.5	353	195.6	393	220.5
274	148.4	314	172.1	354	196.2	394	221.2
275	149.0	315	172.7	355	196.8	395	221.8
276	149.5	316	173.3	356	197.4	396	222.4
277	150.1	317	173.9	357	198.0	397	223.1
278	150.7	318	174.5	358	198.6	398	223.7
279	151.3	319	175.1	359	199.2	399	224.3
280	151.9	320	175.6	360	199.8	400	224.9
281	152.5	321	176.2	361	200.4	401	225.7
282	153.1	322	176.8	362	201.1	402	226.4
283	153.7	323	177.4	363	201.7	403	227.1
284	154.3	324	178.0	364	202.3	404	227.8
285	154.9	325	178.6	365	203.0	405	228.6
286	155.5	326	179.2	366	203.6	406	229.3
287	156.1	327	179.8	367	204.2	407	230.0
288	156.7	328	180.4	368	204.8	408	230.7
289	157.2	329	181.0	369	205.5	409	231.4
290	157.8	330	181.6	370	206.1	410	232.1
291	158.4	331	182.2	371	206.7	411	232.8
292	159.0	332	182.8	372	207.3	412	233.5
293	159.6	333	183.5	373	208.0	413	234.3
294	160.2	334	184.1	374	208.6	414	235.0
295	160.8	335	184.7	375	209.2	415	235.7
296	161.4	336	185.4	376	209.9	416	236.4
297	162.0	337	186.0	377	210.5	417	237.1
298	162.6	338	186.6	378	211.1	418	237.8
299	163.2	339	187.2	379	211.7	419	238.5
300	163.8	340	187.8	380	212.4	420	239.2
301	164.4	341	188.4	381	213.0	421	239.9
302	165.0	342	189.0	382	213.6	422	240.6
303	165.6	343	189.6	383	214.3	423	241.3
304	166.2	344	190.2	384	214.9	424	242.0
305	166.8	345	190.8	385	215.5	425	242.7
306	167.3	346	191.4	386	216.1	426	243.4
307	167.9	347	192.0	387	216.8	427	244.1
308	168.5	348	192.6	388	217.4	428	244.9
309	169.1	349	193.2	389	218.0	429	245.6
						430	246.3

MALTOSE.

42

General Gravimetric Method.—Tentative.

Proceed as directed under 25 and obtain, from 27. the weight of maltose equivalent to the weight of copper reduced.

Wein Method.—Tentative.

43

REAGENTS.

The reagents and solutions used are described under 24.

44

DETERMINATION.

Place 50 cc. of the reagent in a beaker and heat to the boiling point. When boiling briskly, add 25 cc. of the maltose solution containing not more than 0.250 gram of maltose and boil for 4 minutes. Filter immediately through asbestos and determine, by one of the methods given under 26, 29–34 respectively, the amount of copper reduced.

Obtain, from 45, the weight of maltose equivalent to the weight of copper found.

45

TABLE 5.

For the determination of maltose.

[According to Wein.*]

[Expressed in milligrams.]

COPPER	CU- PROUS OXID	MAL- TOSE	COPPER	CU- PROUS OXID	MAL- TOSE	COPPER	CU- PROUS OXID	MAL- TOSE	COPPER	CU- PROUS OXID	MAL- TOSE
31	34.9	26.1	71	79.9	61.0	111	125.0	96.4	151	170.0	132.3
32	36.0	27.0	72	81.1	61.8	112	126.1	97.3	152	171.1	133.2
33	37.2	27.9	73	82.2	62.7	113	127.2	98.1	153	172.3	134.1
34	38.3	28.7	74	83.3	63.6	114	128.3	99.0	154	173.4	135.0
35	39.4	29.6	75	84.4	64.5	115	129.6	99.9	155	174.5	135.9
36	40.5	30.5	76	85.6	65.4	116	130.6	100.8	156	175.6	136.8
37	41.7	31.3	77	86.7	66.2	117	131.7	101.7	157	176.8	137.7
38	42.8	32.2	78	87.8	67.1	118	132.8	102.6	158	177.9	138.6
39	43.9	33.1	79	88.9	68.0	119	134.0	103.5	159	179.0	139.5
40	45.0	33.9	80	90.1	68.9	120	135.1	104.4	160	180.1	140.4
41	46.2	34.8	81	91.2	69.7	121	136.2	105.3	161	181.3	141.3
42	47.3	35.7	82	92.3	70.6	122	137.4	106.2	162	182.4	142.2
43	48.4	36.5	83	93.4	71.5	123	138.5	107.1	163	183.5	143.1
44	49.5	37.4	84	94.6	72.4	124	139.6	108.0	164	184.6	144.0
45	50.7	38.3	85	95.7	73.2	125	140.7	108.9	165	185.8	144.9
46	51.8	39.1	86	96.8	74.1	126	141.9	109.8	166	186.9	145.8
47	52.9	40.0	87	97.9	75.0	127	143.0	110.7	167	188.0	146.7
48	54.0	40.9	88	99.1	75.9	128	144.1	111.6	168	189.1	147.6
49	55.2	41.8	89	100.2	76.8	129	145.2	112.5	169	190.3	148.5
50	56.3	42.6	90	101.3	77.7	130	146.4	113.4	170	191.4	149.4
51	57.4	43.5	91	102.4	78.6	131	147.5	114.3	171	192.5	150.3
52	58.5	44.4	92	103.6	79.5	132	148.6	115.2	172	193.6	151.2
53	59.7	45.2	93	104.7	80.3	133	149.7	116.1	173	194.8	152.0
54	60.8	46.1	94	105.8	81.2	134	150.9	117.0	174	195.9	152.9
55	61.9	47.0	95	107.0	82.1	135	152.0	117.9	175	197.0	153.8
56	63.0	47.8	96	108.1	83.0	136	153.1	118.8	176	198.1	154.7
57	64.2	48.7	97	109.2	83.9	137	154.2	119.7	177	199.3	155.6
58	65.3	49.6	98	110.3	84.8	138	155.4	120.6	178	200.4	156.5
59	66.4	50.4	99	111.5	85.7	139	156.5	121.5	179	201.5	157.4
60	67.6	51.3	100	112.6	86.6	140	157.6	122.4	180	202.6	158.3
61	68.7	52.2	101	113.7	87.5	141	158.7	123.3	181	203.8	159.2
62	69.8	53.1	102	114.8	88.4	142	159.9	124.2	182	204.9	160.1
63	70.9	53.9	103	116.0	89.2	143	161.0	125.1	183	206.0	160.9
64	72.1	54.8	104	117.1	90.1	144	162.1	126.0	184	207.1	161.8
65	73.2	55.7	105	118.2	91.0	145	163.2	126.9	185	208.3	162.7
66	74.3	56.6	106	119.3	91.9	146	164.4	127.8	186	209.4	163.6
67	75.4	57.4	107	120.5	92.8	147	165.5	128.7	187	210.5	164.5
68	76.6	58.3	108	121.6	93.7	148	166.6	129.6	188	211.7	165.4
69	77.7	59.2	109	122.7	94.6	149	167.7	130.5	189	212.8	166.3
70	78.8	60.1	110	123.8	95.5	150	168.9	131.4	190	213.9	167.2

TABLE 5.—Continued.
For the determination of maltose.
[Expressed in milligrams.]

COOPER	CU- PROUS OXID	MAL- TOSE	COPPER	CU- PROUS OXID	MAL- TOSE	COPPER	CU- PROUS OXID	MAL- TOSE	COPPER	CU- PROUS OXID	MAL- TOSE
191	215.0	168.1	221	243.7	194.8	251	282.6	221.7	291	316.4	248.7
192	216.2	169.0	222	249.9	195.7	252	283.7	222.6	282	317.5	249.6
193	217.3	169.8	223	251.0	196.6	253	284.8	223.5	283	318.6	250.4
194	218.4	170.7	224	252.4	197.5	254	286.0	224.4	284	319.7	251.3
195	219.5	171.6	225	253.3	198.4	255	287.1	225.3	285	320.9	252.2
196	220.7	172.5	226	254.4	199.3	256	288.2	226.2	286	322.0	253.1
197	221.8	173.4	227	255.6	200.2	257	289.3	227.1	287	323.1	254.0
198	222.9	174.3	228	256.7	201.1	258	290.5	228.0	288	324.2	254.9
199	224.0	175.2	229	257.8	202.0	259	291.6	228.9	289	325.4	255.8
200	225.2	176.1	230	258.9	202.9	260	292.7	229.8	290	326.5	256.6
201	226.3	177.0	231	260.1	203.8	261	293.8	230.7	291	327.4	257.5
202	227.4	177.9	232	261.2	204.7	262	295.0	231.6	292	328.7	258.4
203	228.5	178.7	233	262.3	205.6	263	296.1	232.5	293	329.9	259.3
204	229.7	179.6	234	263.4	206.5	264	297.2	233.4	294	331.0	260.2
205	230.8	180.5	235	264.6	207.4	265	298.3	234.3	295	332.1	261.1
206	231.9	181.4	236	265.7	208.3	266	299.5	235.2	296	333.2	262.0
207	233.0	182.3	237	266.8	209.1	267	300.6	236.1	297	334.4	262.8
208	234.2	183.2	238	268.0	210.0	268	301.7	237.0	298	335.5	263.7
209	235.3	184.1	239	269.1	210.9	269	302.8	237.9	299	336.6	264.6
210	236.4	185.0	240	270.2	211.8	270	304.0	238.8	300	337.8	265.5
211	237.6	185.9	241	271.3	212.7	271	305.1	239.7			
212	238.7	186.8	242	272.5	213.6	272	306.2	240.6			
213	239.8	187.7	243	273.6	214.5	273	307.3	241.5			
214	240.9	188.6	244	274.7	215.4	274	308.5	242.4			
215	242.1	189.5	245	275.8	216.3	275	309.6	243.3			
216	243.2	190.4	246	277.0	217.2	276	310.7	244.2			
217	244.3	191.2	247	278.1	218.1	277	311.9	245.1			
218	245.4	192.1	248	279.2	219.0	278	313.0	246.0			
219	246.6	193.0	249	280.3	219.9	279	314.1	246.9			
220	247.7	193.9	250	281.5	220.8	280	315.2	247.8			

LACTOSE.

General Gravimetric Method.—Tentative.

Proceed as directed under 25 and obtain, from 27, the weight of lactose equivalent to the weight of copper reduced.

Soxhlet-Wein Method.—Official.

REAGENTS.

The reagents and solutions used are described under 24.

DETERMINATION.

Place 50 cc. of the reagent in a beaker and heat to the boiling point. When boiling briskly, add 100 cc. of the lactose solution containing not more than 0.300 gram of lactose and boil for 6 minutes. Filter immediately through asbestos and determine by one of the methods given under 26, 29–34 inclusive, the amount of copper reduced. Obtain, from 49, the weight of lactose equivalent to the weight of copper found.

[Expressed in milligrams.]

COPPER	LACTOSE	COPPER	LACTOSE	COPPER	LACTOSE	COPPER	LACTOSE	COPPER	LACTOSE
100	71.6	160	116.4	220	161.9	280	208.3	340	255.7
101	72.4	161	117.1	221	162.7	281	209.1	341	256.5
102	73.1	162	117.9	222	163.4	282	209.9	342	257.4
103	73.8	163	118.6	223	164.2	283	210.7	343	258.2
104	74.6	164	119.4	224	164.9	284	211.5	344	259.0
105	75.3	165	120.2	225	165.7	285	212.3	345	259.8
106	76.1	166	120.9	226	166.4	286	213.1	346	260.6
107	76.8	167	121.7	227	167.2	287	213.9	347	261.4
108	77.6	168	122.4	228	167.9	288	214.7	348	262.3
109	78.3	169	123.2	229	168.6	289	215.5	349	263.1
110	79.0	170	123.9	230	169.4	290	216.3	350	263.9
111	79.8	171	124.7	231	170.1	291	217.1	351	264.7
112	80.5	172	125.5	232	170.9	292	217.9	352	265.5
113	81.3	173	126.2	233	171.6	293	218.7	353	266.3
114	82.0	174	127.0	234	172.4	294	219.5	354	267.2
115	82.7	175	127.8	235	173.1	295	220.3	355	268.0
116	83.5	176	128.5	236	173.9	296	221.1	356	268.8
117	84.2	177	129.3	237	174.6	297	221.9	357	269.6
118	85.0	178	130.1	238	175.4	298	222.7	358	270.4
119	85.7	179	130.8	239	176.2	299	223.5	359	271.2
120	86.4	180	131.6	240	176.9	300	224.4	360	272.1
121	87.2	181	132.4	241	177.7	301	225.2	361	272.9
122	87.9	182	133.1	242	178.5	302	225.9	362	273.7
123	88.7	183	133.9	243	179.3	303	226.7	363	274.5
124	89.4	184	134.7	244	180.1	304	227.5	364	275.3
125	90.1	185	135.4	245	180.8	305	228.3	365	276.2
126	90.9	186	136.2	246	181.6	306	229.1	366	277.1
127	91.6	187	137.0	247	182.4	307	229.8	367	277.9
128	92.4	188	137.7	248	183.2	308	230.6	368	278.8
129	93.1	189	138.5	249	184.0	309	231.4	369	279.6
130	93.8	190	139.3	250	184.8	310	232.2	370	280.5
131	94.6	191	140.0	251	185.5	311	232.9	371	281.4
132	95.3	192	140.8	252	186.3	312	233.7	372	282.2
133	96.1	193	141.6	253	187.1	313	234.5	373	283.1
134	96.9	194	142.3	254	187.9	314	235.3	374	283.9
135	97.6	195	143.1	255	188.7	315	236.1	375	284.8
136	98.3	196	143.9	256	189.4	316	236.8	376	285.7
137	99.1	197	144.6	257	190.2	317	237.6	377	286.5
138	99.8	198	145.4	258	191.0	318	238.4	378	287.4
139	100.5	199	146.2	259	191.8	319	239.2	379	288.2
140	101.3	200	146.9	260	192.5	320	240.0	380	289.1
141	102.0	201	147.7	261	193.3	321	240.7	381	289.9
142	102.8	202	148.5	262	194.1	322	241.5	382	290.8
143	103.5	203	149.2	263	194.9	323	242.3	383	291.7
144	104.3	204	150.0	264	195.7	324	243.1	384	292.5
145	105.1	205	150.7	265	196.4	325	243.9	385	293.4
146	105.8	206	151.5	266	197.2	326	244.6	386	294.2
147	106.6	207	152.2	267	198.0	327	245.4	387	295.1
148	107.3	208	153.0	268	198.8	328	246.2	388	296.0
149	108.1	209	153.7	269	199.5	329	247.0	389	296.8
150	108.8	210	154.5	270	200.3	330	247.7	390	297.7
151	109.6	211	155.2	271	201.1	331	248.5	391	298.5
152	110.3	212	156.0	272	201.9	332	249.2	392	299.4
153	111.1	213	156.7	273	202.7	333	250.0	393	300.3
154	111.9	214	157.5	274	203.5	334	250.8	394	301.1
155	112.6	215	158.2	275	204.3	335	251.6	395	302.0
156	113.4	216	159.0	276	205.1	336	252.5	396	302.8
157	114.1	217	159.7	277	205.9	337	253.3	397	303.7
158	114.9	218	160.4	278	206.7	338	254.1	398	304.6
159	115.6	219	161.2	279	207.5	339	254.9	399	305.4
								400	306.3

DEXTROSE.

50 *Approximate Volumetric Method for Rapid Work.—Tentative.*

Proceed as directed under 21. Standardize the reagent against pure dextrose.

51 *Soxhlet Method.—Tentative.*

Proceed as directed under 23. Under these conditions 100 cc. of the reagent require 0.475 gram of anhydrous dextrose for complete reduction and the formula becomes $\frac{100 \times 0.475}{VW} = \text{per cent of dextrose.}$

52 *General Gravimetric Method.—Tentative.*

Proceed as directed under 25 and obtain, from 27, the weight of dextrose equivalent to the weight of copper reduced.

Allihn Gravimetric Method.—Tentative.

53

REAGENT.

Allihn's Modification of Fehling's Solution.—Prepare by mixing, immediately before use, equal volumes of (a) and (b).

(a) *Copper sulphate solution.*—Dissolve 34.639 grams of copper sulphate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) in water and dilute to 500 cc.

(b) *Alkaline tartrate solution.*—Dissolve 173 grams of Rochelle salts and 125 grams of potassium hydroxid in water and dilute to 500 cc.

54

DETERMINATION.

Place 30 cc. of the copper sulphate solution, 30 cc. of the alkaline tartrate solution, and 60 cc. of water in a beaker and heat to boiling. Add 25 cc. of the solution of the material to be examined, prepared so as not to contain more than 0.25 gram of dextrose, and boil for exactly 2 minutes, keeping the beaker covered. Filter immediately through asbestos, and obtain the weight of copper by one of the methods given under 26, 29-34 inclusive. The corresponding weight of dextrose is found in 55.

TABLE 7.—ALLIHN'S TABLE.¹⁰*For the determination of dextrose.*

[Expressed in milligrams.]

COPPER	CU- PROUS OXID	DEX- TROSE	COPPER	CU- PROUS OXID	DEX- TROSE	COPPER	CU- PROUS OXID	DEX- TROSE	COPPER	CU- PROUS OXID	DEX- TROSE
11	12.4	6.6	71	79.9	36.3	131	147.5	66.7	191	215.0	97.8
12	13.5	7.1	72	81.1	36.8	132	148.6	67.2	192	216.2	98.4
13	14.6	7.6	73	82.2	37.3	133	149.7	67.7	193	217.3	98.9
14	15.8	8.1	74	83.3	37.8	134	150.9	68.2	194	218.4	99.4
15	16.9	8.6	75	84.4	38.3	135	152.0	68.8	195	219.5	100.0
16	18.0	9.0	76	85.6	38.8	136	153.1	69.3	196	220.7	100.5
17	19.1	9.5	77	86.7	39.3	137	154.2	69.8	197	221.8	101.0
18	20.3	10.0	78	87.8	39.8	138	155.4	70.3	198	222.9	101.5
19	21.4	10.5	79	88.9	40.3	139	156.5	70.8	199	224.0	102.0
20	22.5	11.0	80	90.1	40.8	140	157.6	71.3	200	225.2	102.6
21	23.6	11.5	81	91.2	41.3	141	158.7	71.8	201	226.3	103.1
22	24.8	12.0	82	92.3	41.8	142	159.9	72.3	202	227.4	103.7
23	25.9	12.5	83	93.4	42.3	143	161.0	72.9	203	228.5	104.2
24	27.0	13.0	84	94.6	42.8	144	162.1	73.4	204	229.7	104.7
25	28.1	13.5	85	95.7	43.4	145	163.2	73.9	205	230.8	105.3
26	29.3	14.0	86	96.8	43.9	146	164.4	74.4	206	231.9	105.8
27	30.4	14.5	87	97.9	44.4	147	165.5	74.9	207	233.0	106.3
28	31.5	15.0	88	99.1	44.9	148	166.6	75.5	208	234.2	106.8
29	32.7	15.5	89	100.2	45.4	149	167.7	76.0	209	235.3	107.4
30	33.8	16.0	90	101.3	45.9	150	168.9	76.5	210	236.4	107.9
31	34.9	16.5	91	102.4	46.4	151	170.0	77.0	211	237.6	108.4
32	36.0	17.0	92	103.6	46.9	152	171.1	77.5	212	238.7	109.0
33	37.2	17.5	93	104.7	47.4	153	172.3	78.1	213	239.8	109.5
34	38.3	18.0	94	105.8	47.9	154	173.4	78.6	214	240.9	110.0
35	39.4	18.5	95	107.0	48.4	155	174.5	79.1	215	242.1	110.6
36	40.5	18.9	96	108.1	48.9	156	175.6	79.6	216	243.2	111.1
37	41.7	19.4	97	109.2	49.4	157	176.8	80.1	217	244.3	111.6
38	42.8	19.9	98	110.3	49.9	158	177.9	80.7	218	245.4	112.1
39	43.9	20.4	99	111.5	50.4	159	179.0	81.2	219	246.6	112.7
40	45.0	20.9	100	112.6	50.9	160	180.1	81.7	220	247.7	113.2
41	46.2	21.4	101	113.7	51.4	161	181.3	82.2	221	248.7	113.7
42	47.3	21.9	102	114.8	51.9	162	182.4	82.7	222	249.9	114.3
43	48.4	22.4	103	116.0	52.4	163	183.5	83.3	223	251.0	114.8
44	49.5	22.9	104	117.1	52.9	164	184.6	83.8	224	252.4	115.3
45	50.7	23.4	105	118.2	53.5	165	185.8	84.3	225	253.3	115.9
46	51.8	23.9	106	119.3	54.0	166	186.9	84.8	226	254.4	116.4
47	52.9	24.4	107	120.5	54.5	167	188.0	85.3	227	255.6	116.9
48	54.0	24.9	108	121.6	55.0	168	189.1	85.9	228	256.7	117.4
49	55.2	25.4	109	122.7	55.5	169	190.3	86.4	229	257.8	118.0
50	56.3	25.9	110	123.8	56.0	170	191.4	86.9	230	258.9	118.5
51	57.4	26.4	111	125.0	56.5	171	192.5	87.4	231	260.1	119.0
52	58.5	26.9	112	126.1	57.0	172	193.6	87.9	232	261.2	119.6
53	59.7	27.4	113	127.2	57.5	173	194.8	88.5	233	262.3	120.1
54	60.8	27.9	114	128.3	58.0	174	195.9	89.0	234	263.4	120.7
55	61.9	28.4	115	129.6	58.6	175	197.0	89.5	235	264.6	121.2
56	63.0	28.8	116	130.6	59.1	176	198.1	90.0	236	265.7	121.7
57	64.2	29.3	117	131.7	59.6	177	199.3	90.5	237	266.8	122.3
58	65.3	29.8	118	132.8	60.1	178	200.4	91.1	238	268.0	122.8
59	66.4	30.3	119	134.0	60.6	179	201.5	91.6	239	269.1	123.4
60	67.6	30.8	120	135.1	61.1	180	202.6	92.1	240	270.2	123.9
61	68.7	31.3	121	136.2	61.6	181	203.8	92.6	241	271.3	124.4
62	69.8	31.8	122	137.4	62.1	182	204.9	93.1	242	272.5	125.0
63	70.9	32.3	123	138.5	62.6	183	206.0	93.7	243	273.6	125.5
64	72.1	32.8	124	139.6	63.1	184	207.1	94.2	244	274.7	126.0
65	73.2	33.3	125	140.7	63.7	185	208.3	94.7	245	275.8	126.6
66	74.3	33.8	126	141.9	64.2	186	209.4	95.2	246	277.0	127.1
67	75.4	34.3	127	143.0	64.7	187	210.5	95.7	247	278.1	127.6
68	76.6	34.8	128	144.1	65.2	188	211.7	96.3	248	279.2	128.1
69	77.7	35.3	129	145.2	65.7	189	212.8	96.8	249	280.3	128.7
70	78.8	35.8	130	146.4	66.2	190	213.9	97.3	250	281.5	129.2

TABLE 7.—ALLIHN'S TABLE.—Continued.

[Expressed in milligrams.]

COPPER	CU- PROUS OXID	DEX- TROSE	COPPER	CU- PROUS OXID	DEX- TROSE	COPPER	CU- PROUS OXID	DEX- TROSE	COPPER	CU- PROUS OXID	DEX- TROSE
251	282.6	129.7	306	344.5	159.8	361	406.4	190.6	416	468.4	222.2
252	283.7	130.3	307	345.6	160.4	362	407.6	191.1	417	469.5	222.8
253	284.8	130.8	308	346.8	160.9	363	408.7	191.7	418	470.6	223.3
254	286.0	131.4	309	347.9	161.5	364	409.8	192.3	419	471.8	223.9
255	287.1	131.9	310	349.0	162.0	365	410.9	192.9	420	472.9	224.5
256	288.2	132.4	311	350.1	162.6	366	412.1	193.4	421	474.0	225.1
257	289.3	133.0	312	351.3	163.1	367	413.2	194.0	422	475.6	225.7
258	290.5	133.5	313	352.4	163.7	368	414.3	194.6	423	476.2	226.3
259	291.6	134.1	314	353.5	164.2	369	415.4	195.1	424	477.4	226.9
260	292.7	134.6	315	354.6	164.8	370	416.6	195.7	425	478.5	227.5
261	293.8	135.1	316	355.8	165.3	371	417.7	196.3	426	479.6	228.0
262	295.0	135.7	317	356.9	165.9	372	418.8	196.8	427	480.7	228.6
263	296.1	136.2	318	358.0	166.4	373	420.0	197.4	428	481.9	229.2
264	297.2	136.8	319	359.1	167.0	374	421.1	198.0	429	483.0	229.8
265	298.3	137.3	320	360.3	167.5	375	422.2	198.6	430	484.1	230.4
266	299.5	137.8	321	361.4	168.1	376	423.3	199.1	431	485.3	231.0
267	300.6	138.4	322	362.5	168.6	377	424.5	199.7	432	486.4	231.6
268	301.7	138.9	323	363.7	169.2	378	425.6	200.3	433	487.5	232.2
269	302.8	139.5	324	364.8	169.7	379	426.7	200.8	434	488.6	232.8
270	304.0	140.0	325	365.9	170.3	380	427.8	201.4	435	489.7	233.4
271	305.1	140.6	326	367.0	170.9	381	429.0	202.0	436	490.9	233.9
272	306.2	141.1	327	368.2	171.4	382	430.1	202.5	437	492.0	234.5
273	307.3	141.7	328	369.3	172.0	383	431.2	203.1	438	493.1	235.1
274	308.5	142.2	329	370.4	172.5	384	432.3	203.7	439	494.3	235.7
275	309.6	142.8	330	371.5	173.1	385	433.5	204.3	440	495.4	236.3
276	310.7	143.3	331	372.7	173.7	386	434.6	204.8	441	496.5	236.9
277	311.9	143.9	332	373.8	174.2	387	435.7	205.4	442	497.6	237.5
278	313.0	144.4	333	374.9	174.8	388	436.8	206.0	443	498.8	238.1
279	314.1	145.0	334	376.0	175.3	389	438.0	206.5	444	499.9	238.7
280	315.2	145.5	335	377.2	175.9	390	439.1	207.1	445	501.0	239.3
281	316.4	146.1	336	378.3	176.5	391	440.2	207.7	446	502.1	239.8
282	317.5	146.6	337	379.4	177.0	392	441.3	208.3	447	503.2	240.4
283	318.6	147.2	338	380.5	177.6	393	442.4	208.8	448	504.4	241.0
284	319.7	147.7	339	381.7	178.1	394	443.6	209.4	449	505.5	241.6
285	320.9	148.3	340	382.8	178.7	395	444.7	210.0	450	506.6	242.2
286	322.0	148.8	341	383.9	179.3	396	445.9	210.6	451	507.8	242.8
287	323.1	149.4	342	385.0	179.8	397	447.0	211.2	452	508.9	243.4
288	324.2	149.9	343	386.2	180.4	398	448.1	211.7	453	510.0	244.0
289	325.4	150.5	344	387.3	180.9	399	449.2	212.3	454	511.1	244.6
290	326.5	151.0	345	388.4	181.5	400	450.3	212.9	455	512.3	245.2
291	327.4	151.6	346	389.6	182.1	401	451.5	213.5	456	513.4	245.7
292	328.7	152.1	347	390.7	182.6	402	452.6	214.1	457	514.5	246.3
293	329.9	152.7	348	391.8	183.2	403	453.7	214.6	458	515.6	246.9
294	331.0	153.2	349	392.9	183.7	404	454.8	215.2	459	516.8	247.5
295	332.1	153.8	350	394.0	184.3	405	456.0	215.8	460	517.9	248.1
296	333.3	154.3	351	395.2	184.9	406	457.1	216.4	461	519.0	248.7
297	334.4	154.9	352	396.3	185.4	407	458.2	217.0	462	520.1	249.3
298	335.5	155.4	353	397.4	186.0	408	459.4	217.5	463	521.3	249.9
299	336.6	156.0	354	398.6	186.6	409	460.5	218.1			
300	337.8	156.5	355	399.7	187.2	410	461.6	218.7			
301	338.9	157.1	356	400.8	187.7	411	462.7	219.3			
302	340.0	157.6	357	401.9	188.3	412	463.8	219.9			
303	341.1	158.2	358	403.1	188.9	413	465.0	220.4			
304	342.3	158.7	359	404.2	189.4	414	466.1	221.0			
305	343.4	159.3	360	405.3	190.0	415	467.2	221.6			

56

REDUCING SUGARS OTHER THAN DEXTROSE.

Proceed as directed under 54 and multiply the weight of dextrose found in 55 by the following factors:

Levulose,	1.093;
Invert sugar,	1.046;
Arabinose,	0.969;
Xylose,	1.017;
Galactose,	1.114.

TOTAL SUGARS.¹¹

(Applicable to cattle foods.)

57

PREPARATION OF SOLUTION.

Place 12 grams of the material in a 300 cc. graduated flask, if the substance has an acid reaction add 1-3 grams of calcium carbonate, and boil on a steam bath for 1 hour with 150 cc. of 50% alcohol by volume, using a small funnel in the neck of the flask to condense the vapor. Cool, and allow the mixture to stand several hours, preferably overnight. Make up to volume with neutral 95% alcohol, mix thoroughly, allow to settle, transfer 200 cc. to a beaker with a pipette, and evaporate on a steam bath to a volume of 20-30 cc.

Do not evaporate to dryness, a little alcohol in the residue doing no harm. Transfer to a 100 cc. graduated flask, and rinse the beaker thoroughly with water, adding the rinsings to the contents of the flask. Add enough saturated neutral lead acetate solution to produce a flocculent precipitate, shake thoroughly and allow to stand 15 minutes. Make up to the mark with water, mix thoroughly, and filter through a dry filter. Add sufficient anhydrous sodium carbonate to the filtrate to precipitate all the lead, again filter through a dry paper and test the filtrate with a little anhydrous sodium carbonate to make sure that all the lead has been removed.

58

DETERMINATION OF REDUCING SUGARS.

Proceed as directed under 26 or 29-34 respectively, employing the Soxhlet modification of Fehling's solution and using 25 cc. of the solution (representing 2 grams of the sample), prepared as directed in 57. Express the results as dextrose or invert sugar.

59

SUCROSE.

Introduce 50 cc. of the solution, prepared as directed in 57, into a 100 cc. graduated flask, add a piece of litmus paper, neutralize with acetic acid, add 5 cc. of concentrated hydrochloric acid and allow the inversion to proceed at room temperature as directed under 14 or 16. When inversion is complete, transfer the solution to a beaker, neutralize with sodium carbonate, return the solution to the 100 cc. flask, dilute to the mark with water, filter if necessary and determine reducing sugars in 50 cc. of the solution (representing 2 grams of the sample) as directed in 58, and calculate the results as invert sugar. Subtract the per cent of reducing sugars before inversion from the per cent of total sugar after inversion, both calculated as invert sugar, and multiply the difference by 0.95 to obtain the per cent of sucrose present.

Since the insoluble material of grain or cattle food occupies some space in the flask as originally made up, it is necessary to correct for this volume. Results of a large number of determinations on various materials have shown the average volume of 12 grams of material to be 9 cc., and therefore to obtain the true amount of sugars present all results must be multiplied by the factor 0.97.

STARCH.

60 *Direct Acid Hydrolysis (Modified Sachsse Method).—Official.*

(In this method there will be included as starch the pentosans and other carbohydrate bodies present which undergo hydrolysis and conversion into reducing sugars on boiling with hydrochloric acid.)

Stir a quantity of the sample, representing 2.5–3 grams of the dry material, in a beaker with 50 cc. of cold water for an hour. Transfer to a filter and wash with 250 cc. of cold water. Heat the insoluble residue for $2\frac{1}{2}$ hours with 200 cc. of water and 20 cc. of hydrochloric acid (sp. gr. 1.125) in a flask provided with a reflux condenser. Cool, and nearly neutralize with sodium hydroxid. Complete the volume to 250 cc., filter, and determine the dextrose in an aliquot of the filtrate as directed under **52** or **54**. The weight of the dextrose obtained multiplied by 0.90 gives the weight of starch.

The factor 0.9 is the theoretical ratio between starch and glucose but, according to Noyes¹² and other investigators, the factor 0.93 more nearly approaches the actual yield.

*Diastase Method with Subsequent Acid Hydrolysis.—Tentative.***61**

REAGENT.

Malt extract.—Digest 10 grams of fresh, finely ground malt for 2–3 hours at ordinary temperature with 200 cc. of water and filter. Determine the amount of dextrose in a given quantity of the filtrate after boiling with acid, etc., as in the starch determination, and make the proper correction in the subsequent determination.

62

DETERMINATION.

Extract a convenient quantity of the substance (ground to an impalpable powder and representing 4–5 grams of the dry material) on a hardened filter with 5 successive portions of 10 cc. of ether; wash with 150 cc. of 10% alcohol and then with a little strong alcohol. Place the residue in a beaker with 50 cc. of water, immerse the beaker in boiling water, and stir constantly for 15 minutes or until all the starch is gelatinized; cool to 55°C., add 20 cc. of malt extract, and maintain at this temperature for an hour. Heat again to boiling for a few minutes, cool to 55°C., add 20 cc. of malt extract, and maintain at this temperature for an hour or until the residue treated with iodine shows no blue color upon microscopic examination. Cool, make up directly to 250 cc., and filter. Place 200 cc. of the filtrate in a flask with 20 cc. of hydrochloric acid (sp. gr. 1.125); connect with a reflux condenser and heat in a boiling water bath for $2\frac{1}{2}$ hours. Cool, nearly neutralize with sodium hydroxid solution, finish the neutralization with sodium carbonate solution, and make up to 500 cc. Mix the solution well, pour through a dry filter, and determine the dextrose in an aliquot as directed under **52** or **54**. Conduct a blank determination upon the same volume of the malt extract as used upon the sample and correct the weight of reduced copper accordingly. The weight of the dextrose obtained multiplied by 0.90 gives the weight of starch.

PENTOSANS.—TENTATIVE.

63

REAGENT.

Phloroglucin.—Dissolve a small quantity of the phloroglucin in a few drops of acetic anhydride, heat almost to boiling, and add a few drops of concentrated sul-

phuric acid. A violet color indicates the presence of diresorcin. A phloroglucin which gives more than a faint coloration may be purified by the following method:

Heat in a beaker about 300 cc. of hydrochloric acid (sp. gr. 1.06) and 11 grams of commercial phloroglucin, added in small quantities at a time, stirring constantly until it has almost entirely dissolved. Pour the hot solution into a sufficient quantity of the same hydrochloric acid (cold) to make the volume 1500 cc. Allow it to stand at least overnight, preferably several days, to permit the diresorcin to crystallize out. Filter immediately before using. A yellow tint does not interfere with its usefulness. In using it, add the volume containing the required amount to the distillate.

64

DETERMINATION.

Place a quantity of the material, 2-5 grams, chosen so that the weight of phloroglucin obtained shall not exceed 0.300 gram, in a 300 cc. distillation flask, together with 100 cc. of 12% hydrochloric acid (sp. gr. 1.06), and several pieces of recently heated pumice stone. Place the flask on a wire gauze, connect with a condenser, and heat, rather gently at first, and regulate so as to distil over 30 cc. in about 10 minutes, the distillate passing through a small filter paper. Replace the 30 cc. distilled by a like quantity of the dilute acid, added by means of a separatory funnel in such a manner as to wash down the particles adhering to the sides of the flask, and continue the process until the distillate amounts to 360 cc. To the total distillate add gradually a quantity of phloroglucin dissolved in 12% hydrochloric acid and stir thoroughly the resulting mixture. The amount of phloroglucin used should be about double that of the furfural expected. The solution turns first yellow, then green, and very soon an amorphous greenish precipitate appears, which grows darker rapidly, till it becomes finally almost black. Make the solution up to 400 cc. with 12% hydrochloric acid, and allow to stand overnight.

Filter the amorphous black precipitate into a tared Gooch crucible through an asbestos mat, wash carefully with 150 cc. of water in such a way that the water is not entirely removed from the crucible until the very last, then dry for 4 hours at the temperature of boiling water, cool and weigh in a weighing bottle, the increase in weight being reckoned as furfural phloroglucid. To calculate the furfural, pentose, or pentosan from the phloroglucid, use the following formulas given by Kröber:

(1) For a weight of phloroglucid, designated by "a" in the following formulas, under 0.03 gram,

$$\text{Furfural} = (a + 0.0052) \times 0.5170.$$

$$\text{Pentoses} = (a + 0.0052) \times 1.0170.$$

$$\text{Pentosans} = (a + 0.0052) \times 0.8949.$$

In the above and also in the following formulas, the factor 0.0052 represents the weight of phloroglucid which remains dissolved in the 400 cc. of acid solution.

(2) For a weight of phloroglucid "a" over 0.300 gram,

$$\text{Furfural} = (a + 0.0052) \times 0.5180.$$

$$\text{Pentoses} = (a + 0.0052) \times 1.0026.$$

$$\text{Pentosans} = (a + 0.0052) \times 0.8824.$$

For a weight of phloroglucid "a" between 0.03 and 0.300 gram use Kröber's table, 65, or the following formulas in which the factors were calculated from Kröber's tables by C. A. Browne,¹³

$$\text{Furfural} = (a + 0.0052) \times 0.5185.$$

$$\text{Pentoses} = (a + 0.0052) \times 1.0075.$$

$$\text{Pentosans} = (a + 0.0052) \times 0.8866.$$

TABLE 8.—KRÖBER'S TABLE.¹⁴
For Determining Pentoses and Pentosans.

[Expressed in grams.]

FURFURAL PHLOROGLUCID	FURFURAL	ARABINOSE	ARABAN	XYLOSE	XYLAN	PENTOSE	PENTOSAN
0.030	0.0182	0.0391	0.0344	0.0324	0.0285	0.0358	0.0315
0.031	0.0188	0.0402	0.0354	0.0333	0.0293	0.0368	0.0324
0.032	0.0193	0.0413	0.0363	0.0342	0.0301	0.0378	0.0333
0.033	0.0198	0.0424	0.0373	0.0352	0.0309	0.0388	0.0341
0.034	0.0203	0.0435	0.0383	0.0361	0.0317	0.0398	0.0350
0.035	0.0209	0.0446	0.0393	0.0370	0.0326	0.0408	0.0359
0.036	0.0214	0.0457	0.0402	0.0379	0.0334	0.0418	0.0368
0.037	0.0219	0.0468	0.0412	0.0388	0.0342	0.0428	0.0377
0.038	0.0224	0.0479	0.0422	0.0398	0.0350	0.0439	0.0386
0.039	0.0229	0.0490	0.0431	0.0407	0.0358	0.0449	0.0395
0.040	0.0235	0.0501	0.0441	0.0416	0.0366	0.0459	0.0404
0.041	0.0240	0.0512	0.0451	0.0425	0.0374	0.0469	0.0413
0.042	0.0245	0.0523	0.0460	0.0434	0.0382	0.0479	0.0422
0.043	0.0250	0.0534	0.0470	0.0443	0.0390	0.0489	0.0431
0.044	0.0255	0.0545	0.0480	0.0452	0.0398	0.0499	0.0440
0.045	0.0260	0.0556	0.0490	0.0462	0.0406	0.0509	0.0448
0.046	0.0266	0.0567	0.0499	0.0471	0.0414	0.0519	0.0457
0.047	0.0271	0.0578	0.0509	0.0480	0.0422	0.0529	0.0466
0.048	0.0276	0.0589	0.0519	0.0489	0.0430	0.0539	0.0475
0.049	0.0281	0.0600	0.0528	0.0498	0.0438	0.0549	0.0484
0.050	0.0286	0.0611	0.0538	0.0507	0.0446	0.0559	0.0492
0.051	0.0292	0.0622	0.0548	0.0516	0.0454	0.0569	0.0501
0.052	0.0297	0.0633	0.0557	0.0525	0.0462	0.0579	0.0510
0.053	0.0302	0.0644	0.0567	0.0534	0.0470	0.0589	0.0519
0.054	0.0307	0.0655	0.0576	0.0543	0.0478	0.0599	0.0528
0.055	0.0312	0.0666	0.0586	0.0553	0.0486	0.0610	0.0537
0.056	0.0318	0.0677	0.0596	0.0562	0.0494	0.0620	0.0546
0.057	0.0323	0.0688	0.0605	0.0571	0.0502	0.0630	0.0555
0.058	0.0328	0.0699	0.0615	0.0580	0.0510	0.0640	0.0564
0.059	0.0333	0.0710	0.0624	0.0589	0.0518	0.0650	0.0573
0.060	0.0338	0.0721	0.0634	0.0598	0.0526	0.0660	0.0581
0.061	0.0344	0.0732	0.0644	0.0607	0.0534	0.0670	0.0590
0.062	0.0349	0.0743	0.0653	0.0616	0.0542	0.0680	0.0599
0.063	0.0354	0.0754	0.0663	0.0626	0.0550	0.0690	0.0608
0.064	0.0359	0.0765	0.0673	0.0635	0.0558	0.0700	0.0617
0.065	0.0364	0.0776	0.0683	0.0644	0.0567	0.0710	0.0625
0.066	0.0370	0.0787	0.0692	0.0653	0.0575	0.0720	0.0634
0.067	0.0375	0.0798	0.0702	0.0662	0.0583	0.0730	0.0643
0.068	0.0380	0.0809	0.0712	0.0672	0.0591	0.0741	0.0652
0.069	0.0385	0.0820	0.0721	0.0681	0.0599	0.0751	0.0661
0.070	0.0390	0.0831	0.0731	0.0690	0.0607	0.0761	0.0670
0.071	0.0396	0.0842	0.0741	0.0699	0.0615	0.0771	0.0679
0.072	0.0401	0.0853	0.0750	0.0708	0.0623	0.0781	0.0688
0.073	0.0406	0.0864	0.0760	0.0717	0.0631	0.0791	0.0697
0.074	0.0411	0.0875	0.0770	0.0726	0.0639	0.0801	0.0706

TABLE 8.—KRÖBER'S TABLE.—Continued.

[Expressed in grams.]

FURFURAL PHLOROGUCID	FURFURAL	ARABINOSE	ARABAN	XYLOSE	XYLAN	PENTOSE	PENTOSAN
0.075	0.0416	0.0886	0.0780	0.0736	0.0647	0.0811	0.0714
0.076	0.0422	0.0897	0.0789	0.0745	0.0655	0.0821	0.0722
0.077	0.0427	0.0908	0.0799	0.0754	0.0663	0.0831	0.0731
0.078	0.0432	0.0919	0.0809	0.0763	0.0671	0.0841	0.0740
0.079	0.0437	0.0930	0.0818	0.0772	0.0679	0.0851	0.0749
0.080	0.0442	0.0941	0.0828	0.0781	0.0687	0.0861	0.0758
0.081	0.0448	0.0952	0.0838	0.0790	0.0695	0.0871	0.0767
0.082	0.0453	0.0963	0.0847	0.0799	0.0703	0.0881	0.0776
0.083	0.0458	0.0974	0.0857	0.0808	0.0711	0.0891	0.0785
0.084	0.0463	0.0985	0.0867	0.0817	0.0719	0.0901	0.0794
0.085	0.0468	0.0996	0.0877	0.0827	0.0727	0.0912	0.0803
0.086	0.0474	0.1007	0.0886	0.0836	0.0735	0.0922	0.0812
0.087	0.0479	0.1018	0.0896	0.0845	0.0743	0.0932	0.0821
0.088	0.0484	0.1029	0.0906	0.0854	0.0751	0.0942	0.0830
0.089	0.0489	0.1040	0.0915	0.0863	0.0759	0.0952	0.0838
0.090	0.0494	0.1051	0.0925	0.0872	0.0767	0.0962	0.0847
0.091	0.0499	0.1062	0.0935	0.0881	0.0775	0.0972	0.0856
0.092	0.0505	0.1073	0.0944	0.0890	0.0783	0.0982	0.0865
0.093	0.0510	0.1084	0.0954	0.0900	0.0791	0.0992	0.0874
0.094	0.0515	0.1095	0.0964	0.0909	0.0800	0.1002	0.0883
0.095	0.0520	0.1106	0.0974	0.0918	0.0808	0.1012	0.0891
0.096	0.0525	0.1117	0.0983	0.0927	0.0816	0.1022	0.0899
0.097	0.0531	0.1128	0.0993	0.0936	0.0824	0.1032	0.0908
0.098	0.0536	0.1139	0.1003	0.0946	0.0832	0.1043	0.0917
0.099	0.0541	0.1150	0.1012	0.0955	0.0840	0.1053	0.0926
0.100	0.0546	0.1161	0.1022	0.0964	0.0848	0.1063	0.0935
0.101	0.0551	0.1171	0.1032	0.0973	0.0856	0.1073	0.0944
0.102	0.0557	0.1182	0.1041	0.0982	0.0864	0.1083	0.0953
0.103	0.0562	0.1193	0.1051	0.0991	0.0872	0.1093	0.0962
0.104	0.0567	0.1204	0.1060	0.1000	0.0880	0.1103	0.0971
0.105	0.0572	0.1215	0.1070	0.1010	0.0888	0.1113	0.0979
0.106	0.0577	0.1226	0.1080	0.1019	0.0896	0.1123	0.0988
0.107	0.0582	0.1237	0.1089	0.1028	0.0904	0.1133	0.0997
0.108	0.0588	0.1248	0.1099	0.1037	0.0912	0.1143	0.1006
0.109	0.0593	0.1259	0.1108	0.1046	0.0920	0.1153	0.1015
0.110	0.0598	0.1270	0.1118	0.1055	0.0928	0.1163	0.1023
0.111	0.0603	0.1281	0.1128	0.1064	0.0936	0.1173	0.1032
0.112	0.0608	0.1292	0.1137	0.1073	0.0944	0.1183	0.1041
0.113	0.0614	0.1303	0.1147	0.1082	0.0952	0.1193	0.1050
0.114	0.0619	0.1314	0.1156	0.1091	0.0960	0.1203	0.1059
0.115	0.0624	0.1325	0.1166	0.1101	0.0968	0.1213	0.1067
0.116	0.0629	0.1336	0.1176	0.1110	0.0976	0.1223	0.1076
0.117	0.0634	0.1347	0.1185	0.1119	0.0984	0.1233	0.1085
0.118	0.0640	0.1358	0.1195	0.1128	0.0992	0.1243	0.1094
0.119	0.0645	0.1369	0.1204	0.1137	0.1000	0.1253	0.1103

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TABLE 8.—KRÖBER'S TABLE.—Continued.

[Expressed in grams.]

FURFURAL PHLOROGLUCID	FURFURAL	ARABINOSE	ARABAN	XYLOSE	XYLAN	PENTOSE	PENTOSAN
0.120	0.0650	0.1380	0.1214	0.1146	0.1008	0.1263	0.1111
0.121	0.0655	0.1391	0.1224	0.1155	0.1016	0.1273	0.1120
0.122	0.0660	0.1402	0.1233	0.1164	0.1024	0.1283	0.1129
0.123	0.0665	0.1413	0.1243	0.1173	0.1032	0.1293	0.1138
0.124	0.0671	0.1424	0.1253	0.1182	0.1040	0.1303	0.1147
0.125	0.0676	0.1435	0.1263	0.1192	0.1049	0.1314	0.1156
0.126	0.0681	0.1446	0.1272	0.1201	0.1057	0.1324	0.1165
0.127	0.0686	0.1457	0.1282	0.1210	0.1065	0.1334	0.1174
0.128	0.0691	0.1468	0.1292	0.1219	0.1073	0.1344	0.1183
0.129	0.0697	0.1479	0.1301	0.1228	0.1081	0.1354	0.1192
0.130	0.0702	1490	0.1311	0.1237	0.1089	0.1364	0.1201
0.131	0.0707	0.1501	0.1321	0.1246	0.1097	0.1374	0.1210
0.132	0.0712	0.1512	0.1330	0.1255	0.1105	0.1384	0.1219
0.133	0.0717	0.1523	0.1340	0.1264	0.1113	0.1394	0.1227
0.134	0.0723	0.1534	0.1350	0.1273	0.1121	0.1404	0.1236
0.135	0.0728	0.1545	0.1360	0.1283	0.1129	0.1414	0.1244
0.136	0.0733	0.1556	0.1369	0.1292	0.1137	0.1424	0.1253
0.137	0.0738	0.1567	0.1379	0.1301	0.1145	0.1434	0.1262
0.138	0.0743	0.1578	0.1389	0.1310	0.1153	0.1444	0.1271
0.139	0.0748	0.1589	0.1398	0.1319	0.1161	0.1454	0.1280
0.140	0.0754	0.1600	0.1408	0.1328	0.1169	0.1464	0.1288
0.141	0.0759	0.1611	0.1418	0.1337	0.1177	0.1474	0.1297
0.142	0.0764	0.1622	0.1427	0.1346	0.1185	0.1484	0.1306
0.143	0.0769	0.1633	0.1437	0.1355	0.1193	0.1494	0.1315
0.144	0.0774	0.1644	0.1447	0.1364	0.1201	0.1504	0.1324
0.145	0.0780	0.1655	0.1457	0.1374	0.1209	0.1515	0.1333
0.146	0.0785	0.1666	0.1466	0.1383	0.1217	0.1525	0.1342
0.147	0.0790	0.1677	0.1476	0.1392	0.1225	0.1535	0.1351
0.148	0.0795	0.1688	0.1486	0.1401	0.1233	0.1545	0.1360
0.149	0.0800	0.1699	0.1495	0.1410	0.1241	0.1555	0.1369
0.150	0.0805	0.1710	0.1505	0.1419	0.1249	0.1565	0.1377
0.151	0.0811	0.1721	0.1515	0.1428	0.1257	0.1575	0.1386
0.152	0.0816	0.1732	0.1524	0.1437	0.1265	0.1585	0.1395
0.153	0.0821	0.1743	0.1534	0.1446	0.1273	0.1595	0.1404
0.154	0.0826	0.1754	0.1544	0.1455	0.1281	0.1605	0.1413
0.155	0.0831	0.1765	0.1554	0.1465	0.1289	0.1615	0.1421
0.156	0.0837	0.1776	0.1563	0.1474	0.1297	0.1625	0.1430
0.157	0.0842	0.1787	0.1573	0.1483	0.1305	0.1635	0.1439
0.158	0.0847	0.1798	0.1583	0.1492	0.1313	0.1645	0.1448
0.159	0.0852	0.1809	0.1592	0.1501	0.1321	0.1655	0.1457
0.160	0.0857	0.1820	0.1602	0.1510	0.1329	0.1665	0.1465
0.161	0.0863	0.1831	0.1612	0.1519	0.1337	0.1675	0.1474
0.162	0.0868	0.1842	0.1621	0.1528	0.1345	0.1685	0.1483
0.163	0.0873	0.1853	0.1631	0.1537	0.1353	0.1695	0.1492
0.164	0.0878	0.1864	0.1640	0.1546	0.1361	0.1705	0.1501

TABLE 8.—KRÖBER'S TABLE.—Continued.

[Expressed in grams.]

FURFURAL PHLOROGLUCID	FURFURAL	ARABINOSE	ARABAN	XYLOSE	XYLAN	PENTOSE	PENTOSAN
0.165	0.0883	0.1875	0.1650	0.1556	0.1369	0.1716	0.1510
0.166	0.0888	0.1886	0.1660	0.1565	0.1377	0.1726	0.1519
0.167	0.0894	0.1897	0.1669	0.1574	0.1385	0.1736	0.1528
0.168	0.0899	0.1908	0.1679	0.1583	0.1393	0.1746	0.1537
0.169	0.0904	0.1919	0.1688	0.1592	0.1401	0.1756	0.1546
0.170	0.0909	0.1930	0.1698	0.1601	0.1409	0.1766	0.1554
0.171	0.0914	0.1941	0.1708	0.1610	0.1417	0.1776	0.1563
0.172	0.0920	0.1952	0.1717	0.1619	0.1425	0.1786	0.1572
0.173	0.0925	0.1963	0.1727	0.1628	0.1433	0.1796	0.1581
0.174	0.0930	0.1974	0.1736	0.1637	0.1441	0.1806	0.1590
0.175	0.0935	0.1985	0.1746	0.1647	0.1449	0.1816	0.1598
0.176	0.0940	0.1996	0.1756	0.1656	0.1457	0.1826	0.1607
0.177	0.0946	0.2007	0.1765	0.1665	0.1465	0.1836	0.1616
0.178	0.0951	0.2018	0.1775	0.1674	0.1473	0.1846	0.1625
0.179	0.0956	0.2029	0.1784	0.1683	0.1481	0.1856	0.1634
0.180	0.0961	0.2039	0.1794	0.1692	0.1489	0.1866	0.1642
0.181	0.0966	0.2050	0.1804	0.1701	0.1497	0.1876	0.1651
0.182	0.0971	0.2061	0.1813	0.1710	0.1505	0.1886	0.1660
0.183	0.0977	0.2072	0.1823	0.1719	0.1513	0.1896	0.1669
0.184	0.0982	0.2082	0.1832	0.1728	0.1521	0.1906	0.1678
0.185	0.0987	0.2093	0.1842	0.1738	0.1529	0.1916	0.1686
0.186	0.0992	0.2104	0.1851	0.1747	0.1537	0.1926	0.1695
0.187	0.0997	0.2115	0.1861	0.1756	0.1545	0.1936	0.1704
0.188	0.1003	0.2126	0.1870	0.1765	0.1553	0.1946	0.1712
0.189	0.1008	0.2136	0.1880	0.1774	0.1561	0.1955	0.1721
0.190	0.1013	0.2147	0.1889	0.1783	0.1569	0.1965	0.1729
0.191	0.1018	0.2158	0.1899	0.1792	0.1577	0.1975	0.1738
0.192	0.1023	0.2168	0.1908	0.1801	0.1585	0.1985	0.1747
0.193	0.1028	0.2179	0.1918	0.1810	0.1593	0.1995	0.1756
0.194	0.1034	0.2190	0.1927	0.1819	0.1601	0.2005	0.1764
0.195	0.1039	0.2201	0.1937	0.1829	0.1609	0.2015	0.1773
0.196	0.1044	0.2212	0.1946	0.1838	0.1617	0.2025	0.1782
0.197	0.1049	0.2222	0.1956	0.1847	0.1625	0.2035	0.1791
0.198	0.1054	0.2233	0.1965	0.1856	0.1633	0.2045	0.1800
0.199	0.1059	0.2244	0.1975	0.1865	0.1641	0.2055	0.1808
0.200	0.1065	0.2255	0.1984	0.1874	0.1649	0.2065	0.1817
0.201	0.1070	0.2266	0.1994	0.1883	0.1657	0.2075	0.1826
0.202	0.1075	0.2276	0.2003	0.1892	0.1665	0.2085	0.1835
0.203	0.1080	0.2287	0.2013	0.1901	0.1673	0.2095	0.1844
0.204	0.1085	0.2298	0.2022	0.1910	0.1681	0.2105	0.1853
0.205	0.1090	0.2309	0.2032	0.1920	0.1689	0.2115	0.1861
0.206	0.1096	0.2320	0.2041	0.1929	0.1697	0.2125	0.1869
0.207	0.1101	0.2330	0.2051	0.1938	0.1705	0.2134	0.1878
0.208	0.1106	0.2341	0.2060	0.1947	0.1713	0.2144	0.1887
0.209	0.1111	0.2352	0.2069	0.1956	0.1721	0.2154	0.1896

TABLE 8.—KRÖBER'S TABLE.—Continued.

[Expressed in grams.]

FURFURAL PHLOROGLUCID	FURFURAL	ARABINOSE	ARABAN	XYLOSE	XYLAN	PENTOSE	PENTOSAN
0.210	0.1116	0.2363	0.2079	0.1965	0.1729	0.2164	0.1904
0.211	0.1121	0.2374	0.2089	0.1975	0.1737	0.2174	0.1913
0.212	0.1127	0.2384	0.2098	0.1984	0.1745	0.2184	0.1922
0.213	0.1132	0.2395	0.2108	0.1993	0.1753	0.2194	0.1931
0.214	0.1137	0.2406	0.2117	0.2002	0.1761	0.2204	0.1940
0.215	0.1142	0.2417	0.2127	0.2011	0.1770	0.2214	0.1948
0.216	0.1147	0.2428	0.2136	0.2020	0.1778	0.2224	0.1957
0.217	0.1152	0.2438	0.2146	0.2029	0.1786	0.2234	0.1966
0.218	0.1158	0.2449	0.2155	0.2038	0.1794	0.2244	0.1974
0.219	0.1163	0.2460	0.2165	0.2047	0.1802	0.2254	0.1983
0.220	0.1168	0.2471	0.2174	0.2057	0.1810	0.2264	0.1992
0.221	0.1173	0.2482	0.2184	0.2066	0.1818	0.2274	0.2001
0.222	0.1178	0.2492	0.2193	0.2075	0.1826	0.2284	0.2010
0.223	0.1183	0.2503	0.2203	0.2084	0.1834	0.2294	0.2019
0.224	0.1189	0.2514	0.2212	0.2093	0.1842	0.2304	0.2028
0.225	0.1194	0.2525	0.2222	0.2102	0.1850	0.2314	0.2037
0.226	0.1199	0.2536	0.2232	0.2111	0.1858	0.2324	0.2046
0.227	0.1204	0.2546	0.2241	0.2121	0.1866	0.2334	0.2054
0.228	0.1209	0.2557	0.2251	0.2130	0.1874	0.2344	0.2063
0.229	0.1214	0.2568	0.2260	0.2139	0.1882	0.2354	0.2072
0.230	0.1220	0.2579	0.2270	0.2148	0.1890	0.2364	0.2081
0.231	0.1225	0.2590	0.2280	0.2157	0.1898	0.2374	0.2089
0.232	0.1230	0.2600	0.2289	0.2166	0.1906	0.2383	0.2097
0.233	0.1235	0.2611	0.2299	0.2175	0.1914	0.2393	0.2106
0.234	0.1240	0.2622	0.2308	0.2184	0.1922	0.2403	0.2115
0.235	0.1245	0.2633	0.2318	0.2193	0.1930	0.2413	0.2124
0.236	0.1251	0.2644	0.2327	0.2202	0.1938	0.2423	0.2132
0.237	0.1256	0.2654	0.2337	0.2211	0.1946	0.2433	0.2141
0.238	0.1261	0.2665	0.2346	0.2220	0.1954	0.2443	0.2150
0.239	0.1266	0.2676	0.2356	0.2229	0.1962	0.2453	0.2159
0.240	0.1271	0.2687	0.2365	0.2239	0.1970	0.2463	0.2168
0.241	0.1276	0.2698	0.2375	0.2248	0.1978	0.2473	0.2176
0.242	0.1281	0.2708	0.2384	0.2257	0.1986	0.2483	0.2185
0.243	0.1287	0.2719	0.2394	0.2266	0.1994	0.2493	0.2194
0.244	0.1292	0.2730	0.2403	0.2275	0.2002	0.2503	0.2203
0.245	0.1297	0.2741	0.2413	0.2284	0.2010	0.2513	0.2212
0.246	0.1302	0.2752	0.2422	0.2293	0.2018	0.2523	0.2220
0.247	0.1307	0.2762	0.2432	0.2302	0.2026	0.2533	0.2229
0.248	0.1312	0.2773	0.2441	0.2311	0.2034	0.2543	0.2238
0.249	0.1318	0.2784	0.2451	0.2320	0.2042	0.2553	0.2247
0.250	0.1323	0.2795	0.2460	0.2330	0.2050	0.2563	0.2256
0.251	0.1328	0.2806	0.2470	0.2339	0.2058	0.2573	0.2264
0.252	0.1333	0.2816	0.2479	0.2348	0.2066	0.2582	0.2272
0.253	0.1338	0.2827	0.2489	0.2357	0.2074	0.2592	0.2281
0.254	0.1343	0.2838	0.2498	0.2366	0.2082	0.2602	0.2290

TABLE 8.—KRÖBER'S TABLE.—Continued.

[Expressed in grams.]

FURFURAL PHLOROGLUCID	FURFURAL	ARABINOSE	ARABAN	XYLOSE	XYLAN	PENTOSE	PENTOSAN
0.255	0.1349	0.2849	0.2508	0.2375	0.2090	0.2612	0.2299
0.256	0.1354	0.2860	0.2517	0.2384	0.2098	0.2622	0.2307
0.257	0.1359	0.2870	0.2526	0.2393	0.2106	0.2632	0.2316
0.258	0.1364	0.2881	0.2536	0.2402	0.2114	0.2642	0.2325
0.259	0.1369	0.2892	0.2545	0.2411	0.2122	0.2652	0.2334
0.260	0.1374	0.2903	0.2555	0.2420	0.2130	0.2662	0.2342
0.261	0.1380	0.2914	0.2565	0.2429	0.2138	0.2672	0.2351
0.262	0.1385	0.2924	0.2574	0.2438	0.2146	0.2681	0.2359
0.263	0.1390	0.2935	0.2584	0.2447	0.2154	0.2691	0.2368
0.264	0.1395	0.2946	0.2593	0.2456	0.2162	0.2701	0.2377
0.265	0.1400	0.2957	0.2603	0.2465	0.2170	0.2711	0.2385
0.266	0.1405	0.2968	0.2612	0.2474	0.2178	0.2721	0.2394
0.267	0.1411	0.2978	0.2622	0.2483	0.2186	0.2731	0.2403
0.268	0.1416	0.2989	0.2631	0.2492	0.2194	0.2741	0.2412
0.269	0.1421	0.3000	0.2641	0.2502	0.2202	0.2751	0.2421
0.270	0.1426	0.3011	0.2650	0.2511	0.2210	0.2761	0.2429
0.271	0.1431	0.3022	0.2660	0.2520	0.2218	0.2771	0.2438
0.272	0.1436	0.3032	0.2669	0.2529	0.2226	0.2781	0.2447
0.273	0.1442	0.3043	0.2679	0.2538	0.2234	0.2791	0.2456
0.274	0.1447	0.3054	0.2688	0.2547	0.2242	0.2801	0.2465
0.275	0.1452	0.3065	0.2698	0.2556	0.2250	0.2811	0.2473
0.276	0.1457	0.3076	0.2707	0.2565	0.2258	0.2821	0.2482
0.277	0.1462	0.3086	0.2717	0.2574	0.2266	0.2830	0.2490
0.278	0.1467	0.3097	0.2726	0.2583	0.2274	0.2840	0.2499
0.279	0.1473	0.3108	0.2736	0.2592	0.2282	0.2850	0.2508
0.280	0.1478	0.3119	0.2745	0.2602	0.2290	0.2861	0.2517
0.281	0.1483	0.3130	0.2755	0.2611	0.2298	0.2871	0.2526
0.282	0.1488	0.3140	0.2764	0.2620	0.2306	0.2880	0.2534
0.283	0.1493	0.3151	0.2774	0.2629	0.2314	0.2890	0.2543
0.284	0.1498	0.3162	0.2783	0.2638	0.2322	0.2900	0.2552
0.285	0.1504	0.3173	0.2793	0.2647	0.2330	0.2910	0.2561
0.286	0.1509	0.3184	0.2802	0.2656	0.2338	0.2920	0.2570
0.287	0.1514	0.3194	0.2812	0.2665	0.2346	0.2930	0.2578
0.288	0.1519	0.3205	0.2821	0.2674	0.2354	0.2940	0.2587
0.289	0.1524	0.3216	0.2831	0.2683	0.2362	0.2950	0.2596
0.290	0.1529	0.3227	0.2840	0.2693	0.2370	0.2960	0.2605
0.291	0.1535	0.3238	0.2850	0.2702	0.2378	0.2970	0.2614
0.292	0.1540	0.3248	0.2859	0.2711	0.2386	0.2980	0.2622
0.293	0.1545	0.3259	0.2868	0.2720	0.2394	0.2990	0.2631
0.294	0.1550	0.3270	0.2878	0.2729	0.2402	0.3000	0.2640
0.295	0.1555	0.3281	0.2887	0.2738	0.2410	0.3010	0.2649
0.296	0.1560	0.3292	0.2897	0.2747	0.2418	0.3020	0.2658
0.297	0.1566	0.3302	0.2906	0.2756	0.2426	0.3030	0.2666
0.298	0.1571	0.3313	0.2916	0.2765	0.2434	0.3040	0.2675
0.299	0.1576	0.3324	0.2925	0.2774	0.2442	0.3050	0.2684
0.300	0.1581	0.3335	0.2935	0.2784	0.2450	0.3060	0.2693

66

GALACTAN.—TENTATIVE.

Extract a convenient quantity of the substance, representing 2.5-3 grams of the dry material, on a hardened filter with 5 successive portions of 10 cc. of ether, place the extracted residue in a beaker, about 5.5 cm. in diameter and 7 cm. deep, together with 60 cc. of nitric acid of 1.15 sp. gr., and evaporate the solution to exactly one third its volume in a water bath at a temperature of 94°-96°C. After standing 24 hours, add 10 cc. of water to the precipitate, and allow it to stand another 24 hours. The mucic acid has in the meantime crystallized, but it is mixed with considerable material only partially oxidized by the nitric acid. Filter the solution through filter paper, wash with 30 cc. of water to remove as much of the nitric acid as possible, and replace the filter and contents in the beaker. Add 30 cc. of ammonium carbonate solution, consisting of 1 part ammonium carbonate, 19 parts water, and 1 part strong ammonium hydroxid, and heat the mixture on a water bath, at 80°C., for 15 minutes, with constant stirring. The ammonium carbonate takes up the mucic acid, forming soluble ammonium mucate. Wash the filter paper and contents several times with hot water by decantation, passing the washings through a filter paper, to which finally transfer the material and thoroughly wash. Evaporate the filtrate to dryness over a water bath, avoiding unnecessary heating which causes decomposition, add 5 cc. of nitric acid of 1.15 sp. gr., stir thoroughly the mixture and allow to stand for 30 minutes. The nitric acid decomposes the ammonium mucate, precipitating the mucic acid; collect this on a tared filter or Gooch, wash with 10-15 cc. of water, then with 60 cc. of alcohol, and a number of times with ether, dry at the temperature of boiling water for 3 hours, and weigh. Multiply the weight of the mucic acid by 1.33, which gives galactose, and multiply this product by 0.9 which gives galactan.

CRUDE FIBER.—OFFICIAL.

67

REAGENTS.

- (a) 1.25% sulphuric acid solution.—Exact strength, determined by titration.
- (b) 1.25% sodium hydroxid solution.—Exact strength, determined by titration.

68

DETERMINATION.

Extract a quantity of the substance, representing about 2 grams of the dry material, with ordinary ether, or use the residue from the determination of the ether extract. To this residue in a 500 cc. flask add 200 cc. of boiling 1.25% sulphuric acid; connect the flask with an inverted condenser, the tube of which passes only a short distance beyond the rubber stopper into the flask, or simply cover a tall conical flask, which is well suited for this determination, with a watch glass or short stemmed funnel, boil at once and continue boiling gently for 30 minutes. A blast of air conducted into the flask will serve to reduce the frothing of the liquid. Filter through linen and wash with boiling water until the washings are no longer acid; rinse the substance back into the flask with 200 cc. of boiling, 1.25% solution of sodium hydroxid, free or nearly free from sodium carbonate boil at once, and continue boiling gently for 30 minutes as directed above for the treatment with acid, filter at once rapidly, and wash with boiling water until the washings are neutral. The last filtration may be performed upon a Gooch crucible, a linen filter, or a tared filter paper. If a linen filter is used, rinse the crude fiber, after washing is completed, into a flat-bottomed platinum dish by means of a jet of water; evaporate to dryness on a steam bath, dry to constant weight at 110°C., weigh, incinerate completely, and weigh again. The loss in weight is considered to be crude fiber. If a tared filter

paper is used, weigh in a weighing bottle. In any case the crude fiber after drying to constant weight at 110°C. must be incinerated and the amount of the ash deducted from the original weight.

69

WATER-SOLUBLE ACIDITY OF FEEDS.—TENTATIVE.

Weigh 10 grams of the sample into a shaking bottle, add 200 cc. of water, and shake for 15 minutes. Filter the extract through a folded filter and take a 20 cc. aliquot (equivalent to 1 gram of sample) for the titration. Dilute with 50 cc. of water and titrate with N/10 sodium hydroxid, using phenolphthalein as indicator.

In reporting the acidity of feeds, state the results in terms of cc. of N/10 sodium hydroxid required for neutralization.

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IX. SACCHARINE PRODUCTS.

1 PREPARATION OF SAMPLE.—TENTATIVE.

(a) *Liquids (molasses, sirups, etc.)*.—Mix materials of this class thoroughly. If crystals of sugar are present, dissolve them either by heating gently or by weighing the whole mass, then adding water, heating until completely dissolved and after cooling, re-weighing. Calculate all results to the weight of the original substance.

(b) *Semisolids (jellies, jams, etc.)*.—Weigh 50 grams of the sample into a 250 cc. graduated flask. Treat with water, fill to the mark and mix thoroughly. If insoluble material remains, mix uniformly by shaking before taking aliquots for the various determinations.

(c) *Solids (sugar, confectionery, etc.)*.—Grind and mix thoroughly materials of this class to secure uniform samples.

MOISTURE.

DRYING METHODS.

2 SUGARS.—OFFICIAL.

Dry 2-5 grams in a flat dish (nickel, platinum, or aluminium) at the temperature of boiling water for 10 hours; cool in a desiccator and weigh; then dry again for an hour or until there is only a slight change in weight.

With some sugars, more especially those of large grain, there is danger of occlusion and retention of water. The International Commission for Unifying Methods of Sugar Analysis prescribe drying at 105°-110°C. for normal beet sugars. This temperature is sufficient to expel the last traces of occluded water and is not attended with sufficient decomposition to affect the weight of the product. The drying temperature should never exceed 110°C¹.

MASSECUITES, MOLASSES, AND OTHER LIQUID AND SEMILIQUID PRODUCTS.

3 Drying upon Pumice Stone.—Tentative.

Prepare pumice stone of two grades of fineness, one of which will pass through a 1 mm. sieve, the other through a 6 mm. sieve. Make the determination in flat metallic dishes or in shallow, flat-bottomed, weighing bottles. Place a layer of the fine pumice stone, 3 mm. in thickness, on the bottom of the dish, then a layer of the coarse pumice stone 6-10 mm. in thickness, dry and weigh. Dilute the sample with a weighed portion of water so that the diluted material shall contain 20-30% of solid matter. Weigh into the dish, prepared as described above, an amount of the diluted sample to yield, approximately, 1 gram of dry matter. If this weighing can not be made rapidly, use a weighing bottle provided with a cork through which a pipette passes. Dry in vacuo at 70°C. to constant weight, making trial weighings at intervals of 2 hours. For substances containing little or no levulose or other readily decomposable substance, the drying may be made in a water oven at the temperature of boiling water.

4 Drying upon Quartz Sand.—Tentative.

Digest pure quartz sand with strong hydrochloric acid, wash, dry, and ignite. Preserve in a stoppered bottle.

Place 6-7 grams of the prepared sand and a short stirring rod in a flat-bottomed dish. Dry thoroughly, cool in a desiccator, and weigh. Then add 3-4 grams of the molasses, mix with the sand (if necessary to thoroughly incorporate the two, add a little water), dry in a water oven at the temperature of boiling water for 8-10 hours, stirring at intervals of an hour, cool in a desiccator, and weigh. Stir, heat again for an hour, cool, and weigh. Repeat the heating and weighing until the loss of water in an hour is not greater than 3 mg.

AREOMETRIC METHODS.

(Not applicable to low-grade sugar products, molasses and other materials containing large amounts of non-sugar solids.)

SPECIFIC GRAVITY, WATER AND TOTAL SOLIDS.

5

By Means of a Spindle.—Official.

The density of juices, sirups, etc., is most conveniently determined by means of the Brix hydrometer. For rough work, or where less accuracy is desired, the Baumé hydrometer may be used. The Brix spindle should be graduated to tenths. The range of degrees recorded by each individual spindle should be as limited as possible. The solution should be as nearly as practicable of the same temperature as the air at the time of reading, and, if the variation from the temperature of the graduation of the spindle amounts to more than 1°, a correction must be applied according to the table under 6. Before taking the density of a juice, allow it to stand in the cylinder until all air bubbles have escaped, and until all fatty or waxy matter has come to the surface and been skimmed off. The cylinder should be large enough in diameter to allow the hydrometer to come to rest without touching the sides. A table of specific gravities at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ and per cent by weight of sucrose is given under 9, and a table for the comparison of specific gravities at $\frac{17.5^{\circ}\text{C.}}{17.5^{\circ}}$, degrees Brix (per cent by weight of sucrose), and degree Baumé is given under 8.

If the sample is too dense to determine the density directly, dilute a weighed portion with a weighed quantity of water, or dissolve a weighed portion and dilute to a known volume with water.

In the first instance the per cent of total solids is calculated by the following formula:

Per cent of solids in the undiluted material = $\frac{WS}{w}$ in which

S = per cent of solids in the diluted material;

W = weight of the diluted material;

w = weight of the sample taken for dilution.

When the dilution is made to a definite volume, the following formula is to be used:

Per cent of solids in the undiluted material = $\frac{VDS}{W}$ in which

V = volume of the diluted solution at a given temperature;

D = specific gravity of the diluted solution at the same temperature;

S = per cent of solids in the diluted solution at the same temperature;

W = weight of the sample taken for dilution at the same temperature.

If the spindle reading be made at any other temperature than 17.5°C., the result should be corrected according to the following:

TABLE 9.

For correction of the readings of the Brix spindle when made at other than the standard temperature, 17.5°C.

(For temperatures below 17.5°C. the correction is to be subtracted.)

TEMPERATURE	DEGREE BRIX OF THE SOLUTION												
	0	5	10	15	20	25	30	35	40	50	60	70	75
°C.													
0	0.17	0.30	0.41	0.52	0.62	0.72	0.82	0.92	0.98	1.11	1.22	1.25	1.29
5	0.23	0.30	0.37	0.44	0.52	0.59	0.65	0.72	0.75	0.80	0.88	0.91	0.94
10	0.20	0.26	0.29	0.33	0.36	0.39	0.42	0.45	0.48	0.50	0.54	0.58	0.61
11	0.18	0.23	0.26	0.28	0.31	0.34	0.36	0.39	0.41	0.43	0.47	0.50	0.53
12	0.16	0.20	0.22	0.24	0.26	0.29	0.31	0.33	0.34	0.36	0.40	0.42	0.46
13	0.14	0.18	0.19	0.21	0.22	0.24	0.26	0.27	0.28	0.29	0.33	0.35	0.39
14	0.12	0.15	0.16	0.17	0.18	0.19	0.21	0.22	0.22	0.23	0.26	0.28	0.32
15	0.09	0.11	0.12	0.14	0.14	0.15	0.16	0.17	0.16	0.17	0.19	0.21	0.25
16	0.06	0.07	0.08	0.09	0.10	0.10	0.11	0.12	0.12	0.12	0.14	0.16	0.18
17	0.02	0.02	0.03	0.03	0.03	0.04	0.04	0.04	0.04	0.04	0.05	0.05	0.06
18	0.02	0.03	0.03	0.03	0.03	0.03	0.03	0.03	0.03	0.03	0.03	0.03	0.02
19	0.06	0.08	0.08	0.09	0.09	0.10	0.10	0.10	0.10	0.10	0.10	0.09	0.06
20	0.11	0.14	0.15	0.17	0.17	0.18	0.18	0.18	0.19	0.19	0.18	0.15	0.11
21	0.16	0.20	0.22	0.24	0.24	0.25	0.25	0.25	0.26	0.26	0.25	0.22	0.18
22	0.21	0.26	0.29	0.31	0.31	0.32	0.32	0.32	0.33	0.34	0.32	0.29	0.25
23	0.27	0.32	0.35	0.37	0.38	0.39	0.39	0.39	0.40	0.42	0.39	0.36	0.33
24	0.32	0.38	0.41	0.43	0.44	0.46	0.46	0.47	0.47	0.50	0.46	0.43	0.40
25	0.37	0.44	0.47	0.49	0.51	0.53	0.54	0.55	0.55	0.58	0.54	0.51	0.48
26	0.43	0.50	0.54	0.56	0.58	0.60	0.61	0.62	0.62	0.66	0.62	0.58	0.55
27	0.49	0.57	0.61	0.63	0.65	0.68	0.68	0.69	0.70	0.74	0.70	0.65	0.62
28	0.56	0.64	0.68	0.70	0.72	0.76	0.76	0.78	0.78	0.82	0.78	0.72	0.70
29	0.63	0.71	0.75	0.78	0.79	0.84	0.84	0.86	0.86	0.90	0.86	0.80	0.78
30	0.70	0.78	0.82	0.87	0.87	0.92	0.92	0.94	0.94	0.98	0.94	0.88	0.86
35	1.10	1.17	1.22	1.24	1.30	1.32	1.33	1.35	1.36	1.39	1.34	1.27	1.25
40	1.50	1.61	1.67	1.71	1.73	1.79	1.79	1.80	1.82	1.83	1.78	1.69	1.65
50	2.65	2.71	2.74	2.78	2.80	2.80	2.80	2.80	2.79	2.70	2.56	2.51
60	3.87	3.88	3.88	3.88	3.88	3.88	3.88	3.90	3.82	3.70	3.43	3.41
70	5.17	5.18	5.20	5.14	5.13	5.10	5.08	5.06	4.90	4.72	4.47	4.35
80	6.62	6.59	6.54	6.46	6.38	6.30	6.26	6.06	5.82	5.50	5.33
90	8.26	8.16	8.06	7.97	7.83	7.71	7.58	7.30	6.96	6.58	6.37
100	10.01	9.87	9.72	9.56	9.39	9.21	9.03	8.64	8.22	7.76	7.42

Example.—A sugar solution shows a reading of 30.2° Brix at 30°C. To find the necessary correction for the conversion of this reading to the reading which would have been obtained if the observation had been made at 17.5°C., find the vertical column in the table headed 30° Brix, which is the nearest to the observed reading. Follow down this column until the number is reached which is opposite to the temperature of observation—in this case 30°. The number found, 0.92, is to be added to the observed reading.

7

By Means of a Pycnometer.—Official.

(a) *By specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$* .—Determine the specific gravity of the solution at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer and ascertain the corresponding per cent by weight of sucrose from 9. When the density of the substance is too high for a direct determination, dilute and calculate the sucrose content of the original material as directed under 5.

(b) *By specific gravity at $\frac{17.5^{\circ}\text{C.}}{17.5^{\circ}}$* .—Proceed as directed under (a), the determinations of specific gravity being made at $\frac{17.5^{\circ}\text{C.}}{17.5^{\circ}}$ instead of at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$. Ascertain the corresponding per cent by weight of sucrose from 8.

The pycnometer determination should not be made at any other temperature than $\frac{17.5^{\circ}\text{C.}}{17.5^{\circ}}$ or $\frac{20^{\circ}\text{C.}}{4^{\circ}}$.

8

TABLE 10.

For the comparison of specific gravities at $\frac{17.5^{\circ}\text{C.}}{17.5^{\circ}}$, degrees Brix and degrees Baumé.

$$\text{Degree Baumé} = 146.78 - \frac{146.78}{\text{sp. gr.}}$$

DEGREE BRIX OR PER CENT BY WEIGHT OF SUCROSE	SPECIFIC GRAVITY	DEGREE BAUMÉ	DEGREE BRIX OR PER CENT BY WEIGHT OF SUCROSE	SPECIFIC GRAVITY	DEGREE BAUMÉ	DEGREE BRIX OR PER CENT BY WEIGHT OF SUCROSE	SPECIFIC GRAVITY	DEGREE BAUMÉ
1.0	1.00388	0.6	33.0	1.14423	18.5	65.0	1.31989	35.6
2.0	1.00779	1.1	34.0	1.14915	19.05	66.0	1.32601	36.1
3.0	1.01173	1.7	35.0	1.15411	19.6	67.0	1.33217	36.6
4.0	1.01570	2.3	36.0	1.15911	20.1	68.0	1.33836	37.1
5.0	1.01970	2.8	37.0	1.16413	20.7	69.0	1.34460	37.6
6.0	1.02373	3.4	38.0	1.16920	21.2	70.0	1.35088	38.1
7.0	1.02779	4.0	39.0	1.17430	21.8	71.0	1.35720	38.6
8.0	1.03187	4.5	40.0	1.17943	22.3	72.0	1.36355	39.1
9.0	1.03599	5.1	41.0	1.18460	22.9	73.0	1.36995	39.6
10.0	1.04014	5.7	42.0	1.18981	23.4	74.0	1.37639	40.1
11.0	1.04431	6.2	43.0	1.19505	23.95	75.0	1.38287	40.6
12.0	1.04852	6.8	44.0	1.20033	24.5	76.0	1.38939	41.1
13.0	1.05276	7.4	45.0	1.20565	25.0	77.0	1.39595	41.6
14.0	1.05703	7.9	46.0	1.21100	25.6	78.0	1.40254	42.1
15.0	1.06133	8.5	47.0	1.21639	26.1	79.0	1.40918	42.6
16.0	1.06566	9.0	48.0	1.22182	26.6	80.0	1.41586	43.1
17.0	1.07002	9.6	49.0	1.22728	27.2	81.0	1.42258	43.6
18.0	1.07441	10.1	50.0	1.23278	27.7	82.0	1.42934	44.1
19.0	1.07884	10.7	51.0	1.23832	28.2	83.0	1.43614	44.6
20.0	1.08329	11.3	52.0	1.24390	28.8	84.0	1.44298	45.1
21.0	1.08778	11.8	53.0	1.24951	29.3	85.0	1.44986	45.5
22.0	1.09231	12.4	54.0	1.25517	29.8	86.0	1.45678	46.0
23.0	1.09686	13.0	55.0	1.26086	30.4	87.0	1.46374	46.5
24.0	1.10145	13.5	56.0	1.26658	30.9	88.0	1.47074	47.0
25.0	1.10607	14.1	57.0	1.27235	31.4	89.0	1.47778	47.45
26.0	1.11072	14.6	58.0	1.27816	31.9	90.0	1.48486	47.9
27.0	1.11541	15.2	59.0	1.28400	32.5	91.0	1.49199	48.5
28.0	1.12013	15.7	60.0	1.28989	33.0	92.0	1.49915	48.9
29.0	1.12488	16.3	61.0	1.29581	33.5	93.0	1.50635	49.4
30.0	1.12967	16.8	62.0	1.30177	34.0	94.0	1.51359	49.8
31.0	1.13449	17.4	63.0	1.30777	34.5	95.0	1.52087	50.3
32.0	1.13934	17.95	64.0	1.31381	35.1			

When the number expressing the specific gravity found by analysis falls between the numbers given in the above table, the exact equivalent in degrees Brix or Baumé is found by a simple calculation.

Example.—The pycnometer shows the specific gravity of a certain sirup to be 1.20909. The table shows that the corresponding degree Brix is between 45.0 and 46.0. Subtracting the specific gravity of a solution of 45° Brix from the corresponding figure for 46°, we have (expressing the specific gravities as whole numbers) 121,100 - 120,565 = 535, the difference in specific gravity for 1° Brix at this point in the table. Subtracting the specific gravity corresponding to 45° from the specific gravity found by analysis, we have $120,909 - 120,565 = 344$; $\frac{344}{535} = 0.64$, the fraction of 1° Brix more than 45°. The degree Brix, corresponding to a sp. gr. of 1.20909, is therefore 45.64.

TABLE 11.

Densities of solutions of cane sugar at 20°C.*

(This table is the basis for standardizing hydrometers indicating per cent of sugar at 20°C.)

PER CENT SUGAR	TENTHS OF PER CENT									
	0	1	2	3	4	5	6	7	8	9
0	0.998234	0.998622	0.999010	0.999398	0.999786	1.000174	1.000563	1.000952	1.001342	1.001731
1	1.002120	1.002509	1.002897	1.003286	1.003675	1.004064	1.004453	1.004844	1.005234	1.005624
2	1.006015	1.006405	1.006796	1.007188	1.007580	1.007972	1.008363	1.008755	1.009148	1.009541
3	1.009934	1.010327	1.010721	1.011115	1.011510	1.011904	1.012298	1.012694	1.013089	1.013485
4	1.013881	1.014277	1.014673	1.015070	1.015467	1.015864	1.016261	1.016659	1.017058	1.017456
5	1.017854	1.018253	1.018652	1.019052	1.019451	1.019851	1.020251	1.020651	1.021053	1.021454
6	1.021855	1.022257	1.022659	1.023061	1.023463	1.023867	1.024270	1.024673	1.025077	1.025481
7	1.025885	1.026289	1.026694	1.027099	1.027504	1.027910	1.028316	1.028722	1.029128	1.029535
8	1.029942	1.030349	1.030757	1.031165	1.031573	1.031982	1.032391	1.032800	1.033209	1.033619
9	1.034029	1.034439	1.034850	1.035260	1.035671	1.036082	1.036494	1.036906	1.037318	1.037730
10	1.038143	1.038556	1.038970	1.039383	1.039797	1.040212	1.040626	1.041041	1.041456	1.041872
11	1.042288	1.042704	1.043121	1.043537	1.043954	1.044370	1.044788	1.045206	1.045625	1.046043
12	1.046462	1.046883	1.047300	1.047720	1.048140	1.048559	1.048980	1.049401	1.049822	1.050243
13	1.050665	1.051087	1.051510	1.051933	1.052356	1.052778	1.053202	1.053626	1.054050	1.054475
14	1.054900	1.055325	1.055751	1.056176	1.056602	1.057029	1.057455	1.057882	1.058310	1.058737
15	1.059165	1.059593	1.060022	1.060451	1.060880	1.061308	1.061738	1.062168	1.062598	1.063029
16	1.063460	1.063892	1.064324	1.064756	1.065188	1.065621	1.066054	1.066487	1.066921	1.067355
17	1.067789	1.068223	1.068658	1.069093	1.069529	1.069964	1.070400	1.070836	1.071273	1.071710
18	1.072147	1.072585	1.073023	1.073461	1.073900	1.074338	1.074777	1.075217	1.075657	1.076097
19	1.076537	1.076978	1.077419	1.077860	1.078302	1.078744	1.079187	1.079629	1.080072	1.080515
20	1.080959	1.081403	1.081848	1.082292	1.082737	1.083182	1.083628	1.084074	1.084520	1.084967
21	1.085414	1.085861	1.086309	1.086757	1.087205	1.087652	1.088101	1.088550	1.089000	1.089450
22	1.089900	1.090351	1.090802	1.091253	1.091704	1.092155	1.092607	1.093060	1.093513	1.093966
23	1.094420	1.094873	1.095328	1.095782	1.096236	1.096691	1.097147	1.097603	1.098058	1.098514
24	1.098971	1.099428	1.099886	1.100344	1.100802	1.101259	1.101718	1.102177	1.102637	1.103097
25	1.103557	1.104017	1.104478	1.104938	1.105400	1.105862	1.106324	1.106786	1.107248	1.107711
26	1.108175	1.108639	1.109103	1.109568	1.110033	1.110497	1.110963	1.111429	1.111895	1.112361
27	1.112828	1.113295	1.113763	1.114229	1.114697	1.115166	1.115635	1.116104	1.116572	1.117042
28	1.117512	1.117982	1.118453	1.118923	1.119395	1.119867	1.120339	1.120812	1.121284	1.121757
29	1.122231	1.122705	1.123179	1.123653	1.124128	1.124603	1.125079	1.125555	1.126030	1.126507
30	1.126994	1.127466	1.127939	1.128417	1.128896	1.129374	1.129853	1.130332	1.130812	1.131292
31	1.131773	1.132254	1.132735	1.133216	1.133698	1.134180	1.134663	1.135146	1.135628	1.136112
32	1.136596	1.137078	1.137565	1.138049	1.138534	1.139020	1.139506	1.139993	1.140479	1.140966
33	1.141453	1.141941	1.142429	1.142916	1.143405	1.143894	1.144384	1.144874	1.145364	1.145854
34	1.146345	1.146836	1.147328	1.147820	1.148313	1.148805	1.149298	1.149792	1.150286	1.150780
35	1.151275	1.151770	1.152265	1.152760	1.153256	1.153752	1.154249	1.154746	1.155242	1.155740
36	1.156238	1.156736	1.157235	1.157733	1.158233	1.158733	1.159233	1.159733	1.160233	1.160734
37	1.161236	1.161738	1.162240	1.162742	1.163245	1.163748	1.164252	1.164756	1.165259	1.165764
38	1.166269	1.166775	1.167281	1.167786	1.168293	1.168800	1.169307	1.169815	1.170322	1.170831
39	1.171340	1.171849	1.172359	1.172869	1.173379	1.173889	1.174400	1.174911	1.175423	1.175935
40	1.176447	1.176960	1.177473	1.177987	1.178501	1.179014	1.179527	1.180044	1.180560	1.181076
41	1.181592	1.182108	1.182625	1.183142	1.183660	1.184178	1.184696	1.185215	1.185734	1.186253
42	1.186773	1.187293	1.187814	1.188335	1.188856	1.189379	1.189901	1.190423	1.190946	1.191469
43	1.191993	1.192517	1.193041	1.193565	1.194090	1.194616	1.195141	1.195667	1.196193	1.196720
44	1.197247	1.197775	1.198303	1.198832	1.199360	1.199890	1.200420	1.200950	1.201480	1.202010
45	1.202540	1.203071	1.203603	1.204136	1.204668	1.205200	1.205733	1.206266	1.206801	1.207335
46	1.207870	1.208405	1.208940	1.209477	1.210013	1.210549	1.211086	1.211623	1.212162	1.212701
47	1.213238	1.213777	1.214317	1.214856	1.215395	1.215936	1.216476	1.217017	1.217559	1.218101
48	1.218643	1.219185	1.219729	1.220272	1.220815	1.221360	1.221904	1.222449	1.222995	1.223540
49	1.224086	1.224632	1.225180	1.225727	1.226274	1.226823	1.227371	1.227919	1.228469	1.229018
50	1.229567	1.230117	1.230668	1.231219	1.231770	1.232322	1.232874	1.233426	1.233979	1.234532
51	1.235085	1.235639	1.236194	1.236748	1.237303	1.237858	1.238414	1.238970	1.239527	1.240084
52	1.240641	1.241198	1.241757	1.242315	1.242873	1.243433	1.243992	1.244552	1.245113	1.245673
53	1.246234	1.246795	1.247355	1.247920	1.248482	1.249046	1.249609	1.250172	1.250737	1.251301
54	1.251866	1.252431	1.252997	1.253563	1.254129	1.254697	1.255264	1.255831	1.256400	1.256967
55	1.257535	1.258104	1.258674	1.259244	1.259815	1.260385	1.260955	1.261527	1.262099	1.262671
56	1.263243	1.263816	1.264390	1.264963	1.265537	1.266112	1.266686	1.267261	1.267837	1.268413
57	1.268989	1.269565	1.270143	1.270720	1.271299	1.271877	1.272455	1.273035	1.273614	1.274194
58	1.274774	1.275354	1.275936	1.276517	1.277098	1.277680	1.278262	1.278844	1.279428	1.280011
59	1.280595	1.281179	1.281764	1.282349	1.282935	1.283521	1.284107	1.284694	1.285281	1.285869

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TABLE 11.—Continued.
Densities of solutions of cane sugar at 20°C.

PER CENT SUGAR	TENTHS OF PER CENT									
	0	1	2	3	4	5	6	7	8	9
60	1.286456	1.287044	1.287633	1.288222	1.288811	1.289401	1.289991	1.290581	1.291172	1.291763
61	1.292354	1.292946	1.293539	1.294131	1.294725	1.295318	1.295911	1.296506	1.297100	1.297696
62	1.298291	1.298886	1.299483	1.300079	1.300677	1.301274	1.301871	1.302470	1.303068	1.303668
63	1.304267	1.304867	1.305467	1.306068	1.306669	1.307271	1.307872	1.308475	1.309077	1.309680
64	1.310282	1.310885	1.311489	1.312093	1.312699	1.313304	1.313909	1.314515	1.315121	1.315728
65	1.316334	1.316941	1.317549	1.318157	1.318766	1.319374	1.319983	1.320593	1.321203	1.321814
66	1.322425	1.323036	1.323648	1.324259	1.324872	1.325484	1.326097	1.326711	1.327325	1.327940
67	1.328554	1.329170	1.329785	1.330401	1.331017	1.331633	1.332250	1.332868	1.333485	1.334103
68	1.334722	1.335342	1.335961	1.336581	1.337200	1.337821	1.338441	1.339063	1.339684	1.340306
69	1.340928	1.341551	1.342174	1.342798	1.343421	1.344046	1.344671	1.345296	1.345922	1.346547
70	1.347174	1.347801	1.348427	1.349055	1.349682	1.350311	1.350939	1.351568	1.352197	1.352827
71	1.353456	1.354087	1.354717	1.355349	1.355980	1.356612	1.357245	1.357877	1.358511	1.359144
72	1.359778	1.360413	1.361047	1.361682	1.362317	1.362953	1.363590	1.364226	1.364864	1.365501
73	1.366139	1.366777	1.367415	1.368054	1.368693	1.369333	1.369973	1.370613	1.371254	1.371894
74	1.372536	1.373178	1.373820	1.374463	1.375105	1.375749	1.376392	1.377036	1.377680	1.378326
75	1.378971	1.379617	1.380262	1.380909	1.381555	1.382203	1.382851	1.383499	1.384148	1.384796
76	1.385446	1.386096	1.386745	1.387396	1.388045	1.388696	1.389347	1.389999	1.390651	1.391303
77	1.391956	1.392610	1.393263	1.393917	1.394571	1.395226	1.395881	1.396536	1.397192	1.397848
78	1.398505	1.399162	1.399819	1.400477	1.401134	1.401793	1.402452	1.403111	1.403771	1.404430
79	1.405091	1.405752	1.406412	1.407074	1.407735	1.408398	1.409061	1.409723	1.410387	1.411051
80	1.411715	1.412380	1.413044	1.413709	1.414374	1.415040	1.415706	1.416373	1.417039	1.417707
81	1.418374	1.419043	1.419711	1.420380	1.421049	1.421719	1.422390	1.423059	1.423730	1.424400
82	1.425072	1.425744	1.426416	1.427089	1.427761	1.428435	1.429109	1.429782	1.430457	1.431131
83	1.431807	1.432483	1.433158	1.433835	1.434511	1.435188	1.435866	1.436543	1.437222	1.437900
84	1.438579	1.439259	1.439938	1.440619	1.441299	1.441980	1.442661	1.443342	1.444024	1.444705
85	1.445388	1.446071	1.446754	1.447438	1.448121	1.448806	1.449491	1.450175	1.450860	1.451545
86	1.452232	1.452919	1.453605	1.454292	1.454980	1.455668	1.456357	1.457045	1.457735	1.458424
87	1.459114	1.459805	1.460495	1.461186	1.461877	1.462568	1.463260	1.463953	1.464645	1.465338
88	1.466032	1.466726	1.467420	1.468115	1.468810	1.469504	1.470200	1.470896	1.471592	1.472289
89	1.472986	1.473684	1.474381	1.475080	1.475779	1.476477	1.477176	1.477876	1.478575	1.479275
90	1.479976	1.480677	1.481378	1.482080	1.482782	1.483484	1.484187	1.484890	1.485593	1.486297
91	1.487002	1.487707	1.488411	1.489117	1.489823	1.490528	1.491234	1.491941	1.492647	1.493355
92	1.494063	1.494771	1.495479	1.496188	1.496897	1.497606	1.498316	1.499026	1.499736	1.500447
93	1.501158	1.501870	1.502582	1.503293	1.504006	1.504719	1.505432	1.506146	1.506859	1.507574
94	1.508289	1.509004	1.509720	1.510435	1.511151	1.511868	1.512585	1.513302	1.514019	1.514737
95	1.515455	1.516174	1.516893	1.517612	1.518332	1.519051	1.519771	1.520492	1.521212	1.521934
96	1.522656	1.523378	1.524100	1.524823	1.525546	1.526269	1.526993	1.527717	1.528441	1.529166
97	1.529891	1.530616	1.531342	1.532068	1.532794	1.533521	1.534248	1.534976	1.535704	1.536432
98	1.537161	1.537889	1.538618	1.539347	1.540076	1.540806	1.541536	1.542267	1.542998	1.543730
99	1.544462	1.545194	1.545926	1.546659	1.547392	1.548127	1.548861	1.549595	1.550329	1.551064
100	1.551800									

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REFRACTOMETER METHOD.—TENTATIVE.

Determine the refractive index of the solution at 28°C. and obtain the corresponding percentage of dry substance from 11. If the refractive index is obtained at a temperature other than 28°C., correct the result as indicated in 12. If the solution is too dark to be read in the instrument, dilute with a concentrated sugar solution. Water should never be used for this purpose. Mix weighed amounts of the solution under examination and a solution of pure sugar of about the same strength, and obtain the amount of dry substance in the former by the following formula:

$$x = \frac{(A + B) C - BD}{A} \text{ in which}$$

x = per cent of dry substance to be found;

A = weight in grams of the material mixed with B;

B = weight in grams of pure sugar solution employed in the dilution;

C = per cent of dry substance in the mixture of A and B obtained from the refractive index;

D = per cent of dry substance in the pure sugar solution obtained from its refractive index.

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TABLE 12.—GEERLIGS³ TABLE.

For dry substance in sugar-house products by the Abbe refractometer, at 28°C.

INDEX	PER CENT DRY SUB- STANCE	DECIMALS TO BE ADDED FOR FRACTIONAL READINGS*	INDEX	PER CENT DRY SUB- STANCE	DECIMALS TO BE ADDED FOR FRACTIONAL READINGS*	INDEX	PER CENT DRY SUB- STANCE	DECIMALS TO BE ADDED FOR FRACTIONAL READINGS*
1.3335	1	0.0001 = 0.05	1.3484	11	0.0001 = 0.05	1.3746	27	0.0001 = 0.05
1.3349	2	0.0002 = 0.1	1.3500	12	0.0002 = 0.1	1.3761	28	0.0002 = 0.1
1.3364	3	0.0003 = 0.2	1.3516	13	0.0003 = 0.2	1.3782	29	0.0003 = 0.15
1.3379	4	0.0004 = 0.25	1.3530	14	0.0004 = 0.25	1.3800	30	0.0004 = 0.2
1.3394	5	0.0005 = 0.3	1.3546	15	0.0005 = 0.3	1.3818	31	0.0005 = 0.25
1.3409	6	0.0006 = 0.4	1.3562	16	0.0006 = 0.4	1.3836	32	0.0006 = 0.3
1.3424	7	0.0007 = 0.5	1.3578	17	0.0007 = 0.45	1.3854	33	0.0007 = 0.35
1.3439	8	0.0008 = 0.6	1.3594	18	0.0008 = 0.5	1.3872	34	0.0008 = 0.4
1.3454	9	0.0009 = 0.7	1.3611	19	0.0009 = 0.6	1.3890	35	0.0009 = 0.45
1.3469	10	0.0010 = 0.75	1.3627	20	0.0010 = 0.65	1.3909	36	0.0010 = 0.5
		0.0011 = 0.8	1.3644	21	0.0011 = 0.7	1.3928	37	0.0011 = 0.55
		0.0012 = 0.85	1.3661	22	0.0012 = 0.75	1.3947	38	0.0012 = 0.6
		0.0013 = 0.9	1.3678	23	0.0013 = 0.8	1.3966	39	0.0013 = 0.65
		0.0014 = 0.95	1.3695	24	0.0014 = 0.85	1.3984	40	0.0014 = 0.7
		0.0015 = 1.0	1.3712	25	0.0015 = 0.9	1.4003	41	0.0015 = 0.75
			1.3729	26	0.0016 = 0.95			0.0016 = 0.8
								0.0017 = 0.85
								0.0018 = 0.9
								0.0019 = 0.95
								0.0020 = 1.0
								0.0021 = 1.0
1.4023	42	0.0001 = 0.05	1.4292	55	0.0001 = 0.05	1.4711	73	0.0001 = 0.0
1.4043	43	0.0002 = 0.1	1.4314	56	0.0002 = 0.1	1.4736	74	0.0002 = 0.05
1.4063	44	0.0003 = 0.15	1.4337	57	0.0003 = 0.1	1.4761	75	0.0003 = 0.1
1.4083	45	0.0004 = 0.2	1.4359	58	0.0004 = 0.15	1.4786	76	0.0004 = 0.15
1.4104	46	0.0005 = 0.25	1.4382	59	0.0005 = 0.2	1.4811	77	0.0005 = 0.2
1.4124	47	0.0006 = 0.3	1.4405	60	0.0006 = 0.25	1.4836	78	0.0006 = 0.2
1.4145	48	0.0007 = 0.35	1.4428	61	0.0007 = 0.3	1.4862	79	0.0007 = 0.25
1.4166	49	0.0008 = 0.4	1.4451	62	0.0008 = 0.35	1.4888	80	0.0008 = 0.3
1.4186	50	0.0009 = 0.45	1.4474	63	0.0009 = 0.4	1.4914	81	0.0009 = 0.35
1.4207	51	0.0010 = 0.5	1.4497	64	0.0010 = 0.45	1.4940	82	0.0010 = 0.35
1.4228	52	0.0011 = 0.55	1.4520	65	0.0011 = 0.5	1.4966	83	0.0011 = 0.4
1.4249	53	0.0012 = 0.6	1.4543	66	0.0012 = 0.55	1.4992	84	0.0012 = 0.45
1.4270	54	0.0013 = 0.65	1.4567	67	0.0013 = 0.6	1.5019	85	0.0013 = 0.5
		0.0014 = 0.7	1.4591	68	0.0014 = 0.65	1.5046	86	0.0014 = 0.55
		0.0015 = 0.75	1.4615	69	0.0015 = 0.7	1.5073	87	0.0015 = 0.6
		0.0016 = 0.8	1.4639	70	0.0016 = 0.75	1.5100	88	0.0016 = 0.65
		0.0017 = 0.85	1.4663	71	0.0017 = 0.8	1.5127	89	0.0017 = 0.7
		0.0018 = 0.9	1.4687	72	0.0018 = 0.85	1.5155	90	0.0018 = 0.75
		0.0019 = 0.95			0.0019 = 0.9			0.0019 = 0.8
		0.0020 = 1.0			0.0020 = 0.95			0.0020 = 0.85
		0.0021 = 1.0			0.0021 = 1.0			0.0021 = 0.9
					0.0022 = 0.95			0.0022 = 0.95
					0.0023 = 1.0			0.0023 = 1.0
					0.0024 = 1.0			0.0024 = 0.9
								0.0025 = 0.9
								0.0026 = 0.95
								0.0027 = 1.0
								0.0028 = 1.0

* Find in the table the refractive index which is next lower than the reading actually made and note the corresponding whole number for the per cent of dry substance. Subtract the refractive index obtained from the table from the observed reading; the decimal corresponding to this difference, as given in the column so marked, is added to the whole per cent of dry substance as first obtained.

12

TABLE 13.
Corrections for temperature.

TEMPERATURE OF THE PRISMS IN °C.	DRY SUBSTANCE												
	0	5	10	15	20	25	30	40	50	60	70	80	90
	Subtract—												
20	0.53	0.54	0.55	0.56	0.57	0.58	0.60	0.62	0.64	0.62	0.61	0.60	0.58
21	0.46	0.47	0.48	0.49	0.50	0.51	0.52	0.54	0.56	0.54	0.53	0.52	0.50
22	0.40	0.41	0.42	0.42	0.43	0.44	0.45	0.47	0.48	0.47	0.46	0.45	0.44
23	0.33	0.33	0.34	0.35	0.36	0.37	0.38	0.39	0.40	0.39	0.38	0.38	0.38
24	0.26	0.26	0.27	0.28	0.28	0.29	0.30	0.31	0.32	0.31	0.31	0.30	0.30
25	0.20	0.20	0.21	0.21	0.22	0.22	0.23	0.23	0.24	0.23	0.23	0.23	0.22
26	0.12	0.12	0.13	0.14	0.14	0.15	0.15	0.16	0.16	0.16	0.15	0.15	0.14
27	0.07	0.07	0.07	0.07	0.07	0.07	0.08	0.08	0.08	0.08	0.08	0.08	0.07
	Add—												
	0	5	10	15	20	25	30	40	50	60	70	80	90
29	0.07	0.07	0.07	0.07	0.07	0.07	0.08	0.08	0.08	0.08	0.08	0.08	0.07
30	0.12	0.12	0.13	0.14	0.14	0.14	0.15	0.15	0.16	0.16	0.16	0.15	0.14
31	0.20	0.20	0.21	0.21	0.22	0.22	0.23	0.23	0.24	0.23	0.23	0.23	0.22
32	0.26	0.26	0.27	0.28	0.28	0.29	0.30	0.31	0.32	0.31	0.31	0.30	0.30
33	0.33	0.33	0.34	0.35	0.36	0.37	0.38	0.39	0.40	0.39	0.38	0.38	0.38
34	0.40	0.41	0.42	0.42	0.43	0.44	0.45	0.47	0.48	0.47	0.46	0.45	0.44
35	0.46	0.47	0.48	0.49	0.50	0.51	0.52	0.54	0.56	0.54	0.53	0.52	0.50

ASH.

13

Method I.—Official.

Heat 5-10 grams of the sample in a 50-100 cc. platinum dish at 100°C. until the water is expelled, add a few drops of pure olive oil, and heat slowly over a flame until swelling ceases. Then place the dish in a muffle and heat at low redness until a white ash is obtained.

14

Method II.—Official.

Carbonize the mass at a low heat, dissolve the soluble salts in hot water, burn the residual mass as directed in 13, add the solution of soluble salts, and evaporate to dryness at 100°C., ignite gently, cool in a desiccator, and weigh.

15

Method III.—Official.

Saturate the sample with sulphuric acid, dry, ignite gently, then burn in a muffle at low redness. Deduct one tenth of the weight of the ash, and calculate the per cent.

16

QUANTITATIVE ANALYSIS OF THE ASH.—OFFICIAL.

Proceed as directed under III.

17

SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.

Ash the material as directed under 13 or 14. Add water to the ash in the platinum dish, heat nearly to boiling, filter through an ashless filter paper, and wash with hot water until the combined filtrate and washings measure about 60 cc. Re

turn the filter paper and contents to the platinum dish, ignite carefully, and weigh. Calculate the percentages of water-soluble and water-insoluble ash.

18 ALKALINITY OF THE SOLUBLE ASH.—TENTATIVE.

Cool the filtrate from 17 and titrate with N/10 hydrochloric acid, using methyl orange as an indicator.

Express the alkalinity in terms of the number of cc. of N/10 acid per 1 gram of the sample.

19 ALKALINITY OF THE INSOLUBLE ASH.—TENTATIVE.

Add an excess of N/10 hydrochloric acid (usually 10–15 cc.) to the ignited insoluble ash in the platinum dish, under 17, heat to boiling over an asbestos plate, cool, and titrate the excess of hydrochloric acid with N/10 sodium hydroxid, using methyl orange as an indicator.

Express the alkalinity in terms of the number of cc. of N/10 acid per 1 gram of the sample.

20 MINERAL ADULTERANTS IN THE ASH.—TENTATIVE.

Mix 100 grams of molasses, sirup, honey, or the confectionery solution prepared as directed under 1 (b) and evaporate to a sirupy consistency, with about 35 grams of concentrated sulphuric acid in a large porcelain evaporating dish. Pass an electric current through it while stirring by placing one platinum electrode in the bottom of the dish near one side and attaching the other to the lower end of the glass rod with which the contents are stirred. Begin with a current of about 1 ampere and gradually increase to 4 (modified from method of Budde and Schou⁴ for determining nitrogen electrolytically). In 10–15 minutes the mass is reduced to a fine dry char, which may be readily burnt to a white ash in the original dish over a free flame or in a muffle.

This method⁵ is preferred to the ordinary method of heating with sulphuric acid, especially in the case of molasses, because, if properly manipulated, it comes quietly into the form of a very finely divided char or powder, especially adapted for subsequent quick ignition.

If an electric current is not available, treat in a large porcelain dish 100 grams of the saccharine solution, evaporated to a sirupy consistency, with sufficient concentrated sulphuric acid to thoroughly carbonize the mass and ignite in the usual manner.

The following adulterants may be present: salts of tin, used in molasses to bleach; mineral pigments, such as chromate of lead in yellow confectionery; oxid of iron, sometimes used to simulate the color of chocolate; and copper. These elements may be detected by the usual qualitative tests.

21 NITROGEN.—TENTATIVE.

Determine nitrogen in 5 grams of the material as directed under I, 18, 21 or 23, using a larger quantity of the sulphuric acid if necessary for complete digestion.

SUCROSE.

22 Method I.—Tentative.

(Substances in which the volume of the combined insoluble matter and precipitate from clarifying agents is less than 1 cc. from 26 grams.)

Determine sucrose by polarization before and after inversion, as directed under VIII, 14.

All products which contain dextrose or other reducing sugars in the crystalline form, or in supersaturated solution, exhibit the phenomenon of birotation. The constant rotation only should be employed in the Clerget formula, and to obtain this the solutions prepared for direct polarization should be allowed to stand overnight before making the reading. If it is desired to make the direct reading immediately, the birotation may be destroyed by heating the neutral solution to boiling for a few minutes or by adding a few drops of strong ammonium hydroxid before completing the volume.

23

Method II. (Double dilution method.⁶)—Tentative.

(Substances in which the volume of the combined insoluble matter and precipitate from clarifying agents is more than 1 cc. from 26 grams.)

Weigh out a half normal weight of the sample and make up the solution to 100 cc., employing the appropriate clarifier (basic lead acetate for dark colored confectionery or molasses and alumina cream for light colored confectionery). Also weigh out a normal weight of the sample and make up a second solution with the clarifier to 100 cc. Filter and obtain direct polariscopic readings of both solutions. Invert each solution as directed in 22 and obtain its invert reading.

The true direct polarization of the sample is the product of the two direct readings divided by their difference.

The true invert polarization is the product of the two invert readings divided by their difference.

Calculate the sucrose from the true polarizations thus obtained by the formula given under VIII, 14.

COMMERCIAL GLUCOSE (APPROXIMATE).

24

Method I.—Tentative.

(Substances containing little or no invert sugar.)

Commercial glucose can not be determined accurately owing to the varying amounts of dextrin, maltose, and dextrose present in this product. However, in sirups, in which the amount of invert sugar is so small as not to appreciably affect the result, commercial glucose may be estimated approximately by the following formula:⁷

$$G = \frac{(a - S) 100}{175} \text{ in which}$$

G = per cent of commercial glucose;

a = direct polarization;

S = per cent of cane sugar.

Express the results in terms of commercial glucose polarizing $+175^{\circ}\text{V}$.

Method II.—Tentative.

25

(Substances containing invert sugar.⁷)

Prepare an inverted half normal solution of the substance as directed under VIII, 14 except that after inversion cool the solution, make neutral to phenolphthalein with sodium hydroxid solution, slightly acidify with hydrochloric acid, and treat with 5-10 cc. of alumina cream before making up to the mark. Filter and polarize at 87°C . in a 200 mm. jacketed tube. Multiply the reading by 200 and divide by the factor 163 to express the amount of glucose present in terms of glucose polarizing $+175^{\circ}\text{V}$.

26

REDUCING SUGARS.—TENTATIVE.

Determine either as dextrose or invert sugar as directed under VIII, 50, 51, 52, 54, or 21, 23, 25, 36 or 39.

27

STARCH.—TENTATIVE.

Measure 25 cc. of a solution or uniform mixture, prepared as directed in 1 (b), (representing 5 grams of the sample) into a 300 cc. beaker, or introduce 5 grams of the finely ground sample (previously extracted with ether if the sample contains much fat) into the beaker, add sufficient water to make the volume 100 cc., heat to about 60°C. (avoiding if possible gelatinizing the starch) and allow to stand for about an hour, stirring frequently to secure complete solution of the sugars. Transfer to a stout wide-mouthed bottle, rinse the beaker with a little warm water, cool, add an equal volume of 95% alcohol, mix, and allow to stand at least an hour. Centrifugalize until the precipitate is closely packed on the bottom of the bottle and decant the supernatant liquid through a hardened filter. Wash the precipitate with successive 50 cc. portions of 50% alcohol by centrifugalizing and decanting through the filter until 3 or 4 drops of the washings give no test for sugar with alphanaphthol as described under 68. Transfer the residue from the bottle and the hardened filter to a large flask and determine starch as directed under VIII, 60.

ETHER EXTRACT IN CONFECTIONERY.

28

Continuous Extraction.—Tentative.

(1) Measure 25 cc. of a 20% mixture or solution, prepared as directed under 1 (b), into a very thin, readily frangible, glass evaporating shell (*Hofmeister Schälchen*), containing 5–7 grams of freshly ignited asbestos fiber; or (2) If impossible to obtain a uniform sample, weigh 5 grams of the mixed finely divided sample into a dish, and wash with water upon the asbestos in the evaporating shell, using, if necessary, a small portion of the asbestos fiber on a stirring rod to transfer the last traces of the sample from the dish to the shell. Dry to constant weight at 100°C., cool, wrap loosely in smooth paper, crush into rather small fragments between the fingers, transfer carefully the crushed mass, exclusive of the paper, to an extraction tube or a fat extraction cartridge. A thin lead disk (bottle cap) may be substituted for the *Schälchen*. The disk may then be cut into small pieces and placed in the extraction tube. Extract with anhydrous ether or petroleum ether (b. p. 45°–60°C. and without weighable residue) in a continuous extraction apparatus for at least 25 hours. In most cases it is advisable to remove the substance from the extractor after the first 12 hours, grind with sand to a fine powder, and re-extract for the remaining 13 hours. Transfer the extract to a tared flask, evaporate the solvent, dry to constant weight in an oven at 100°C.

29

Roese-Gottlieb Method.—Tentative.

Substances such as butter-scotch, invariably yield extremely inaccurate results by the above method. In such cases introduce 4 grams of the material, or an amount of a uniform solution equivalent to this amount of the dry substance, into a Röhrig tube or similar apparatus, make up to a volume of 10 cc. with water, add 1.25 cc. of concentrated ammonium hydroxid and mix thoroughly. Add 10 cc. of 95% alcohol and mix. Then add 25 cc. of washed ether and shake vigorously for half a minute; then add 25 cc. of petroleum ether (b. p. below 60°C.), and shake again for half a minute. Allow to stand for 20 minutes or until the separation between the

liquids is complete. Draw off as much as possible of the ether-fat solution (usually 0.5–0.8 cc. will be left) into a weighed flask through a small, rapid filter. The flask should be weighed with a similar one as a counterpoise. Again extract the liquid remaining in the tube, this time with 15 cc. each of ether and petroleum ether, shake vigorously half a minute with each, and allow to settle. Proceed as above, washing the tip of the spigot and the filter with a few cc. of a mixture of equal parts of the 2 ethers (previously mixed and free from deposited water). For absolutely exact results the extraction must be repeated. This third extraction usually yields not more than about 1 mg. of fat, if the previous ether-fat solutions have been drawn off closely, or an amount averaging about 0.02% on a 4 gram charge. Evaporate the ether slowly on a steam bath, then dry the fat in a boiling water oven until the loss in weight ceases. Test the purity of the fat by dissolving in a little petroleum ether. Should a residue remain, wash the fat out completely with petroleum ether, dry the residue, weigh, and deduct the weight.

30**PARAFFIN IN CONFECTIONERY.—TENTATIVE.**

Add to the ether extract in the flask, as above obtained, 10 cc. of 95% alcohol and 2 cc. of sodium hydroxid solution (1 to 1), connect the flask with a reflux condenser, and heat for an hour on the water bath, or until saponification is complete. Remove the condenser and allow the flask to remain on the bath until the alcohol is evaporated and the residue is dry. Dissolve the residue as completely as possible in about 40 cc. of water and heat on the bath, shaking frequently. Wash into a separatory funnel, cool, and extract with 4 successive portions of petroleum ether, which are collected in a tared flask or capsule. Evaporate the petroleum ether and dry in the oven to constant weight.

Any phytosterol or cholesterol present in the fat would be extracted with the paraffin. The amount is so insignificant that it may be disregarded generally. The character of the final residue should, however, be confirmed by determining its melting point, specific gravity, and refractive index.

31 ALCOHOL IN SIRUPS USED IN CONFECTIONERY ("BRANDY DROPS").—TENTATIVE.

Collect in a beaker the sirup from a sufficient number of pieces to yield 30–50 grams of sirup. Strain the sirup into a tared beaker and weigh. Introduce the sirup into a 250–300 cc. distilling flask, dilute with half its volume of water, attach the flask to a vertical condenser and distil almost 50 cc., or as much of the liquid as possible without causing charring. Foaming may be prevented by adding a little tannin, or a piece of paraffin about the size of a pea, to the contents of the distillation flask. Cool the distillate, make up to volume with water, mix well, and ascertain the specific gravity of the liquid by means of a pycnometer, and obtain the corresponding weight of alcohol in the 50 cc. of distillate from **XVI, 5**. Calculate the per cent by weight of alcohol in the candy filling.

32**COLORING MATTER.—TENTATIVE.**

Proceed as directed under **XI**.

33**METALS.—TENTATIVE.**

Proceed as directed under **XII**.

HONEY.⁸

34

PREPARATION OF SAMPLE.—TENTATIVE.

(a) *Liquid or strained honey*.—If the sample is free from granulation, mix thoroughly by stirring or shaking before drawing weighed portions for the analytical determination. If the honey is granulated, place the container, having the stopper loose, in a water bath, and heat at a temperature not exceeding 50°C. until the sugar crystals dissolve; mix thoroughly, cool, and weigh portions for the analytical determinations. If sediment such as particles of comb, wax, sticks, bees, etc., are present, heat the sample to 40°C. in a water bath and filter through cheese-cloth before weighing portions for analysis.

(b) *Comb honey*.—Cut across the top of the comb, if sealed, and separate completely from the comb by straining through a 40 mesh sieve. When portions of the comb or wax pass through the sieve, heat the sample as in (a) and strain through cloth. If the honey is granulated in the comb, heat until the wax is liquified, stir, cool, remove the wax and take the clear liquid for analysis.

35

MOISTURE.

Weigh 2 grams of the sample into a tared, flat-bottomed aluminium dish, having a diameter of about 60 mm. and containing 10–15 grams of fine quartz sand, which has been previously washed, dried and ignited, and a small glass stirring rod; add 5–10 cc. of water and thoroughly incorporate with the sand and honey mixture by means of the rod; dry the dish and its contents to constant weight in a vacuum oven at a temperature not exceeding 70°C.

36

ASH.—OFFICIAL.

Weigh 5–10 grams of honey into a platinum dish, add a few drops of pure olive oil to prevent spattering, and heat carefully until swelling ceases and then ignite at a temperature not above dull redness until a white ash is obtained.

37

SOLUBLE ASH.—TENTATIVE.

Proceed as directed under 17.

38

ALKALINITY OF THE SOLUBLE ASH.—TENTATIVE.

Proceed as directed under 18.

POLARIZATION.

39

Direct Polarization.—Tentative.

(a) *Immediate direct polarization*.—Transfer 26 grams of the honey to a 100 cc. flask with water, add 5 cc. of alumina cream, dilute to the mark with water at 20°C., filter, and polarize immediately in a 200 mm. tube.

(b) *Constant direct polarization*.—Pour the solution from the tube used in reading (a) back into the flask, stopper, and allow to stand for 24 hours. At the end of this time again polarize the solution at 20°C. in a 200 mm. tube.

(c) *Birotation*.—The difference between (a) and (b) gives the birotation.

(d) *Direct polarization at 87°C.*—Polarize the solution, obtained in (b), at 87°C. in a jacketed 200 mm. tube.

40

Invert Polarization.—Tentative.

(a) *At 20°C.*—Invert 50 cc. of the solution obtained in **39** as directed under **VIII, 14** or **16**, and polarize at 20°C. in a 200 mm. tube.

(b) *At 87°C.*—Polarize the solution, obtained as directed in (a), at 87°C. in a 200 mm. jacketed tube.

41

REDUCING SUGARS.—TENTATIVE.

Dilute 10 cc. of the solution, used for direct polarization, **39**, to 250 cc. and determine reducing sugars in 25 cc. of this solution by one of the methods given under **VIII, 25, 36, 39** or **56**, respectively. Calculate the result to per cent of invert sugar.

42

SUCROSE.—TENTATIVE.

Proceed as directed under **VIII, 18**. Determine reducing sugars after inversion by diluting 10 cc. of the solution obtained in **40**, with a small amount of water, neutralizing with sodium carbonate, and making up to 250 cc. with water. Employ 50 cc. of this solution for the determination, using the same method as in **41**.

43

LEVULOSE.—TENTATIVE.

Multiply the direct reading at 87°C., **39 (d)**, by 1.0315 and subtract the product from the constant direct polarization at 20°C., **39 (b)**: divide the difference by 2.3919 to obtain the grams of levulose in a normal weight of the honey. From this figure calculate the per cent of levulose in the original sample.

44

DEXTROSE.—TENTATIVE.

Subtract the per cent of levulose, obtained in **43**, from the per cent of invert sugar, found in **41**, to obtain the approximate per cent of dextrose.

The dextrose can be determined more accurately by multiplying the per cent of levulose, as found in **43**, by the factor 0.915, which gives its dextrose equivalent in copper reducing power. Subtract this figure from that of the reducing sugars, **41**, calculated as dextrose, to obtain the percentage of dextrose in the sample. (Owing to the difference in the reducing powers of different sugars, the sum of the dextrose thus found and the levulose as obtained in **43** will be greater than the amount of invert sugar obtained in **41**).

45

DEXTRIN (APPROXIMATE).—TENTATIVE.

Transfer 8 grams of the sample (4 grams in the case of dark colored honey-dew honey) to a 100 cc. flask (using not more than 4 cc. of water) by allowing the sample to drain from the weighing dish into the flask and then dissolving the residue in 2 cc. of water. After adding this solution to the contents of the flask, rinse the weighing dish with two 1 cc. portions of water to which a little alcohol is added subsequently. Fill the flask to the mark with absolute alcohol, shaking constantly. Set the flask aside until the dextrin has collected on the sides and bottom and the liquid is clear. Decant the clear liquid through a filter paper and wash the residue in the flask with 10 cc. of 95% alcohol, pouring the washings through the same filter. Dissolve the dextrin in the flask with boiling water and filter through the filter paper already used, receiving the filtrate in a tared dish, prepared as directed under **4**. Rinse the flask and wash the filter a number of times with small portions of hot water, evaporate on a water bath and dry to constant weight in vacuo at 70°C.

After determining the weight of the alcohol precipitate, dissolve the latter in water and make up to definite volume, using 50 cc. of water for each 0.5 gram of precipitate or part thereof.

Determine reducing sugars in the solution both before and after inversion as directed under VIII, 18, expressing the results as invert sugar. Calculate sucrose from the results thus obtained and subtract the sum of the reducing sugars before inversion and sucrose from the weight of the total alcoholic precipitate to obtain the weight of the dextrin.

46

FREE ACID.—TENTATIVE.

Dissolve 10 grams of the honey in water and titrate with N/10 sodium hydroxid using phenolphthalein as an indicator. Express the results in terms of cc. of N/10 sodium hydroxid required to neutralize 100 grams of the sample.

47

GLUCOSE.—TENTATIVE.

Qualitative test.—Dilute the honey with water in the proportion of 1 to 1, then add a few cc. of iodine solution (1 gram of iodine, 3 grams of potassium iodide, 50 cc. of water). In the presence of glucose the solution turns red or violet, the depth and character of the color depending upon the quality and nature of the glucose employed. A blank test with a pure honey of about the same color should be made in order to secure an accurate color comparison. Should the honey be dark and the percentage of glucose very small, precipitate the dextrin which may be present by adding several volumes of 95% alcohol. Allow to stand until the precipitate settles (do not filter), decant the liquid, dissolve the residue of dextrins in hot water, cool and apply the above test to this solution. A negative result is not proof of the absence of glucose as some glucose, especially of high conversion, does not give any reaction with iodine.⁹

Quantitative test.—An approximate determination can be made by Browne's formula as follows: Multiply the difference in the polarizations of the invert solution at 20°C. and 87°C. by 77 and divide this product by the percentage of invert sugar after inversion found in the sample. Multiply the quotient by 100 and divide the product by 26.7, to obtain the percentage of honey in the sample; 100 per cent minus the per cent of honey gives the percentage of glucose.

COMMERCIAL INVERT SUGAR.¹⁰

QUALITATIVE TESTS.

Fiehe Test (Bryan Modification¹¹).—Tentative.

48

REAGENT.

Resorcin solution.—Dissolve 1 gram of resorcin in 100 cc. of hydrochloric acid, sp. gr. 1.19.

49

MANIPULATION.

Introduce 10 cc. of a 50% honey solution into a test tube and add 5 cc. of ether. Shake gently and allow to stand for some time until the ether layer is clear. Transfer 2 cc. of this clear ether solution to a small test tube and add a large drop of the resorcin solution. Shake and note the color immediately. In the presence of artificial invert sugar, the resorcin assumes immediately an orange-red color turning to dark red.

*Feder Anilin Chlorid Test.*¹²—*Tentative.*

50

REAGENT.

Anilin chlorid solution.—To 100 cc. of C. P. anilin add 30 cc. of 25% hydrochloric acid.

51

MANIPULATION.

Introduce 5 grams of the honey into a porcelain dish and add 2.5 cc. of the anilin reagent. A bright red color indicates the presence of commercial invert sugar.

52

DIASTASE.¹³

Mix 1 part of honey with 2 parts of sterile water. Treat 10 cc. of this solution with 1 cc. of 1% soluble starch solution and digest at 45°C. for an hour. At the end of this time test the mixture with 1 cc. of iodine solution (1 gram of iodine, 2 grams of potassium iodide, 300 cc. of water). Treat another 10 cc. portion of the honey solution, mixed with 1 cc. of the soluble starch solution, without heating to 45°C., with the reagent and compare the colors produced. If the original honey had not been heated sufficiently to kill the diastase, an olive-green or brown coloration will be produced in the mixture that has been heated at 45°C. Heated or artificial honey becomes blue.

MAPLE PRODUCTS.

53

PREPARATION OF SAMPLE.—TENTATIVE.

(a) *Maple sirup.*—Determine the moisture by the method given under 54 (a). If the moisture is less than 35%, and there is some mineral sediment, pour the clear sirup into a beaker, washing the sediment also into the beaker with water. Then concentrate the sirup by boiling to a moisture content of about 35% (b. p. 104°C.). Set aside until cool, or preferably let the covered material stand overnight, and pour off the clear liquid for the analytical work. Where no sediment is present the sample is ready for analysis after careful mixing. Where sugar has crystallized out, warm to dissolve the sugar before starting the analysis. It is desirable in order to compare results upon different samples, to reduce all results other than moisture to a dry substance basis as determined in the clear sirup.

(b) *Maple sugar, maple cream, maple wax, etc.*—Determine moisture, by the method given under 54 (b), in the sample in its original condition by thoroughly mixing, if semi-plastic, or by rubbing up in a mortar representative portions of the product if solid. For all other analytical determinations use a solution prepared as follows: Weigh roughly 100 grams of the product into a beaker and dissolve by boiling with 200 cc. of water. Decant the resulting sirup while hot through a muslin filter, concentrate by boiling to a moisture content of 35% (b. p. 104°C.), cool, or preferably let the covered material stand overnight, set aside until clear, and use this clear sirup for analysis. It is desirable, in order to compare results upon different samples, that all results except moisture be expressed upon a dry basis.

54

MOISTURE.—TENTATIVE.

(a) *Maple sirup.*—Proceed as directed under 35 or 10.

(b) *Maple sugar, maple cream, etc.*—Proceed as directed under 35.

55

POLARIZATION.—TENTATIVE.

(a) *Direct at 20°C.*—Proceed as directed under VIII, 14.

(b) *Invert at 20°C.*—Proceed as directed under VIII, 14.

(c) *Invert at 87°C.*—Proceed as directed under 25 to detect commercial glucose.

56 **REDUCING SUGARS AS INVERT SUGAR.—TENTATIVE.**

(a) *Before inversion.*—Proceed as directed under **VIII, 25**, using an aliquot of the solution used for direct polarization, **55 (a)**, and only neutral lead acetate for clarification.

(b) *After inversion.*—Proceed as directed under **VIII, 25**, using an aliquot of the solution used for the invert polarization, **55 (b)**, and only neutral lead acetate for clarification.

SUCROSE.**57** *By Polarization.—Tentative.*

Proceed as directed under **VIII, 14** or **16**.

58 *By Reducing Sugars Before and After Inversion.—Tentative.*

Proceed as directed under **VIII, 18**.

59 **TOTAL ASH.—TENTATIVE.**

Proceed as directed under **13**.

60 **SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.**

Proceed as directed under **17**.

61 **ALKALINITY OF THE SOLUBLE ASH.—TENTATIVE.**

Proceed as directed under **18**.

62 **ALKALINITY OF THE INSOLUBLE ASH.—TENTATIVE.**

Proceed as directed under **19**.

LEAD NUMBER (WINTON).—TENTATIVE.**63** **REAGENTS.**

Standard basic lead acetate solution.—Boil 430 grams of normal lead acetate and 130 grams of litharge, for 30 minutes, or boil 560 grams of Horne's dry basic lead acetate with 1 liter of water, cool, allow to settle and dilute the supernatant liquid to 1.25 sp. gr. To a measured amount of this solution add 4 volumes of water and filter if not perfectly clear. The solution should be standardized each time a set of determinations is made.

If the directions for preparing the basic lead acetate are not carried out carefully, the use of Horne's dry basic lead acetate is preferable.

64 **DETERMINATION OF LEAD IN THE BLANK.**

Transfer 25 cc. of the standard basic lead acetate to a 100 cc. flask, add a few drops of acetic acid, and make up to the mark with water. Shake and determine lead sulphate in 10 cc. of the solution as directed under **65**. The use of the acid is imperative in this case to keep the lead in solution, when diluted with water.

65 **DETERMINATION.**

Transfer 25 grams of the sample to a 100 cc. flask by means of water. Add 25 cc. of the standard basic lead acetate and shake, fill to the mark, shake, and allow to stand for at least 3 hours before filtering. Pipette 10 cc. of the clear filtrate into a 250 cc. beaker, add 40 cc. of water and 1 cc. of concentrated sulphuric acid, shake and add 100 cc. of 95% alcohol. Allow to stand overnight, filter on a tared Gooch,

wash with 95% alcohol, dry in a water oven, and ignite in a muffle or over a Bunsen burner, applying the heat gradually at first, and avoiding a reducing flame. Cool and weigh. Subtract the weight of lead sulphate so found from the weight of lead sulphate found in the blank, 64, and multiply by the factor 27.325. The use of this factor gives the lead number directly without the various calculations otherwise required.

MALIC-ACID VALUE.

66

*Cowles Method.*¹⁴—*Tentative.*

Weigh 6.7 grams of the sample into a 200 cc. beaker, add 5 cc. of water, then 2 cc. of a 10% calcium acetate solution and stir. Add gradually, and with constant stirring, 100 cc. of 95% alcohol, and agitate the solution until the precipitate settles, or let stand, until the supernatant liquid is clear. Filter off the precipitate and wash with 75 cc. of 85% alcohol. Dry the filter paper and ignite in a platinum dish. Add 10 cc. of N/10 hydrochloric acid and warm gently until all the lime dissolves. Cool and titrate back with N/10 sodium hydroxid, using methyl orange as an indicator. The difference in cc. divided by 10 represents the malic acid value of the sample. Previous to use the reagents should be tested by a blank determination and any necessary corrections applied.

67

METALS.—TENTATIVE.

Proceed as directed under XII.

SUGAR HOUSE PRODUCTS.

SUCROSE IN BEETS.

68

*Alcohol Extraction Method (Herzfeld Modification)*¹⁵.—*Tentative.*

Weigh 26 grams of the beet pulp and transfer to a 100 cc. flask with about 50 cc. of 90% alcohol and 3–5 cc. of basic lead acetate solution. Connect a reflux condenser to the flask and place on a boiling water bath for 10–15 minutes. Then pour the whole into a Soxhlet extractor, washing out the flask with fresh portions of 90% alcohol. Connect the same 100 cc. flask to the extractor, and fit the latter with a return condenser. Add 90% alcohol until the siphon is started and the flask is about three fourths full. Place the flask in a covered water bath kept at a heat that will allow the alcohol to boil freely. Continue the extraction for 1–4 hours, or until a test of the alcohol in the extractor gives no color with alpha-naphthol solution when tested as follows: Introduce into a test tube a few drops of the alcohol coming from the extractor, add 4 or 5 drops of a 20% alcoholic alpha-naphthol solution and 2 cc. of water. Shake well, tip the tube, and allow 2–5 cc. of colorless concentrated sulphuric acid to flow down the side of the tube; then hold the tube upright and, if sucrose is present, a color varying from a faint to a deep violet will be noted at the junction of the two liquids. On shaking, the whole solution becomes a blue violet color. This test is suitable for this work, but it must be remembered that other substances besides sucrose give this color reaction.

Remove the flask, transfer to a 100 cc. graduated flask, cool to the standard temperature, dilute to the mark with 90% alcohol, shake and filter, keeping the funnel covered with a watch glass. Polarize in a 200 mm. tube.

Avoid evaporation and changes of temperature and also use a minimum amount of basic acetate for clarification, 3 cc. rather than 5 cc. By digesting the beet pulp with the alcohol before extraction, the time of extraction is greatly shortened the pulp becomes thoroughly impregnated with the alcohol, and all the air is removed, resulting in a good extraction of the whole material. If the pulp is fine

and tends to clog the siphon, alcohol-washed cotton may be used as a plug in the extractor before adding the beet pulp, and a fine mesh screen may be placed over the pulp to keep the whole compact in the extractor.

69 *Pellet Aqueous Method*¹⁶ (*Hot Digestion*).—*Tentative.*

Weigh 52 grams of the beet cuttings and transfer them with water to a wide-mouthed flask graduated to a content of 201.2 cc.; add 5–10 cc. of basic lead acetate solution, fill the flask to the mark with hot water, and shake. Immerse the flask in a water bath at 80°C. and rotate at intervals. Add water from time to time so that at the end of the heating (about 30 minutes) the water in the flask is a little above the mark. Remove the flask from the water bath and allow it to cool to standard temperature. Add sufficient concentrated acetic acid to make the solution very slightly acid (generally less than 0.5 cc.) and a few drops of ether to break the foam. Make up to the mark, mix thoroughly, filter, and polarize in a 200 mm. tube.

The fineness of the pulp governs the time of heating. Add enough water at the start and maintain this volume during the extraction, so that not more than 5 cc. of water will be necessary to complete the volume after cooling. The proportion of pulp to water must not be increased beyond the prescribed amount, for when smaller proportions of water to pulp are used and then a large quantity of water is added at the last to make up to volume, the sugar does not become equally diffused and the results are too low. Differences of over 1% in sugar content may be caused by lack of care in this particular.

70 *Hot Water Digestion Method*.—*Tentative.*

(*Herzfeld Modification of the Sachs Le Docte Method*¹⁷.)

There are needed nickel-plated sheet iron vessels, 11 cm. high, 6 cm. body diameter, and 4 cm. mouth diameter, also stoppers covered with tin foil to fit the same.

Weigh 26 grams of the beet pulp on a watch glass (small enough to go into the neck of the beaker) and transfer to the metal beaker, add 177 cc. of dilute basic lead acetate solution (5 parts of basic lead acetate solution (sp. gr. 1.25) to 100 parts of water), shake and stopper lightly. Submerge the beaker in a water bath at 75°–80°C. for 30 minutes, shaking intermittently. When all the air has been expelled (generally after 5 minutes), tighten the stopper. After 30 minutes, shake, cool to standard temperature, filter, add a drop of acetic acid to the filtrate and polarize in a 400 mm. tube. The reading is the per cent of sugar in the beet pulp.

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- ¹⁴ J. Am. Chem. Soc., 1908, 30: 1285.
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X. FOOD PRESERVATIVES.—TENTATIVE.

SALICYLIC ACID.

1

PREPARATION OF SAMPLE.

(a) *Non-alcoholic liquids*.—Many liquids may be extracted directly as described in 2 or 4 without further treatment. If gums or mucilaginous substances are present, pipette 100 cc. into a 250 cc. volumetric flask, add about 5 grams of sodium chlorid, shake until the latter is dissolved, make up to the mark with alcohol, shake vigorously, allow the mixture to stand for 10 minutes with occasional shaking, filter through a dry folded filter and treat an aliquot of the filtrate as directed under (b).

(b) *Alcoholic liquids*.—Make 200 cc. of the sample alkaline with sodium hydroxid solution, using litmus as an indicator, and evaporate on a steam bath to about one third its original volume. Dilute to the original volume with water and filter, if necessary, through a dry filter.

(c) *Solid or semi-solid substances*.—Grind the sample and mix thoroughly. Transfer a convenient quantity (50–200 grams according to the consistency of the sample) to a 500 cc. volumetric flask, add sufficient water to make a volume of about 400 cc., shake until the mixture becomes uniform, add 2–5 grams of calcium chlorid, shake until the latter is dissolved, render distinctly alkaline with sodium hydroxid solution, using litmus as an indicator, fill to the mark with water, shake thoroughly, allow to stand for at least 2 hours shaking frequently and filter through a large folded filter.

DETECTION AND ESTIMATION.

2

Ferric Chlorid Test.—Qualitative.

Introduce 50 cc. of the sample or an equivalent amount of an aqueous extract, prepared as directed under 1, into a separatory funnel, add one tenth its volume of dilute hydrochloric acid (1 to 3) and extract with 50 cc. of ether. If the mixture emulsifies, add 10–15 cc. of petroleum ether (b. p. below 60°C.) and shake. If this treatment fails to break the emulsion whirl the mixture in a centrifuge, or allow it to stand until a considerable portion of the aqueous layer has separated, run off the latter, shake vigorously and again allow to separate. Wash the ether layer with two 5 cc. portions of water, evaporate the greater portion of the ether in a porcelain dish on a steam bath, allow the remainder to evaporate spontaneously and add a drop of 0.5% ferric chlorid solution. A violet color indicates salicylic acid.

If coloring matter or other interfering substances are present in the residue left after evaporation of the ether, purify the salicylic acid by one of the following methods:

(a) Dissolve the residue from the ether extract, obtained as directed above, in about 25 cc. of ether, transfer the latter to a separatory funnel and shake with an equal quantity of water, made distinctly alkaline with several drops of ammonium hydroxid. Allow to separate, filter the aqueous layer through a wet filter into a porcelain dish, evaporate almost to dryness, and test the residue as directed above.

(b) Dry the residue from the ether extract, obtained as directed above, in a desiccator over sulphuric acid and extract with several 10 cc. portions of carbon

disulphid or petroleum ether (b. p. below 60°C.), rubbing the contents of the dish with a glass rod, and filtering the successive portions of the solvent through a dry paper into a second porcelain dish. Evaporate the greater portion of the solvent on a steam bath, allow the remainder to evaporate spontaneously and test the residue as directed above.

(c) Transfer the residue from the ether extract, obtained as directed above, to a small porcelain crucible by means of a few cc. of ether and allow the solvent to evaporate spontaneously. Cut a hole in a piece of asbestos board sufficiently large to admit about two thirds of the crucible, cover the latter with a small, round-bottomed flask filled with cold water, and heat over a small Bunsen flame until any salicylic acid present has sublimed and condensed upon the bottom of the flask. Test the sublimate as directed above.

3

Jorissen's Test.—Qualitative.

Dissolve the residue from the ether extract, obtained as directed under 2, or, in case impurities are present, the purified material obtained as directed under 2 (a), (b) or (c) in a little hot water. Cool 10 cc. of the solution in a test tube, add 4 or 5 drops of 10% potassium nitrite solution, 4 or 5 drops of 50% acetic acid and 1 drop of 10% cupric sulphate solution, mix thoroughly and heat to boiling. Boil for half a minute and allow to stand for 1-2 minutes. In the presence of salicylic acid a blood red color will develop.

Colorimetric Method.—Quantitative.

4

EXTRACTION.

Pipette a convenient portion of the sample (100 cc. or a volume representing not less than 20 grams of the original sample) or a solution, prepared as in 1, into a separatory funnel, make the solution neutral to litmus with dilute hydrochloric acid (1 to 3) and add an excess of concentrated hydrochloric acid equivalent to 2 cc. of acid for each 100 cc. of solution. Extract with 4 separate portions of ether, using for each extraction a volume of ether equivalent to half the volume of the aqueous layer. If an emulsion forms on shaking, this may usually be broken by adding a little (one fifth the volume of the ether layer) petroleum ether (b. p. below 60°C.) and shaking again or by centrifugalizing. If an emulsion still persists, allow it to remain with the aqueous layer. If an emulsion remains after the fourth extraction, separate it from the clear ether and the clear aqueous layer and extract it separately with 2-3 small portions of ether. Combine the ether extracts, wash with one tenth their volume of water, allow the layers to separate and reject the aqueous layer. Wash in this way until the aqueous layer after separation yields a yellow color upon the addition of methyl orange and 2 drops of N/10 sodium hydroxid. Distil slowly the greater part of the ether, transfer the remainder to a porcelain dish and allow the ether to evaporate spontaneously. If there are no interfering substances present, proceed as directed in 5. If such interfering substances are present, purify the residue by one of the following methods:

(a) Dry thoroughly the residue in vacuo over sulphuric acid and extract with 10 portions of 10-15 cc. each of carbon disulphid or petroleum ether (b. p. below 60°C.), rub the contents of the dish with a glass rod and filter the successive portions of the solvent through a dry filter into a porcelain dish. Test the extracted residue with a drop of ferric alum solution and, if it gives a reaction for salicylic acid, dissolve it in water and reextract with ether, proceeding as directed above. Distil the greater portion of the carbon disulphid or petroleum ether and allow the remainder to evaporate spontaneously. Proceed as directed in 5.

(b) Dissolve the residue in 40-50 cc. of ether. Transfer the ether solution to a separatory funnel and extract with 3 successive 15 cc. portions of 1% ammonium hydroxid. (If fat is known to be present in the original ether extract, extract the latter directly with 4 portions of the ammonium hydroxid instead of 3.) Combine the alkaline aqueous extracts, acidify, again extract with ether and wash the combined ether extracts as directed above. Distil slowly the greater portion of the ether, allow the remainder to evaporate spontaneously and proceed as directed in 5.

5

DETERMINATION.

Dissolve the residue, obtained in 4, in a small amount of hot water and, after cooling, dilute to a definite volume (usually 50-100 cc.), dependent on the amount of salicylic acid present. If the solution is not clear, filter through a dry filter. Dilute aliquots of the solution and treat with a few drops of 0.5% ferric chlorid solution or 2% ferric alum solution.

The ferric alum solution should be boiled until a precipitate appears, allowed to settle, and filtered. The acidity of the solution is slightly increased in this manner, but it remains clear for a considerable time, and the turbidity caused by its dilution with water is much less and does not appear as soon as when the unboiled solution is used. This turbidity interferes with the exact matching of the color.

Compare the colors developed with that obtained when a standard salicylic acid solution (containing 1 mg. of salicylic acid in 50 cc.) is similarly treated, using Nessler tubes or a colorimeter. In either case, and especially with ferric chlorid, avoid an excess of the reagent, although an excess of 0.5 cc. of 2% ferric alum solution may be added to 50 cc. of the comparison solution of salicylic acid without impairing the results.

BENZOIC ACID.

PREPARATION OF SAMPLE.

6

General Method.

If solid or semi-solid, grind the sample, and mix thoroughly. Transfer about 150 grams to a 500 cc. graduated flask, add enough pulverized sodium chlorid to saturate the water in the sample, render alkaline with sodium hydroxid solution or milk of lime, and dilute to the mark with a saturated salt solution. Allow to stand for at least 2 hours, with frequent shaking, and filter. If the sample contains large amounts of matter precipitable by salt solution, it is advisable to follow a method similar to that given under 7 (d). When alcohol is present, follow the method given under 7 (c). When large amounts of fats are present, make an alkaline extraction of the filtrate before proceeding as directed under 11.

7

Special Methods.

(a) *Ketchup*.—Saturate the water in 150 grams of ketchup by adding 15 grams of pulverized sodium chlorid. Transfer the mixture to a 500 cc. graduated flask, rinsing with about 150 cc. of saturated sodium chlorid solution. Make slightly alkaline to litmus paper with strong sodium hydroxid solution and fill to the mark with saturated salt solution. Allow to stand for at least 2 hours, shaking frequently. Squeeze through a heavy muslin bag and then filter through a large folded filter.

(b) *Jellies, jams, preserves and marmalades*.—Dissolve 150 grams of the sample in about 300 cc. of saturated salt solution. Add 15 grams of pulverized sodium chlorid. Make alkaline to litmus paper with milk of lime. Transfer to a 500 cc. graduated flask and dilute to the mark with saturated salt solution. Allow to

stand for at least 2 hours, shaking frequently, centrifugalize if necessary, and filter through a large folded filter.

(c) *Cider containing alcohol, and similar products.*—Make 250 cc. of the sample alkaline to litmus paper with sodium hydroxid solution and evaporate on the steam bath to about 100 cc. Transfer the sample to a 250 cc. graduated flask, add 30 grams of pulverized sodium chlorid and shake until dissolved. Dilute to the original volume, 250 cc., with saturated salt solution, allow to stand for at least 2 hours, shaking frequently, and filter through a folded filter.

(d) *Salted or dried fish.*—Wash 50 grams of the ground sample into a 500 cc. graduated flask with water. Make slightly alkaline to litmus paper with strong sodium hydroxid solution and dilute to the mark with water. Allow to stand for at least 2 hours, shaking frequently, and then filter through a folded filter. Pipette accurately as large a portion of the filtrate as possible (at least 300 cc.) into a second 500 cc. flask. Add 30 grams of the pulverized sodium chlorid for each 100 cc. of solution. Shake until the salt has dissolved and dilute to the mark with saturated salt solution. Mix thoroughly and filter off the precipitated protein matter on a folded filter.

8

DETECTION AND ESTIMATION.

Extract benzoic acid as directed under 2 or 4. If benzoic acid is present in considerable quantity, it will crystallize from the ether in shining leaflets having a characteristic odor on heating. Dissolve the residue in hot water, divide into 2 portions, and test according to 9 or 10.

9

Ferric Chlorid Test.—Qualitative.

Make the solution from 8 alkaline with ammonium hydroxid, expel the excess of ammonia by evaporation, dissolve the residue in water, and add a few drops of a neutral 0.5% ferric chlorid solution. A brownish precipitate of ferric benzoate indicates the presence of benzoic acid.

10

*Modified Mohler Test.*²—Qualitative.

Add to the water solution, prepared as described under 8, 1–3 cc. of N/3 sodium hydroxid and evaporate to dryness. To the residue, add 5–10 drops of concentrated sulphuric acid and a small crystal of potassium nitrate. Heat for 10 minutes in a glycerol bath at 120°–130°C., or for 20 minutes in a boiling water bath. The temperature must not exceed 130°C. After cooling add 1 cc. of water and make distinctly ammoniacal; boil the solution to decompose any ammonium nitrite which may have been formed. Cool and add a drop of fresh, colorless ammonium sulphid, without allowing the layers to mix. A red-brown ring indicates benzoic acid. On mixing, the color diffuses through the whole liquid and, on heating, finally changes to greenish yellow. This differentiates benzoic acid from salicylic acid or cinnamic acid. The last two form colored compounds, which are not destroyed by heating. The presence of phenolphthalein interferes with this test.

11

Quantitative Method.

Pipette a convenient portion (100–200 cc.) of the filtrate, obtained in 6 or 7, into a separatory funnel. Neutralize the solution to litmus paper with hydrochloric acid (1 to 3) and add an excess of 5 cc. of the same acid. In the case of salted fish a precipitation of protein matter usually occurs on acidifying, but the precipitate does not interfere with the extraction. Extract carefully with chloroform, using successive portions of 70, 50, 40, and 30 cc. To avoid an emulsion, shake cautiously

each time. The chloroform layer usually separates readily after standing a few minutes. If an emulsion forms, break it: (1) by stirring the chloroform layer with a glass rod; (2) by drawing it off into a second funnel and giving 1 or 2 sharp shakes from one end of the funnel to the other; or (3) by centrifugalizing for a few moments. As this is a progressive extraction, draw off carefully as much of the clear chloroform solution as possible after each extraction, but do not draw off any of the emulsion with the chloroform layer. If this precaution is taken, the chloroform extract need not be washed.

Transfer the combined chloroform extracts to a porcelain evaporating dish, rinse the container several times with a few cc. of chloroform, and evaporate to dryness at room temperature in a current of air dried over calcium chlorid.

The extract may also be transferred from the separatory funnel to a 300 cc. Erlenmeyer flask, rinsing the separatory funnel 3 times with 5-10 cc. of chloroform. Distil very carefully to about one fourth the original volume, keeping the temperature down so that the chloroform comes over in drops, not in a steady stream. Then transfer the residue to a porcelain evaporating dish, rinsing the flask 3 times with 5-10 cc. portions of chloroform, and allow to evaporate to dryness spontaneously.

Dry the residue overnight (or until no odor of acetic acid can be detected if the product is a ketchup) in a desiccator containing sulphuric acid. Dissolve the residue of benzoic acid in 30-50 cc. of neutral alcohol, add about one fourth this volume of water, 1 or 2 drops of phenolphthalein, and titrate with N/20 sodium hydroxid (1 cc. is equivalent to 0.0072 gram of anhydrous sodium benzoate).

SACCHARIN.

12

Qualitative Test.

Extract with ether (after maceration and exhaustion with water, if necessary), as directed in 1 and 4. Allow the ether extract to evaporate spontaneously and note the taste of the residue. The presence of saccharin, to the extent of 20 mg. per liter, is indicated by a sweet taste. Confirm by heating with sodium hydroxid, as described below, and detecting the salicylic acid formed thereby. A sweet taste, suggesting the presence of a trace of saccharin, has been obtained frequently in saccharin-free wines, due to the so-called "false saccharin".

Acidify 50 cc. of a liquid food or the aqueous extract of 50 grams of a solid or semi-solid, prepared as directed in 1 (c), and extract with ether as directed in 13. Dissolve the residue, remaining after evaporation of the ether, in a little hot water and test a small portion of this solution for salicylic acid as directed under 2 or 3. Dilute the remainder of the solution to about 10 cc., and add 2 cc. of sulphuric acid (1 to 3). Heat to boiling and add a slight excess of 5% potassium permanganate solution, drop by drop; partly cool the solution, dissolve a piece of sodium hydroxid in it, and filter the mixture into a silver dish (silver crucible lids are well adapted to the purpose); evaporate to dryness and heat for 20 minutes at 210°-215°C. Dissolve the residue in water, acidify with hydrochloric acid and test the ether extract for salicylic acid as directed under 2 or 3. By this method all the so-called "false saccharin" and the salicylic acid naturally present (also added salicylic acid when not present in too large an amount) are destroyed, while 5 mg. of saccharin per liter are detected with certainty.

13

Quantitative Method.

Pipette 100 cc. of the sample, or a convenient portion of a solution, prepared as directed under 1, representing not less than 20 grams of the sample, into a sepa-

ratory funnel, make the solution neutral to litmus with dilute hydrochloric acid (1 to 3) and then add concentrated hydrochloric acid in the proportion of 5 cc. for each 100 cc. of solution. Extract with 4 separate portions of ether using, for each extraction, a volume of ether equivalent to half the volume of the aqueous layer. If the mixture emulsifies on shaking, this difficulty may be overcome as directed under 4. Wash the combined ether extracts with two 5 cc. portions of water, remove the ether by distillation, and transfer the residue into a platinum crucible by means of a small amount of ether. Evaporate the ether on a steam bath, add about 2-3 cc. of 10% sodium carbonate solution to the residue, rotate so that all of the residue is brought into contact with the solution, and evaporate to dryness on a steam bath. Add 4 grams of a mixture of equal parts of anhydrous sodium and potassium carbonates, heat gently at first, and then to complete fusion for 30 minutes over an alcohol or other sulphur-free flame. Cool, dissolve the melt in water, acidify with hydrochloric acid and determine the sulphate present as barium sulphate. Correct the result thus obtained for any sulphur present in the fusion mixture as found in a blank determination. Calculate the amount of saccharin in the sample by multiplying the weight of barium sulphate by 0.7845.

BORIC ACID AND BORATES.

14

*Qualitative Test.*³

Preliminary test.—Immerse a strip of turmeric paper in the sample acidified with hydrochloric acid in the proportion of 7 cc. of concentrated acid to each 100 cc. of sample, and allow the paper to dry spontaneously. If borax or boric acid is present, the paper will acquire a peculiar red color, changed by ammonium hydroxid to a dark blue-green but restored by acid. Solid or pasty samples may be heated with enough water to make them sufficiently fluid, concentrated hydrochloric acid added in about the proportion of 1 to 13 and the liquid tested in the same way.

Confirmatory test.—Make about 25 grams of the sample decidedly alkaline with lime water and evaporate to dryness on a water bath. Ignite the residue to destroy organic matter. Digest with about 15 cc. of water, add concentrated hydrochloric acid, drop by drop, until the ignited residue is dissolved, and then add 1 cc. in excess. Saturate a piece of turmeric paper with the solution, and allow it to dry without the aid of heat. In the presence of borax or boric acid, the color change will be the same as given above.

15

*Quantitative Method.*⁴

Make 10-100 grams of the sample (depending upon the nature of the sample and the amount of boric acid present) distinctly alkaline with sodium hydroxid solution and evaporate to dryness in a platinum dish. Ignite the residue until organic matter is destroyed, avoiding an intense red heat, cool, digest with about 20 cc. of hot water, and add hydrochloric acid, drop by drop, until the reaction is distinctly acid. Filter into a 100 cc. flask, and wash with a little hot water, the volume of the filtrate not to exceed 50-60 cc. Return the filter containing any unburned carbon to the platinum dish, make alkaline by wetting thoroughly with lime water, dry on a steam bath and ignite to a white ash. Dissolve the ash in a few cc. of dilute hydrochloric acid and add to the liquid in the 100 cc. flask, rinsing the dish with a few cc. of water. To the combined solutions, add 0.5 gram of calcium chlorid and a few drops of phenolphthalein, then 10% sodium hydroxid solution until a permanent light pink color is produced, and finally dilute to

the mark with lime water. Mix and filter through a dry filter. To 50 cc. of the filtrate add N/1 sulphuric acid until the pink color disappears, then add methyl orange, and continue the addition of the acid until the yellow color is changed to pink. Boil for about 1 minute to expel carbon dioxide. Cool, and carefully add N/5 sodium hydroxid until the liquid assumes a yellow tinge, avoiding an excess of the alkali. All the boric acid is now in a free state with no uncombined sulphuric acid present. Add a little phenolphthalein, and an equal volume of neutral glycerol. Titrate with N/5 sodium hydroxid until a permanent pink color is produced. About 10 grams of mannitol may be substituted for the glycerol in this determination. At the end of the titration add an additional 2 grams and continue the titration if the pink color is discharged. Repeat the alternate addition of mannitol and alkali until a permanent end point is reached.

One cc. of N/5 sodium hydroxid is equivalent to 0.0124 gram of boric acid.

FORMALDEHYDE.

16

PREPARATION OF SAMPLE.

If solid or semi-solid, macerate 200–300 grams of the material with about 100 cc. of water in a mortar. Transfer to a short-necked, 500–800 cc. copper or glass distillation flask and make distinctly acid with phosphoric acid, connect with a condenser and distil 40–50 cc. In the case of highly colored liquids, the same method of preparation should be employed.

In the case of meats and fats, extract the formaldehyde with alcohol and use the filtrate. In the case of fat, heat the mixture above the melting point of the fat to insure thorough extraction. In the case of milk, shake with an equal volume of strong alcohol and use the filtrate. Shake other liquids with an equal volume of strong alcohol and filter from any insoluble matter.

QUALITATIVE TESTS.

17

*Phenylhydrazin Hydrochlorid Method.*⁵

Mix 5 cc. of the distillate, as prepared under 16, or of an alcoholic solution or extract obtained as directed above, with 0.03 gram of phenylhydrazin hydrochlorid, and 4 or 5 drops of a 1% ferric chlorid solution. Add slowly and with agitation, in a bath of cold water to prevent heating the liquid, 1–2 cc. of concentrated sulphuric acid. Dissolve the precipitate by the addition either of concentrated sulphuric acid (keeping the mixture cool) or alcohol. In the presence of formaldehyde a red color develops.

This method gives reliable reactions for formaldehyde in solutions of formaldehyde varying from 1 part in 50,000 to 1 part in 150,000. Acetaldehyde and benzaldehyde give no reaction when treated by this method and do not interfere with the reaction given by formaldehyde.

18

*Hehner Method.*⁶

Mix about 5 cc. of the distillate, obtained in 16, with an equal volume of pure milk, or a 1–2% solution of egg albumen, in a test tube and underlay with strong commercial sulphuric acid without mixing. A violet or blue color at the junction of the two liquids indicates formaldehyde. This color is given only in the presence of a trace of ferric chlorid or other oxidizing agent. As pointed out by Hehner, milk may be treated directly by this method and gives positive tests in the presence of 1 or more parts of formaldehyde per 10,000. Some other articles of food rich in proteins, for example, egg albumen, give the reaction in the presence of water without the addition of milk.

19

Leach Method.

Mix about 5 cc. of the distillate, obtained under **16**, with an equal volume of pure milk in a porcelain casserole and add about 10 cc. of concentrated hydrochloric acid, containing 1 cc. of 10% ferric chlorid solution, to each 500 cc. of acid. Heat to 80°–90°C. directly over the gas flame, rotating the casserole to break up the curd. A violet coloration indicates formaldehyde.

Rimini Method.⁷

20

Phenylhydrazin Hydrochlorid and Sodium Nitro-prussid Test.

This method may be applied directly to liquid foods, to an aqueous or alcoholic extract of solid foods, or to the distillate prepared as directed in **16**. In the case of milk, apply the method directly. In the case of meat, comminute the sample, extract with 2 volumes of hot water, and employ the expressed liquid for the test. Heat fats above their melting point with 10 cc. of alcohol, shake thoroughly, cool, filter through a moistened filter, and use the filtrate for the test.

Dissolve a lump of phenylhydrazin hydrochlorid about the size of a pea in 3–5 cc. of the liquid to be tested, add 2–4 drops (not more) of a 5–10% sodium nitro-prussid solution and 8–12 drops of an approximately 12% sodium hydroxid solution. If formaldehyde is present, a green or blue color develops depending upon the amount. When formaldehyde is present to the extent of more than 1 part in 70,000–80,000 in the solution tested, a distinct green or bluish green reaction is obtained. In more dilute solutions the green tint becomes less marked and a yellow tinge tending toward greenish brown develops.

With this method acetaldehyde and benzaldehyde give a color varying from red to brown, according to the strength of the solution. A reaction may therefore be obtained with these aldehydes similar to that obtained with formaldehyde in solutions more dilute than 1 part in 70,000. The presence of acetaldehyde or benzaldehyde together with formaldehyde gives a yellowish or yellowish green tinge. The reaction for formaldehyde may therefore be masked by the presence of other aldehydes, but is characteristic when a clear green color is obtained.

21

Phenylhydrazin Hydrochlorid and Potassium Ferricyanid Test.

Proceed as directed in **20**, substituting a solution of potassium ferricyanid for the sodium nitro-prussid. Formaldehyde gives a red color. Alcoholic extracts from foods must be diluted with water to prevent the precipitation of potassium ferricyanid. The test is not applicable in the presence of the coloring matter of blood.

22

Phenylhydrazin Hydrochlorid and Ferric Chlorid Test.

Treat 15 cc. of milk or other liquid food or of the distillate, prepared as directed under **16**, with 1 cc. of a dilute phenylhydrazin hydrochlorid solution, then with a few drops of dilute ferric chlorid solution and, finally, with concentrated hydrochloric acid. The presence of formaldehyde is indicated by the formation of a red color, which changes after some time to orange yellow.

Milk may be examined directly by this method, but more delicate tests may be obtained from the distillate from milk or from milk serum. Acetaldehyde or benzaldehyde does not interfere with the reaction.

23

*Phloroglucol Method.*³

To 10 cc. of milk or other liquid food under examination in a test tube add, by means of a pipette, 2 cc. of phloroglucol reagent (1 gram of phloroglucol, 20 grams of sodium hydroxid and water to make 100 cc.), placing the end of the pipette on the bottom of the tube in such a manner that the reagent will form a separate layer.

If form^{the color} be present, a bright red coloration (not purple) forms at the zone of co^l. This solution gives a yellow color in the presence of some aldehydes, and, if it is used for the detection of aldehyde formed by the oxidation of methyl alcohol after the destruction of ethyl aldehyde with hydrogen peroxid, an orange yellow color will slowly appear when an insufficient amount of hydrogen peroxid has been employed. On the other hand, if the excess of hydrogen peroxid is not fully destroyed before the use of this reagent, a purple color develops slowly. The clear, red color given by the use of this reagent forms quickly, and, in the presence of but a small amount of formaldehyde fades rapidly.

FLUORIDS.

QUALITATIVE TESTS.

24

*Method I.—Modified Method of Blarez.*⁹

Thoroughly mix the sample and boil 150 cc. (in the case of solid foods an aqueous extract may be employed provided the fluorids are in a soluble form). Add to the boiling liquid 5 cc. of 10% potassium sulphate solution and 10 cc. of 10% barium acetate solution. Collect the precipitate in a compact mass (a centrifuge may be used advantageously) and wash upon a small filter. Transfer to a platinum crucible and ignite.

Dip a carefully cleaned glass plate, while hot, in a mixture of equal parts of Carnaüba wax and paraffin and allow to cool. Make, with a sharp instrument, a distinctive mark through the wax, taking care not to scratch the surface of the glass.

Add a few drops of concentrated sulphuric acid to the residue in the crucible and cover with the waxed plate, having the mark nearly over the center and making sure that the edge of the crucible is in close contact with it. Keep the top surface of the plate cool by means of a suitable device and heat the crucible for an hour at as high a temperature as practicable without melting the wax (an electric stove gives the most satisfactory form of heat).

If fluorids be present, a distinct etching will be apparent on the glass where it was exposed.

25

Method II.

The preceding method may be varied by mixing a small amount of precipitated silica with the precipitated barium fluorid and applying the method for the detection of fluosilicates, under 27 or 28.

This method is of value in the case of foods whose ash contains a considerable amount of silica. Under these circumstances, concentrated sulphuric acid liberates silicon fluorid, which would escape detection under 24.

FLUOBORATES AND FLUOSILICATES.

26

PREPARATION OF SAMPLE.

Make about 200 grams of the sample alkaline with lime water, evaporate to dryness, and incinerate. Extract the crude ash with water, to which sufficient acetic

acid has been added to decompose carbonates, filter, ignite the insoluble portion, extract with dilute acetic acid, and again filter. The insoluble portion now contains calcium silicate and fluorid, while the filtrate will contain all the boric acid present.

QUALITATIVE TESTS.

27

Method I.¹⁰

ak up

Incinerate the filter, from 26, containing the insoluble portion, ~~mix~~ with a little precipitated silica, transfer to a short test tube, attached to a small U-tube containing a few drops of water and add 1-2 cc. of concentrated sulphuric acid. Keep the test tube in a beaker of water on the steam bath for 30-40 minutes. If any fluorin be present, the silicon fluorid generated will be decomposed by the water in the U-tube and will form a gelatinous deposit on the walls of the tube.

Next test the filtrate as directed under 14. If both hydrofluoric and boric acids be present, it is probable that they are combined as borofluorid. If, however, silicon fluorid is detected and not boric acid, the operation should be repeated without the introduction of the silica, in which case the formation of the silicon skeleton is conclusive evidence of the presence of fluosilicate. In an ash containing an appreciable amount of silica, sulphuric acid will liberate silicon fluorid rather than hydrofluoric acid. The presence of a fluosilicate is indicated, therefore, and not the presence of a fluorid.

28

Method II.

Incinerate the filter, from 26, containing the insoluble portion, in a platinum crucible, mix with a little precipitated silica, and add 1 cc. of concentrated sulphuric acid. Cover the crucible with a watch glass, from the underside of which a drop of water is suspended, and heat for an hour at 70°-80°C., keeping the watch glass cooled. The silicon fluorid which is formed is decomposed by the water, leaving a gelatinous deposit of silica and etching a ring at the periphery of the drop of water. Test the filtrate for boric acid as directed under 14.

SULPHUROUS ACID.

29

Qualitative Test.¹¹

Add some sulphur-free zinc, and several cc. of hydrochloric acid to about 25 grams of the sample (with the addition of water, if necessary) in a 200 cc. Erlenmeyer flask. In the presence of sulphites, hydrogen sulphid will be generated and may be detected with lead acetate paper. Traces of metallic sulphids are occasionally present in vegetables, and will give the same reaction as sulphites under the conditions of the above test. Positive results obtained by this method should be verified by the distillation method under 30.

It is always advisable to make the quantitative determination of sulphites, owing to the danger that the test may be due to traces of sulphids. A trace is not to be considered sufficient indication of the presence of sulphur dioxid either as a bleaching agent or as a preservative.

TOTAL SULPHUROUS ACID.

30

Method I.—Distillation Method.

Distil 20-100 grams of the sample (adding recently boiled water if necessary) in a current of carbon dioxid, after the addition of about 5 cc. of a 20% glacial phos-

phoric acid solution, until 150 cc. have passed over. Collect the distillate in about 100 cc. of nearly saturated bromin water, allowing the end of the condenser to dip below the surface. The method and apparatus may be simplified without material loss in accuracy by omitting the current of carbon dioxid, adding 10 cc. of phosphoric acid instead of 5 cc., and dropping into the distillation flask, immediately before attaching the condenser, a piece of sodium bicarbonate weighing not more than 1 gram. The carbon dioxid liberated is not sufficient to expel the air entirely from the apparatus, but will prevent oxidation to a large extent. When the distillation is finished, boil off the excess of bromin, dilute the solution to about 250 cc., add 5 cc. of hydrochloric acid (1 to 3), heat to boiling, and precipitate the sulphuric acid with 10% barium chlorid solution. Boil for a few minutes longer, allow to stand overnight in a warm place, filter on a weighed Gooch, wash with hot water, ignite at a dull red heat, and weigh as barium sulphate.

31*Method II.—Direct Titration Method.*

In the examination of wine, fairly accurate results may be obtained by the following method:

Place 25 cc. of 5.6% potassium hydroxid solution in a 200 cc. flask. Introduce 50 cc. of the sample, mix with the potassium hydroxid solution, and allow the mixture to stand for 15 minutes with occasional agitation. Add 10 cc. of sulphuric acid (1 to 3) and a few cc. of starch solution, and titrate the mixture with N/50 iodine solution. Introduce the iodine solution as rapidly as possible and continue the addition until the blue color persists for several minutes. One cc. of N/50 iodine is equivalent to 0.00064 gram of sulphur dioxide.

DETERMINATION OF FREE SULPHUROUS ACID.**32***(Especially Adapted to Wine.)*

Treat 50 cc. of the sample in a 200 cc. flask with about 5 cc. of sulphuric acid (1 to 3) add about 0.5 gram of sodium carbonate to expel the air, and titrate the sulphurous acid with N/50 iodine, as directed under **31**.

BETA-NAPHTHOL.**33***Qualitative Test.*

Extract 200 cc. of the sample, or of its aqueous extract, prepared as directed under **1 (C)**, with 10 cc. of chloroform in a separatory funnel. To the chloroform extract in a test tube add a few drops of alcoholic potash, and place in a boiling water bath for 2 minutes. The presence of beta-naphthol is indicated by the formation of a deep blue color, which changes to green and then to yellow.

ABRASTOL.**QUALITATIVE TESTS.****34***Sinibaldi Method.¹²*

Make 50 cc. of the sample alkaline with a few drops of ammonium hydroxid and extract with 10 cc. of amyl alcohol, adding ethyl alcohol if an emulsion is formed. Decant the amyl alcohol, filter if turbid, and evaporate to dryness. Add to the residue 2 cc. of nitric acid (1 to 1), heat on the water bath until half of the liquid is evaporated, and transfer to a test tube with the addition of 1 cc. of water. Add about 0.2 gram of ferrous sulphate and an excess of ammonium hydroxid, drop by

drop, with constant shaking. If the resultant precipitate is of a reddish color, dissolve it in a few drops of sulphuric acid, and add ferrous sulphate and ammonium hydroxid as before. As soon as a dark colored or greenish precipitate is obtained, introduce 5 cc. of alcohol, dissolve the precipitate in sulphuric acid, shake well and filter. In the absence of abrastol this method gives a colorless or light yellow liquid, while a red color is produced in the presence of 0.01 gram of abrastol.

35 *Sanglé-Ferrière Method.*¹³

Boil 200 cc. of the sample with 8 cc. of concentrated hydrochloric acid for an hour in a flask fitted with a reflux condenser. Abrastol is thus converted into betanaphthol and is detected as directed under 33.

SUCROL OR DULCIN.

QUALITATIVE TESTS.

36 *Morpurgo Method.*¹⁴

Evaporate about 100 cc. of the sample, or of the aqueous extract prepared as directed under 1 (C) and neutralized with acetic acid, to a sirupy consistency after the addition of about 5 grams of lead carbonate, and extract the residue several times with 90% alcohol. Evaporate the alcoholic extract to dryness, extract the residue with ether, and allow the ether to evaporate spontaneously in a porcelain dish. Add 2 or 3 drops each of phenol and concentrated sulphuric acid and heat for about 5 minutes on the water bath, cool, transfer to a test tube and overlay with ammonium hydroxid or sodium hydroxid solution with the least possible mixing. The presence of dulcin is indicated by the formation of a blue color at the zone of contact.

37 *Jorissen Method.*¹⁵

Suspend the residue from the ether extract obtained as directed above in about 5 cc. of water; add 2-4 cc. of an approximately 10% solution of mercuric nitrate, and heat for 5-10 minutes on the water bath. In the presence of sucrol a violet blue color is formed, which is changed to a deep violet on the addition of lead peroxid.

FORMIC ACID.

*Quantitative Method.*¹⁶

38

REAGENTS.

(a) *Sodium acetate solution.*—Dissolve 50 grams of dry sodium acetate in sufficient water to make 100 cc. and filter.

(b) *Mercuric chlorid reagent.*—Dissolve 100 grams of mercuric chlorid and 150 grams of sodium chlorid in sufficient water to make 1 liter and filter.

(c) *Tartaric acid.*

(d) *Barium carbonate.*

39

APPARATUS.

The apparatus required (Fig. 6) consists of a steam generator (S), a 300 cc. flask (A) in which the sample is placed, a 500 cc. flask (B), containing a suspension of barium carbonate, a spray trap (T), a condenser, and a 1 liter graduated flask (C). The tip of the tube (D), leading into (B), consists of a bulb containing a number of small holes to break the vapor into small bubbles.

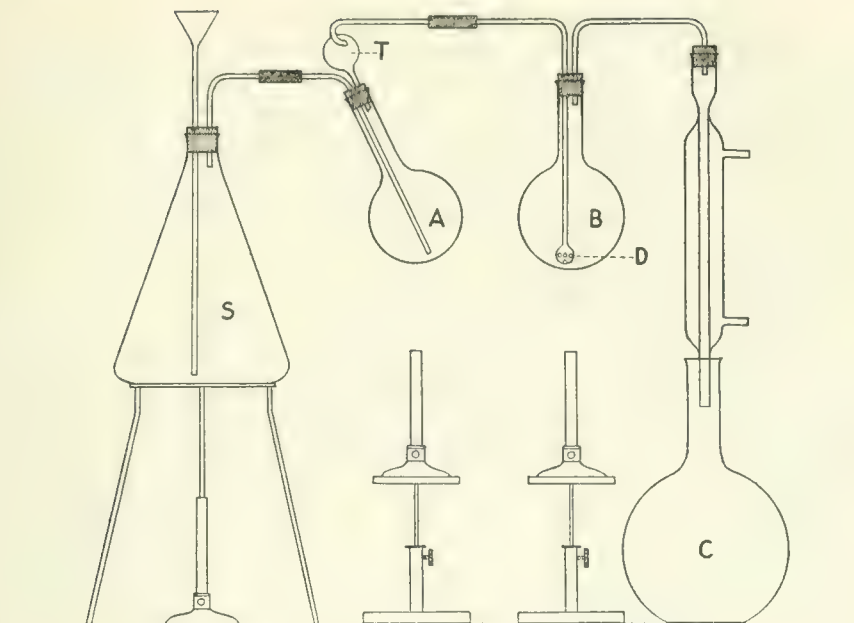


FIG. 6. APPARATUS FOR DETECTION OF FORMIC ACID.

40

DETERMINATION.

For thin liquids like fruit juices, use 50 cc. For heavy liquids and semi-solids like sirups and jams, use 50 grams diluted with 50 cc. of water. Place the sample in the flask (A), add 1 gram of tartaric acid, and connect as shown in Fig. 6, the flask (B) having been charged previously with a suspension of 2 grams of barium carbonate in 100 cc. of water. If much acetic acid is present, sufficient barium carbonate must be used so that at least 1 gram remains at the end of the operation. Heat the contents of flasks (A) and (B) to boiling and distil with steam from the generator (S), the vapor passing first through the sample in flask (A), then through the boiling suspension of barium carbonate in (B), after which it is condensed, and measured in the graduated flask (C). Continue the distillation until 1 liter of distillate is collected, maintaining the volume of the liquids in the flasks (A) and (B) as nearly constant as possible by heating with small Bunsen flames, and avoiding charring of the sample in the flask (A). After 1 liter of distillate has been collected, disconnect the apparatus and filter the contents of flask (B) while hot, washing the barium carbonate with a little hot water. The filtrate and washings should now measure about 150 cc. If not they should be boiled down to that volume. Then add 10 cc. of the sodium acetate, 2 cc. of 10% hydrochloric acid, and 25 cc. of the mercuric chlorid reagent. Mix thoroughly and immerse the container in a boiling water bath or steam bath for 2 hours. Then filter on a tared Gooch, wash the precipitate thoroughly with cold water and finally with a little alcohol. Dry in a boiling water oven for 30 minutes, cool, weigh, and calculate the weight of formic acid present by multiplying the weight of the precipitate by 0.0975. If the weight of mercurous chlorid obtained exceeds 1.5 grams, the determination must be repeated, using more mercuric chlorid reagent or a smaller amount of sample. A blank

test should be conducted with each new lot of reagents employed in the reduction, using 150 cc. of water, 1 cc. of 10% barium chlorid solution, 2 cc. of 10% hydrochloric acid, 10 cc. of the sodium acetate, and 25 cc. of the mercuric chlorid reagent, heating the mixture in a boiling water bath or steam bath for 2 hours. The weight of mercurous chlorid obtained in this blank test must be deducted from that obtained in the regular determination.

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XI. COLORING MATTERS IN FOODS.—TENTATIVE.

(An italicized number, following immediately the name of a dye, is the number by which that dye is designated in "A Systematic Survey of the Organic Colouring Matters", 1904, by Arthur Green, based on the German of Schultz and Julius.)

1

PIGMENTS.

The insoluble pigments, ultramarine, lampblack, etc., are most commonly used as facings and may be separated by washing the sample with water and allowing the washings to settle. The particles of coloring matter can be identified by microscopic examination and by treatment of the residue or purified coloring matter with chemical reagents. A large proportion of the common pigments other than lakes, such as the yellow, brown and red ochres and umbers, are derivatives of the heavy metals and contain iron, manganese, etc. Others, such as various green and blue compounds, including the green chlorophyll derivatives, contain copper. These pigments may be identified by the usual tests for the respective metals. The analytical properties of the insoluble coloring matters are described in various standard works, some of which are listed in the bibliography, especially "Farbstofftabellen by Schultz¹."

SOLUBLE COLORING MATTERS AND THEIR LAKES.

COAL TAR DYES.

2

Wool dyeing test².

(a) *Wines, fruit juices, distilled liquors, flavoring extracts, vinegars, beers, sirups, non-alcoholic beverages and similar products.*—Dilute 20–200 cc. of the sample with 1–3 volumes of water and boil or heat on the steam bath with a small piece of white woolen cloth (nun's veiling). When the mixture contains much alcohol, heat until most of the alcohol has been removed; in other cases, take out the wool after 5–15 minutes and rinse with water. Then treat the liquid with 3 or 4 drops of concentrated hydrochloric acid for each 100 cc. and warm again for 10–20 minutes with a clean piece of wool. The basic dyes go on the fiber best from neutral or faintly ammoniacal solutions and, if present, will appear on the first piece of wool. Acid colors dye from neutral solutions but more readily from those containing free acid. If the wool takes up any considerable amount of coloring matter in either case, the presence of coal tar dyes is indicated. The lichen colors³ (Archil, Cudbear, Litmus) go readily on wool, however, and many other natural colors, such as Turmeric, will dye the fiber, if present in considerable amount. On the other hand, a few coal tar dyes, especially Auramine O and Naphthol Green B, are quite unstable and, if present in small amounts, may give no distinct dyeing. Acid dyes are much more frequently used than basic dyes and in most cases may be removed from wool without much decomposition by "stripping" the latter with dilute ammonia⁴. By the action of the alkali, many natural colors are destroyed, while others remain for the most part on the fiber. If the behavior with wool in neutral and acid solutions indicates the presence of acid dyes, rinse the colored cloth thoroughly with water, cover with 2% ammonium hydroxid solution in a casserole, boil for a few minutes, remove the cloth and squeeze out the adhering liquid. Boil the ammoniacal solu-

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tion to remove the excess of ammonia, drop in a piece of clean, wet wool, make distinctly but not strongly acid with hydrochloric acid and boil again. If acid coal tar dyes are present, they will usually give a fairly clean, bright dyeing on the second piece of wool. A further purification may be carried out by repeating the stripping and re-dyeing, though generally accompanied by corresponding loss of dye.

(b) *Candies and similar colored sugar products*.—Dissolve about 20 grams of the sample in 100 cc. of water and treat the solution as directed under (a). When the coloring matter is on the surface of the candy, pour off the solution before the colorless inner portion has dissolved.

(c) *Jams and jellies*.—Boil a mixture of 10–20 grams of the sample and 100 cc. of water with wool in neutral and also in acid solution as directed under (a). For thick jams it is usually better, though less easy, first to extract the coloring substances by treating the product as directed under (d).

(d) *Canned and preserved fruits and vegetables, sausage casings, smoked fish, coffee, spices, etc.*—Macerate 20–200 grams of the sample with 4–5 times its weight of 80% alcohol. After standing a few hours pour off the solvent as completely as possible and repeat the extraction, using 70% alcohol containing about 1% of ammonia. (1) Examine separately the filtered alcoholic extracts as directed under (a); or, (2) Boil the ammoniacal solution until practically neutral, complete the neutralization with acetic acid, add the neutral 80% alcohol extract, continue the evaporation until most of the alcohol is removed, and boil with wool as directed under (a).

(e) *Cocoa and chocolate products*.—Treat cocoa as directed under (d). The alcoholic extract will contain a large amount of natural coloring matter and several dyeings and strippings may be necessary to get rid of this in order to show the presence of coal tar dyes.

Chocolate may be treated similarly but the following procedure is preferable: Wash 20–200 grams of the well divided sample with gasoline on a filter until most of the fat has been removed; if the gasoline is colored, reserve for the examination of oil-soluble dyes as directed under 3. Remove most of the adherent solvent from the residue by evaporation or pressure between layers of absorbent paper and digest with alcohol as directed under (d).

Coal tar dyes may also be detected in chocolate and cocoa products by mixing directly with 3–4 times their weight of hot water and immediately boiling the magma with wool, as directed under (a). Because of the presence of large amounts of fatty and protein materials, this method is not very satisfactory.

(f) *Cereal products*.—Proceed as directed under (d), in most cases working with a large amount of the sample, 200–500 grams, and a relatively smaller amount of alcohol. Where tests are to be made only for the acid dyes, the extraction with neutral 80% alcohol may be omitted advantageously.

3

OIL-SOLUBLE DYES⁵.

Prepare an alcoholic solution of the oil-soluble dye by one of the following methods which are to be applied to the oil or fat obtained by extraction with ether or gasoline if the nature of the substance requires it:

(a) Shake the oil or melted fat with an equal volume of 90% alcohol. The alcohol after separation will contain Aniline Yellow, Butter Yellow, Aminoazotoluene and Auramine, if present.

(b) Saponify 20–200 grams of the oil or fat with alcoholic potash and, after removal of most of the alcohol on the steam bath, extract the soap with ether or gasoline. Most of the common dyes are removed by this treatment, though the digestion with strong alkali may cause some decomposition and make the extraction rather troublesome.

(c) Dilute 20–200 grams of the oil or melted fat with 1–2 volumes of gasoline and shake out successively with 2–4% potassium or sodium hydroxid solution, 12–15% hydrochloric acid, and phosphoric-sulphuric acid mixture, prepared by mixing 85% phosphoric acid with about 10–20% by volume of concentrated sulphuric acid.

The dilute alkali extracts Sudan G and Annatto. The dilute hydrochloric acid extracts Aniline Yellow (7), Aminoazotoluene, and Butter Yellow (16), the first 2 forming orange-red, the latter cherry-red solutions in this solvent. Benzeneazo-beta-naphthylamin and homologues also come in this group, though they are not extracted very readily and decompose rapidly on standing in strongly acid solution. The phosphoric acid mixture is necessary for the extraction of Sudan I (11), Sudan II (49), Sudan III (143), and the homologue of the last, Sudan IV. The procedure is not very suitable in the presence of Auramine but this dye is seldom found in oils. Neutralize the alkaline and dilute hydrochloric acid solutions; dilute the phosphoric acid mixture and partially neutralize, cooling the liquid during this operation; and extract the dyes by shaking with ether or gasoline.

For the direct dyeing test use the alcoholic solution, obtained as directed in (a). Evaporate to dryness the ether or gasoline solutions, obtained as directed in (b) and (c), and dissolve the residue in 10–20 cc. of strong alcohol. To the alcoholic solution add some strands of white silk and a little water and evaporate on the steam bath until the alcohol has been removed or until the dye is taken up by the silk. The dyeing test is sometimes unsatisfactory and in all cases a small portion of the alcoholic solution should be tested by treating with an equal volume of concentrated hydrochloric acid and stannous chlorid solution. The common oil-soluble coal tar dyes are rendered more red or blue by the acid and are decolorized by the reducing agent. Most of the natural coloring matters become slightly paler with the acid and are little changed by the stannous chlorid solution.

SEPARATION OF COLORING MATTERS IN PURE CONDITION BY MEANS OF IMMISCIBLE SOLVENTS⁶.

4

PREPARATION OF SAMPLE.

(a) *Water-soluble colors*.—Proceed as directed under 2, omitting the fixation of the color on wool, and obtain an aqueous solution as free as practicable from suspended matter, alcohol, acids, alkalies and salts.

(b) *Water-insoluble lakes*.—If the sample is in solid form, treat the well divided material with sufficient water to form a paste. Liquids require no preparation except the removal of alcohol when present.

5 MIXTURES OF ORANGE I, ERYTHROSINE, INDIGO CARMINE, AMARANTH, TARTRAZINE, NAPHTHOL YELLOW S, PONCEAU 3R AND LIGHT GREEN S F YELLOWISH.

The use of immiscible solvents for the separation of mixtures of coloring matters usually involves a systematic fractionation since many of the dyes used do not differ very greatly in their solubilities in the various solvents. When it seems probable that only the 8 coal tar dyes permitted under the Federal Food and Drugs Act⁷ are present, the following abridged procedure may be used for their separation. For this procedure the concentration of the dye solution should lie preferably between 0.01–0.05%. The solutions obtained in the examination of colored food products practically never require further dilution but with commercial food colors care must be taken that the concentration is not too high. Treat the sample, prepared as directed in 4, with one half its volume of concentrated hydrochloric acid and extract a few times with amyl alcohol. The use of the centrifuge is sometimes necessary

to separate the layers. Designate the residual aqueous layer as *A*. Combine the amyl alcohol extracts and wash with 4-5 portions of $N/4$ hydrochloric acid or until this solvent extracts very little color. These washings will contain any Indigo Carmine, Amaranth and Tartrazine present, the Indigo Carmine being removed in somewhat larger proportion in the first washings than the other 2. With ordinary concentration very little Ponceau will be removed. Designate these combined washings as *B*.

6**ORANGE I AND ERYTHROSINE.**

Measure, if necessary, the amyl alcohol extract, under **5**, then (1) Dilute with an equal volume of petroleum ether or low boiling gasoline, and again wash several times with $N/4$ hydrochloric acid to extract Ponceau 3R and Naphthol Yellow S; or, (2) Without dilution with gasoline, wash with 5% salt solution until these 2 dyes are extracted. Designate these washings as *C*. The Ponceau and Yellow having been removed, the amyl alcohol, containing an equal volume of gasoline, is washed a few times with water which will extract Orange I. This dye having been removed, shake the solution, although the latter may appear almost colorless, with very dilute sodium hydroxid solution to remove Erythrosine. If considerable Orange I is present, some of it may contaminate the washings containing the Ponceau 3R and Naphthol Yellow S, especially when these have been separated by means of $N/4$ hydrochloric acid after the addition of gasoline.

7**INDIGO CARMINE, AMARANTH AND TARTRAZINE.**

The presence of 2 or all 3 of these dyes is usually indicated by the appearance of the $N/4$ hydrochloric acid washings, *B*, under **5**. Evaporate the combined $N/4$ hydrochloric acid washings to dryness to remove the excess of hydrochloric acid and dissolved amyl alcohol. Dissolve the residue in water, divide the solution and identify the constituent colors in the portions. To a portion of the slightly acidified solution add a few decigrams of urea, warm and add 1 or 2 drops of sodium nitrite solution. Indigo Carmine is converted into the pale yellow isatin sulphonate while the other dyes are but little affected. The isatin compound is not ordinarily present in sufficient concentration to tint the solution but it differs from Tartrazine also in being much less readily extracted by amyl alcohol from strong acid solutions (less than one half from $4N$ acid). The solution now contains the Amaranth or Tartrazine, or both, practically unaffected. Amaranth is much more quickly attacked by most reducing agents than Tartrazine. Treat the solution, which should be neutral or faintly acid (in the presence of sodium carbonate the reduction of the tartrazine takes place still more slowly), at room temperature with a dilute solution of sodium hyposulphite ($Na_2S_2O_4$), adding the latter very carefully, drop by drop, and allowing sufficient time after the addition of each drop for the reduction to take place. When the color shows that the Amaranth has been destroyed completely, shake the mixture at once with air to oxidize the slight excess of hyposulphite before it can react further on the Tartrazine.

To separate the Indigo Carmine heat to boiling another portion of the solution, which should be neutral or faintly acid, and add dilute sodium hyposulphite solution, drop by drop, until all the dyes are reduced. On shaking with air the Indigo Carmine is quickly re-formed.

8**NAPHTHOL YELLOW S AND PONCEAU 3R.**

Treat the $N/4$ acid solution or the salt solution, *C*, under **6**, as the case may be, containing the Ponceau and Naphthol Yellow S, with enough hydrochloric acid to make it about $2N$ and shake out a few times with washed ethyl acetate^a. Remove

the Yellow from the combined ethyl acetate extracts by shaking with water. Naphthol Yellow S is almost colorless in strongly acid solutions, and its absence in washings, etc., must never be assumed until these have been made alkaline. Finally separate the Ponceau 3R from the acid solution by shaking with amyl alcohol, and then wash out the dye from this extract with a few small portions of water. If, in the case of mixtures containing Orange I, the washings of the ethyl acetate, which should contain only Naphthol Yellow S, become more red upon the addition of alkalies, combine, then (1) Make N/4 with hydrochloric acid and remove the contaminating Orange by shaking with amyl alcohol-gasoline mixture (1 to 1); or, (2) Treat the combined washings with one fifth their volume of concentrated hydrochloric acid, extract the dyes by shaking once with amyl alcohol, and remove the Yellow by washing with several portions of 5% salt solution.

9

LIGHT GREEN S F YELLOWISH.

The original mixture, A, under 5, from which the above mentioned 7 colors have been removed by adding acid and shaking out with amyl alcohol, may still contain Light Green S F Yellowish, which will be colorless or nearly so in the acid solution. To separate this dye treat the mixture with strong ammonia or potassium hydroxide solution until slightly alkaline, and neutralize with acetic acid. Any Green present will now be apparent by the color of the mixture. Extract the color by shaking with a few small portions of dichlorhydrin. Wash the dichlorhydrin extract with a little water, then dilute with several volumes of benzene or carbon tetrachloride, and remove the dye by shaking with water.

When coal tar dyes other than the 8 mentioned above are present, the solutions obtained in this procedure will be found to contain a coloring matter which does not correspond exactly in properties to one of the dyes named above. When coal tar dyes other than these 8 are present, reference should be made to the larger works².

COAL TAR DYES IN GENERAL.

10

BASIC DYES.

Most basic dyes may be separated from mixtures by making alkaline with sodium hydroxide and shaking with ether¹⁰. Use the sample, prepared as in 4, for this purpose. Separate the ether layer, which may or may not be colored, and shake with 2-5% acetic acid, which will take up any dye present, forming a colored solution. Although the common basic colors undergo some alteration by this treatment¹¹, it may be used for the qualitative detection and separation of Methyl Violet B (451), Magenta (448), Bismarck Brown (197), Malachite Green (427), and Rhodamine B (504). With car Auramine (425) may also be separated in this way though it is quickly decomposed on standing in alkaline solution.

11

ACID DYES.

The following short procedure is often convenient for the examination of mixtures of acid dyes: Make the sample, prepared as in 4, the color concentration of which does not vary greatly from 0.01-0.05%, strongly acid by adding one half its volume of concentrated hydrochloric acid and shake with amyl alcohol. Separate the amyl alcohol solution and wash by shaking with successive portions of one half its volume of water, reserving the portions in separate test tubes or beakers. Because of the acid dissolved in the amyl alcohol these washings will show a regular decrease in acidity and the coloring matters will appear in *maximum* amount in the different fractions according to their respective solubilities. Ponceau 6R (108) is washed out chiefly while the acidity is still high, N/1 or above. Amaranth (107), Brilliant

Scarlet (106) and Tartrazine (94) appear when the washings have an acidity between N/1 and N/4; Orange G (14) and Soluble Blue (480) between N/2 and N/16; Palatine Scarlet (53), Ponceau 2R (55) and 3R (56), Naphthol Yellow S (4), Cochineal (706), Crystal Ponceau (64) and Azorubine A (103) between N/16 and N/256. When the acid is practically all removed, Orange I (85), Orange II (86) and Croceine Orange (13) begin to wash out, and less readily, Orange IV (88) and Metanil Yellow (95). Finally the unsulphonated coloring matters, such as Erythrosine G (516), Erythrosine (517) and the Rose Bengals (520 and 523) are removed very slowly by water or not at all when all traces of acid have been removed. Acid Yellow (8) and Brilliant Yellow S (89) are not very uniform in composition. They are partially taken up by amyl alcohol from acid solution and appear chiefly in the first washings. Indigo Carmine (692) behaves somewhat similarly.

IDENTIFICATION OF THE COAL TAR DYES¹².

12

GENERAL.

The most widely used tests for the identification of coal tar dyes refer to the changes produced with acids and alkalies. Other tests, based upon the behavior with reducing agents, followed perhaps by treatment with oxidants or by separation and identification of the reduction products¹³, and tests based upon oxidation of the dye and treatment of the oxidation products¹⁴ are generally applicable. Spectroscopic methods are also used¹⁵.

13

COLOR CHANGES PRODUCED WITH ACIDS AND ALKALIES.

Transfer the separated coloring matter to wool (or silk in the case of oil-soluble dyes) by boiling as directed in **2(a)** or **3**. Care should be taken that the final dyeing is made in a solution fairly free from foreign matter such as sugar or aromatic substances, which, adhering to the fiber, may modify the reaction. In most cases the amount of color available is small and should not be dyed on too large a piece of wool (or silk). Rinse thoroughly the dyed fibre in running water, dry, cut into small pieces and place separately in the depressions of a white porcelain spot plate. Moisten the pieces with the respective reagents employed. (For many coloring matters the hue upon treatment with acids or alkalies varies markedly with the concentration of the reagents and amount of dye present; therefore the unknown dye should be compared with dyeings of known colors of approximately the same dye concentration as shown by this appearance.)

The table under **14** shows the color changes produced by concentrated hydrochloric and sulphuric acids, 10% sodium hydroxid and 12% ammonium hydroxid solutions on wool dyed with 0.1–0.5% of the respective coloring matters. Included also are the reactions of the oil-soluble colors but these refer to dyeings on silk. The dyes are arranged approximately according to hue. Brown is classed with orange, black (gray) with violet.

TABLE 14.

Color reactions produced on dyed fibers by various reagents.

COLORING MATTER		HYDROCHLORIC ACID	SULPHURIC ACID	SODIUM HYDROXID	AMMONIUM HYDROXID
Rhodamine B	504	Orange	Yellow	Bluer *	Bluer
Rose Bengal	523	Almost decolorized	Orange	No change	No change
Archil	710	Red	Dull brown	Violet	Violet
Magenta	448	Yellowish brown	Dull brown	Decolorized	Paler
Acid Magenta	462	Almost decolorized	Yellow	Decolorized	Decolorized
Palatine Red	62	Darker	Violet	Dull brown	Little change
Bordeaux B	65	Violet	Blue	Brown	Little change
Amaranth	107	Slightly darker	Violet to brownish	Dull brownish	Little change
Azorubine A	103	Little change	Violet	Red	Red
Erythrosine	517	Orange-yellow	Orange-yellow	No change	No change
Ponceau 6RB	169	Blue	Blue	Dull violet-red	Little change
Ponceau 6R	108	Violet-red	Violet	Brown	Orange-red
Crystal	64	Violet-red	Violet	Dull brown	Little change
Ponceau					
Ponceau 3R	56	Little change	Little change	Dull orange	Little change
Sudan III*	143	Violet, then brown	Green	Violet-red	Little change
Safranine	584	Greenish blue	Green	Red	Red
Brilliant	106	Red	Violet-red	Yellowish brown	Orange-red
Scarlet					
Ponceau 2R	55	Little change	Little change	Brownish yellow	No change
Palatine	53	Darker	Violet-red	Brownish yellow	No change
Scarlet					
Erythrosine G	516	Yellow-orange	Yellow-orange	No change	No change
Sudan II*	49	Red	Violet-red	Little change	No change
Sudan I*	11	Orange-red	Red	Redder	No change
Cochineal	706	Little change	Little change	Violet-red	Violet-red
Bismarck	197	Redder, darker	Browner	Yellower	Yellower
Brown					
Bismarck	201	Redder, darker	Browner	Yellower	Yellower
Brown R					
Orange I	85	Violet	Violet	Red, dark	Red, dark
Orange II	86	Red	Red	Dull red	No change
Croceine	13	Orange-red	Orange	Slightly darker	No change
Orange					
Orange G	14	Little change	Orange	Dull, brownish red	No change
Orthotoluene-azo-beta-naphthyl-amine*		Red	Violet	Little change	No change
Sudan G*	10	Orange-yellow	Brownish yellow	Orange-yellow	No change
Butter Yellow*	16	Violet-red	Orange-yellow	No change	No change
Aniline Yellow*	7	Brownish red	Orange-yellow	Little change	No change
Aminoazo-ortho-toluene*		Dull orange	Orange-yellow	Little change	No change

* Oil-soluble.

14

TABLE 14.—Continued.

COLORING MATTER		HYDROCHLORIC ACID	SULPHURIC ACID	SODIUM HYDROXID	AMMONIUM HYDROXID
Fluoresceine	510	Little change	Little change	Green flu- orescent	Green flu- orescent
Metanil Yellow	95	Violet-red	Violet	No change	No change
Azoflavine	92	Violet-red	Violet-red	Dull brown	Little change
Acid Yellow	8	Red	Orange	Little change	No change
Brilliant Yellow S	89	Violet-red	Violet-red	Little change	Little change
Tartrazine	94	Slightly darker	Slightly darker	Little change	Little change
Naphthol Yellow S	4	Almost decolorized	Very pale, dull brown	No change	No change
Auramine	425	Decolorized	Almost decolorized	Decolorized	Paler
Turmeric	707	Red	Reddish brown	Orange	Orange
Quinoline Yellow	667	Slightly darker	Brownish yellow	Slightly paler	Little change
Naphthol Green B	398	Yellowish	Brownish yellow	No change	No change
Guinea Green B	433	Pale orange- yellow	Pale, dull yellow	Decolorized	Decolorized
Light Green S F Yellowish	435	Pale orange- yellow	Pale, dull yellow	Decolorized	Decolorized
Night Green 2B	438	Pale orange- yellow	Pale, dull yellow	Decolorized	Paler
Malachite Green	427	Almost decolorized	Almost decolorized	Decolorized	Decolorized
Erioglaucine A	436	Yellow	Pale, dull yellow or brown	Slightly darker	Little change
Patent Blue A	442	Pale orange- yellow	Pale or dull brown	Little change	Little change
Soluble Blue	480	Paler	Brown	Pale reddish	Almost decol- orized
Indigo Carmine	692	Slightly darker	Slightly darker	Greenish yel- low	Greenish blue
Formyl Violet	468	Pale orange- yellow	Pale, dull orange	Decolorized	Decolorized
Methyl Violet B	451	Yellowish	Yellowish	Decolorized	Almost decol- orized
Nigrosine, sol- uble	602	Dull bluish	Dull greenish	Brownish red, paler	Pale reddish

15 SPECIAL TESTS FOR COAL TAR DYES PERMITTED⁷ UNDER THE FEDERAL FOOD AND DRUGS ACT.

The dyes, given in 5, are sufficiently characterized in most cases by the solubilities shown in their separation and by the color changes given by acids and alkalies on the dyed fiber. This is especially true with Amaranth, Tartrazine, and Orange I. By treatment with reducing agents such as stannous chlorid, titanous chlorid, zinc dust or sodium hyposulphite in acid solution, Indigo Carmine, Amaranth, Tartrazine, Ponceau 3R and Orange I are decolorized. With Indigo Carmine the color returns on shaking with air, most readily on warming, or on the addition of oxidizing agents such as ferric chlorid or potassium persulphate. Excess of the reducing agents must of course be avoided. With the last 4 named dyes the color is not restored. Dilute solutions of Light Green S F Yellowish, Naphthol Yellow S and

Erythrosine become paler or colorless with acids so that the effects of acid reducing agents are not so readily apparent. Neutral solutions of Naphthol Yellow S are decolorized by sodium hyposulphite and other reducing agents, the color not returning with air or oxidants. An evanescent deepening of the shade may take place immediately upon the addition of the hyposulphite. Erythrosine and Light Green S F Yellowish become paler with sodium hyposulphite, the color being partially restored upon the addition of potassium persulphate.

In hot solutions containing an excess of sodium tartrate the dyes named are readily decolorized by titanium trichlorid¹⁴. In the case of Indigo Carmine if the reducing agent has been added carefully and an excess avoided, the blue color readily returns on shaking with air. With Erythrosine and Light Green S F Yellowish the color is scarcely restored by air but on cooling and adding potassium persulphate returns imperfectly. The reduction products of the other dyes do not give colored solutions again on oxidation disregarding a slight yellowish or brownish tint that may sometimes appear.

Indigo Carmine is extracted in small proportions from slightly acid solutions by shaking with dichlorhydrin. Most of the other common bluish dyes are triphenylmethane derivatives and are relatively more soluble in this liquid than in the aqueous layer. A small portion (1 cc.) of the solution obtained in the separation, 5, may be used directly.

Ponceau 3R gives in neutral or faintly acid solutions a bluish red, flocculent precipitate with barium chlorid or acetate, practically all of the dye being removed from solution. Some of the solution obtained in the separation, 5, may be used in this test, first neutralizing the free hydrochloric acid with sodium acetate; or better, it may be evaporated to dryness on the steam bath to remove the acid and the residue taken up with a little water. The solution should contain 0.005% or more of the dye.

Naphthol Yellow S, in solutions containing an excess of ammonia or sodium carbonate, becomes intensely rose-red on the addition of sodium hyposulphite, the color gradually fading again as complete reduction takes place.

Erythrosine differs from most of the common dyes by containing iodine. To test for this, acidify the solution with sulphuric acid, shake with ether, separate the ether solution of the color and evaporate to dryness in a platinum dish after the addition of a few drops of sodium carbonate solution or sufficient to form the deep red sodium salt. Hold the dish containing the residue in the Bunsen flame until organic matter is destroyed, take up the residue with water, acidify with sulphuric acid and test for iodine in one of the usual ways, such as with chlorin water and carbon disulphid or tetrachlorid, or with starch paste and an oxidizing agent. It is useless to test for iodine with very small amounts of dye but in most cases sufficient coloring matter can be separated from the food product to give satisfactory results.

16

NATURAL COLORING MATTERS.

The natural coloring matters as a class show much less tendency to dye animal fiber than do the common synthetic colors. In many cases the crude products used contain a number of colored substances and a complete separation can scarcely be attempted. Most of the natural coloring matters, in dilute solution, are sensitive to alkalies, some to acids, hence such reagents must be used with care.

SEPARATION OF NATURAL COLORING MATTERS.

17

Extraction with Ether from Neutral Solutions.

From neutral solutions ether extracts Carotin, Xanthophyll (the pigment found in leaves, fats and oils, egg yolk, carrots, etc.), the coloring matter of tomatoes and paprika and green Chlorophyll. The coloring matter remains in the ether solution on shaking with dilute sodium hydroxid solution or dilute hydrochloric acid, no apparent change taking place although chemically the substances may be altered more or less by this treatment.

18

Extraction with Ether from Acid Solutions.

From slightly acid solutions ether extracts very readily and completely the coloring matter of Alkanet, Annatto, Turmeric, and the red dyewoods, Sandalwood, Camwood and Barwood. It extracts in large proportion the flavone coloring matters of Fustic, Persian Berries (after hydrolysis), and Quercitron as well as the coloring matter of Brazilwood and the green derivatives formed from Chlorophyll by alkaline treatment. It extracts in relatively small amount the coloring matters of Logwood, Archil, Saffron and Cochineal. The coloring matters of this group are readily removed from ether by shaking with alkaline solutions but in most cases rapidly undergo chemical change.

19

Extraction with Amyl Alcohol from Acid Solutions.

From slightly acid solutions amyl alcohol extracts largely the coloring matters of Logwood, Archil, Saffron and Cochineal. [From ammoniacal Cochineal (Carmine) the ordinary coloring matter is readily re-formed upon standing with hydrochloric acid.] Amyl alcohol extracts in relatively small proportions Caramel and the Anthocyanins constituting the red coloring matter of the most common fruits.

IDENTIFICATION OF NATURAL COLORING MATTERS.

20

REAGENTS.

- (a) *Hydrochloric acid*.—Sp. gr. 1.20.
- (b) *10% sodium or potassium hydroxid solution*.
- (c) *Sodium hyposulphite solution*.—A freshly prepared 5% solution of "Blankite", sodium hyposulphite ($\text{Na}_2\text{S}_2\text{O}_4$).
- (d) *0.5% ferric chlorid solution*.—Freshly prepared but may be made by diluting a 10% stock solution.
- (e) *10% potassium or ammonium alum solution*.
- (f) *5% uranium or sodium uranium acetate solution*.
- (g) *Sulphuric acid*.—Sp. gr. 1.84.

21

PROCEDURE.

Relatively few good tests are known for the common natural colors. Some of their most useful analytical properties¹⁷ are tabulated in 22. In general these tests should be applied to the somewhat purified solutions of the coloring matter obtained as indicated in 17, 18 or 19.

Evaporate ether solutions to dryness, warm the residue with a little alcohol and dilute the alcoholic solution with water. Apply the reagents as stated below:

Hydrochloric acid.—Add concentrated acid (sp. gr. 1.20) to the solution, first 1 or 2 drops, then a large excess, equal to 3–4 times the volume of the solution.

Sodium hydroxid (potassium hydroxid).—Make the solution slightly alkaline by adding a drop of the 10% sodium hydroxid solution. A 10% solution of potassium hydroxid in methyl alcohol must be used for the “brown phase reaction” for chlorophyll, described in **23**, and may also be employed for the other tests.

Sodium hyposulphite.—Add the sodium hyposulphite solution drop by drop.

Ferric chlorid.—Add a small amount of the 0.5% ferric chlorid solution to the solution to be tested. The reagent must be added very carefully, a small drop at a time, as the colorations are not obtained in all cases when an excess is used.

Alum.—Add to the test solution one fifth its volume of the 10% potassium or ammonium alum solution.

Uranium acetate.—Add the 5% uranium acetate solution drop by drop to the solution to be tested.

Concentrated sulphuric acid on the dry color.—Evaporate a small amount of the solution or of the coloring matter in a porcelain dish. Cool thoroughly and treat the dry residue with 1 or 2 drops of cold, concentrated sulphuric acid. The colorations are in some cases extremely fugitive and may be observed only the instant the acid wets the residue.

The properties of pure preparations of the various natural coloring matters are described, for the most part, by Rupe¹⁸. Properties of the Chlorophylls and Carotinoids are given by Willstätter and Stoll¹⁹. Those of the coloring matters of the Corn Flower, Rose, Pelargona Flower, Larkspur, Cranberry, Whortleberry and Purple Grape are described by Willstätter²⁰.

TABLE 15.
Behavior of certain natural coloring matters with common reagents.

COLORING MATTER	HYDROCHLORIC ACID	SODIUM HYDROXID SOLUTION	SODIUM HYOSULPHITE SOLUTION	FERRIC CHLORID SOLUTION	ALUM SOLUTION	CRANIC ACETATE SOLUTION	CONCENTRATED SULPHURIC ACID ON DRY COLOR
Logwood	Deep red with excess of acid	Violet to violet-blue	Almost decolorized, color returning imperfectly by reoxidation	Dark shades of violet, brown or black (the first hue of ten evanescent)	Rose-red (change rather slow)	Violet, quickly fading	Red, changing to yellow
Redwoods (Brazilwood, Sandalwood, Camwood and Barwood)	Deep red with excess of acid	Violet-red	Dark shades of violet, brown or black (the first hue of ten evanescent)	Rose-red (change rather slow)
Anthocyanins of red fruit colors	Change to green, dull blue or slate color, usually very quickly, becoming browner by oxidation	Anthrocyanidins derived by hydrolysis, almost completely decolorized
Alkanet	Deep blue	Violet-blue
Archil	Little or no change	Blue	Decolorized, color returning when shaken with air. Reaction more easily seen in alkaline solution	Yellowish green	Violet-blue

Cochineal	Little or no change Remains orange. Little change	Violet	No marked change Little affected	Slightly darker No marked change Perhaps somewhat browner	Green	Blue
Annatto	Orange-red or carmine-red on addition of several volumes of concentrated acid	Orange-brown	Little affected	No marked change Perhaps somewhat browner	Somewhat browner	Red
Turmeric (solution in ether or alcohol characterized by pure yellow color and light green fluorescence)	Becomes intensely yellow with 2-4 volumes of concentrated acid	Bright-yellow	Little affected	Olive-green or black colorations	Orange colorations	Yellow to orange
Flavone colors of Fustic, Persian Berries, Quercitron, etc.	Little or no change	Remains yellow	Little affected	No marked change. Perhaps somewhat browner	Not affected	Blue
Saffron	Little change. Perhaps slightly paler	Little or no change	Little affected			Blue, reaction obtained with difficulty
Carotin and Xanthophyll	More brownish	"Brown phase reaction", ²³ Little change or slightly deeper brown	Slightly paler	No change		
Green Chlorophyll	Little or no change					
Caramel						

SPECIAL TESTS FOR NATURAL COLORING MATTERS.

23

CHLOROPHYLL.

The "brown phase reaction"²¹ may be useful for the characterization of Chlorophyll, when this has not been previously treated with alkalis. Treat the green ether or petroleum ether solution of the coloring matter with a small amount of 10% solution of potassium hydroxid in methyl alcohol. The color becomes brown, returning to green in a few minutes.

ANNATTO.

24

*Leach Test*²².

Pour on a moistened filter an alkaline solution of the color obtained by shaking out the oil or melted and filtered fat with warm, dilute sodium hydroxid solution. If Annatto is present, the filter paper will absorb the color so that, when washed with a gentle stream of water, it will remain dyed a straw color. Dry the filter and add a drop of stannous chlorid solution. If the color turns pink the presence of Annatto is confirmed.

25

TURMERIC.

Carry out the highly characteristic reaction of Curcumine (Turmeric) with boric acid as follows: Treat the aqueous or dilute alcoholic solution of the color with hydrochloric acid until the shade just begins to appear slightly orange. Divide the mixture into 2 parts and add some boric acid powder or crystals to 1 portion. A marked reddening will be quickly apparent, best seen by comparison with the portion to which the boric acid has not been added. The test may also be made by dipping a piece of filter paper in the alcoholic solution of the coloring matter, drying at 100°C., then moistening with a weak solution of boric acid to which a few drops of hydrochloric acid have been added. On drying again a cherry-red color will be developed.

26

COCHINEAL.

When the presence of Cochineal is suspected, acidify the mixture with one third its volume of concentrated hydrochloric acid and shake with amyl alcohol. Wash the amyl alcohol solution of the coloring matter 2-4 times with equal volumes of water to remove hydrochloric acid, etc. Dilute the amyl alcohol with 1-2 volumes of gasoline and shake with a few small portions of water to remove the color. Separate the solution into 2 portions. To the first add, drop by drop, 5% uranium acetate solution, shaking thoroughly after each addition. In the presence of cochineal a characteristic emerald-green color is produced²³. The green coloration with uranium salts is not developed in the presence of much free acid. Therefore add a little sodium acetate before making this test or a correspondingly large amount of uranium acetate must be added. To the second portion add a drop or so of ammonium hydroxid, and, in the presence of Cochineal, a violet coloration results. This, however, is not so sensitive to small amounts as the first test and many fruit colors give tests hardly to be distinguished.

As cochineal lakes very often contain tin, further examination for this metal should always be made when water-insoluble cochineal compounds appear to be present.

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XII. METALS IN FOODS.

ARSENIC¹.—TENTATIVE.

1

REAGENTS.

(a) *Nitric and sulphuric acids, arsenic-free*.—Specific gravities 1.42 and 1.84, respectively.

(b) *Sulphuric acid (1 to 2)*.

(c) *Zinc, arsenic-free*.—Stick zinc broken into pieces approximately 1 cm. in length.

(d) *Lead acetate paper*.—Heavy filter paper soaked in 20% lead acetate solution, dried and cut into pieces about 4.5 by 16 cm.

(e) *Lead acetate cotton*.—Absorbent cotton soaked in 5% lead acetate solution.

(f) *Mercuric bromid paper*.—Cut heavy, close-textured drafting paper (similar to Whatman's cold pressed) into strips exactly 2.5 mm. wide and about 12 cm. long. Soak for an hour in a 5% solution of mercuric bromid in 95% alcohol, squeeze out the excess of solution and dry on glass rods. Cut off the ends of the strips before using.

(g) *20% potassium iodid solution*.

(h) *Stannous chlorid solution*.—Forty grams of stannous chlorid crystals made up to 100 cc. with concentrated hydrochloric acid.

(i) *Standard arsenic solution*.—Dissolve 1 gram of arsenious oxid in 25 cc. of 20% sodium hydroxid solution, neutralize with dilute sulphuric acid, add 10 cc. of the concentrated sulphuric acid and dilute to 1 liter with recently boiled water. One cc. of this solution contains 1 mg. of arsenious oxid (As_2O_3).

Dilute 20 cc. of this solution to 1 liter. Fifty cc. of the latter solution when diluted to 1 liter give a dilute standard solution containing 0.001 mg. of arsenious oxid (As_2O_3) per cc. which is used to prepare the standard stains. The dilute solutions must be freshly prepared immediately before use.

2

APPARATUS.

Use a 2 ounce wide-mouthed bottle as a generator. Fit this by means of a perforated rubber stopper with a glass tube, diameter 1 cm. and 6 cm. long, containing a piece of the lead acetate paper rolled into a cylinder. Connect this tube by means of a perforated rubber stopper with a similar tube filled with the lead acetate cotton, squeezed to remove excess of the solution. The cotton in all tubes used should be uniformly moist to obtain comparative stains. Connect the second tube by means of a perforated rubber stopper with a narrow glass tube, internal diameter 3 mm. and 12 cm. long, containing a strip of the mercuric bromid paper. See Fig. 7. Rubber stoppers used for connections must be free from any white coating.

3

PREPARATION OF SOLUTION.

Weigh 5–50 grams of the finely divided and well mixed sample into a porcelain casserole, the amount selected depending upon the character of the material and the ease with which it is oxidized. With dry, highly nitrogenous substances employ 5 grams; pulped vegetables, 25 grams; liquids with low solid content like beer or

vinegar, 50 grams. Add 10–15 cc. of the nitric acid, cover the casserole by setting a watch glass inside the rim, convex side upward, heat until vigorous action is over, cool and add 10 cc. of the concentrated sulphuric acid. Heat on a wire gauze over a flame until the mixture turns dark brown or black, then add more nitric acid in 5 cc. portions, heating between each addition until the liquid remains colorless or yellow when evaporated until sulphur trioxid fumes are evolved. To remove

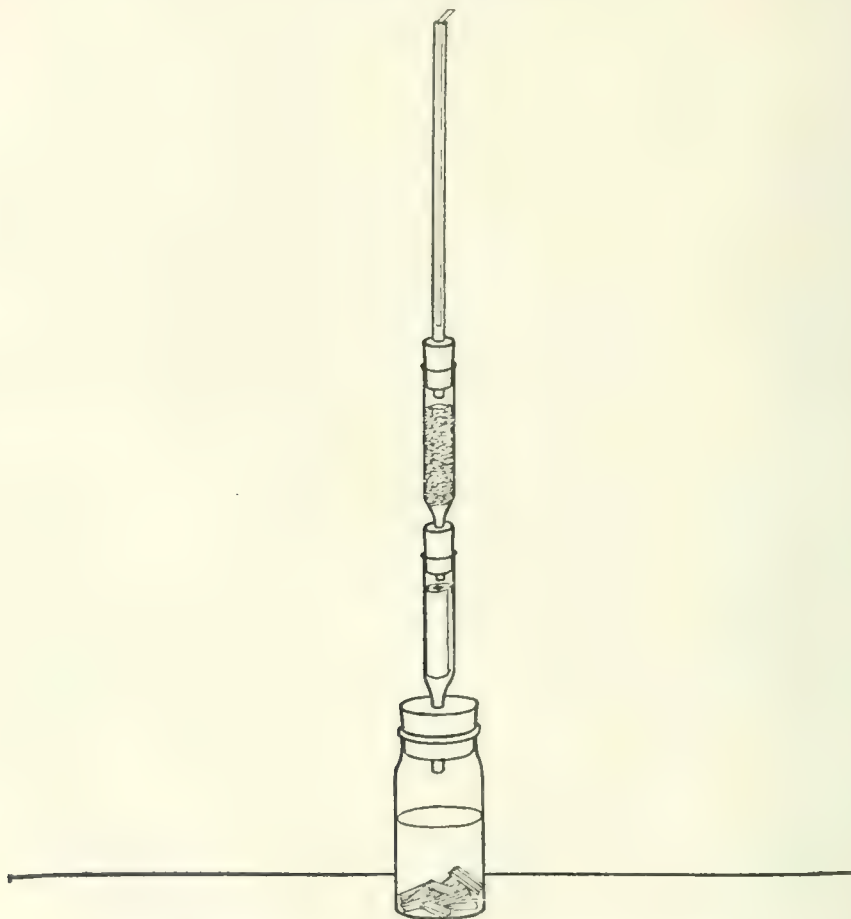


FIG. 7. APPARATUS FOR THE DETERMINATION OF ARSENIC.

completely all nitric or nitrous acid, evaporate to about 5 cc., cool, dilute with 10–15 cc. of water and again evaporate until white fumes are evolved. Cool, dilute with water, again cool, and make up with water to a definite volume (usually 25–100 cc., depending upon the amount of sample taken and its arsenic content).

4

DETERMINATION.

Introduce 20 cc. of the solution or, if the amount of arsenic is large, an aliquot containing not more than 0.03 mg. of As_2O_3 , prepared as directed in 3, into the

generator of the apparatus described in 2 and add 20 cc. of the dilute sulphuric acid. If the total volume is less than 40 cc., dilute to that volume with water and add 4 cc. of the 20% potassium iodid solution. Heat to about 90°C., add 3 drops of the stannous chlorid solution and heat for 10 minutes. Cool the generator and its contents in a pan containing water and ice; when cold add about 15 grams of the stick zinc and connect the entire apparatus as described in 2. Keep the bottles in ice water for 15 minutes, then remove from the bath and allow the evolution of gas to proceed for an hour longer. Remove the sensitized paper and compare the stain with similar ones produced under like conditions with known amounts of arsenic, using portions of the standard arsenic solution, containing 0.001, 0.002, 0.005, 0.010, 0.015, 0.025 and 0.030 mg. of arsenious oxid (As_2O_3), and adding such quantities of water and sulphuric acid that the same volume and acid strength are maintained as above.

TIN².

5

Gravimetric Method.—Tentative.

Weigh 50–100 grams of the sample (depending upon the amount of dry substance present and the relative ease with which the organic matter is oxidized) into an 800 cc. Kjeldahl flask and add 100 cc. of concentrated nitric acid. Allow to stand overnight (this procedure being preferred if much fat or sugar is present) or else place the flask on a wire gauze over a free flame and heat until the contents boil quietly. Add 25–50 cc. of concentrated sulphuric acid (depending upon the amount of dry substance present in the sample), and heat until white fumes are generated, cool somewhat, then add 5–10 cc. of concentrated nitric acid and continue heating as before. Repeat the addition of nitric acid until the solution remains clear after boiling off the nitric acid and fumes of sulphur trioxid appear.

Add 200 cc. of water to the digested sample, prepared as directed above, and pour into a 600 cc. beaker. Rinse out the Kjeldahl flask with 3 portions of boiling water so that the total volume of the solution is about 400 cc. Cool, add concentrated ammonia until just alkaline and then hydrochloric or sulphuric acid until the acidity is about 2%. Place the beaker, covered, on a hot plate, heat to about 95°C. and pass in a slow stream of hydrogen sulphid for another hour. Digest on the hot plate for an hour and allow to stand 1–2 hours longer.

Filter the tin sulphid on an 11 cm. filter, similar in quality to No. 590, white ribbon, S. & S. Wash alternately with 3 portions each of wash solution (100 cc. of saturated ammonium acetate solution, 50 cc. of glacial acetic acid and 850 cc. of water) and hot water. Digest the filter and precipitate in a 50 cc. beaker with 3 successive portions of ammonium polysulphid, heat to boiling each time and filter through a 9 cm. filter. Wash the precipitate on the filter with hot water. Acidify the filtrate with acetic acid, digest on a hot plate for an hour, allow to stand overnight and filter through a double 11 cm. filter. Wash alternately with 2 portions each of the wash solution and hot water and dry thoroughly in a weighed porcelain crucible. Ignite over a Bunsen flame, very gently at first and later at full heat. The crucible, partly covered, is then heated strongly with a large or Méker burner. Stannic sulphid must be roasted gently to the oxid, which may be heated strongly without loss by volatilization. Weigh as stannic oxid and calculate to metallic tin.

Volumetric Method³.—Tentative.

6

REAGENTS.

(a) *Air-free wash solution.*—Dissolve 20 grams of sodium bicarbonate in 2 liters of boiled water and add 40 cc. of concentrated hydrochloric acid. This solution should be freshly prepared before use.

(b) *N/100 iodine*.—The solution must be standardized frequently against (d), containing asbestos, and treated as described in 7, omitting the precipitation and boiling with hydrochloric acid and potassium chlorate. To obtain exact results the tin solution used for standardization should contain about the same amount of tin as is found in the sample under examination.

(c) *N/100 sodium thiosulphate*.

(d) *Standard tin solution*.—Dissolve 1 gram of tin in about 500 cc. of concentrated hydrochloric acid. Make up to 1 liter with water. One cc. contains 1 mg. of tin.

(e) *Sheet aluminium*.—Use sheet aluminium, about 30 gauge, free from tin.

7

DETERMINATION.

Proceed as directed in 5 to "Digest on the hot plate for an hour and allow to stand 1-2 hours longer".

Filter the precipitate of tin sulphid upon asbestos in a Gooch crucible with a detachable bottom, using suction. Wash the precipitate a few times and then transfer the detachable bottom, asbestos pad, and tin precipitate to a 300 cc. Erlenmeyer flask. Remove all traces of the precipitate from the inside of the crucible by means of a jet of hot water and a policeman, using a minimum amount of water for washing.

Add 100 cc. of concentrated hydrochloric acid and 0.5 gram of potassium chlorate to the flask. Boil for about 15 minutes, making about 4 more additions of smaller amounts of potassium chlorate as chlorin is boiled out of the solution. Wash the particles of potassium chlorate down from the neck of the flask with water and finally boil to remove chlorin. Then add about 1 gram of the sheet aluminium to dispel the last traces of chlorin.

Attach the flasks, in duplicate, as described below, to a large carbon dioxid generator. Pass the carbon dioxid through a scrubber containing water and then divide into 2 streams by means of a Y-tube, each stream of carbon dioxid entering one of the flasks by means of a long rubber tube connected with a bulbed tube, passed through the rubber stopper of the flask and having its lower end near the surface of the liquid in the flask. The carbon dioxid leaves the flask by a second bulbed tube, the opening of which is near the top of the flask. This glass tube is connected by a long rubber tube to a second glass tube about 10 inches long which is immersed in a cylinder containing water. This gives a water-seal to the delivery tube and a pressure against which the current of carbon dioxid must work. It also restrains any strong flow of gas when not desired and permits a gas pressure in the Erlenmeyer flask.

After the flasks are connected, raise the tubes in the water-seal cylinders so that the generator has practically no pressure to overcome. Allow the carbon dioxid to run for a few minutes. Drop the tubes to the bottom of the cylinders, creating pressure in the flasks. Lift the rubber stoppers of the flasks alternately about a dozen times, in order to force out any air remaining in the flasks. Slightly raise the stopper on one of the flasks and quickly drop about 2 grams of sheet aluminium into the flask. The aluminium should be folded into a strip about 1 cm. wide and slightly bent so as to prevent it from striking directly on the bottom of the flask. After the aluminium has entirely dissolved, raise the tubes in the water-seal cylinders so as to allow carbon dioxid to pass through, place the flasks upon hot plates, and heat to boiling. After boiling for a few minutes, remove the flasks from the hot plates and cool in ice water (or cold running water), still maintaining within them an

atmosphere of carbon dioxid. Lower the tubes in the cylinder. When cool, disconnect the flasks one at a time, putting a glass plug into the carbon dioxid inflow. Wash the tubes, rubber stopper and sides of the flask with the air-free wash solution, add starch paste and titrate at once with the N/100 iodine.

If it is desired to titrate by the excess method, run an excess of the N/100 iodine into the flask while it is still connected with the carbon dioxid stream. Then wash out the tubes and titrate the excess of iodine with the N/100 sodium thiosulphate.

The rubber connections should be washed with water after each determination.

8

COPPER.—TENTATIVE.

Destroy organic matter as directed in 5. Concentrate the sulphuric acid residue by continued digestion to a volume of 10–15 cc., cool, dilute with a little water, transfer to a 400 cc. beaker, rinse the Kjeldahl flask with water, adding the rinsings to the contents of the beaker, dilute to about 200 cc. and boil to expel nitrous fumes. Cool, render the solution slightly alkaline with ammonium hydroxid and boil to expel the excess of ammonia. Add 5 cc. of concentrated hydrochloric acid for each 100 cc. of solution, heat to incipient boiling and saturate the solution with hydrogen sulphid. Allow to stand on a steam bath for a few minutes until the sulphid flocculates, filter and wash the precipitate with hydrogen sulphid water. Protect the precipitate from contact with air as much as possible, use only hydrogen sulphid water for washing and carry out this operation without interruption. Reserve the filtrate for the determination of zinc, if necessary. Place the filter containing the copper sulphid precipitate in a small flask, add 4–5 cc. of concentrated sulphuric acid and the same amount of nitric acid and heat until white fumes appear. Continue the oxidation, adding a little nitric acid from time to time, until the liquid remains colorless upon heating to the appearance of white fumes. Cool, dilute with about 30 cc. of water, add an excess of bromine water and boil until all bromine is expelled. Determine the copper as directed in VIII, 29, using N/100 sodium thiosulphate for the titration.

9

ZINC.—TENTATIVE.

Proceed as directed in 8 to the point indicated by the sentence "Reserve the filtrate for the determination of zinc, if necessary". Boil the filtrate, containing the zinc, to expel hydrogen sulphid and to reduce the volume to about 250–300 cc., add a drop of methyl orange and 5 grams of ammonium chlorid and make alkaline with ammonium hydroxid. Add dilute hydrochloric acid, drop by drop, until the reaction is faintly acid, then add 10–15 cc. of 50% sodium or ammonium acetate solution and pass in hydrogen sulphid for a few minutes until precipitation is complete. Allow the precipitate to settle, filter, refilter, if necessary, until the filtrate is clear, and wash the precipitate twice with hydrogen sulphid water. Dissolve the precipitate on the filter with a little hydrochloric acid (1 to 3), wash the filter with water, boil the filtrate and washings to expel hydrogen sulphid, cool and add a distinct excess of bromine water. Then add 5 grams of ammonium chlorid and ammonium hydroxid until the color, caused by free bromine, disappears. Add hydrochloric acid (1 to 3), drop by drop, until the bromine color just reappears, then add 10–15 cc. of 50% sodium or ammonium acetate solution and 0.5 cc. of 10% ferric chlorid solution, or enough to precipitate all the phosphates. Boil until all the iron is precipitated. Filter while hot and wash the precipitate with water containing a little sodium acetate. Pass hydrogen sulphid into the combined filtrate and washings

until all the zinc sulphid, which should be pure white, is precipitated, filter upon a tared Gooch and wash with hydrogen sulphid water, containing a little ammonium nitrate. Dry the crucible and its contents in an oven, ignite at a bright red heat, cool and weigh as zinc oxid. Calculate the weight of metallic zinc.

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² J. Assoc. Official Agri. Chemists, 1915, **1**: 257.

³ Proc. Eighth Intern. Cong. Appl. Chem., 1912, **18**: 35.

XIII. FRUITS AND FRUIT PRODUCTS.

1

PREPARATION OF SAMPLE.—TENTATIVE.

All samples received in open packages (i.e., not in sterile condition) must be transferred without delay to glass-stoppered containers and kept in a cool place. The determination of alcohol, total and volatile acids, solids and sugars, particularly in the case of fruit juices and fresh fruits, should be made at once as fermentation is liable to begin very soon. Portions for the determinations of sucrose and reducing sugar may be weighed and, after adding a slight excess of neutral lead acetate solution, kept without fermenting for several days if desired. The various products are prepared as directed below.

(a) *Juices*.—Prepare the fresh juice by pressing the well pulped fruit in a jelly bag and filtering through muslin.

(b) *Jellies and sirups*.—Mix thoroughly to insure uniformity in sampling. Weigh 60 grams into a 300 cc. flask, add water, dissolve by frequent shaking, then make up to the mark with water, and use aliquots for the various determinations. If the jelly contains starch or other insoluble material, mix thoroughly before taking the aliquots.

(c) *Fresh and dried fruits*.—Pulp the whole, well cleaned fruit in a large mortar or by means of a food chopper and mix thoroughly. In the case of stone fruits, remove the pits and determine their proportion in a weighed sample.

(d) *Jams, marmalades, preserves and canned fruits*.—Pulp thoroughly the entire contents of the jar or can, as directed under (c); with stone fruits remove the pits and, if desired, determine their proportion in a weighed sample. In the examination of canned fruits it is often sufficient merely to examine the sirups in which the fruits are preserved. In such cases the liquor may be separated and treated as prescribed for juices.

2

ALCOHOL.—TENTATIVE.

Determine alcohol in 50 grams of the original material as directed under **IX, 31**.

3

TOTAL SOLIDS.—TENTATIVE.

(a) *Juices, jellies and sirups containing no insoluble matter*.—Proceed as directed in **IX, 3, 5, 7 or 10**, employing the sample prepared as directed in **1 (a) or (b)**.

(b) *Fresh and dried fruits, jams, marmalades, preserves, canned foods and other products containing insoluble matter*.—Weigh about 20 grams of pulped fresh fruit, or such an amount of fruit products as will give not more than 3-4 grams of dried material; if necessary to secure a thin layer of the material, add a few cc. of water, mix thoroughly, and dry as directed in **IX, 3 or 4**.

It is to be noted that certain State and Federal regulations require the moisture in dried apples to be determined by drying for 4 hours at the temperature of boiling water.

INSOLUBLE SOLIDS.

4

Direct Method.—Tentative.

Transfer 50 grams of the sample to a mortar by means of warm water and macerate thoroughly; then transfer to a muslin filter and wash thoroughly with about 500 cc. of warm water, stirring the pulp thoroughly on each addition of water. This amount of water is usually sufficient to remove all soluble material. In extreme cases increase

the washings to 1000 cc. Transfer the insoluble residue to an evaporating dish, dry and weigh. If it is desired to determine the alcohol precipitate, **18**, cool the filtrate, make up to a definite volume and reserve for this determination.

5*Indirect Method.—Tentative.*

Transfer 25 grams of the fruit product to a 250–500 cc. graduated flask, the size of the flask depending upon the volume of insoluble matter present, add water, shake thoroughly and make up to volume. Allow to settle and either filter or decant the supernatant liquid. Determine the soluble solids in an aliquot, as directed in **3 (a)**. The fruit must be macerated thoroughly; the use of a mechanical shaker is advisable. The percentage of insoluble solids is the difference between the percentage of the total solids and the percentage of soluble solids.

6**TOTAL ASH.—OFFICIAL.**

Determine the ash as directed under **VIII, 4**, using 50 cc. of the solution of the jelly or diluted sirup, **1 (b)**, evaporated to dryness, or 25 grams of juice or of fresh or canned fruit, or 10 grams of jam, marmalade, preserves, or dried fruit.

7**ALKALINITY OF THE ASH.—TENTATIVE.**

Into the platinum dish containing the ash introduce a measured excess of N/5 nitric acid, heat to boiling, cool and add a few drops of methyl orange. Carefully rub up the ash with a rubber-tipped stirring rod and titrate the excess of acid with N/10 potassium or sodium hydroxid. Express the result as the number of cc. of N/10 acid required to neutralize the ash from 100 grams of the sample.

8**SULPHATE AND CHLORID.—TENTATIVE.**

Wash the solution of the ash, obtained in **7**, into a 50 cc. flask and make up to the mark with water. Evaporate 25 cc. of this solution to dryness several times with concentrated hydrochloric acid, take up the final residue in a small amount of hot water, filter, wash the paper with hot water, acidify the filtrate with a few drops of hydrochloric acid and determine the sulphate by precipitation with barium chlorid solution. From the weight of barium sulphate calculate the sulphate present as per cent of potassium sulphate.

In the other portion of the solution determine the chlorin as directed under **III, 15**. The nitric acid added before making the titration will, if it contain enough nitrous oxid, completely destroy the red color of the methyl orange and leave a clear solution for the titration. Calculate the chlorin as per cent of sodium chlorid.

9**TOTAL ACIDITY.—TENTATIVE.**

Dilute 25 cc. of the solution of jelly or diluted sirup, **1 (b)**, or 10 grams of juice or fresh fruit, with recently boiled water to about 250 cc., or less if the sample be not highly colored; titrate the acid with N/10 alkali, using phenolphthalein as an indicator. In the case of highly colored products employ azolitmin solution or phenolphthalein powder [**XVI, 25**] on a spot plate instead of phenolphthalein solution. Calculate the results as malic, citric or tartaric acid, specifying the acid used and expressing the results in grams per 100 cc.

10**VOLATILE ACIDS.—TENTATIVE.**

Dissolve 10 grams of the sample, dilute to 25 cc. and distil in a current of steam, as directed under **XVI, 27**. Each cc. of N/10 alkali is equivalent to 0.0060 gram of acetic acid.

11 FREE MINERAL ACIDS.—TENTATIVE.

Proceed as directed under **XIX, 26, 27** or **28**.

12 PROTEIN.—OFFICIAL.

Proceed as directed under **I, 18, 21** or **23**, using 5 grams of jelly or other fruit product containing a large amount of sugar, or 10 grams of juice or fresh fruit and a larger quantity of the sulphuric acid if necessary for complete digestion. Multiply the percentage of nitrogen by 6.25 to obtain the percentage of protein.

SUCROSE.

13 *By Polarization.—Official.*

Determine by polarizing before and after inversion, as directed under **IX, 22** or **23**.

14 *By Reducing Sugars Before and After Inversion.—Tentative.*

Proceed as directed under **VIII, 18**.

15 REDUCING SUGARS.—TENTATIVE.

Proceed as directed under **VIII, 25**, expressing the results as invert sugar.

16 COMMERCIAL GLUCOSE.—TENTATIVE.

Proceed as directed under **IX, 25**.

17 DEXTRIN.—TENTATIVE.

Dissolve 10 grams of the sample in a 100 cc. flask, add 20 mg. of potassium fluorid, and then about one fourth of a cake of compressed yeast. Allow the fermentation to proceed below 25°C. for 2-3 hours to prevent excessive foaming, and then incubate at 27°-30°C. for 5 days. At the end of that time, clarify with basic lead acetate solution and alumina cream, make up to 100 cc. and polarize in a 200 mm. tube. A pure fruit jelly will show a dextro or laevo-rotation of not more than a few tenths of a degree. If a polariscope having the Ventzke scale be used and a 10% solution polarized in a 200 mm. tube, the number of degrees read on the sugar scale of the instrument multiplied by 0.8755 will give the percentage of dextrin, or the following formula may be used:

$$\text{Percentage of dextrin} = \frac{C \times 100}{198 \times L \times W} \text{ in which}$$

C = degrees of circular rotation;

L = length of tube in decimeters;

W = weight of sample in 1 cc.

18 ALCOHOL PRECIPITATE.—TENTATIVE.

Evaporate 100 cc. of a 20% solution of jelly or diluted sirup, **1 (b)**, or of the washings from the determination of insoluble solids, **4**, to 20 cc.; add slowly, with constant stirring, 200 cc. of 95% alcohol by volume and allow the mixture to stand overnight. Filter and wash with 80% alcohol by volume. Wash the precipitate from the filter paper with hot water into a platinum dish; evaporate to dryness; dry at 100°C. for several hours and weigh; then burn off the organic matter and weigh the residue as ash. Designate the loss in weight upon ignition as the alcohol precipitate.

The ash should be chiefly lime and not more than 5% of the total weight of the alcohol precipitate. If it is greater than this, some of the salts of the organic acids have been brought down. Titrate the water-soluble portion of this ash with N/10 acid, as any potassium bitartrate precipitated by the alcohol can thus be estimated.

STARCH.

19

Qualitative Test.—Tentative.

First destroy the color of the jelly by treatment with sulphuric acid and potassium permanganate solution and then test with iodine solution. Bring the solution of jelly nearly to boiling, add several cc. of dilute sulphuric acid and then potassium permanganate solution until all color is destroyed. The starch remains unaffected by this treatment. The presence of starch is not necessarily an indication of its addition as an adulterant. It is usually present in small amount in the apple, and occasionally in other fruits, and unless it is found in the fruit product in considerable amount its presence may be due to these natural sources.

GELATIN¹.

20

Qualitative Test.—Tentative.

The presence of gelatin in jellies and jams is shown by the increased content of nitrogen. Precipitate a concentrated solution of jelly or jam with 10 volumes of absolute alcohol and determine nitrogen in the dried precipitate as directed under I, 18, 21 or 23.

AGAR AGAR.

QUALITATIVE TESTS.

21

By Microscopic Examination².—Tentative.

Heat the jelly with 5% sulphuric acid, add a crystal of potassium permanganate and allow to settle. If agar agar is present the sediment will be rich in diatoms which can be detected by the use of the microscope.

22

By Precipitation³.—Tentative.

Cover 30 grams of the jam or jelly with 270 cc. of hot water, stir until thoroughly disintegrated and boil for 3 minutes. Filter immediately, while still boiling hot, through a filter paper of texture similar to No. 597, S. & S. In the presence of agar agar a precipitate will form upon standing not longer than 24 hours. Filter, wash with cold water and dissolve from the paper by means of a very small amount of boiling water. Upon chilling this hot water solution a firm jelly will be formed that can be examined by the touch. This method will detect 0.2% of agar agar with certainty if the proportions of jam or jelly and water are strictly observed.

TARTARIC ACID.—TENTATIVE.

23

PREPARATION OF SOLUTION.

Filter fruit juices and employ the filtrate directly. In the case of jellies filter the solution, prepared as directed in 1 (b), and employ the filtrate. In the case of sirups or substances containing insoluble matter like pulped fruit, jams, marmalades, etc., weigh 50–100 grams, the amount selected being dependent upon the content of solids, of the sample, prepared as directed in 1 (c) or (d), introduce into a 200 cc. graduated flask, make up to the mark with water, allow to stand for an hour, shake at frequent intervals, filter through a dry paper and use the filtrate.

24

DETERMINATION.

Determine the tartaric acid in 100 cc. of fruit juice or the same amount of a solution of the sample, prepared as directed in 23, employing the method given under XVI, 29, except that 20 cc. of alcohol are used in the precipitation instead of 15 cc.

MALIC ACID.

25

Method I.—Tentative.

(For fruit juices and similar products containing no tartaric acid and not over 15% of sugars and in which the color does not interfere with polarization.)

Filter the sample, if necessary to secure a solution which can be readily polarized, and polarize with white light and a dichromate cell, using a 200 mm. tube if possible.

If the sample contains free mineral acid, transfer a measured portion (75 cc. is a convenient volume) to a 100 cc. graduated flask, add enough standard alkali, calculated from the acidity as determined in 9, to neutralize the total acidity, dilute to the mark, mix well and filter. If no free mineral acids are present, it is unnecessary to neutralize the sample. If neutralized, proper correction must be made for dilution in making the final calculation.

Transfer 25 cc. of the sample, or a neutralized solution, to a flask graduated at 25 and 27.5 cc., add about 2.5 grams of powdered uranyl acetate, and shake vigorously at frequent intervals for 3 hours, keeping the mixture well protected from light. If all of the uranyl acetate dissolves, add more so that a small amount remains undissolved at the end of 3 hours. Dilute the solution to the 27.5 cc. mark with saturated uranyl acetate solution, mix well and filter, if necessary, through a folded filter. Polarize, if possible, in a 200 mm. tube. If the solution is too dark to polarize in a 200 mm. tube, a 100 or 50 mm. tube may be used. Multiply the reading by 1.1 to correct for the dilution.

Multiply the algebraic difference in degrees Ventzke between the 2 readings calculated to the basis of a 200 mm. tube by the factor 0.036 to obtain the weight of malic acid in the sample in grams per 100 cc.

Make all polarizations at the same room temperature with white light and with a dichromate cell. Make at least 6 readings in each case and take an average of these.

In the case of dark colored fruit juices which cannot be polarized readily, approximately quantitative results may be obtained by adding to the solutions a few drops of bromin, shaking thoroughly and filtering just before polarization.

Method II.—Tentative.

(Approximate determination for fruit juices and similar products containing no tartaric acid and more than 15% of sugars.)

26

PREPARATION OF SOLUTION¹.

Weigh out 25 grams of the sample and transfer to a 600 cc. beaker with a little 95% alcohol by volume. Add alcohol a little at a time until 200 cc. have been added, stirring the mixture well, and warming, if necessary, to insure solution of all alcohol-soluble substances. Filter on a Büchner funnel, using suction, and thoroughly wash the precipitated peetins and insoluble matter with 95% alcohol, disregarding any slight turbidity which may appear in the filtrate after the washings have been added. From 9, calculate the amount of N/4 barium hydroxid required nearly to neutralize the acidity in the 25 grams of sample taken. To the combined filtrate and washings in an Erlenmeyer flask add the calculated quantity of barium hydroxid solution, stir until reaction is complete and then add 3-5 drops, or more if required, of 50% barium acetate solution to insure an excess of barium. Make up the volume of the mixture to about 375 cc. (not less) with alcohol, and reflux until the precipitate settles readily after being shaken. This may require 3-4

hours. Filter with suction and thoroughly wash the precipitate in the flask and on the paper with 95% alcohol by volume. Transfer the portion on the filter to the original flask, rinsing the paper with a jet of hot water. Digest the precipitate with hot water, containing 2 grams of sodium sulphate in solution, until the reaction is complete, and boil until the barium sulphate precipitate settles readily. Concentrate by evaporation, if necessary, and transfer to a 100 cc. volumetric flask with a little hot water, cool, make up to volume with water and filter.

27

DETERMINATION.

Transfer 25 cc. of the filtrate, obtained in **26**, to a flask graduated at 25 and 27.5 cc., add about 2.5 grams of pulverized uranyl acetate and shake vigorously at frequent intervals for 3 hours, keeping the solution well protected from light. If all the uranyl acetate dissolves, add more so that a small amount remains undissolved at the end of 3 hours. Dilute the solution to the 27.5 cc. mark with saturated uranyl acetate solution, mix well, filter if necessary, and polarize in a 200 mm. tube, using the same precautions as described in **25**. Multiply the reading, calculated to the basis of a 200 mm. tube, by 1.1 to correct for the dilution.

Polarize another portion of the filtrate, obtained in **26**, which has not been treated with uranyl acetate. Multiply the algebraic difference in degrees Ventzke between the 2 readings, calculated to the basis of a 200 mm. tube, by the factor 0.036 to obtain the weight of malic acid in grams per 100 cc. in the solution as obtained in **26**.

Method III.—Tentative.

(Approximate determination for products containing tartaric acid.)

28PREPARATION OF SOLUTION⁵.

Prepare the sample as directed under **26** up to the point of filtration and washing of the barium malate precipitate, then dry the precipitate thoroughly and transfer the portion on the filter to the original flask, rinsing the paper with a jet of hot water. Digest the precipitate with hot water, transfer to a 100 cc. volumetric flask with a little hot water, cool, make up to volume with water and filter to remove insoluble barium tartrate. This amount of water is sufficient to dissolve barium malate up to amounts as large as approximately 0.9 gram in 100 cc. More than 100 cc. of water must be used when more than 0.9 gram of barium malate is present. The amount of barium tartrate dissolved by hot water is so small as to affect only slightly the polarization after treatment with uranyl acetate.

29

DETERMINATION.

Proceed as directed in **27**, using the solution prepared as directed in **28**.

CITRIC ACID⁶.—TENTATIVE.

(Applicable in the presence of sugar and malic and tartaric acids.)

30

REAGENTS.

- (a) *Barium hydroxid solution*.—Approximately N/4.
- (b) *50% barium acetate solution*.
- (c) *Sulphuric acid (1 to 1) and (1 to 5)*.
- (d) *Potassium or sodium bromid solution*.—Dissolve 15 grams of potassium bromid in 40 cc. of water or 16 grams of sodium bromid in 50 cc. of water.

- (e) 5% potassium permanganate solution.
(f) Ferrous sulphate solution.—Dissolve 20 grams of ferrous sulphate in 100 cc. of water containing 1 cc. of concentrated sulphuric acid.
(g) Bromin water.—Freshly prepared, saturated solution.

31

DETERMINATION.

Proceed as directed in 26 up to "Filter with suction and thoroughly wash the precipitate in the flask and on the paper with 95% alcohol by volume". Transfer the precipitate from the filter to the flask with a jet of hot water, boil until alcohol can no longer be detected by odor, and add enough of the sulphuric acid (1 to 5) to precipitate all the barium originally added and to allow 2 cc. in excess. Evaporate by careful boiling to a volume of 60-70 cc., cool and add 5 cc. of freshly prepared saturated bromin water, or enough to show a distinct excess. Transfer with water to a 100 cc. volumetric flask and dilute to the mark at standard temperature. Mix thoroughly, allow the precipitate to settle and filter through a dry paper. The precipitate may be separated by centrifugalizing and the supernatant liquid decanted, if necessary. Pipette an aliquot of the filtrate, containing not more than 250 mg. of citric acid, calculated from the total acidity of the sample, into a 300 cc. Erlenmeyer flask. If possible, the amount of citric acid in the aliquot should exceed 50 mg. Add 10 cc. of the sulphuric acid (1 to 1) and 5 cc. of the potassium or sodium bromid solution, mix, warm the flask in a water bath to 48°-50°C. and allow it to remain in the bath for 5 minutes. After removing from the bath add rapidly from a pipette, drop by drop with frequent interruptions, 25 cc. of the 5% potassium permanganate solution and shake vigorously, avoiding a temperature during oxidation exceeding 55°C. Set the flask aside until the hydrated peroxid of manganese begins to settle. The supernatant liquid should be dark brown, showing an excess of permanganate; if an excess is not indicated, add more permanganate. Shake, again set aside to settle and repeat this operation until the precipitate assumes a yellow color and most of it has dissolved. Finally, while the solution is still warm, remove the last undissolved portion of hydrated peroxid of manganese precipitate and also the excess of bromin by adding, drop by drop, the clear ferrous sulphate solution. Allow the solution to cool, shaking occasionally. If the operations have been properly conducted, a heavy white precipitate of pentabromacetone is obtained which becomes crystalline on occasional shaking and in this condition is entirely insoluble in water. Allow the mixture to stand overnight, collect it by means of gentle suction on a tared Gooch crucible provided with a thin pad of asbestos, previously dried over sulphuric acid in a vacuum desiccator, wash with water slightly acidified with sulphuric acid and finally wash twice with water. Dry the precipitate to constant weight over sulphuric acid in a vacuum desiccator, protecting the precipitate from strong light. The weight of pentabromacetone multiplied by the factor 0.424 gives the equivalent weight of anhydrous citric acid ($\text{H}_3\text{C}_6\text{H}_5\text{O}_7$). Occasionally the pentabromacetone is first obtained in the form of oily droplets. These become crystalline on standing or on cooling and are usually discolored by negligible traces of manganese or iron.

The above method may be applied directly to the sample without previous precipitation of the citric acid as the barium salt when the amount of sugar or other permanganate reducing substances is not excessive. In this case begin the determination with the addition of 2 cc. of sulphuric acid (1 to 5) and the treatment with bromin water.

32

METALS.—TENTATIVE.

Proceed as directed under XII.

33

PRESERVATIVES.—TENTATIVE.

Proceed as directed under **X**.

34

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI**.

35

SWEETENING SUBSTITUTES.—TENTATIVE.

Proceed as directed under **X, 12, 36** or **37**.

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XIV. CANNED VEGETABLES.

1 PHYSICAL EXAMINATION¹.—TENTATIVE.

Note carefully the external appearance of the packages to detect the presence of "leakers", "swells" or "springers". In general the ends of sound tins of canned vegetables are slightly concave. On opening the package note the relative proportion of solid and liquid contents and the level of the solids and of the total contents in the tin. Note the general appearance, odor, flavor, color and size of the vegetables; appearance of the liquor or brine, whether clear or turbid, and the condition of the inner walls of the container, especially as to blackening and corrosion. In all instances the analyst should familiarize himself with the normal appearance, odor, color, flavor and other properties of the product under examination. Careful macroscopic or microscopic examination should be made for worm infestation, mold, dirt, or other evidence of decomposition or filth.

2 PREPARATION OF SAMPLE.—TENTATIVE.

The preparation of the sample for analysis depends upon the character of the product and the determinations to be made. Samples in which only the solid or liquid portion is required should be treated as follows: Weigh the full can, open, pour off the liquid, allow the solid portion to drain for a minute, re-weigh the can and drained vegetables, then remove the solid portion and weigh the dry, empty can. The method selected for draining the vegetables is dependent upon the nature and condition of the sample. In most cases it is sufficient to cut around the cover and before turning it back allow the liquor to drain through the slit. Whenever a portion of the solid material would escape with the liquor by this procedure, drain upon a piece of cheese-cloth. From the weights thus obtained determine the percentage of liquid and solid contents. If only the solid portion is required, separate in a similar manner and grind thoroughly the drained vegetables in a mortar or food chopper. If a composite of the solid and liquid portion is required, grind thoroughly the contents of the can in a mortar or food chopper. In all cases mix thoroughly the portion used and preserve the balance in glass-stoppered containers. Unless the analysis is to be completed in a reasonably short time, determine the moisture in a portion of the sample prepared as above and, in order to prevent decomposition, dry the remainder and then expose to air until it becomes air-dry, grind, mix thoroughly and preserve in glass-stoppered containers. A second moisture determination is required in this procedure.

3 MOISTURE.—TENTATIVE.

Dry a quantity of the sample, representing about 2 grams of dry material, as directed in **IX, 2**.

4 ASH.—OFFICIAL.

Determine total ash as directed in **VIII, 4**.

5 SALT.—OFFICIAL.

Determine chlorin as directed under **III, 15**, and express the result in terms of sodium chlorid.

6

SUGARS.—TENTATIVE.

Determine reducing sugars and sucrose as directed in **VIII**, **58** and **59**, varying the weight of the sample employed according to its sugar content.

7

TOTAL ACIDS.—TENTATIVE.

Proceed as directed in **XVI**, **25**. Express the result as citric acid; 1 cc. of N/10 alkali is equivalent to 0.0070 gram of crystallized citric acid.

8

VOLATILE ACIDS.—TENTATIVE.

Proceed as directed in **XVI**, **27**. Express the results as acetic acid; 1 cc. of N/10 alkali is equivalent to 0.0060 gram of acetic acid.

9

PRESERVATIVES.—TENTATIVE.

Proceed as directed under **X**.

10

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI**.

11

METALS.—TENTATIVE.

Proceed as directed under **XII**.

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¹ U. S. Bur. Chem. Bulls. 125 and 151; U. S. Dept. Agr. Bull. 196; U. S. Bur. Chem. Circ. 54; Research Laboratory, National Canners Association, Bull. 2.

XV. CEREAL FOODS.

WHEAT FLOUR.

1 MOISTURE.—OFFICIAL.

Determine moisture as directed in **VIII, 2**.

2 ASH.—OFFICIAL.

Determine ash as directed in **VIII, 4**, using 5 grams of the flour.

3 CRUDE FAT OR ETHER EXTRACT.—OFFICIAL.

Determine the ether extract as directed in **VIII, 10**. With fine flour the addition of an equal weight of clean, dry sand is frequently necessary.

4 CRUDE FIBER.—OFFICIAL.

Determine crude fiber as directed in **VIII, 68**.

5 ACIDITY OF WATER EXTRACT.—TENTATIVE.

Weigh 18 grams of the flour into a 500 cc. Erlenmeyer flask and add 200 cc. of carbon dioxid-free water. Place the flask, loosely stoppered, for an hour in a water bath kept at 40°C., shaking occasionally. Filter upon a dry, folded filter, returning the first 10–15 cc. of the filtrate to the filter. Titrate 100 cc. of the clear filtrate with N/20 sodium hydroxid, using phenolphthalein as an indicator. Each cc. of N/20 sodium hydroxid is equivalent to 0.05% acidity as lactic acid.

6 SUGARS.—TENTATIVE.

Determine reducing sugars and sucrose as directed in **VIII, 58** and **59**.

7 PROTEIN.—OFFICIAL.

Determine nitrogen as directed in **I, 18, 21** or **23**. Multiply the percentage of nitrogen by 5.7 to obtain the percentage of protein.

ALCOHOL-SOLUBLE PROTEIN.

8 *Method I. (By nitrogen determination)—Tentative.*

Transfer 4 grams of the flour to a 150–200 cc. bottle or Erlenmeyer flask and add 100 cc. of 70% alcohol by volume, taking care that none of the material sticks to the bottom of the container. Shake thoroughly 10–12 times at intervals of 30 minutes at room temperature, or shake continuously in a shaking machine for an hour, and then set aside overnight. Shake thoroughly once more, allow to settle and filter through a dry, folded filter, returning the first runnings to the filter until a clear filtrate is obtained. Pipette 50 cc. of the filtrate, equivalent to 2 grams of the sample, into a Kjeldahl flask, dilute with 100 cc. of water to prevent frothing during digestion and determine nitrogen as directed in **I, 18, 21** or **23**.

9

Method II. (By Polarization)—Tentative.

REAGENT.

Millon's reagent.—Dissolve metallic mercury in an equal weight of concentrated nitric acid and dilute the solution with an equal volume of water. The freshly prepared solution must be used.

10

DETERMINATION.

Weigh 15.97 grams of the flour into a 300 cc. flask and add 100 cc. of alcohol (sp. gr. 0.90). Shake at 30 minute intervals for 3 hours and then let stand overnight. Filter through a dry, folded filter and polarize in a 200 mm. tube. Precipitate the proteins in 50 cc. of the filtrate by the addition of 5 cc. of Millon's reagent. Shake, filter and polarize the filtrate in a 200 mm. tube. Multiply the reading in degrees Ventzke by 1.1 to correct for the dilution and deduct the product from the first reading. This difference multiplied by 0.2 gives the per cent of gliadin¹ nitrogen.

11 PROTEIN SOLUBLE IN 5 PER CENT POTASSIUM SULPHATE SOLUTION.—TENTATIVE.

Weigh 6 grams of the flour into a 200 cc. flask and introduce exactly 100 cc. of 5% potassium sulphate solution. Shake at 30 minute intervals for 3 hours and let stand overnight or, better still, agitate at moderate speed in a shaker for 3 hours, let settle 30 minutes, filter and determine the nitrogen in 50 cc. of the filtrate as directed in **I, 18, 21** or **23**.

12 GLOBULIN AND ALBUMIN (EDESTIN AND LEUCOSIN) AND AMINO NITROGEN².—TENTATIVE.

Weigh 10 grams of the flour into a 500 cc. Erlenmeyer flask, add 250 cc. of 1% sodium chlorid solution, stopper the flask and shake thoroughly. Let stand, with occasional shaking, for 3 hours, filter through dry paper and evaporate 100 cc. of the filtrate to a small volume in a Kjeldahl digestion flask with 5 cc. of concentrated sulphuric acid. Add the remainder of the sulphuric acid and determine the nitrogen as directed in **I, 18, 21** or **23**. To a second 100 cc. of the filtrate add 5 cc. of 20% phosphotungstic acid solution, shake thoroughly, allow to settle and filter by decantation. Wash slightly with water, concentrate the filtrate with 5 cc. of sulphuric acid in a Kjeldahl flask and determine the nitrogen (amino) as directed in **I, 18, 21** or **23**. Deduct the amino nitrogen from the nitrogen found in the first fraction to obtain the nitrogen as globulin and albumin³.

13

GLUTENIN.—TENTATIVE.

Deduct the sum of the potassium sulphate-soluble nitrogen, **11**, and the alcohol-soluble nitrogen, **8**, from the total nitrogen, **7**, and multiply the difference by 5.7.

14

COLD WATER-SOLUBLE EXTRACT.—TENTATIVE.

Weigh 20 grams of the flour into a 500 cc. Erlenmeyer flask and add gradually 200 cc. of water at 10°C., shake vigorously when about 50 cc. of water have been added and continue shaking during the addition of the remainder. Allow to stand at 10°C. for 40 minutes, shaking occasionally. Filter through a large, dry, coarse filter paper, returning the first runnings to the filter until a clear filtrate is obtained. Pipette 20 cc. of the clear filtrate into a tared dish, evaporate to dryness on a steam bath, and dry to constant weight in an oven at 100°C. for periods of 30 minutes.

GLUTEN.

15 *Bamihl Test. (Qualitative)—Tentative.*

Place a very small quantity (about 1.5 mg.) of the flour on a microscope slide, add a drop of water, containing 0.2 gram of water-soluble eosin in 1 liter, and mix by means of a cover-glass, holding the latter at first in such a manner that it is raised slightly above the slide, and taking care that none of the flour escapes from beneath it. Finally allow the cover-glass to rest on the slide and rub it back and forth until the gluten has collected into rolls. The operation should be carried out on a white paper so that the formation of gluten rolls can be noted. Wheat flour, or other flours containing gluten, show by this treatment a copious amount of gluten, which absorbs the eosin with avidity, assuming a carmine color. Rye and corn flour yield only a trace of gluten, and buckwheat flour no appreciable amount. The preparations are best examined with the naked eye, thus gaining an idea of the amount of gluten present. If the flour is coarse, or contains a considerable amount of bran elements, as is true of buckwheat flour and low-grade wheat flour, the test should be made after bolting, as the bran particles and coarse lumps interfere with the formation of gluten rolls.

16 *Quantitative Method.—Tentative.*

Weigh 25 grams of the flour into a cup or porcelain mortar, add sufficient tap water (about 15 cc.) to form a firm dough ball and work into a dough with a spatula or pestle, taking care that none of the material adheres to the utensil employed. Allow the dough to stand in water at room temperature for an hour, then knead gently in a stream of tap water until the starch and all soluble matters are removed. This operation requires approximately 12 minutes and should be performed over bolting cloth or a horsehair sieve. To determine if the gluten is starch-free let 1 or 2 drops of the wash water, obtained by squeezing the gluten, fall into a beaker containing perfectly clear water. If starch is present a cloudiness appears. Allow the gluten thus obtained to stand in water for an hour, then press as dry as possible between the hands, roll into a ball, place in a tared, flat-bottomed dish and weigh as moist gluten. Transfer to an oven, dry to constant weight at 100°C. (about 24 hours), cool and weigh as dry gluten.

CHLORIN.

17 *Qualitative Test. (Chlorin-Bleached Flours)—Tentative.*

Extract 30 grams of the flour with gasoline and allow the latter to evaporate. A small amount of oil remains. Heat a piece of copper wire in a colorless gas flame until it is black and no longer colors the flame green. Dip the hot end of the wire into the oil and again bring into the flame. If chlorin or bromin has been used as a bleaching agent, a green or blue coloration is produced.

18 *Quantitative Method. (Added Chlorin in Chlorin-Bleached Flours)—Tentative.*

Weigh 20 grams of the flour into a flat-bottomed aluminium dish, 8-10 cm. in diameter, and dry 5 hours in a boiling water or steam oven, transfer, with as little exposure to the air as possible, to a continuous fat extractor, and extract for 16 hours with anhydrous alcohol-free ether, which is also free from chlorin. Transfer the ether extract to a nickel dish and add 25 cc. of a solution containing 25 grams of sodium hydroxid and 15 grams of sodium nitrate per liter. Place the dish on a steam bath, evaporate to dryness and ignite in a muffle at a dull red heat until

the contents are thoroughly charred. Extract the charred mass with 25 cc. of 1% nitric acid and filter. Return the residue to the dish, char and again extract with 25 cc. of 1% nitric acid, filter, wash with hot water, return to the dish and ignite to a white ash. Dissolve the ash in 5% nitric acid and add the solution to the filtrates previously obtained. Determine the chlorin in the combined filtrates either gravimetrically, as directed in **I, 16 (a)**, or volumetrically, as directed in **III, 15**, using N/50 solutions for greater accuracy.

NITRITE NITROGEN.—TENTATIVE.

19

REAGENTS.

(a) *Sulphanilic acid solution*.—Dissolve 0.5 gram of sulphanilic acid in 150 cc. of 20% acetic acid.

(b) *Alpha-naphthylamin hydrochlorid solution*.—Dissolve, by heating, 0.2 gram of the salt in 150 cc. of 20% acetic acid.

(c) *Standard nitrite solution*.—Dissolve 0.1097 gram of dry C. P. silver nitrite in about 20 cc. of hot water, add 0.10 gram of C. P. sodium chlorid, shake until the silver chlorid flocculates and make up to 1 liter. Draw off 10 cc. of the clear solution and dilute to 1 liter. Each cc. of the last solution is equivalent to 0.0001 mg. of nitrogen as nitrite. [Cf. **IV, 12 (d)**]

The silver nitrite may be prepared as follows: To a cold solution of about 2 grams of sodium or potassium nitrite in 50 cc. of water, add a solution of silver nitrate as long as a precipitate appears. Decant the liquid and thoroughly wash the precipitate with cold water. Dissolve in boiling water. On cooling the silver nitrite crystallizes out. Dry the crystals in the dark at ordinary temperature (preferably in a vacuum).

20

DETERMINATION.

(1) Select a series of 100 cc. volumetric flasks of uniform dimensions and color. Place 2 grams of high-grade, nitrite-free flour in each; add approximately 70 cc. of nitrite-free water and shake until the flour is thoroughly moistened. Add to these flasks varying amounts of the standard sodium nitrite solution, so that a series of comparison standards will be obtained having a range covering the probable nitrite content of the unknown sample. Reserve 1 flask for a blank test. In order to avoid making a large series of standards it is well to make a preliminary test to ascertain the approximate nitrite content of the unknown. Where the quantity of nitrite present is small, the nitrite solution in the flasks may be increased by 0.4 cc. each. Where bleaching is excessive, 1 gram of flour may be used throughout, or the standards may be given a wider variation in nitrite content.

To each of 2 similar flasks add 2 grams of the flour and 90 cc. of water; shake thoroughly and digest all the flasks, including the blank, in a water bath at 40°C. for at least 15 minutes; add 2 cc. each of the sulphanilic acid and alpha-naphthylamin hydrochlorid solutions to each flask. Continue the digestion at 40°C. for an additional 20 minutes. The color must be developed in all the flasks under conditions as nearly uniform as possible. Make up to the marks with nitrite-free water and compare the unknown with the series of standards. This may be done in a large, white, enameled pan; the effect of the turbidity, due to the flour, being minimized by the white background. The solutions should be allowed to subside and should not be shaken during comparison; or,

(2) Weigh 20 grams of the flour into a 500 cc. Erlenmeyer flask, add 200 cc. of nitrite-free water, previously warmed to 40°C., and close the flask with a rubber stopper. Shake vigorously for 5 minutes and digest for an hour in a water bath,

keeping the temperature of the liquid in the flask at 40°C. and shaking at 10 minute intervals. Finally filter on a dry, nitrite-free, folded filter. Return the first runnings to the filter until a clear filtrate is obtained. Pipette 50 cc. of the filtrate and 50 cc. of the standard nitrite solution into small flasks; add to each, 50 cc. of water, 2 cc. each of the sulphanilic acid and alpha-naphthylamin hydrochlorid solutions, shake and allow to stand an hour to bring out the color. Compare the 2 solutions in a colorimeter. Divide the height of the column of the standard solution by that of the solution of the sample to obtain the parts of nitrogen as nitrous acid (free and combined) per million of flour.

21

GASOLINE COLOR VALUE.—TENTATIVE.

Place 20 grams of the flour in a wide-mouthed, glass-stoppered 120 cc. bottle and add 100 cc. of colorless gasoline. Stopper tightly and shake vigorously for 5 minutes. After standing 16 hours, shake again for a few seconds until the flour has been loosened from the bottom of the bottle and thoroughly mixed with the gasoline, then filter immediately on a dry 11 cm. paper into an Erlenmeyer flask, keeping the funnel covered with a watch glass to prevent evaporation. In order to secure a clear filtrate, a certain quantity of the flour should be allowed to pass over onto the paper and the first portion of the filtrate passed through a second time. It will be found convenient to fit the filter paper to the funnel by means of water and dry thoroughly either by standing overnight in a well-ventilated place or by heating.

Determine the color value of the clear gasoline solution in a Schreiner or similar colorimeter, using for comparison a 0.005% potassium chromate solution. This solution corresponds to a gasoline number of 1.0 and is conveniently prepared by diluting 10 cc. of a 0.5% solution to 1 liter. The colorimeter tube, containing the gasoline solution, should first be adjusted so as to read 50 mm., then the tube containing the standard chromate solution raised or lowered until the shades of yellow in both tubes match. The reading of the chromate solution, divided by the reading of the gasoline solution, gives the gasoline color value. The color value may be determined also in Nessler tubes, using for comparison potassium chromate solutions of various dilutions prepared from a 0.5% solution and filling the tubes in all cases to the height of 50 mm.

BIBLIOGRAPHY.

¹ U. S. Bur. Chem. Bull. 152, p. 104.

² Ibid., 81, p. 124.

³ Ibid., 122, p. 54.

XVI. WINES.

1

PHYSICAL EXAMINATION.—TENTATIVE.

Note the following: whether the container is "bottle full"; the appearance of the wine, whether there is any sediment and if it is bright or turbid; condition when opened, whether still, gaseous or carbonated; color and depth of color; odor, whether vinous, acetous, pleasant or foreign; and taste, whether vinous, acetous, sweet, dry or foreign.

2

PREPARATION OF SAMPLE.—TENTATIVE.

If gas is contained in the wine, remove it by pouring back and forth in beakers.

Filter the wine, regardless of appearance, before analysis and determine immediately the specific gravity and such ingredients as alcohol, acids and sugars as are liable to change through exposure.

3

SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ} \text{C.}}{4^{\circ}}$ by means of a pycnometer.

4

ALCOHOL.—TENTATIVE.

(a) *By volume*.—Measure 100 cc. of the liquid at 20°C. into a 300-500 cc. distillation flask, add 50 cc. of water, attach the flask to a vertical condenser by means of a bent tube and distil almost 100 cc., making up to 100 cc. volume when cooled to 20°C. Foaming, which sometimes occurs, especially with young wines, may be prevented by the addition of a small amount of tannin. To determine the alcohol in wines which have undergone acetous fermentation and contain an abnormal amount of acetic acid, exactly neutralize the portion taken with sodium hydroxid solution before distilling. This is unnecessary, however, in wines of normal taste and odor. Determine the specific gravity of the distillate at $\frac{20^{\circ} \text{C.}}{4^{\circ}}$ and obtain the corresponding percentage of alcohol by volume from 5.

(b) *Grams per 100 cc.*—From the specific gravity of the distillate, obtained in (a), ascertain from 5 the corresponding alcohol content in grams per 100 cc.

(c) *By weight*.—Divide the number of grams in the 100 cc. of distillate, as obtained in (b), by the weight of the sample as calculated from its specific gravity.

(d) *By immersion refractometer*.—The percentages of alcohol, as determined in (a) and (c), may be verified by determining the immersion refractometer reading of the distillate and obtaining, from 6, the corresponding percentages of alcohol.

5

TABLE 16.—ALCOHOL TABLE.

(Calculated by the U. S. Bureau of Standards from its experimental results¹.)*For calculating the percentages of alcohol in mixtures of ethyl alcohol and water from their specific gravities.*

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.99823	0.00	0.00	0.00	0.99492	2.25	1.79	1.78	0.99174	4.50	3.58	3.55
0.99815	0.05	0.04	0.04	0.99485	2.30	1.82	1.81	0.99168	4.55	3.62	3.59
0.99808	0.10	0.08	0.08	0.99477	2.35	1.86	1.85	0.99161	4.60	3.66	3.63
0.99800	0.15	0.12	0.12	0.99470	2.40	1.90	1.89	0.99154	4.65	3.70	3.67
0.99793	0.20	0.16	0.16	0.99463	2.45	1.94	1.93	0.99147	4.70	3.74	3.71
0.99785	0.25	0.20	0.20	0.99456	2.50	1.98	1.97	0.99140	4.75	3.78	3.75
0.99778	0.30	0.24	0.24	0.99449	2.55	2.02	2.01	0.99133	4.80	3.82	3.79
0.99770	0.35	0.28	0.28	0.99442	2.60	2.06	2.05	0.99127	4.85	3.86	3.83
0.99763	0.40	0.32	0.32	0.99434	2.65	2.10	2.09	0.99120	4.90	3.90	3.87
0.99755	0.45	0.36	0.36	0.99427	2.70	2.14	2.13	0.99113	4.95	3.94	3.91
0.99748	0.50	0.40	0.40	0.99420	2.75	2.18	2.17	0.99106	5.00	3.98	3.95
0.99741	0.55	0.44	0.44	0.99413	2.80	2.22	2.21	0.99100	5.05	4.02	3.99
0.99734	0.60	0.47	0.47	0.99405	2.85	2.26	2.25	0.99093	5.10	4.06	4.03
0.99726	0.65	0.51	0.51	0.99398	2.90	2.30	2.29	0.99087	5.15	4.10	4.07
0.99719	0.70	0.55	0.55	0.99391	2.95	2.34	2.33	0.99080	5.20	4.14	4.10
0.99711	0.75	0.59	0.59	0.99384	3.00	2.38	2.37	0.99073	5.25	4.18	4.14
0.99704	0.80	0.63	0.63	0.99377	3.05	2.42	2.41	0.99066	5.30	4.22	4.18
0.99697	0.85	0.67	0.67	0.99370	3.10	2.46	2.45	0.99060	5.35	4.26	4.22
0.99690	0.90	0.71	0.71	0.99362	3.15	2.50	2.49	0.99053	5.40	4.30	4.26
0.99682	0.95	0.75	0.75	0.99355	3.20	2.54	2.53	0.99047	5.45	4.34	4.30
0.99675	1.00	0.79	0.79	0.99348	3.25	2.58	2.57	0.99040	5.50	4.38	4.34
0.99667	1.05	0.83	0.83	0.99341	3.30	2.62	2.60	0.99033	5.55	4.42	4.38
0.99660	1.10	0.87	0.87	0.99334	3.35	2.66	2.64	0.99026	5.60	4.46	4.42
0.99652	1.15	0.91	0.91	0.99327	3.40	2.70	2.68	0.99020	5.65	4.50	4.46
0.99645	1.20	0.95	0.95	0.99320	3.45	2.74	2.72	0.99013	5.70	4.54	4.50
0.99638	1.25	0.99	0.99	0.99313	3.50	2.78	2.76	0.99006	5.75	4.58	4.54
0.99631	1.30	1.03	1.03	0.99306	3.55	2.82	2.80	0.98999	5.80	4.62	4.58
0.99623	1.35	1.07	1.07	0.99299	3.60	2.86	2.84	0.98993	5.85	4.66	4.62
0.99616	1.40	1.11	1.11	0.99292	3.65	2.90	2.88	0.98986	5.90	4.70	4.66
0.99608	1.45	1.15	1.15	0.99285	3.70	2.94	2.92	0.98980	5.95	4.74	4.70
0.99601	1.50	1.19	1.19	0.99278	3.75	2.98	2.96	0.98973	6.00	4.78	4.74
0.99594	1.55	1.23	1.23	0.99271	3.80	3.02	3.00	0.98967	6.05	4.82	4.78
0.99587	1.60	1.27	1.26	0.99264	3.85	3.06	3.04	0.98960	6.10	4.87	4.82
0.99579	1.65	1.31	1.30	0.99257	3.90	3.10	3.08	0.98954	6.15	4.91	4.86
0.99572	1.70	1.35	1.34	0.99250	3.95	3.14	3.12	0.98947	6.20	4.95	4.89
0.99564	1.75	1.39	1.38	0.99243	4.00	3.18	3.16	0.98941	6.25	4.99	4.93
0.99557	1.80	1.43	1.42	0.99236	4.05	3.22	3.20	0.98934	6.30	5.03	4.97
0.99550	1.85	1.47	1.46	0.99229	4.10	3.26	3.24	0.98928	6.35	5.07	5.01
0.99543	1.90	1.51	1.50	0.99222	4.15	3.30	3.28	0.98921	6.40	5.11	5.05
0.99535	1.95	1.55	1.54	0.99215	4.20	3.34	3.32	0.98915	6.45	5.15	5.09
0.99528	2.00	1.59	1.58	0.99208	4.25	3.38	3.36	0.98908	6.50	5.19	5.13
0.99520	2.05	1.63	1.62	0.99201	4.30	3.42	3.39	0.98902	6.55	5.23	5.17
0.99513	2.10	1.67	1.66	0.99195	4.35	3.46	3.43	0.98895	6.60	5.27	5.21
0.99506	2.15	1.71	1.70	0.99188	4.40	3.50	3.47	0.98889	6.65	5.31	5.25
0.99499	2.20	1.75	1.74	0.99181	4.45	3.54	3.51	0.98882	6.70	5.35	5.29

5

TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.98876	6.75	5.39	5.33	0.98566	9.25	7.41	7.30	0.98267	11.75	9.44	9.28
0.98870	6.80	5.43	5.37	0.98560	9.30	7.45	7.34	0.98261	11.80	9.48	9.31
0.98864	6.85	5.47	5.41	0.98554	9.35	7.49	7.38	0.98255	11.85	9.52	9.35
0.98857	6.90	5.51	5.45	0.98549	9.40	7.53	7.42	0.98250	11.90	9.56	9.39
0.98851	6.95	5.55	5.49	0.98543	9.45	7.57	7.46	0.98244	11.95	9.60	9.43
0.98845	7.00	5.59	5.53	0.98537	9.50	7.61	7.50	0.98238	12.00	9.64	9.47
0.98839	7.05	5.63	5.57	0.98531	9.55	7.65	7.54	0.98232	12.05	9.68	9.51
0.98832	7.10	5.67	5.60	0.98524	9.60	7.69	7.58	0.98226	12.10	9.72	9.55
0.98826	7.15	5.71	5.64	0.98518	9.65	7.73	7.62	0.98220	12.15	9.76	9.59
0.98820	7.20	5.75	5.68	0.98512	9.70	7.77	7.66	0.98214	12.20	9.80	9.63
0.98813	7.25	5.79	5.72	0.98506	9.75	7.81	7.70	0.98208	12.25	9.84	9.67
0.98806	7.30	5.83	5.76	0.98501	9.80	7.85	7.73	0.98203	12.30	9.89	9.71
0.98800	7.35	5.87	5.80	0.98495	9.85	7.89	7.77	0.98197	12.35	9.93	9.75
0.98794	7.40	5.91	5.84	0.98488	9.90	7.93	7.81	0.98191	12.40	9.97	9.79
0.98788	7.45	5.95	5.88	0.98482	9.95	7.97	7.85	0.98185	12.45	10.01	9.83
0.98781	7.50	5.99	5.92	0.98476	10.00	8.02	7.89	0.98180	12.50	10.05	9.87
0.98775	7.55	6.03	5.96	0.98470	10.05	8.06	7.93	0.98174	12.55	10.09	9.91
0.98769	7.60	6.07	6.00	0.98463	10.10	8.10	7.97	0.98168	12.60	10.13	9.95
0.98763	7.65	6.11	6.04	0.98457	10.15	8.14	8.01	0.98162	12.65	10.17	9.99
0.98756	7.70	6.15	6.08	0.98452	10.20	8.18	8.05	0.98156	12.70	10.21	10.03
0.98750	7.75	6.19	6.12	0.98446	10.25	8.22	8.09	0.98150	12.75	10.25	10.07
0.98744	7.80	6.24	6.16	0.98441	10.30	8.26	8.13	0.98145	12.80	10.29	10.10
0.98738	7.85	6.28	6.20	0.98435	10.35	8.30	8.17	0.98139	12.85	10.33	10.14
0.98731	7.90	6.32	6.24	0.98428	10.40	8.34	8.21	0.98132	12.90	10.38	10.18
0.98725	7.95	6.36	6.28	0.98422	10.45	8.38	8.25	0.98127	12.95	10.42	10.22
0.98718	8.00	6.40	6.32	0.98416	10.50	8.42	8.29	0.98122	13.00	10.46	10.26
0.98712	8.05	6.44	6.36	0.98410	10.55	8.46	8.33	0.98116	13.05	10.50	10.30
0.98706	8.10	6.48	6.39	0.98404	10.60	8.50	8.37	0.98111	13.10	10.54	10.34
0.98700	8.15	6.52	6.43	0.98398	10.65	8.54	8.41	0.98105	13.15	10.58	10.38
0.98694	8.20	6.56	6.47	0.98391	10.70	8.58	8.45	0.98100	13.20	10.62	10.42
0.98688	8.25	6.60	6.51	0.98385	10.75	8.62	8.49	0.98094	13.25	10.66	10.46
0.98682	8.30	6.64	6.55	0.98379	10.80	8.66	8.52	0.98089	13.30	10.70	10.50
0.98676	8.35	6.68	6.59	0.98373	10.85	8.70	8.56	0.98083	13.35	10.74	10.54
0.98670	8.40	6.72	6.63	0.98368	10.90	8.75	8.60	0.98077	13.40	10.78	10.58
0.98664	8.45	6.76	6.67	0.98362	10.95	8.79	8.64	0.98071	13.45	10.82	10.62
0.98658	8.50	6.80	6.71	0.98356	11.00	8.83	8.68	0.98066	13.50	10.86	10.66
0.98652	8.55	6.84	6.75	0.98350	11.05	8.87	8.72	0.98060	13.55	10.90	10.70
0.98646	8.60	6.88	6.79	0.98344	11.10	8.91	8.76	0.98054	13.60	10.95	10.74
0.98640	8.65	6.92	6.83	0.98338	11.15	8.95	8.80	0.98048	13.65	10.99	10.78
0.98633	8.70	6.96	6.87	0.98332	11.20	8.99	8.84	0.98043	13.70	11.03	10.81
0.98627	8.75	7.00	6.91	0.98326	11.25	9.03	8.88	0.98037	13.75	11.07	10.85
0.98620	8.80	7.04	6.95	0.98320	11.30	9.07	8.92	0.98031	13.80	11.11	10.89
0.98614	8.85	7.08	6.99	0.98314	11.35	9.11	8.96	0.98025	13.85	11.15	10.93
0.98608	8.90	7.12	7.03	0.98308	11.40	9.15	9.00	0.98020	13.90	11.19	10.97
0.98602	8.95	7.16	7.07	0.98302	11.45	9.19	9.04	0.98014	13.95	11.23	11.01
0.98596	9.00	7.20	7.10	0.98296	11.50	9.23	9.08	0.98009	14.00	11.28	11.05
0.98590	9.05	7.24	7.14	0.98290	11.55	9.27	9.12	0.98003	14.05	11.32	11.09
0.98584	9.10	7.29	7.18	0.98285	11.60	9.32	9.16	0.97998	14.10	11.36	11.13
0.98578	9.15	7.33	7.22	0.98279	11.65	9.36	9.20	0.97992	14.15	11.40	11.17
0.98572	9.20	7.37	7.26	0.98273	11.70	9.40	9.24	0.97986	14.20	11.44	11.21

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TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.97980	14.25	11.48	11.25	0.97704	16.75	13.53	13.22	0.97438	19.25	15.59	15.20
0.97975	14.30	11.52	11.29	0.97699	16.80	13.57	13.26	0.97433	19.30	15.64	15.23
0.97969	14.35	11.56	11.33	0.97694	16.85	13.61	13.30	0.97428	19.35	15.68	15.27
0.97964	14.40	11.60	11.37	0.97689	16.90	13.66	13.34	0.97423	19.40	15.72	15.31
0.97958	14.45	11.64	11.41	0.97683	16.95	13.70	13.38	0.97417	19.45	15.76	15.35
0.97953	14.50	11.68	11.44	0.97678	17.00	13.74	13.42	0.97412	19.50	15.80	15.39
0.97947	14.55	11.72	11.48	0.97672	17.05	13.78	13.46	0.97407	19.55	15.84	15.43
0.97942	14.60	11.77	11.52	0.97667	17.10	13.82	13.50	0.97402	19.60	15.88	15.47
0.97936	14.65	11.81	11.56	0.97661	17.15	13.86	13.54	0.97396	19.65	15.92	15.51
0.97930	14.70	11.85	11.60	0.97656	17.20	13.90	13.58	0.97391	19.70	15.97	15.55
0.97924	14.75	11.89	11.64	0.97650	17.25	13.94	13.62	0.97386	19.75	16.01	15.59
0.97919	14.80	11.93	11.68	0.97645	17.30	13.98	13.66	0.97381	19.80	16.05	15.63
0.97913	14.85	11.97	11.72	0.97639	17.35	14.02	13.70	0.97375	19.85	16.09	15.67
0.97908	14.90	12.01	11.76	0.97634	17.40	14.07	13.74	0.97370	19.90	16.13	15.71
0.97902	14.95	12.05	11.80	0.97629	17.45	14.11	13.78	0.97364	19.95	16.17	15.75
0.97897	15.00	12.09	11.84	0.97624	17.50	14.15	13.81	0.97359	20.00	16.21	15.79
0.97891	15.05	12.13	11.88	0.97618	17.55	14.19	13.85	0.97354	20.05	16.25	15.83
0.97885	15.10	12.18	11.92	0.97613	17.60	14.23	13.89	0.97349	20.10	16.30	15.87
0.97879	15.15	12.22	11.96	0.97607	17.65	14.27	13.93	0.97344	20.15	16.34	15.91
0.97874	15.20	12.26	12.00	0.97602	17.70	14.31	13.97	0.97339	20.20	16.38	15.95
0.97868	15.25	12.30	12.04	0.97596	17.75	14.35	14.01	0.97333	20.25	16.42	15.99
0.97863	15.30	12.34	12.08	0.97591	17.80	14.40	14.05	0.97328	20.30	16.46	16.02
0.97857	15.35	12.38	12.12	0.97586	17.85	14.44	14.09	0.97322	20.35	16.50	16.06
0.97852	15.40	12.42	12.16	0.97581	17.90	14.48	14.13	0.97317	20.40	16.55	16.10
0.97846	15.45	12.46	12.20	0.97575	17.95	14.52	14.17	0.97311	20.45	16.59	16.14
0.97841	15.50	12.50	12.23	0.97570	18.00	14.56	14.21	0.97306	20.50	16.63	16.18
0.97835	15.55	12.54	12.27	0.97564	18.05	14.60	14.25	0.97300	20.55	16.67	16.22
0.97830	15.60	12.59	12.31	0.97559	18.10	14.64	14.29	0.97295	20.60	16.71	16.26
0.97824	15.65	12.63	12.35	0.97553	18.15	14.68	14.33	0.97289	20.65	16.75	16.30
0.97819	15.70	12.67	12.39	0.97548	18.20	14.73	14.37	0.97284	20.70	16.80	16.34
0.97813	15.75	12.71	12.43	0.97542	18.25	14.77	14.41	0.97278	20.75	16.84	16.38
0.97808	15.80	12.75	12.47	0.97538	18.30	14.81	14.45	0.97273	20.80	16.88	16.42
0.97802	15.85	12.79	12.51	0.97532	18.35	14.85	14.49	0.97268	20.85	16.92	16.46
0.97797	15.90	12.83	12.55	0.97527	18.40	14.89	14.52	0.97263	20.90	16.96	16.50
0.97791	15.95	12.87	12.59	0.97522	18.45	14.93	14.56	0.97257	20.95	17.00	16.54
0.97786	16.00	12.92	12.63	0.97517	18.50	14.97	14.60	0.97252	21.00	17.04	16.58
0.97780	16.05	12.96	12.67	0.97512	18.55	15.01	14.64	0.97247	21.05	17.08	16.62
0.97775	16.10	13.00	12.71	0.97507	18.60	15.06	14.68	0.97242	21.10	17.13	16.66
0.97769	16.15	13.04	12.75	0.97501	18.65	15.10	14.72	0.97237	21.15	17.17	16.70
0.97764	16.20	13.08	12.79	0.97496	18.70	15.14	14.76	0.97232	21.20	17.21	16.73
0.97758	16.25	13.12	12.83	0.97490	18.75	15.18	14.80	0.97227	21.25	17.25	16.77
0.97753	16.30	13.16	12.87	0.97485	18.80	15.22	14.84	0.97222	21.30	17.29	16.81
0.97747	16.35	13.20	12.91	0.97479	18.85	15.26	14.88	0.97216	21.35	17.33	16.85
0.97742	16.40	13.24	12.95	0.97474	18.90	15.30	14.92	0.97210	21.40	17.37	16.89
0.97737	16.45	13.28	12.99	0.97469	18.95	15.34	14.96	0.97204	21.45	17.42	16.93
0.97732	16.50	13.33	13.02	0.97464	19.00	15.39	15.00	0.97199	21.50	17.46	16.97
0.97726	16.55	13.37	13.06	0.97459	19.05	15.43	15.04	0.97193	21.55	17.50	17.01
0.97721	16.60	13.41	13.10	0.97454	19.10	15.47	15.08	0.97188	21.60	17.54	17.05
0.97715	16.65	13.45	13.14	0.97449	19.15	15.51	15.12	0.97183	21.65	17.58	17.09
0.97710	16.70	13.49	13.18	0.97444	19.20	15.55	15.16	0.97178	21.70	17.63	17.13

TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.97172	21.75	17.67	17.17	0.96896	24.25	19.75	19.14	0.96612	26.75	21.85	21.12
0.97167	21.80	17.71	17.21	0.96891	24.30	19.80	19.18	0.96606	26.80	21.90	21.16
0.97161	21.85	17.75	17.25	0.96885	24.35	19.84	19.22	0.96600	26.85	21.94	21.20
0.97156	21.90	17.79	17.29	0.96880	24.40	19.88	19.26	0.96595	26.90	21.98	21.23
0.97150	21.95	17.83	17.33	0.96874	24.45	19.92	19.30	0.96589	26.95	22.02	21.27
0.97145	22.00	17.88	17.37	0.96869	24.50	19.96	19.34	0.96583	27.00	22.07	21.31
0.97139	22.05	17.92	17.41	0.96863	24.55	20.00	19.38	0.96577	27.05	22.11	21.35
0.97134	22.10	17.96	17.45	0.96857	24.60	20.05	19.42	0.96571	27.10	22.15	21.39
0.97128	22.15	18.00	17.49	0.96851	24.65	20.09	19.46	0.96565	27.15	22.19	21.43
0.97123	22.20	18.04	17.52	0.96846	24.70	20.13	19.50	0.96559	27.20	22.24	21.47
0.97118	22.25	18.08	17.56	0.96840	24.75	20.17	19.54	0.96553	27.25	22.28	21.51
0.97113	22.30	18.13	17.60	0.96835	24.80	20.22	19.58	0.96548	27.30	22.32	21.55
0.97107	22.35	18.17	17.64	0.96829	24.85	20.26	19.62	0.96542	27.35	22.36	21.59
0.97102	22.40	18.21	17.68	0.96823	24.90	20.30	19.66	0.96536	27.40	22.40	21.63
0.97096	22.45	18.25	17.72	0.96817	24.95	20.34	19.70	0.96530	27.45	22.44	21.67
0.97091	22.50	18.29	17.76	0.96812	25.00	20.38	19.73	0.96525	27.50	22.49	21.71
0.97085	22.55	18.33	17.80	0.96806	25.05	20.42	19.77	0.96519	27.55	22.53	21.75
0.97080	22.60	18.38	17.84	0.96801	25.10	20.47	19.81	0.96513	27.60	22.57	21.79
0.97074	22.65	18.42	17.88	0.96795	25.15	20.51	19.85	0.96507	27.65	22.61	21.83
0.97069	22.70	18.46	17.92	0.96789	25.20	20.55	19.89	0.96501	27.70	22.66	21.87
0.97063	22.75	18.50	17.96	0.96783	25.25	20.59	19.93	0.96495	27.75	22.70	21.91
0.97058	22.80	18.54	18.00	0.96778	25.30	20.64	19.97	0.96489	27.80	22.74	21.94
0.97052	22.85	18.58	18.04	0.96772	25.35	20.68	20.01	0.96483	27.85	22.78	21.98
0.97047	22.90	18.63	18.08	0.96766	25.40	20.72	20.05	0.96477	27.90	22.83	22.02
0.97041	22.95	18.67	18.12	0.96760	25.45	20.76	20.09	0.96471	27.95	22.87	22.06
0.97036	23.00	18.71	18.16	0.96755	25.50	20.80	20.13	0.96465	28.00	22.91	22.10
0.97030	23.05	18.75	18.20	0.96749	25.55	20.84	20.17	0.96459	28.05	22.95	22.14
0.97025	23.10	18.79	18.24	0.96744	25.60	20.89	20.21	0.96454	28.10	23.00	22.18
0.97019	23.15	18.83	18.28	0.96738	25.65	20.93	20.25	0.96448	28.15	23.04	22.22
0.97013	23.20	18.88	18.31	0.96733	25.70	20.97	20.29	0.96442	28.20	23.08	22.26
0.97007	23.25	18.92	18.35	0.96727	25.75	21.01	20.33	0.96436	28.25	23.12	22.30
0.97002	23.30	18.96	18.39	0.96722	25.80	21.06	20.37	0.96430	28.30	23.17	22.34
0.96996	23.35	19.00	18.43	0.96716	25.85	21.10	20.41	0.96424	28.35	23.21	22.38
0.96991	23.40	19.04	18.47	0.96710	25.90	21.14	20.44	0.96418	28.40	23.25	22.42
0.96985	23.45	19.08	18.51	0.96704	25.95	21.18	20.48	0.96412	28.45	23.29	22.46
0.96980	23.50	19.13	18.55	0.96699	26.00	21.22	20.52	0.96406	28.50	23.33	22.50
0.96974	23.55	19.17	18.59	0.96693	26.05	21.26	20.56	0.96400	28.55	23.37	22.54
0.96969	23.60	19.21	18.63	0.96687	26.10	21.31	20.60	0.96393	28.60	23.42	22.57
0.96963	23.65	19.25	18.67	0.96681	26.15	21.35	20.64	0.96387	28.65	23.46	22.61
0.96958	23.70	19.29	18.71	0.96675	26.20	21.39	20.68	0.96381	28.70	23.51	22.65
0.96952	23.75	19.33	18.75	0.96669	26.25	21.43	20.72	0.96375	28.75	23.55	22.69
0.96947	23.80	19.38	18.79	0.96664	26.30	21.48	20.76	0.96369	28.80	23.59	22.73
0.96941	23.85	19.42	18.83	0.96658	26.35	21.52	20.80	0.96363	28.85	23.63	22.77
0.96936	23.90	19.46	18.87	0.96653	26.40	21.56	20.84	0.96357	28.90	23.67	22.81
0.96930	23.95	19.50	18.91	0.96647	26.45	21.60	20.88	0.96351	28.95	23.71	22.85
0.96925	24.00	19.55	18.94	0.96641	26.50	21.64	20.92	0.96346	29.00	23.76	22.89
0.96919	24.05	19.59	18.98	0.96635	26.55	21.68	20.96	0.96340	29.05	23.80	22.93
0.96913	24.10	19.63	19.02	0.96630	26.60	21.73	21.00	0.96334	29.10	23.84	22.97
0.96907	24.15	19.67	19.06	0.96624	26.65	21.77	21.04	0.96328	29.15	23.88	23.01
0.96902	24.20	19.71	19.10	0.96618	26.70	21.81	21.08	0.96322	29.20	23.93	23.05

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TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.96316	29.25	23.97	23.09	0.96005	31.75	26.10	25.06	0.95669	34.25	28.26	27.03
0.96310	29.30	24.01	23.13	0.95998	31.80	26.15	25.10	0.95662	34.30	28.30	27.07
0.96304	29.35	24.05	23.17	0.95992	31.85	26.19	25.14	0.95655	34.35	28.34	27.11
0.96297	29.40	24.10	23.21	0.95985	31.90	26.23	25.18	0.95648	34.40	28.39	27.15
0.96291	29.45	24.14	23.25	0.95979	31.95	26.27	25.22	0.95641	34.45	28.43	27.19
0.96285	29.50	24.18	23.29	0.95972	32.00	26.32	25.26	0.95634	34.50	28.48	27.23
0.96279	29.55	24.22	23.33	0.95965	32.05	26.36	25.30	0.95627	34.55	28.52	27.27
0.96273	29.60	24.27	23.36	0.95958	32.10	26.41	25.34	0.95619	34.60	28.56	27.31
0.96267	29.65	24.31	23.40	0.95952	32.15	26.45	25.38	0.95612	34.65	28.60	27.35
0.96261	29.70	24.35	23.44	0.95945	32.20	26.49	25.42	0.95605	34.70	28.65	27.39
0.96255	29.75	24.39	23.48	0.95939	32.25	26.53	25.46	0.95598	34.75	28.69	27.43
0.96248	29.80	24.44	23.52	0.95932	32.30	26.58	25.50	0.95591	34.80	28.74	27.47
0.96242	29.85	24.48	23.56	0.95926	32.35	26.62	25.54	0.95584	34.85	28.78	27.51
0.96236	29.90	24.52	23.60	0.95920	32.40	26.66	25.58	0.95577	34.90	28.82	27.55
0.96230	29.95	24.56	23.64	0.95913	32.45	26.70	25.61	0.95570	34.95	28.86	27.59
0.96224	30.00	24.61	23.68	0.95906	32.50	26.75	25.64	0.95563	35.00	28.91	27.63
0.96218	30.05	24.65	23.72	0.95900	32.55	26.79	25.68	0.95556	35.05	28.95	27.67
0.96211	30.10	24.69	23.76	0.95893	32.60	26.83	25.72	0.95549	35.10	29.00	27.71
0.96205	30.15	24.73	23.80	0.95887	32.65	26.87	25.76	0.95542	35.15	29.04	27.75
0.96199	30.20	24.78	23.84	0.95880	32.70	26.92	25.80	0.95535	35.20	29.08	27.79
0.96193	30.25	24.82	23.88	0.95873	32.75	26.96	25.84	0.95528	35.25	29.12	27.82
0.96187	30.30	24.87	23.92	0.95866	32.80	27.01	25.89	0.95521	35.30	29.17	27.86
0.96181	30.35	24.91	23.96	0.95859	32.85	27.05	25.93	0.95513	35.35	29.21	27.90
0.96175	30.40	24.95	24.00	0.95852	32.90	27.09	25.97	0.95506	35.40	29.26	27.94
0.96169	30.45	24.99	24.04	0.95846	32.95	27.13	26.01	0.95499	35.45	29.30	27.98
0.96163	30.50	25.04	24.08	0.95839	33.00	27.18	26.05	0.95492	35.50	29.34	28.02
0.96157	30.55	25.08	24.12	0.95833	33.05	27.22	26.09	0.95485	35.55	29.38	28.06
0.96150	30.60	25.12	24.15	0.95826	33.10	27.27	26.13	0.95478	35.60	29.43	28.10
0.96144	30.65	25.16	24.19	0.95819	33.15	27.31	26.17	0.95470	35.65	29.47	28.14
0.96138	30.70	25.21	24.23	0.95812	33.20	27.35	26.21	0.95463	35.70	29.52	28.18
0.96132	30.75	25.25	24.27	0.95806	33.25	27.39	26.25	0.95456	35.75	29.56	28.22
0.96125	30.80	25.30	24.31	0.95799	33.30	27.44	26.29	0.95449	35.80	29.61	28.26
0.96119	30.85	25.34	24.35	0.95792	33.35	27.48	26.33	0.95441	35.85	29.65	28.30
0.96112	30.90	25.38	24.39	0.95785	33.40	27.52	26.36	0.95434	35.90	29.69	28.34
0.96106	30.95	25.42	24.43	0.95778	33.45	27.56	26.40	0.95426	35.95	29.73	28.38
0.96100	31.00	25.46	24.47	0.95771	33.50	27.61	26.44	0.95419	36.00	29.78	28.42
0.96094	31.05	25.50	24.51	0.95764	33.55	27.65	26.48	0.95412	36.05	29.82	28.46
0.96088	31.10	25.55	24.55	0.95757	33.60	27.70	26.52	0.95405	36.10	29.87	28.49
0.96082	31.15	25.59	24.59	0.95751	33.65	27.74	26.56	0.95397	36.15	29.91	28.53
0.96075	31.20	25.63	24.63	0.95745	33.70	27.78	26.60	0.95390	36.20	29.95	28.57
0.96069	31.25	25.67	24.67	0.95738	33.75	27.82	26.64	0.95382	36.25	29.99	28.61
0.96062	31.30	25.72	24.71	0.95731	33.80	27.87	26.68	0.95375	36.30	30.04	28.65
0.96056	31.35	25.76	24.75	0.95724	33.85	27.91	26.72	0.95367	36.35	30.09	28.69
0.96049	31.40	25.81	24.79	0.95717	33.90	27.96	26.76	0.95360	36.40	30.13	28.73
0.96043	31.45	25.85	24.83	0.95710	33.95	28.00	26.80	0.95353	36.45	30.17	28.77
0.96036	31.50	25.89	24.86	0.95703	34.00	28.04	26.84	0.95346	36.50	30.22	28.81
0.96030	31.55	25.93	24.90	0.95696	34.05	28.08	26.88	0.95338	36.55	30.26	28.85
0.96024	31.60	25.98	24.94	0.95689	34.10	28.13	26.92	0.95331	36.60	30.31	28.89
0.96018	31.65	26.02	24.98	0.95682	34.15	28.17	26.96	0.95323	36.65	30.35	28.93
0.96011	31.70	26.06	25.02	0.95675	34.20	28.22	26.99	0.95315	36.70	30.39	28.97

TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.95308	36.75	<i>30.43</i>	29.01	0.94926	39.25	<i>32.63</i>	30.99	0.94519	41.75	<i>34.86</i>	32.96
0.95301	36.80	<i>30.48</i>	29.05	0.94918	39.30	<i>32.68</i>	31.02	0.94510	41.80	<i>34.91</i>	33.00
0.95294	36.85	<i>30.52</i>	29.09	0.94910	39.35	<i>32.72</i>	31.06	0.94502	41.85	<i>34.95</i>	33.04
0.95287	36.90	<i>30.57</i>	29.13	0.94901	39.40	<i>32.77</i>	31.10	0.94494	41.90	<i>35.00</i>	33.07
0.95279	36.95	<i>30.61</i>	29.17	0.94893	39.45	<i>32.81</i>	31.14	0.94486	41.95	<i>35.04</i>	33.11
0.95272	37.00	<i>30.66</i>	29.21	0.94885	39.50	<i>32.86</i>	31.18	0.94477	42.00	<i>35.09</i>	33.15
0.95264	37.05	<i>30.70</i>	29.25	0.94877	39.55	<i>32.90</i>	31.22	0.94469	42.05	<i>35.13</i>	33.19
0.95257	37.10	<i>30.74</i>	29.29	0.94869	39.60	<i>32.95</i>	31.26	0.94460	42.10	<i>35.18</i>	33.23
0.95249	37.15	<i>30.78</i>	29.33	0.94861	39.65	<i>32.99</i>	31.30	0.94452	42.15	<i>35.22</i>	33.27
0.95242	37.20	<i>30.83</i>	29.36	0.94853	39.70	<i>33.04</i>	31.34	0.94443	42.20	<i>35.27</i>	33.31
0.95234	37.25	<i>30.87</i>	29.40	0.94845	39.75	<i>33.08</i>	31.38	0.94435	42.25	<i>35.31</i>	33.35
0.95227	37.30	<i>30.92</i>	29.44	0.94837	39.80	<i>33.13</i>	31.42	0.94427	42.30	<i>35.36</i>	33.39
0.95219	37.35	<i>30.96</i>	29.48	0.94829	39.85	<i>33.17</i>	31.46	0.94419	42.35	<i>35.40</i>	33.43
0.95211	37.40	<i>31.01</i>	29.52	0.94821	39.90	<i>33.22</i>	31.50	0.94410	42.40	<i>35.45</i>	33.47
0.95203	37.45	<i>31.05</i>	29.56	0.94813	39.95	<i>33.26</i>	31.54	0.94402	42.45	<i>35.49</i>	33.51
0.95196	37.50	<i>31.09</i>	29.60	0.94805	40.00	<i>33.30</i>	31.57	0.94393	42.50	<i>35.54</i>	33.55
0.95188	37.55	<i>31.13</i>	29.64	0.94797	40.05	<i>33.34</i>	31.61	0.94385	42.55	<i>35.58</i>	33.59
0.95181	37.60	<i>31.18</i>	29.68	0.94789	40.10	<i>33.39</i>	31.65	0.94376	42.60	<i>35.63</i>	33.63
0.95173	37.65	<i>31.22</i>	29.72	0.94781	40.15	<i>33.43</i>	31.69	0.94368	42.65	<i>35.67</i>	33.67
0.95166	37.70	<i>31.27</i>	29.76	0.94773	40.20	<i>33.48</i>	31.73	0.94359	42.70	<i>35.72</i>	33.71
0.95158	37.75	<i>31.31</i>	29.80	0.94765	40.25	<i>33.52</i>	31.77	0.94351	42.75	<i>35.76</i>	33.75
0.95151	37.80	<i>31.36</i>	29.84	0.94757	40.30	<i>33.57</i>	31.81	0.94342	42.80	<i>35.81</i>	33.78
0.95143	37.85	<i>31.40</i>	29.88	0.94749	40.35	<i>33.61</i>	31.85	0.94334	42.85	<i>35.85</i>	33.82
0.95135	37.90	<i>31.45</i>	29.92	0.94741	40.40	<i>33.66</i>	31.89	0.94325	42.90	<i>35.90</i>	33.86
0.95127	37.95	<i>31.49</i>	29.96	0.94733	40.45	<i>33.70</i>	31.93	0.94317	42.95	<i>35.94</i>	33.90
0.95120	38.00	<i>31.53</i>	29.99	0.94725	40.50	<i>33.75</i>	31.97	0.94308	43.00	<i>35.99</i>	33.94
0.95112	38.05	<i>31.57</i>	30.03	0.94717	40.55	<i>33.79</i>	32.01	0.94300	43.05	<i>36.03</i>	33.98
0.95104	38.10	<i>31.62</i>	30.07	0.94708	40.60	<i>33.84</i>	32.05	0.94291	43.10	<i>36.08</i>	34.02
0.95096	38.15	<i>31.66</i>	30.11	0.94700	40.65	<i>33.88</i>	32.09	0.94283	43.15	<i>36.12</i>	34.06
0.95089	38.20	<i>31.71</i>	30.15	0.94692	40.70	<i>33.93</i>	32.13	0.94274	43.20	<i>36.17</i>	34.10
0.95081	38.25	<i>31.75</i>	30.19	0.94684	40.75	<i>33.97</i>	32.17	0.94265	43.25	<i>36.21</i>	34.14
0.95074	38.30	<i>31.80</i>	30.23	0.94676	40.80	<i>34.02</i>	32.20	0.94256	43.30	<i>36.26</i>	34.18
0.95066	38.35	<i>31.84</i>	30.27	0.94668	40.85	<i>34.06</i>	32.24	0.94248	43.35	<i>36.30</i>	34.22
0.95058	38.40	<i>31.89</i>	30.31	0.94659	40.90	<i>34.11</i>	32.28	0.94239	43.40	<i>36.35</i>	34.26
0.95050	38.45	<i>31.93</i>	30.35	0.94651	40.95	<i>34.15</i>	32.32	0.94231	43.45	<i>36.39</i>	34.30
0.95043	38.50	<i>31.97</i>	30.39	0.94643	41.00	<i>34.19</i>	32.36	0.94222	43.50	<i>36.44</i>	34.34
0.95035	38.55	<i>32.01</i>	30.43	0.94635	41.05	<i>34.23</i>	32.40	0.94214	43.55	<i>36.48</i>	34.38
0.95027	38.60	<i>32.06</i>	30.47	0.94627	41.10	<i>34.28</i>	32.44	0.94205	43.60	<i>36.53</i>	34.42
0.95019	38.65	<i>32.10</i>	30.51	0.94619	41.15	<i>34.32</i>	32.48	0.94197	43.65	<i>36.57</i>	34.46
0.95011	38.70	<i>32.15</i>	30.55	0.94610	41.20	<i>34.37</i>	32.52	0.94188	43.70	<i>36.62</i>	34.49
0.95003	38.75	<i>32.19</i>	30.59	0.94602	41.25	<i>34.41</i>	32.56	0.94179	43.75	<i>36.66</i>	34.53
0.94996	38.80	<i>32.24</i>	30.63	0.94594	41.30	<i>34.46</i>	32.60	0.94170	43.80	<i>36.71</i>	34.57
0.94988	38.85	<i>32.28</i>	30.67	0.94586	41.35	<i>34.50</i>	32.64	0.94161	43.85	<i>36.75</i>	34.61
0.94980	38.90	<i>32.33</i>	30.71	0.94577	41.40	<i>34.55</i>	32.68	0.94152	43.90	<i>36.80</i>	34.65
0.94972	38.95	<i>32.37</i>	30.75	0.94569	41.45	<i>34.59</i>	32.72	0.94144	43.95	<i>36.84</i>	34.69
0.94964	39.00	<i>32.42</i>	30.79	0.94560	41.50	<i>34.64</i>	32.76	0.94135	44.00	<i>36.89</i>	34.73
0.94956	39.05	<i>32.46</i>	30.83	0.94552	41.55	<i>34.68</i>	32.80	0.94126	44.05	<i>36.94</i>	34.77
0.94949	39.10	<i>32.51</i>	30.87	0.94544	41.60	<i>34.73</i>	32.84	0.94117	44.10	<i>36.99</i>	34.81
0.94941	39.15	<i>32.55</i>	30.91	0.94536	41.65	<i>34.77</i>	32.88	0.94108	44.15	<i>37.03</i>	34.85
0.94934	39.20	<i>32.59</i>	30.95	0.94527	41.70	<i>34.82</i>	32.92	0.94099	44.20	<i>37.08</i>	34.89

5 TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.94091	44.25	37.12	34.93	0.93638	46.75	39.41	36.90	0.93164	49.25	41.72	38.87
0.94082	44.30	37.17	34.97	0.93629	46.80	39.46	36.94	0.93155	49.30	41.77	38.91
0.94073	44.35	37.21	35.01	0.93619	46.85	39.50	36.98	0.93145	49.35	41.82	38.95
0.94064	44.40	37.26	35.05	0.93610	46.90	39.55	37.02	0.93136	49.40	41.87	38.99
0.94055	44.45	37.30	35.09	0.93600	46.95	39.59	37.06	0.93126	49.45	41.91	39.03
0.94046	44.50	37.35	35.13	0.93591	47.00	39.64	37.10	0.93116	49.50	41.96	39.07
0.94037	44.55	37.39	35.17	0.93582	47.05	39.68	37.14	0.93106	49.55	42.01	39.11
0.94028	44.60	37.44	35.20	0.93573	47.10	39.73	37.18	0.93096	49.60	42.06	39.15
0.94020	44.65	37.48	35.24	0.93563	47.15	39.77	37.22	0.93086	49.65	42.10	39.19
0.94011	44.70	37.53	35.28	0.93554	47.20	39.82	37.26	0.93076	49.70	42.15	39.23
0.94002	44.75	37.57	35.32	0.93545	47.25	39.87	37.30	0.93066	49.75	42.19	39.27
0.93993	44.80	37.62	35.36	0.93536	47.30	39.92	37.34	0.93056	49.80	42.24	39.31
0.93984	44.85	37.66	35.40	0.93526	47.35	39.96	37.38	0.93046	49.85	42.29	39.35
0.93975	44.90	37.71	35.44	0.93517	47.40	40.01	37.42	0.93036	49.90	42.34	39.39
0.93966	44.95	37.75	35.48	0.93507	47.45	40.05	37.46	0.93026	49.95	42.38	39.43
0.93957	45.00	37.80	35.52	0.93498	47.50	40.10	37.49	0.93017	50.00	42.43	39.47
0.93948	45.05	37.85	35.56	0.93488	47.55	40.14	37.53	0.93007	50.05	42.47	39.51
0.93939	45.10	37.90	35.60	0.93479	47.60	40.19	37.57	0.92997	50.10	42.52	39.55
0.93931	45.15	37.94	35.64	0.93470	47.65	40.24	37.61	0.92987	50.15	42.57	39.59
0.93922	45.20	37.99	35.68	0.93461	47.70	40.29	37.65	0.92977	50.20	42.62	39.63
0.93912	45.25	38.03	35.72	0.93451	47.75	40.33	37.69	0.92967	50.25	42.66	39.67
0.93903	45.30	38.08	35.76	0.93442	47.80	40.38	37.73	0.92957	50.30	42.71	39.70
0.93894	45.35	38.12	35.80	0.93432	47.85	40.42	37.77	0.92947	50.35	42.76	39.74
0.93885	45.40	38.17	35.84	0.93423	47.90	40.47	37.81	0.92938	50.40	42.81	39.78
0.93876	45.45	38.21	35.88	0.93413	47.95	40.51	37.85	0.92928	50.45	42.85	39.82
0.93867	45.50	38.26	35.92	0.93404	48.00	40.56	37.89	0.92918	50.50	42.90	39.86
0.93858	45.55	38.30	35.96	0.93394	48.05	40.61	37.93	0.92908	50.55	42.94	39.90
0.93849	45.60	38.35	35.99	0.93385	48.10	40.66	37.97	0.92898	50.60	42.99	39.94
0.93840	45.65	38.39	36.03	0.93375	48.15	40.70	38.01	0.92888	50.65	43.04	39.98
0.93831	45.70	38.44	36.07	0.93366	48.20	40.75	38.05	0.92879	50.70	43.09	40.02
0.93822	45.75	38.49	36.11	0.93356	48.25	40.79	38.09	0.92869	50.75	43.13	40.06
0.93813	45.80	38.54	36.15	0.93347	48.30	40.84	38.13	0.92859	50.80	43.18	40.10
0.93803	45.85	38.58	36.19	0.93337	48.35	40.89	38.17	0.92849	50.85	43.23	40.14
0.93794	45.90	38.63	36.23	0.93328	48.40	40.94	38.21	0.92839	50.90	43.28	40.18
0.93785	45.95	38.67	36.27	0.93318	48.45	40.98	38.25	0.92829	50.95	43.32	40.22
0.93776	46.00	38.72	36.31	0.93308	48.50	41.03	38.29	0.92818	51.00	43.37	40.26
0.93767	46.05	38.76	36.35	0.93298	48.55	41.07	38.33	0.92808	51.05	43.42	40.30
0.93758	46.10	38.81	36.39	0.93289	48.60	41.12	38.36	0.92798	51.10	43.47	40.34
0.93749	46.15	38.85	36.43	0.93279	48.65	41.16	38.40	0.92788	51.15	43.51	40.38
0.93740	46.20	38.90	36.47	0.93270	48.70	41.21	38.44	0.92778	51.20	43.56	40.42
0.93730	46.25	38.95	36.51	0.93260	48.75	41.26	38.48	0.92768	51.25	43.60	40.46
0.93721	46.30	39.00	36.55	0.93251	48.80	41.31	38.52	0.92759	51.30	43.65	40.49
0.93712	46.35	39.04	36.59	0.93241	48.85	41.35	38.56	0.92749	51.35	43.70	40.53
0.93703	46.40	39.09	36.63	0.93232	48.90	41.40	38.60	0.92739	51.40	43.75	40.57
0.93693	46.45	39.13	36.67	0.93222	48.95	41.44	38.64	0.92729	51.45	43.79	40.61
0.93684	46.50	39.18	36.70	0.93213	49.00	41.49	38.68	0.92719	51.50	43.84	40.65
0.93675	46.55	39.22	36.74	0.93203	49.05	41.54	38.72	0.92709	51.55	43.89	40.69
0.93666	46.60	39.27	36.78	0.93194	49.10	41.59	38.76	0.92699	51.60	43.94	40.73
0.93656	46.65	39.31	36.82	0.93184	49.15	41.63	38.80	0.92689	51.65	43.98	40.77
0.93647	46.70	39.36	36.86	0.93174	49.20	41.68	38.83	0.92678	51.70	44.03	40.81

TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.92668	51.75	44.08	40.85	0.92157	54.25	46.46	42.82	0.91629	56.75	48.89	44.80
0.92658	51.80	44.13	40.89	0.92147	54.30	46.51	42.86	0.91618	56.80	48.94	44.83
0.92648	51.85	44.17	40.93	0.92137	54.35	46.56	42.90	0.91608	56.85	48.98	44.87
0.92637	51.90	44.22	40.97	0.92126	54.40	46.61	42.94	0.91597	56.90	49.03	44.91
0.92627	51.95	44.26	41.01	0.92116	54.45	46.66	42.98	0.91586	56.95	49.08	44.95
0.92617	52.00	44.31	41.05	0.92105	54.50	46.71	43.02	0.91575	57.00	49.13	44.99
0.92607	52.05	44.36	41.09	0.92095	54.55	46.75	43.06	0.91565	57.05	49.18	45.03
0.92597	52.10	44.41	41.13	0.92084	54.60	46.80	43.10	0.91554	57.10	49.23	45.07
0.92587	52.15	44.46	41.17	0.92074	54.65	46.85	43.14	0.91543	57.15	49.28	45.11
0.92577	52.20	44.51	41.20	0.92063	54.70	46.90	43.18	0.91532	57.20	49.33	45.15
0.92567	52.25	44.55	41.24	0.92053	54.75	46.94	43.22	0.91521	57.25	49.38	45.19
0.92557	52.30	44.60	41.28	0.92042	54.80	46.99	43.26	0.91510	57.30	49.43	45.23
0.92547	52.35	44.65	41.32	0.92032	54.85	47.04	43.30	0.91500	57.35	49.47	45.27
0.92537	52.40	44.70	41.36	0.92021	54.90	47.09	43.34	0.91489	57.40	49.52	45.31
0.92527	52.45	44.74	41.40	0.92011	54.95	47.14	43.38	0.91478	57.45	49.57	45.35
0.92516	52.50	44.79	41.44	0.92000	55.00	47.19	43.42	0.91467	57.50	49.62	45.39
0.92506	52.55	44.84	41.48	0.91990	55.05	47.24	43.46	0.91457	57.55	49.67	45.43
0.92496	52.60	44.89	41.52	0.91979	55.10	47.29	43.49	0.91446	57.60	49.72	45.47
0.92486	52.65	44.93	41.56	0.91969	55.15	47.33	43.53	0.91435	57.65	49.77	45.51
0.92476	52.70	44.98	41.60	0.91958	55.20	47.38	43.57	0.91424	57.70	49.82	45.55
0.92466	52.75	45.03	41.64	0.91948	55.25	47.43	43.61	0.91414	57.75	49.87	45.59
0.92456	52.80	45.08	41.68	0.91937	55.30	47.48	43.65	0.91403	57.80	49.92	45.63
0.92445	52.85	45.12	41.72	0.91927	55.35	47.53	43.69	0.91392	57.85	49.96	45.67
0.92434	52.90	45.17	41.76	0.91916	55.40	47.58	43.73	0.91381	57.90	50.01	45.70
0.92424	52.95	45.22	41.80	0.91906	55.45	47.62	43.77	0.91370	57.95	50.06	45.74
0.92414	53.00	45.27	41.83	0.91895	55.50	47.67	43.81	0.91359	58.00	50.11	45.78
0.92404	53.05	45.31	41.87	0.91885	55.55	47.72	43.85	0.91348	58.05	50.16	45.82
0.92394	53.10	45.36	41.91	0.91874	55.60	47.77	43.89	0.91337	58.10	50.21	45.86
0.92384	53.15	45.41	41.95	0.91864	55.65	47.82	43.93	0.91326	58.15	50.26	45.90
0.92373	53.20	45.46	41.99	0.91853	55.70	47.87	43.97	0.91315	58.20	50.31	45.94
0.92363	53.25	45.51	42.03	0.91842	55.75	47.91	44.01	0.91304	58.25	50.36	45.98
0.92353	53.30	45.56	42.07	0.91831	55.80	47.96	44.04	0.91293	58.30	50.41	46.02
0.92343	53.35	45.60	42.11	0.91821	55.85	48.01	44.08	0.91282	58.35	50.46	46.06
0.92332	53.40	45.65	42.15	0.91810	55.90	48.06	44.12	0.91271	58.40	50.51	46.10
0.92322	53.45	45.70	42.19	0.91800	55.95	48.11	44.16	0.91261	58.45	50.55	46.14
0.92312	53.50	45.75	42.23	0.91789	56.00	48.16	44.20	0.91250	58.50	50.60	46.17
0.92302	53.55	45.79	42.27	0.91779	56.05	48.20	44.24	0.91239	58.55	50.65	46.21
0.92291	53.60	45.84	42.31	0.91768	56.10	48.25	44.28	0.91228	58.60	50.70	46.25
0.92281	53.65	45.89	42.35	0.91758	56.15	48.30	44.32	0.91217	58.65	50.75	46.29
0.92271	53.70	45.94	42.39	0.91747	56.20	48.35	44.36	0.91206	58.70	50.80	46.33
0.92261	53.75	45.98	42.43	0.91736	56.25	48.40	44.40	0.91194	58.75	50.85	46.37
0.92250	53.80	46.03	42.47	0.91725	56.30	48.45	44.44	0.91183	58.80	50.90	46.41
0.92240	53.85	46.08	42.51	0.91715	56.35	48.50	44.48	0.91171	58.85	50.95	46.45
0.92230	53.90	46.13	42.55	0.91704	56.40	48.55	44.52	0.91160	58.90	51.00	46.49
0.92220	53.95	46.18	42.59	0.91694	56.45	48.59	44.56	0.91149	58.95	51.05	46.53
0.92209	54.00	46.23	42.62	0.91683	56.50	48.64	44.60	0.91138	59.00	51.10	46.57
0.92199	54.05	46.27	42.66	0.91672	56.55	48.69	44.64	0.91127	59.05	51.15	46.61
0.92188	54.10	46.32	42.70	0.91661	56.60	48.74	44.68	0.91116	59.10	51.20	46.65
0.92178	54.15	46.36	42.74	0.91650	56.65	48.79	44.72	0.91104	59.15	51.25	46.69
0.92167	54.20	46.41	42.78	0.91639	56.70	48.84	44.76	0.91093	59.20	51.30	46.73

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TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.91082	59.25	51.35	46.77	0.90520	61.75	53.85	48.74	0.89942	64.25	56.39	50.72
0.91071	59.30	51.40	46.81	0.90509	61.80	53.90	48.78	0.89930	64.30	56.44	50.76
0.91060	59.35	51.45	46.85	0.90497	61.85	53.95	48.82	0.89918	64.35	56.49	50.80
0.91049	59.40	51.50	46.89	0.90486	61.90	54.00	48.86	0.89907	64.40	56.54	50.83
0.91038	59.45	51.55	46.93	0.90474	61.95	54.05	48.90	0.89895	64.45	56.59	50.87
0.91027	59.50	51.60	46.97	0.90463	62.00	54.10	48.94	0.89884	64.50	56.64	50.91
0.91016	59.55	51.65	47.01	0.90451	62.05	54.15	48.98	0.89872	64.55	56.70	50.95
0.91005	59.60	51.70	47.05	0.90440	62.10	54.20	49.02	0.89861	64.60	56.75	50.99
0.90993	59.65	51.74	47.09	0.90428	62.15	54.25	49.06	0.89849	64.65	56.80	51.03
0.90982	59.70	51.79	47.12	0.90417	62.20	54.30	49.10	0.89837	64.70	56.85	51.07
0.90971	59.75	51.84	47.16	0.90406	62.25	54.35	49.14	0.89825	64.75	56.90	51.11
0.90960	59.80	51.89	47.20	0.90395	62.30	54.40	49.18	0.89814	64.80	56.95	51.15
0.90949	59.85	51.94	47.24	0.90383	62.35	54.45	49.22	0.89802	64.85	57.00	51.19
0.90938	59.90	51.99	47.28	0.90372	62.40	54.50	49.25	0.89791	64.90	57.05	51.23
0.90926	59.95	52.04	47.32	0.90360	62.45	54.55	49.29	0.89779	64.95	57.11	51.27
0.90915	60.00	52.09	47.36	0.90349	62.50	54.60	49.33	0.89767	65.00	57.16	51.31
0.90904	60.05	52.14	47.40	0.90337	62.55	54.66	49.37	0.89755	65.05	57.21	51.35
0.90893	60.10	52.19	47.44	0.90326	62.60	54.71	49.41	0.89744	65.10	57.26	51.39
0.90882	60.15	52.24	47.48	0.90314	62.65	54.76	49.45	0.89732	65.15	57.31	51.43
0.90871	60.20	52.29	47.52	0.90302	62.70	54.81	49.49	0.89720	65.20	57.36	51.47
0.90859	60.25	52.34	47.56	0.90290	62.75	54.86	49.53	0.89708	65.25	57.41	51.51
0.90848	60.30	52.39	47.60	0.90279	62.80	54.91	49.57	0.89696	65.30	57.46	51.55
0.90837	60.35	52.44	47.64	0.90267	62.85	54.96	49.61	0.89684	65.35	57.52	51.59
0.90826	60.40	52.49	47.68	0.90256	62.90	55.01	49.65	0.89672	65.40	57.57	51.63
0.90814	60.45	52.54	47.72	0.90244	62.95	55.06	49.69	0.89660	65.45	57.62	51.67
0.90803	60.50	52.59	47.76	0.90233	63.00	55.11	49.73	0.89649	65.50	57.67	51.71
0.90792	60.55	52.64	47.80	0.90221	63.05	55.16	49.77	0.89637	65.55	57.72	51.75
0.90781	60.60	52.69	47.84	0.90210	63.10	55.21	49.81	0.89626	65.60	57.77	51.78
0.90769	60.65	52.74	47.88	0.90198	63.15	55.26	49.85	0.89614	65.65	57.83	51.82
0.90758	60.70	52.79	47.91	0.90187	63.20	55.31	49.89	0.89602	65.70	57.88	51.86
0.90747	60.75	52.84	47.95	0.90175	63.25	55.37	49.93	0.89590	65.75	57.93	51.90
0.90736	60.80	52.89	47.99	0.90163	63.30	55.42	49.97	0.89578	65.80	57.98	51.94
0.90724	60.85	52.94	48.03	0.90151	63.35	55.47	50.01	0.89566	65.85	58.04	51.98
0.90713	60.90	52.99	48.07	0.90140	63.40	55.52	50.04	0.89554	65.90	58.09	52.02
0.90701	60.95	53.04	48.11	0.90128	63.45	55.57	50.08	0.89542	65.95	58.14	52.06
0.90690	61.00	53.09	48.15	0.90117	63.50	55.62	50.12	0.89531	66.00	58.19	52.10
0.90678	61.05	53.14	48.19	0.90105	63.55	55.67	50.16	0.89519	66.05	58.24	52.14
0.90667	61.10	53.19	48.23	0.90094	63.60	55.72	50.20	0.89507	66.10	58.29	52.18
0.90656	61.15	53.24	48.27	0.90082	63.65	55.77	50.24	0.89495	66.15	58.35	52.22
0.90645	61.20	53.29	48.31	0.90070	63.70	55.82	50.28	0.89483	66.20	58.40	52.26
0.90633	61.25	53.34	48.35	0.90059	63.75	55.88	50.32	0.89471	66.25	58.45	52.30
0.90622	61.30	53.39	48.39	0.90048	63.80	55.93	50.36	0.89459	66.30	58.50	52.33
0.90610	61.35	53.44	48.43	0.90036	63.85	55.98	50.40	0.89447	66.35	58.55	52.37
0.90599	61.40	53.49	48.47	0.90025	63.90	56.03	50.44	0.89435	66.40	58.60	52.41
0.90588	61.45	53.55	48.51	0.90013	63.95	56.08	50.48	0.89423	66.45	58.66	52.45
0.90577	61.50	53.60	48.55	0.90001	64.00	56.13	50.52	0.89411	66.50	58.71	52.49
0.90565	61.55	53.65	48.59	0.89989	64.05	56.18	50.56	0.89399	66.55	58.76	52.53
0.90554	61.60	53.70	48.62	0.89978	64.10	56.23	50.60	0.89387	66.60	58.81	52.57
0.90543	61.65	53.75	48.66	0.89966	64.15	56.29	50.64	0.89375	66.65	58.87	52.61
0.90532	61.70	53.80	48.70	0.89954	64.20	56.34	50.68	0.89363	66.70	58.92	52.65

TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.89351	66.75	58.97	52.69	0.88744	69.25	61.60	54.66	0.88120	71.75	64.27	56.64
0.89339	66.80	59.02	52.73	0.88732	69.30	61.65	54.70	0.88107	71.80	64.32	56.68
0.89327	66.85	59.07	52.77	0.88720	69.35	61.70	54.74	0.88094	71.85	64.38	56.72
0.89315	66.90	59.12	52.81	0.88707	69.40	61.75	54.78	0.88081	71.90	64.43	56.75
0.89303	66.95	59.18	52.85	0.88695	69.45	61.81	54.82	0.88069	71.95	64.49	56.79
0.89291	67.00	59.23	52.89	0.88682	69.50	61.86	54.86	0.88056	72.00	64.54	56.83
0.89279	67.05	59.28	52.93	0.88670	69.55	61.92	54.90	0.88044	72.05	64.60	56.87
0.89267	67.10	59.33	52.97	0.88658	69.60	61.97	54.94	0.88031	72.10	64.65	56.91
0.89255	67.15	59.39	53.01	0.88646	69.65	62.02	54.98	0.88018	72.15	64.71	56.95
0.89243	67.20	59.44	53.04	0.88633	69.70	62.07	55.02	0.88005	72.20	64.76	56.99
0.89231	67.25	59.49	53.08	0.88621	69.75	62.13	55.06	0.87993	72.25	64.82	57.03
0.89219	67.30	59.54	53.12	0.88608	69.80	62.18	55.10	0.87980	72.30	64.87	57.07
0.89207	67.35	59.60	53.16	0.88596	69.85	62.24	55.14	0.87967	72.35	64.93	57.11
0.89195	67.40	59.65	53.20	0.88583	69.90	62.29	55.18	0.87954	72.40	64.98	57.15
0.89183	67.45	59.70	53.24	0.88571	69.95	62.34	55.22	0.87942	72.45	65.03	57.19
0.89171	67.50	59.75	53.28	0.88558	70.00	62.39	55.25	0.87929	72.50	65.08	57.23
0.89159	67.55	59.81	53.32	0.88546	70.05	62.45	55.29	0.87916	72.55	65.14	57.27
0.89147	67.60	59.86	53.36	0.88533	70.10	62.50	55.33	0.87903	72.60	65.19	57.31
0.89135	67.65	59.91	53.40	0.88521	70.15	62.56	55.37	0.87891	72.65	65.25	57.35
0.89122	67.70	59.96	53.44	0.88508	70.20	62.61	55.41	0.87878	72.70	65.30	57.38
0.89110	67.75	60.02	53.48	0.88496	70.25	62.66	55.45	0.87865	72.75	65.36	57.42
0.89098	67.80	60.07	53.52	0.88484	70.30	62.71	55.49	0.87852	72.80	65.41	57.46
0.89086	67.85	60.12	53.56	0.88472	70.35	62.77	55.53	0.87839	72.85	65.47	57.50
0.89074	67.90	60.17	53.60	0.88459	70.40	62.82	55.57	0.87826	72.90	65.52	57.54
0.89062	67.95	60.23	53.64	0.88447	70.45	62.87	55.61	0.87813	72.95	65.58	57.58
0.89050	68.00	60.28	53.68	0.88434	70.50	62.92	55.65	0.87800	73.00	65.63	57.62
0.89038	68.05	60.33	53.72	0.88422	70.55	62.98	55.69	0.87788	73.05	65.69	57.66
0.89026	68.10	60.38	53.75	0.88409	70.60	63.03	55.73	0.87775	73.10	65.74	57.70
0.89014	68.15	60.44	53.79	0.88397	70.65	63.09	55.77	0.87762	73.15	65.80	57.74
0.89001	68.20	60.49	53.83	0.88384	70.70	63.14	55.81	0.87749	73.20	65.85	57.78
0.88989	68.25	60.54	53.87	0.88372	70.75	63.20	55.85	0.87737	73.25	65.91	57.82
0.88977	68.30	60.59	53.91	0.88359	70.80	63.25	55.89	0.87724	73.30	65.96	57.86
0.88965	68.35	60.65	53.95	0.88347	70.85	63.31	55.93	0.87711	73.35	66.02	57.90
0.88952	68.40	60.70	53.99	0.88334	70.90	63.36	55.97	0.87698	73.40	66.07	57.94
0.88940	68.45	60.75	54.03	0.88322	70.95	63.41	56.01	0.87685	73.45	66.13	57.98
0.88928	68.50	60.80	54.07	0.88309	71.00	63.46	56.04	0.87672	73.50	66.18	58.02
0.88916	68.55	60.86	54.11	0.88297	71.05	63.52	56.08	0.87659	73.55	66.23	58.06
0.88904	68.60	60.91	54.15	0.88284	71.10	63.57	56.12	0.87646	73.60	66.28	58.10
0.88892	68.65	60.96	54.19	0.88272	71.15	63.63	56.16	0.87633	73.65	66.34	58.14
0.88879	68.70	61.01	54.23	0.88259	71.20	63.68	56.20	0.87620	73.70	66.39	58.17
0.88867	68.75	61.07	54.27	0.88246	71.25	63.74	56.24	0.87607	73.75	66.45	58.21
0.88854	68.80	61.12	54.31	0.88233	71.30	63.79	56.28	0.87594	73.80	66.50	58.25
0.88842	68.85	61.17	54.35	0.88221	71.35	63.84	56.32	0.87581	73.85	66.56	58.29
0.88830	68.90	61.22	54.39	0.88208	71.40	63.89	56.36	0.87568	73.90	66.61	58.33
0.88818	68.95	61.28	54.43	0.88196	71.45	63.95	56.40	0.87555	73.95	66.67	58.37
0.88805	69.00	61.33	54.47	0.88183	71.50	64.00	56.44	0.87542	74.00	66.72	58.41
0.88793	69.05	61.39	54.51	0.88171	71.55	64.06	56.48	0.87529	74.05	66.78	58.45
0.88781	69.10	61.44	54.54	0.88158	71.60	64.11	56.52	0.87516	74.10	66.83	58.49
0.88769	69.15	61.49	54.58	0.88145	71.65	64.17	56.56	0.87504	74.15	66.89	58.53
0.88756	69.20	61.54	54.62	0.88132	71.70	64.22	56.60	0.87491	74.20	66.94	58.57

5 TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.87478	74.25	67.00	58.61	0.86818	76.75	69.78	60.58	0.86137	79.25	72.63	62.56
0.87465	74.30	67.05	58.65	0.86805	76.80	69.84	60.62	0.86124	79.30	72.68	62.60
0.87452	74.35	67.11	58.69	0.86791	76.85	69.90	60.66	0.86110	79.35	72.74	62.64
0.87439	74.40	67.16	58.73	0.86778	76.90	69.95	60.70	0.86096	79.40	72.80	62.67
0.87426	74.45	67.22	58.77	0.86764	76.95	70.01	60.74	0.86082	79.45	72.86	62.71
0.87413	74.50	67.27	58.81	0.86751	77.00	70.06	60.78	0.86069	79.50	72.91	62.75
0.87400	74.55	67.33	58.85	0.86737	77.05	70.12	60.82	0.86055	79.55	72.97	62.79
0.87387	74.60	67.38	58.89	0.86724	77.10	70.18	60.86	0.86041	79.60	73.03	62.83
0.87373	74.65	67.44	58.93	0.86710	77.15	70.24	60.90	0.86027	79.65	73.09	62.87
0.87360	74.70	67.49	58.97	0.86697	77.20	70.29	60.94	0.86013	79.70	73.14	62.91
0.87347	74.75	67.55	59.01	0.86684	77.25	70.35	60.98	0.85999	79.75	73.20	62.95
0.87334	74.80	67.61	59.04	0.86671	77.30	70.40	61.02	0.85984	79.80	73.26	62.99
0.87321	74.85	67.67	59.08	0.86657	77.35	70.46	61.06	0.85970	79.85	73.32	63.03
0.87308	74.90	67.72	59.12	0.86644	77.40	70.51	61.10	0.85956	79.90	73.37	63.07
0.87295	74.95	67.78	59.16	0.86630	77.45	70.57	61.14	0.85942	79.95	73.43	63.11
0.87282	75.00	67.83	59.20	0.86617	77.50	70.63	61.18	0.85928	80.00	73.49	63.15
0.87269	75.05	67.89	59.24	0.86603	77.55	70.69	61.22	0.85914	80.05	73.55	63.19
0.87256	75.10	67.94	59.28	0.86589	77.60	70.74	61.25	0.85901	80.10	73.60	63.23
0.87243	75.15	68.00	59.32	0.86575	77.65	70.80	61.29	0.85887	80.15	73.66	63.27
0.87230	75.20	68.05	59.36	0.86562	77.70	70.85	61.33	0.85873	80.20	73.72	63.30
0.87217	75.25	68.11	59.40	0.86548	77.75	70.91	61.37	0.85859	80.25	73.78	63.34
0.87204	75.30	68.16	59.44	0.86535	77.80	70.97	61.41	0.85846	80.30	73.83	63.38
0.87190	75.35	68.22	59.48	0.86521	77.85	71.03	61.45	0.85832	80.35	73.89	63.42
0.87177	75.40	68.27	59.52	0.86508	77.90	71.08	61.49	0.85818	80.40	73.95	63.46
0.87164	75.45	68.33	59.56	0.86494	77.95	71.14	61.53	0.85804	80.45	74.01	63.50
0.87151	75.50	68.38	59.60	0.86480	78.00	71.19	61.57	0.85789	80.50	74.06	63.54
0.87138	75.55	68.44	59.64	0.86466	78.05	71.25	61.61	0.85775	80.55	74.12	63.58
0.87125	75.60	68.49	59.67	0.86453	78.10	71.31	61.65	0.85761	80.60	74.18	63.62
0.87111	75.65	68.55	59.71	0.86439	78.15	71.37	61.69	0.85747	80.65	74.24	63.66
0.87098	75.70	68.60	59.75	0.86426	78.20	71.42	61.73	0.85733	80.70	74.30	63.70
0.87084	75.75	68.66	59.79	0.86412	78.25	71.48	61.77	0.85719	80.75	74.36	63.74
0.87071	75.80	68.72	59.83	0.86399	78.30	71.54	61.81	0.85705	80.80	74.42	63.78
0.87058	75.85	68.78	59.87	0.86385	78.35	71.60	61.85	0.85691	80.85	74.48	63.82
0.87045	75.90	68.83	59.91	0.86371	78.40	71.65	61.88	0.85677	80.90	74.53	63.86
0.87032	75.95	68.89	59.95	0.86357	78.45	71.71	61.92	0.85663	80.95	74.59	63.90
0.87019	76.00	68.94	59.99	0.86344	78.50	71.76	61.96	0.85648	81.00	74.65	63.94
0.87005	76.05	69.00	60.03	0.86330	78.55	71.82	62.00	0.85634	81.05	74.71	63.98
0.86992	76.10	69.05	60.07	0.86316	78.60	71.88	62.04	0.85620	81.10	74.77	64.02
0.86979	76.15	69.11	60.11	0.86302	78.65	71.94	62.08	0.85606	81.15	74.83	64.06
0.86966	76.20	69.16	60.15	0.86289	78.70	71.99	62.12	0.85592	81.20	74.88	64.09
0.86952	76.25	69.22	60.19	0.86275	78.75	72.05	62.16	0.85578	81.25	74.94	64.13
0.86939	76.30	69.27	60.23	0.86261	78.80	72.11	62.20	0.85564	81.30	75.00	64.17
0.86925	76.35	69.33	60.27	0.86247	78.85	72.17	62.24	0.85550	81.35	75.06	64.21
0.86912	76.40	69.39	60.31	0.86234	78.90	72.22	62.28	0.85536	81.40	75.12	64.25
0.86898	76.45	69.45	60.35	0.86220	78.95	72.28	62.32	0.85522	81.45	75.18	64.29
0.86885	76.50	69.50	60.39	0.86206	79.00	72.34	62.36	0.85507	81.50	75.24	64.33
0.86872	76.55	69.56	60.43	0.86192	79.05	72.40	62.40	0.85493	81.55	75.30	64.37
0.86859	76.60	69.61	60.47	0.86179	79.10	72.45	62.44	0.85478	81.60	75.35	64.41
0.86845	76.65	69.67	60.51	0.86165	79.15	72.51	62.48	0.85464	81.65	75.41	64.45
0.86832	76.70	69.72	60.54	0.86151	79.20	72.57	62.52	0.85450	81.70	75.47	64.49

TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.85436	81.75	75.53	64.53	0.84713	84.25	78.50	66.50	0.83957	86.75	81.56	68.48
0.85422	81.80	75.59	64.57	0.84698	84.30	78.56	66.54	0.83942	86.80	81.62	68.52
0.85408	81.85	75.65	64.61	0.84683	84.35	78.62	66.58	0.83927	86.85	81.68	68.56
0.85393	81.90	75.71	64.65	0.84668	84.40	78.68	66.62	0.83912	86.90	81.75	68.60
0.85379	81.95	75.77	64.69	0.84654	84.45	78.74	66.66	0.83896	86.95	81.81	68.64
0.85364	82.00	75.82	64.73	0.84639	84.50	78.80	66.70	0.83881	87.00	81.87	68.68
0.85350	82.05	75.88	64.77	0.84624	84.55	78.86	66.74	0.83865	87.05	81.93	68.72
0.85336	82.10	75.94	64.81	0.84609	84.60	78.93	66.78	0.83850	87.10	81.99	68.76
0.85322	82.15	76.00	64.85	0.84594	84.65	78.99	66.82	0.83834	87.15	82.05	68.80
0.85307	82.20	76.06	64.88	0.84579	84.70	79.05	66.86	0.83818	87.20	82.12	68.84
0.85293	82.25	76.12	64.92	0.84564	84.75	79.11	66.90	0.83802	87.25	82.18	68.88
0.85279	82.30	76.18	64.96	0.84549	84.80	79.17	66.94	0.83787	87.30	82.24	68.91
0.85265	82.35	76.24	65.00	0.84534	84.85	79.23	66.98	0.83771	87.35	82.30	68.95
0.85250	82.40	76.30	65.04	0.84519	84.90	79.29	67.02	0.83756	87.40	82.37	68.99
0.85236	82.45	76.36	65.08	0.84504	84.95	79.35	67.06	0.83740	87.45	82.43	69.03
0.85222	82.50	76.41	65.12	0.84489	85.00	79.41	67.09	0.83725	87.50	82.49	69.07
0.85207	82.55	76.47	65.16	0.84474	85.05	79.47	67.13	0.83709	87.55	82.55	69.11
0.85192	82.60	76.53	65.20	0.84459	85.10	79.53	67.17	0.83694	87.60	82.62	69.15
0.85178	82.65	76.59	65.24	0.84444	85.15	79.59	67.21	0.83678	87.65	82.68	69.19
0.85164	82.70	76.65	65.28	0.84429	85.20	79.65	67.25	0.83663	87.70	82.74	69.23
0.85150	82.75	76.71	65.32	0.84414	85.25	79.71	67.29	0.83647	87.75	82.80	69.27
0.85135	82.80	76.77	65.36	0.84399	85.30	79.78	67.33	0.83632	87.80	82.87	69.30
0.85121	82.85	76.83	65.40	0.84384	85.35	79.84	67.37	0.83616	87.85	82.93	69.34
0.85106	82.90	76.89	65.44	0.84369	85.40	79.90	67.41	0.83601	87.90	82.99	69.38
0.85092	82.95	76.95	65.48	0.84354	85.45	79.96	67.45	0.83585	87.95	83.05	69.42
0.85077	83.00	77.01	65.51	0.84339	85.50	80.02	67.49	0.83569	88.00	83.12	69.46
0.85063	83.05	77.07	65.55	0.84323	85.55	80.08	67.53	0.83553	88.05	83.18	69.50
0.85049	83.10	77.13	65.59	0.84308	85.60	80.14	67.57	0.83537	88.10	83.25	69.54
0.85035	83.15	77.19	65.63	0.84293	85.65	80.20	67.61	0.83521	88.15	83.31	69.58
0.85020	83.20	77.24	65.67	0.84278	85.70	80.27	67.65	0.83505	88.20	83.37	69.62
0.85006	83.25	77.30	65.71	0.84263	85.75	80.33	67.69	0.83489	88.25	83.43	69.66
0.84991	83.30	77.36	65.75	0.84248	85.80	80.39	67.73	0.83473	88.30	83.50	69.70
0.84977	83.35	77.42	65.79	0.84233	85.85	80.45	67.77	0.83457	88.35	83.56	69.74
0.84962	83.40	77.48	65.83	0.84218	85.90	80.51	67.80	0.83442	88.40	83.63	69.78
0.84948	83.45	77.54	65.87	0.84203	85.95	80.57	67.84	0.83426	88.45	83.69	69.82
0.84933	83.50	77.60	65.91	0.84188	86.00	80.63	67.88	0.83410	88.50	83.75	69.86
0.84918	83.55	77.66	65.95	0.84172	86.05	80.69	67.92	0.83394	88.55	83.81	69.90
0.84903	83.60	77.72	65.99	0.84157	86.10	80.76	67.96	0.83379	88.60	83.88	69.94
0.84889	83.65	77.78	66.03	0.84141	86.15	80.82	68.00	0.83363	88.65	83.94	69.98
0.84874	83.70	77.84	66.07	0.84126	86.20	80.88	68.04	0.83347	88.70	84.00	70.01
0.84859	83.75	77.90	66.11	0.84110	86.25	80.94	68.08	0.83331	88.75	84.06	70.05
0.84844	83.80	77.96	66.15	0.84095	86.30	81.00	68.12	0.83315	88.80	84.13	70.09
0.84830	83.85	78.02	66.19	0.84080	86.35	81.06	68.16	0.83299	88.85	84.19	70.13
0.84815	83.90	78.08	66.23	0.84065	86.40	81.13	68.20	0.83283	88.90	84.26	70.17
0.84801	83.95	78.14	66.27	0.84049	86.45	81.19	68.24	0.83267	88.95	84.32	70.21
0.84786	84.00	78.20	66.30	0.84034	86.50	81.25	68.28	0.83251	89.00	84.39	70.25
0.84772	84.05	78.26	66.34	0.84018	86.55	81.31	68.32	0.83235	89.05	84.45	70.29
0.84757	84.10	78.32	66.38	0.84003	86.60	81.37	68.36	0.83219	89.10	84.51	70.33
0.84742	84.15	78.38	66.42	0.83987	86.65	81.43	68.40	0.83203	89.15	84.57	70.37
0.84727	84.20	78.44	66.46	0.83972	86.70	81.50	68.44	0.83186	89.20	84.64	70.41

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TABLE 16.—ALCOHOL TABLE.—Continued.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.83170	89.25	84.70	70.45	0.82332	91.75	87.96	72.42	0.81432	94.25	91.86	74.40
0.83154	89.30	84.77	70.49	0.82315	91.80	88.03	72.46	0.81413	94.30	91.43	74.44
0.83138	89.35	84.83	70.53	0.82298	91.85	88.09	72.50	0.81394	94.35	91.50	74.48
0.83121	89.40	84.90	70.57	0.82281	91.90	88.16	72.54	0.81375	94.40	91.57	74.52
0.83105	89.45	84.96	70.61	0.82263	91.95	88.22	72.58	0.81356	94.45	91.64	74.56
0.83089	89.50	85.03	70.65	0.82246	92.00	88.29	72.62	0.81337	94.50	91.71	74.59
0.83073	89.55	85.09	70.69	0.82229	92.05	88.36	72.66	0.81318	94.55	91.78	74.63
0.83056	89.60	85.15	70.72	0.82212	92.10	88.43	72.70	0.81299	94.60	91.85	74.67
0.83040	89.65	85.21	70.76	0.82194	92.15	88.49	72.74	0.81280	94.65	91.92	74.71
0.83024	89.70	85.28	70.80	0.82177	92.20	88.56	72.78	0.81260	94.70	91.99	74.75
0.83008	89.75	85.34	70.84	0.82159	92.25	88.63	72.82	0.81241	94.75	92.06	74.79
0.82991	89.80	85.41	70.88	0.82141	92.30	88.70	72.86	0.81222	94.80	92.13	74.83
0.82975	89.85	85.47	70.92	0.82123	92.35	88.76	72.90	0.81202	94.85	92.20	74.87
0.82958	89.90	85.54	70.96	0.82106	92.40	88.83	72.94	0.81183	94.90	92.27	74.91
0.82942	89.95	85.60	71.00	0.82088	92.45	88.89	72.98	0.81163	94.95	92.34	74.95
0.82925	90.00	85.67	71.04	0.82071	92.50	88.96	73.02	0.81144	95.00	92.41	74.99
0.82909	90.05	85.73	71.08	0.82053	92.55	89.03	73.06	0.81124	95.05	92.48	75.03
0.82892	90.10	85.80	71.12	0.82035	92.60	89.10	73.09	0.81105	95.10	92.55	75.07
0.82876	90.15	85.86	71.16	0.82017	92.65	89.16	73.13	0.81086	95.15	92.63	75.11
0.82859	90.20	85.93	71.20	0.82000	92.70	89.23	73.17	0.81067	95.20	92.70	75.15
0.82843	90.25	85.99	71.24	0.81982	92.75	89.30	73.21	0.81047	95.25	92.77	75.19
0.82826	90.30	86.06	71.28	0.81964	92.80	89.37	73.25	0.81028	95.30	92.84	75.23
0.82810	90.35	86.12	71.32	0.81946	92.85	89.43	73.29	0.81008	95.35	92.91	75.27
0.82793	90.40	86.19	71.36	0.81929	92.90	89.50	73.33	0.80988	95.40	92.98	75.30
0.82776	90.45	86.25	71.40	0.81911	92.95	89.57	73.37	0.80968	95.45	93.05	75.34
0.82759	90.50	86.32	71.44	0.81893	93.00	89.64	73.41	0.80949	95.50	93.12	75.38
0.82742	90.55	86.38	71.48	0.81875	93.05	89.71	73.45	0.80929	95.55	93.20	75.42
0.82725	90.60	86.45	71.52	0.81856	93.10	89.78	73.49	0.80909	95.60	93.27	75.46
0.82708	90.65	86.51	71.56	0.81838	93.15	89.84	73.53	0.80889	95.65	93.34	75.50
0.82691	90.70	86.58	71.59	0.81821	93.20	89.91	73.57	0.80869	95.70	93.41	75.54
0.82674	90.75	86.64	71.63	0.81803	93.25	89.98	73.61	0.80849	95.75	93.48	75.58
0.82657	90.80	86.71	71.67	0.81784	93.30	90.05	73.65	0.80829	95.80	93.55	75.62
0.82640	90.85	86.77	71.71	0.81766	93.35	90.12	73.69	0.80809	95.85	93.63	75.66
0.82624	90.90	86.84	71.75	0.81748	93.40	90.19	73.72	0.80789	95.90	93.70	75.70
0.82607	90.95	86.90	71.79	0.81730	93.45	90.25	73.76	0.80769	95.95	93.77	75.74
0.82590	91.00	86.97	71.83	0.81711	93.50	90.32	73.80	0.80749	96.00	93.84	75.78
0.82573	91.05	87.03	71.87	0.81693	93.55	90.39	73.84	0.80729	96.05	93.92	75.82
0.82556	91.10	87.10	71.91	0.81675	93.60	90.46	73.88	0.80709	96.10	93.99	75.86
0.82539	91.15	87.17	71.95	0.81657	93.65	90.53	73.92	0.80689	96.15	94.06	75.90
0.82522	91.20	87.24	71.99	0.81638	93.70	90.60	73.96	0.80668	96.20	94.13	75.94
0.82505	91.25	87.30	72.03	0.81620	93.75	90.67	74.00	0.80648	96.25	94.21	75.98
0.82488	91.30	87.37	72.07	0.81601	93.80	90.74	74.04	0.80627	96.30	94.28	76.01
0.82470	91.35	87.43	72.11	0.81582	93.85	90.80	74.08	0.80607	96.35	94.35	76.05
0.82453	91.40	87.50	72.15	0.81563	93.90	90.87	74.12	0.80586	96.40	94.42	76.09
0.82436	91.45	87.56	72.19	0.81545	93.95	90.94	74.16	0.80566	96.45	94.50	76.13
0.82419	91.50	87.63	72.23	0.81526	94.00	91.01	74.20	0.80545	96.50	94.57	76.17
0.82401	91.55	87.69	72.27	0.81507	94.05	91.08	74.24	0.80525	96.55	94.65	76.21
0.82384	91.60	87.76	72.30	0.81488	94.10	91.15	74.28	0.80504	96.60	94.72	76.25
0.82367	91.65	87.83	72.34	0.81469	94.15	91.22	74.32	0.80483	96.65	94.79	76.29
0.82350	91.70	87.90	72.38	0.81450	94.20	91.29	74.36	0.80462	96.70	94.86	76.33

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TABLE 16.—ALCOHOL TABLE.—Concluded.

SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL			SPECIFIC GRAVITY 20° C. 4°	ALCOHOL		
	Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.		Per cent by volume at 20° C.	Per cent by weight	Grams per 100 cc.
0.80442	96.75	94.94	76.37	0.79900	98.00	96.82	77.36	0.79311	99.25	98.78	78.34
0.80421	96.80	95.01	76.41	0.79878	98.05	96.90	77.40	0.79286	99.30	98.86	78.38
0.80400	96.85	95.09	76.45	0.79855	98.10	96.97	77.43	0.79262	99.35	98.94	78.42
0.80379	96.90	95.16	76.49	0.79832	98.15	97.05	77.47	0.79237	99.40	99.02	78.46
0.80358	96.95	95.24	76.53	0.79809	98.20	97.12	77.51	0.79213	99.45	99.10	78.50
0.80337	97.00	95.31	76.57	0.79786	98.25	97.20	77.55	0.79188	99.50	99.18	78.54
0.80315	97.05	95.39	76.61	0.79763	98.30	97.28	77.59	0.79163	99.55	99.26	78.58
0.80294	97.10	95.46	76.65	0.79740	98.35	97.36	77.63	0.79138	99.60	99.34	78.62
0.80273	97.15	95.53	76.69	0.79717	98.40	97.43	77.67	0.79113	99.65	99.42	78.66
0.80252	97.20	95.60	76.72	0.79695	98.45	97.51	77.71	0.79088	99.70	99.51	78.70
0.80230	97.25	95.68	76.76	0.79672	98.50	97.59	77.75	0.79062	99.75	99.59	78.74
0.80208	97.30	95.75	76.80	0.79648	98.55	97.67	77.79	0.79037	99.80	99.67	78.78
0.80186	97.35	95.83	76.84	0.79625	98.60	97.75	77.83	0.79011	99.85	99.75	78.82
0.80164	97.40	95.91	76.88	0.79601	98.65	97.83	77.87	0.78986	99.90	99.83	78.86
0.80143	97.45	95.98	76.92	0.79577	98.70	97.90	77.91	0.78960	99.95	99.91	78.90
0.80122	97.50	96.05	76.96	0.79553	98.75	97.98	77.95	0.78934	100.00	100.00	78.93
0.80100	97.55	96.13	77.00	0.79529	98.80	98.06	77.99				
0.80078	97.60	96.21	77.04	0.79505	98.85	98.14	78.03				
0.80056	97.65	96.29	77.08	0.79481	98.90	98.22	78.07				
0.80034	97.70	96.36	77.12	0.79457	98.95	98.30	78.11				
0.80012	97.75	96.44	77.16	0.79432	99.00	98.38	78.14				
0.79990	97.80	96.52	77.20	0.79408	99.05	98.46	78.18				
0.79968	97.85	96.60	77.24	0.79384	99.10	98.54	78.22				
0.79945	97.90	96.68	77.28	0.79360	99.15	98.62	78.26				
0.79923	97.95	96.75	77.32	0.79335	99.20	98.70	78.30				

TABLE 17.—ALCOHOL
For calculating the percentages of alcohol in mixtures of ethyl alcohol and

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
13.2
13.3
13.4
13.5
13.6
13.7
13.8
13.9
14.0
14.1
14.2	0.04
14.3	0.13
14.4	0.31
14.5	0.08	0.06	0.29
14.6	0.16	0.13	0.38
14.7	0.05	0.04	0.25	0.20	0.46
14.8	0.14	0.11	0.34	0.27	0.55
14.9	0.01	0.01	0.23	0.18	0.43	0.34	0.64
15.0	0.00	0.00	0.10	0.08	0.31	0.24	0.52	0.41	0.73
15.1	0.09	0.07	0.19	0.15	0.39	0.31	0.60	0.48	0.82
15.2	0.17	0.13	0.27	0.21	0.48	0.38	0.69	0.55	0.91
15.3	0.25	0.20	0.35	0.28	0.57	0.45	0.77	0.61	0.99
15.4	0.34	0.27	0.44	0.35	0.65	0.51	0.85	0.68	1.07
15.5	0.43	0.34	0.53	0.42	0.73	0.58	0.94	0.75	1.16
15.6	0.51	0.40	0.60	0.48	0.82	0.65	1.03	0.82	1.24
15.7	0.59	0.47	0.69	0.55	0.91	0.72	1.12	0.89	1.32
15.8	0.68	0.54	0.78	0.62	0.99	0.79	1.21	0.96	1.40
15.9	0.76	0.60	0.85	0.68	1.08	0.86	1.28	1.02	1.47
16.0	0.84	0.67	0.94	0.75	1.17	0.93	1.36	1.08	1.55
16.1	0.93	0.74	1.03	0.82	1.24	0.99	1.44	1.14	1.62
16.2	1.02	0.81	1.12	0.89	1.32	1.05	1.51	1.20	1.70
16.3	1.10	0.87	1.19	0.95	1.40	1.11	1.59	1.26	1.77
16.4	1.18	0.94	1.29	1.02	1.47	1.17	1.66	1.32	1.85
16.5	1.26	1.00	1.36	1.08	1.55	1.23	1.74	1.38	1.92
16.6	1.34	1.06	1.43	1.13	1.62	1.29	1.81	1.44	2.00
16.7	1.41	1.12	1.50	1.19	1.70	1.35	1.89	1.50	2.07
16.8	1.49	1.18	1.57	1.25	1.77	1.41	1.96	1.56	2.15
16.9	1.56	1.24	1.65	1.31	1.85	1.47	2.04	1.62	2.22
17.0	1.63	1.30	1.72	1.37	1.92	1.53	2.11	1.68	2.30
17.1	1.70	1.35	1.80	1.43	1.99	1.58	2.19	1.74	2.37
17.2	1.77	1.41	1.87	1.49	2.06	1.64	2.26	1.80	2.45
17.3	1.85	1.47	1.94	1.54	2.14	1.70	2.34	1.86	2.52
17.4	1.92	1.53	2.01	1.60	2.21	1.76	2.41	1.92	2.59
17.5	2.00	1.59	2.09	1.66	2.29	1.82	2.49	1.98	2.66
17.6	2.07	1.65	2.16	1.72	2.36	1.88	2.56	2.04	2.74
17.7	2.14	1.70	2.24	1.78	2.44	1.94	2.62	2.09	2.81
17.8	2.21	1.76	2.31	1.84	2.51	2.00	2.70	2.15	2.89
17.9	2.29	1.82	2.38	1.89	2.59	2.06	2.77	2.21	2.96

* Calculated and arranged by B. H. St. John from the data of Doroshevskii and Dvorzhanchik.²

TABLE.

water from their Zeiss immersion refractometer readings at 17.5°-25°C.*

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21° C.	22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
.....	0 00	0 00	13.2
.....	0 09	0 07	13.3
.....	0 18	0 14	13.4
.....	0 05	0 04	0 26	0 21	13.5
.....	0 14	0 11	0 35	0 28	13.6
.....	0 01	0 01	0 23	0 18	0 44	0 35	13.7
.....	0 10	0 08	0 31	0 25	0 53	0 42	13.8
.....	0 19	0 15	0 40	0 32	0 62	0 49	13.9
.....	0 08	0 06	0 28	0 22	0 49	0 39	0 70	0 56	14.0
.....	0 16	0 13	0 36	0 29	0 58	0 46	0 79	0 63	14.1
0 03	0 24	0 19	0 45	0 36	0 67	0 53	0 88	0 70	14.2
0 10	0 33	0 26	0 54	0 43	0 75	0 60	0 97	0 77	14.3
0 17	0 41	0 33	0 63	0 50	0 84	0 67	1 06	0 85	14.4
0 23	0 50	0 40	0 72	0 57	0 93	0 74	1 15	0 92	14.5
0 30	0 59	0 47	0 80	0 64	1 02	0 81	1 24	0 99	14.6
0 37	0 68	0 54	0 89	0 71	1 11	0 88	1 32	1 05	14.7
0 44	0 77	0 61	0 98	0 78	1 19	0 95	1 40	1 11	14.8
0 51	0 85	0 68	1 07	0 85	1 28	1 02	1 47	1 17	14.9
0 58	0 94	0 75	1 16	0 92	1 36	1 08	1 55	1 23	15.0
0 65	1 03	0 82	1 24	0 99	1 44	1 14	1 63	1 29	15.1
0 72	1 12	0 89	1 32	1 05	1 51	1 20	1 71	1 36	15.2
0 79	1 21	0 96	1 40	1 11	1 59	1 26	1 79	1 42	15.3
0 85	1 29	1 02	1 47	1 17	1 66	1 32	1 86	1 48	15.4
0 92	1 36	1 08	1 55	1 23	1 74	1 38	1 94	1 54	15.5
0 99	1 44	1 15	1 62	1 29	1 82	1 44	2 01	1 60	15.6
1 05	1 52	1 21	1 70	1 35	1 90	1 51	2 09	1 66	15.7
1 11	1 60	1 27	1 77	1 41	1 97	1 57	2 17	1 72	15.8
1 17	1 67	1 33	1 85	1 47	2 05	1 63	2 25	1 79	15.9
1 23	1 75	1 39	1 92	1 53	2 12	1 69	2 33	1 85	16.0
1 29	1 82	1 45	2 00	1 59	2 20	1 75	2 40	1 91	16.1
1 35	1 90	1 51	2 08	1 65	2 27	1 81	2 48	1 97	16.2
1 41	1 97	1 57	2 16	1 72	2 35	1 87	2 55	2 03	16.3
1 47	2 05	1 63	2 24	1 78	2 43	1 93	2 62	2 09	16.4
1 53	2 12	1 69	2 31	1 84	2 50	1 99	2 70	2 15	16.5
1 59	2 20	1 75	2 39	1 90	2 57	2 05	2 77	2 21	16.6
1 65	2 27	1 81	2 46	1 96	2 65	2 11	2 85	2 27	16.7
1 71	2 35	1 87	2 53	2 02	2 72	2 17	2 92	2 33	16.8
1 77	2 43	1 93	2 61	2 08	2 80	2 23	2 99	2 38	16.9
1 83	2 50	1 99	2 69	2 14	2 87	2 29	3 06	2 44	17.0
1 89	2 57	2 05	2 76	2 20	2 95	2 35	3 14	2 50	17.1
1 95	2 65	2 11	2 82	2 25	3 02	2 41	3 21	2 56	17.2
2 01	2 72	2 17	2 90	2 31	3 10	2 47	3 29	2 62	17.3
2 07	2 79	2 23	2 97	2 37	3 17	2 53	3 36	2 68	17.4
2 12	2 86	2 28	3 04	2 43	3 25	2 59	3 43	2 74	17.5
2 18	2 94	2 34	3 12	2 49	3 32	2 65	3 51	2 80	17.6
2 24	3 01	2 40	3 20	2 55	3 39	2 70	3 58	2 86	17.7
2 30	3 09	2 46	3 27	2 61	3 46	2 76	3 66	2 92	17.8
2 36	3 16	2 52	3 35	2 67	3 53	2 82	3 73	2 98	17.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
18.0	2.36	1.88	2.45	1.95	2.66	2.12	2.85	2.27	3.04
18.1	2.43	1.94	2.52	2.01	2.74	2.18	2.92	2.33	3.11
18.2	2.50	2.00	2.60	2.07	2.81	2.24	3.00	2.39	3.19
18.3	2.57	2.05	2.67	2.13	2.89	2.30	3.07	2.45	3.26
18.4	2.65	2.11	2.75	2.19	2.96	2.36	3.15	2.51	3.34
18.5	2.72	2.17	2.82	2.25	3.03	2.41	3.22	2.57	3.41
18.6	2.80	2.23	2.90	2.31	3.10	2.47	3.30	2.63	3.48
18.7	2.87	2.29	2.97	2.37	3.17	2.53	3.37	2.69	3.55
18.8	2.95	2.35	3.05	2.43	3.25	2.59	3.45	2.75	3.63
18.9	3.02	2.41	3.12	2.49	3.32	2.65	3.52	2.81	3.70
19.0	3.10	2.47	3.19	2.54	3.40	2.71	3.59	2.86	3.77
19.1	3.17	2.53	3.26	2.60	3.47	2.77	3.66	2.92	3.85
19.2	3.25	2.59	3.34	2.66	3.55	2.83	3.73	2.98	3.92
19.3	3.32	2.65	3.41	2.72	3.62	2.89	3.81	3.04	4.00
19.4	3.39	2.70	3.48	2.78	3.70	2.95	3.88	3.10	4.07
19.5	3.46	2.76	3.56	2.84	3.77	3.01	3.96	3.16	4.14
19.6	3.53	2.82	3.63	2.90	3.84	3.06	4.03	3.22	4.22
19.7	3.61	2.88	3.71	2.96	3.91	3.12	4.10	3.27	4.29
19.8	3.68	2.94	3.78	3.02	3.98	3.18	4.17	3.33	4.37
19.9	3.76	3.00	3.86	3.08	4.06	3.24	4.25	3.39	4.44
20.0	3.83	3.06	3.93	3.13	4.13	3.30	4.32	3.45	4.52
20.1	3.90	3.12	4.00	3.19	4.20	3.35	4.39	3.51	4.59
20.2	3.97	3.17	4.07	3.25	4.27	3.41	4.47	3.57	4.66
20.3	4.04	3.23	4.14	3.31	4.34	3.47	4.54	3.63	4.74
20.4	4.12	3.29	4.22	3.37	4.42	3.53	4.61	3.68	4.82
20.5	4.19	3.35	4.29	3.43	4.49	3.59	4.68	3.74	4.89
20.6	4.26	3.41	4.36	3.49	4.56	3.65	4.75	3.80	4.96
20.7	4.33	3.46	4.43	3.54	4.63	3.70	4.83	3.86	5.03
20.8	4.41	3.52	4.51	3.60	4.70	3.76	4.90	3.92	5.10
20.9	4.48	3.58	4.58	3.66	4.78	3.82	4.97	3.98	5.17
21.0	4.56	3.64	4.65	3.72	4.85	3.88	5.04	4.03	5.24
21.1	4.63	3.70	4.73	3.78	4.92	3.94	5.11	4.09	5.31
21.2	4.70	3.76	4.80	3.84	4.99	3.99	5.19	4.15	5.39
21.3	4.77	3.81	4.87	3.89	5.06	4.05	5.26	4.21	5.46
21.4	4.84	3.87	4.94	3.95	5.14	4.11	5.33	4.26	5.53
21.5	4.92	3.93	5.01	4.01	5.21	4.17	5.40	4.32	5.60
21.6	4.99	3.99	5.09	4.07	5.28	4.22	5.47	4.38	5.67
21.7	5.06	4.05	5.16	4.13	5.35	4.28	5.54	4.44	5.75
21.8	5.13	4.10	5.23	4.18	5.43	4.34	5.61	4.49	5.82
21.9	5.20	4.16	5.30	4.24	5.50	4.40	5.69	4.55	5.89
22.0	5.27	4.22	5.37	4.30	5.57	4.45	5.76	4.61	5.96
22.1	5.34	4.27	5.44	4.35	5.64	4.51	5.83	4.67	6.03
22.2	5.41	4.33	5.51	4.41	5.71	4.57	5.90	4.72	6.11
22.3	5.49	4.39	5.58	4.47	5.78	4.63	5.97	4.78	6.18
22.4	5.56	4.45	5.65	4.53	5.85	4.68	6.05	4.84	6.25
22.5	5.63	4.51	5.72	4.58	5.92	4.74	6.12	4.90	6.32
22.6	5.70	4.56	5.80	4.64	6.00	4.80	6.19	4.95	6.39
22.7	5.77	4.62	5.87	4.70	6.07	4.86	6.26	5.01	6.46
22.8	5.85	4.68	5.94	4.75	6.14	4.91	6.33	5.07	6.53
22.9	5.92	4.74	6.01	4.81	6.21	4.97	6.40	5.13	6.60

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	
2.42	3.23	2.58	3.42	2.73	3.61	2.88	3.81	3.04	4.18	18.0
2.48	3.30	2.63	3.50	2.79	3.68	2.94	3.88	3.10	4.26	18.1
2.54	3.37	2.69	3.57	2.85	3.76	3.00	3.96	3.16	4.33	18.2
2.60	3.45	2.75	3.64	2.91	3.83	3.06	4.03	3.22	4.41	18.3
2.66	3.52	2.81	3.71	2.96	3.91	3.12	4.11	3.28	4.48	18.4
2.72	3.59	2.87	3.78	3.02	3.98	3.18	4.18	3.34	4.56	18.5
2.78	3.66	2.92	3.86	3.08	4.06	3.24	4.26	3.40	4.63	18.6
2.83	3.73	2.98	3.93	3.14	4.13	3.30	4.33	3.46	4.70	18.7
2.89	3.81	3.04	4.01	3.20	4.21	3.36	4.41	3.52	4.78	18.8
2.95	3.88	3.10	4.08	3.26	4.28	3.42	4.48	3.58	4.85	18.9
3.01	3.96	3.16	4.16	3.32	4.36	3.48	4.56	3.64	4.93	19.0
3.07	4.03	3.22	4.23	3.38	4.43	3.54	4.63	3.70	5.00	19.1
3.13	4.11	3.28	4.31	3.44	4.51	3.60	4.70	3.76	5.08	19.2
3.19	4.18	3.34	4.38	3.50	4.58	3.66	4.78	3.82	5.15	19.3
3.25	4.26	3.40	4.46	3.56	4.65	3.72	4.85	3.88	5.22	19.4
3.31	4.33	3.46	4.53	3.62	4.73	3.78	4.93	3.94	5.30	19.5
3.37	4.41	3.52	4.61	3.68	4.80	3.84	5.00	4.00	5.38	19.6
3.43	4.48	3.58	4.68	3.74	4.88	3.90	5.08	4.06	5.46	19.7
3.49	4.56	3.64	4.75	3.80	4.95	3.96	5.15	4.12	5.54	19.8
3.55	4.63	3.70	4.83	3.86	5.03	4.02	5.22	4.17	5.62	19.9
3.61	4.72	3.77	4.90	3.92	5.10	4.08	5.29	4.23	5.70	20.0
3.67	4.79	3.83	4.98	3.98	5.17	4.13	5.36	4.29	5.78	20.1
3.73	4.87	3.89	5.05	4.04	5.24	4.19	5.44	4.35	5.86	20.2
3.79	4.94	3.95	5.13	4.10	5.31	4.25	5.51	4.41	5.94	20.3
3.85	5.01	4.01	5.20	4.16	5.38	4.31	5.58	4.47	6.02	20.4
3.91	5.08	4.06	5.27	4.21	5.45	4.37	5.65	4.52	6.10	20.5
3.97	5.15	4.12	5.34	4.27	5.52	4.42	5.72	4.58	6.18	20.6
4.02	5.22	4.18	5.41	4.33	5.60	4.48	5.80	4.64	6.26	20.7
4.08	5.29	4.24	5.48	4.39	5.67	4.54	5.87	4.70	6.34	20.8
4.14	5.36	4.29	5.55	4.45	5.75	4.60	5.95	4.76	6.42	20.9
4.20	5.44	4.35	5.62	4.50	5.82	4.66	6.02	4.81	6.50	21.0
4.25	5.51	4.41	5.70	4.56	5.89	4.72	6.09	4.87	6.58	21.1
4.31	5.58	4.47	5.77	4.62	5.96	4.77	6.16	4.93	6.66	21.2
4.37	5.65	4.52	5.84	4.68	6.03	4.83	6.23	4.99	6.74	21.3
4.43	5.72	4.58	5.91	4.73	6.11	4.89	6.30	5.05	6.82	21.4
4.48	5.80	4.64	5.98	4.79	6.18	4.95	6.37	5.10	6.90	21.5
4.54	5.87	4.70	6.06	4.85	6.25	5.01	6.44	5.16	6.98	21.6
4.60	5.94	4.75	6.13	4.91	6.32	5.06	6.52	5.22	7.06	21.7
4.66	6.01	4.81	6.20	4.97	6.39	5.12	6.59	5.28	7.14	21.8
4.71	6.08	4.87	6.27	5.02	6.47	5.18	6.66	5.34	7.22	21.9
4.77	6.15	4.93	6.34	5.08	6.54	5.24	6.73	5.39	7.30	22.0
4.83	6.22	4.98	6.42	5.14	6.61	5.29	6.80	5.45	7.38	22.1
4.89	6.29	5.04	6.49	5.20	6.68	5.35	6.87	5.51	7.46	22.2
4.95	6.36	5.10	6.56	5.25	6.75	5.41	6.94	5.57	7.54	22.3
5.00	6.43	5.15	6.63	5.31	6.82	5.47	7.01	5.62	7.62	22.4
5.06	6.50	5.21	6.70	5.37	6.89	5.52	7.08	5.68	7.70	22.5
5.11	6.57	5.27	6.77	5.43	6.96	5.58	7.16	5.74	7.78	22.6
5.17	6.64	5.33	6.84	5.48	7.03	5.64	7.23	5.80	7.86	22.7
5.23	6.71	5.38	6.91	5.54	7.10	5.70	7.31	5.86	7.94	22.8
5.29	6.78	5.44	6.99	5.60	7.17	5.75	7.38	5.91	8.02	22.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
23.0	5.99	4.79	6.08	4.87	6.28	5.03	6.47	5.18	6.67
23.1	6.06	4.85	6.15	4.93	6.35	5.09	6.54	5.24	6.74
23.2	6.13	4.91	6.22	4.98	6.42	5.14	6.61	5.30	6.81
23.3	6.20	4.97	6.29	5.04	6.49	5.20	6.68	5.36	6.88
23.4	6.27	5.02	6.36	5.10	6.56	5.26	6.75	5.41	6.95
23.5	6.34	5.08	6.43	5.15	6.63	5.31	6.83	5.47	7.02
23.6	6.41	5.14	6.50	5.21	6.70	5.37	6.90	5.53	7.09
23.7	6.48	5.19	6.57	5.27	6.78	5.43	6.97	5.59	7.16
23.8	6.55	5.25	6.64	5.32	6.85	5.49	7.04	5.64	7.23
23.9	6.62	5.30	6.71	5.38	6.92	5.54	7.11	5.70	7.31
24.0	6.69	5.36	6.78	5.44	6.99	5.60	7.18	5.76	7.38
24.1	6.76	5.42	6.85	5.49	7.06	5.66	7.25	5.82	7.45
24.2	6.83	5.47	6.92	5.55	7.13	5.71	7.32	5.87	7.52
24.3	6.90	5.53	6.99	5.61	7.20	5.77	7.39	5.93	7.59
24.4	6.97	5.59	7.06	5.66	7.27	5.83	7.46	5.99	7.66
24.5	7.04	5.64	7.13	5.72	7.34	5.89	7.53	6.04	7.73
24.6	7.11	5.70	7.20	5.78	7.41	5.94	7.60	6.10	7.80
24.7	7.18	5.76	7.27	5.83	7.48	6.00	7.67	6.15	7.86
24.8	7.25	5.81	7.35	5.89	7.55	6.06	7.74	6.21	7.93
24.9	7.32	5.87	7.42	5.95	7.62	6.11	7.81	6.26	8.00
25.0	7.39	5.93	7.49	6.01	7.68	6.16	7.88	6.32	8.06
25.1	7.46	5.98	7.56	6.06	7.75	6.22	7.94	6.37	8.13
25.2	7.53	6.04	7.63	6.12	7.82	6.27	8.01	6.43	8.20
25.3	7.59	6.09	7.69	6.17	7.89	6.33	8.07	6.48	8.27
25.4	7.66	6.15	7.76	6.23	7.95	6.38	8.14	6.54	8.34
25.5	7.73	6.20	7.83	6.28	8.02	6.44	8.21	6.59	8.41
25.6	7.80	6.26	7.90	6.34	8.09	6.49	8.28	6.65	8.48
25.7	7.87	6.31	7.96	6.39	8.16	6.55	8.35	6.70	8.55
25.8	7.94	6.37	8.03	6.44	8.22	6.60	8.42	6.76	8.62
25.9	8.00	6.42	8.10	6.50	8.29	6.66	8.48	6.81	8.69
26.0	8.07	6.48	8.16	6.55	8.36	6.71	8.55	6.87	8.75
26.1	8.14	6.53	8.23	6.61	8.43	6.77	8.62	6.92	8.82
26.2	8.21	6.59	8.30	6.66	8.50	6.82	8.69	6.98	8.89
26.3	8.27	6.64	8.37	6.72	8.57	6.88	8.75	7.03	8.96
26.4	8.34	6.70	8.44	6.78	8.63	6.93	8.82	7.09	9.03
26.5	8.41	6.75	8.50	6.83	8.70	6.99	8.89	7.15	9.10
26.6	8.48	6.81	8.57	6.88	8.77	7.04	8.96	7.20	9.16
26.7	8.55	6.86	8.64	6.94	8.84	7.10	9.03	7.26	9.23
26.8	8.62	6.92	8.71	6.99	8.91	7.15	9.10	7.31	9.30
26.9	8.68	6.97	8.78	7.05	8.98	7.21	9.17	7.37	9.37
27.0	8.75	7.03	8.85	7.11	9.05	7.27	9.23	7.42	9.44
27.1	8.82	7.08	8.91	7.16	9.11	7.32	9.30	7.48	9.51
27.2	8.89	7.14	8.98	7.22	9.18	7.38	9.37	7.54	9.58
27.3	8.95	7.19	9.05	7.27	9.25	7.43	9.44	7.59	9.65
27.4	9.02	7.25	9.12	7.33	9.32	7.49	9.51	7.65	9.71
27.5	9.09	7.30	9.19	7.38	9.38	7.54	9.58	7.70	9.78
27.6	9.16	7.36	9.26	7.44	9.45	7.60	9.65	7.76	9.85
27.7	9.22	7.41	9.32	7.49	9.52	7.65	9.72	7.82	9.91
27.8	9.29	7.47	9.39	7.55	9.59	7.71	9.79	7.87	9.98
27.9	9.36	7.52	9.46	7.60	9.65	7.76	9.86	7.93	10.05

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight		
5.34	6.86	5.50	7.06	5.66	7.24	5.81	7.45	5.97	23.0	
5.40	6.93	5.56	7.13	5.71	7.32	5.87	7.52	6.03	23.1	
5.46	7.00	5.61	7.20	5.77	7.39	5.93	7.59	6.08	23.2	
5.51	7.07	5.67	7.27	5.83	7.46	5.98	7.66	6.14	23.3	
5.57	7.14	5.73	7.34	5.89	7.53	6.04	7.73	6.20	23.4	
5.63	7.21	5.78	7.41	5.94	7.60	6.10	7.80	6.25	23.5	
5.69	7.28	5.84	7.48	6.00	7.67	6.15	7.87	6.31	23.6	
5.74	7.35	5.89	7.55	6.06	7.74	6.21	7.94	6.37	23.7	
5.80	7.42	5.95	7.62	6.11	7.81	6.27	8.00	6.42	23.8	
5.86	7.49	6.01	7.69	6.17	7.88	6.32	8.07	6.48	23.9	
5.92	7.56	6.07	7.76	6.22	7.95	6.38	8.14	6.53	24.0	
5.97	7.63	6.12	7.83	6.28	8.02	6.44	8.21	6.59	24.1	
6.03	7.70	6.18	7.90	6.34	8.09	6.49	8.28	6.65	24.2	
6.08	7.77	6.24	7.97	6.39	8.16	6.55	8.35	6.70	24.3	
6.14	7.84	6.29	8.04	6.45	8.23	6.60	8.42	6.76	24.4	
6.20	7.91	6.35	8.10	6.50	8.30	6.66	8.48	6.81	24.5	
6.25	7.98	6.41	8.17	6.56	8.37	6.72	8.55	6.87	24.6	
6.31	8.05	6.46	8.24	6.62	8.44	6.77	8.62	6.93	24.7	
6.36	8.12	6.52	8.31	6.67	8.51	6.83	8.69	6.98	24.8	
6.42	8.19	6.58	8.38	6.73	8.58	6.89	8.76	7.04	24.9	
6.47	8.26	6.63	8.45	6.79	8.64	6.94	8.84	7.10	25.0	
6.53	8.33	6.69	8.52	6.84	8.71	7.00	8.91	7.15	25.1	
6.59	8.40	6.75	8.59	6.90	8.78	7.06	8.98	7.21	25.2	
6.64	8.47	6.80	8.66	6.96	8.85	7.11	9.05	7.27	25.3	
6.70	8.54	6.86	8.73	7.01	8.92	7.17	9.12	7.33	25.4	
6.75	8.61	6.92	8.80	7.07	8.99	7.23	9.19	7.38	25.5	
6.81	8.68	6.97	8.86	7.12	9.06	7.28	9.26	7.44	25.6	
6.87	8.75	7.03	8.93	7.18	9.13	7.34	9.33	7.50	25.7	
6.92	8.82	7.08	9.00	7.23	9.20	7.40	9.39	7.55	25.8	
6.98	8.89	7.14	9.07	7.29	9.27	7.45	9.46	7.61	25.9	
7.03	8.95	7.19	9.14	7.35	9.34	7.51	9.53	7.67	26.0	
7.09	9.02	7.25	9.21	7.40	9.41	7.56	9.60	7.73	26.1	
7.14	9.09	7.30	9.28	7.46	9.48	7.62	9.67	7.78	26.2	
7.20	9.16	7.36	9.35	7.51	9.55	7.68	9.74	7.84	26.3	
7.25	9.22	7.41	9.42	7.57	9.61	7.73	9.81	7.90	26.4	
7.31	9.29	7.47	9.49	7.63	9.68	7.79	9.88	7.95	26.5	
7.36	9.36	7.52	9.55	7.68	9.75	7.85	9.95	8.01	26.6	
7.42	9.43	7.58	9.62	7.74	9.82	7.90	10.02	8.07	26.7	
7.47	9.49	7.63	9.69	7.79	9.89	7.96	10.09	8.12	26.8	
7.53	9.56	7.69	9.76	7.85	9.96	8.02	10.16	8.18	26.9	
7.59	9.63	7.74	9.83	7.91	10.03	8.07	10.23	8.24	27.0	
7.65	9.70	7.80	9.90	7.96	10.10	8.13	10.30	8.29	27.1	
7.70	9.76	7.85	9.97	8.02	10.17	8.18	10.37	8.35	27.2	
7.76	9.83	7.91	10.03	8.07	10.24	8.24	10.44	8.40	27.3	
7.81	9.90	7.96	10.10	8.13	10.31	8.30	10.51	8.46	27.4	
7.86	9.97	8.02	10.17	8.18	10.38	8.35	10.58	8.52	27.5	
7.92	10.03	8.07	10.24	8.24	10.45	8.41	10.65	8.57	27.6	
7.97	10.10	8.13	10.31	8.30	10.51	8.46	10.72	8.63	27.7	
8.03	10.17	8.18	10.38	8.35	10.58	8.52	10.79	8.69	27.8	
8.08	10.24	8.24	10.45	8.41	10.65	8.58	10.86	8.74	27.9	

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
28 0	9.43	7.58	9.53	7.66	9.72	7.82	9.92	7.98	10.12
28 1	9.50	7.64	9.59	7.71	9.79	7.87	9.99	8.04	10.18
28 2	9.57	7.69	9.66	7.77	9.86	7.93	10.06	8.09	10.25
28 3	9.64	7.75	9.73	7.82	9.92	7.98	10.13	8.15	10.32
28 4	9.70	7.80	9.80	7.88	9.99	8.04	10.19	8.20	10.39
28 5	9.77	7.86	9.86	7.93	10.06	8.09	10.26	8.26	10.45
28 6	9.84	7.91	9.93	7.99	10.13	8.15	10.32	8.31	10.52
28 7	9.91	7.97	10.00	8.04	10.19	8.20	10.39	8.36	10.59
28 8	9.97	8.02	10.07	8.10	10.26	8.26	10.46	8.42	10.66
28 9	10.04	8.08	10.13	8.15	10.32	8.31	10.52	8.47	10.73
29 0	10.10	8.13	10.19	8.20	10.39	8.36	10.59	8.53	10.79
29 1	10.17	8.18	10.26	8.26	10.46	8.42	10.66	8.58	10.86
29 2	10.24	8.24	10.23	8.31	10.52	8.47	10.73	8.64	10.93
29 3	10.30	8.29	10.40	8.37	10.59	8.53	10.79	8.69	11.00
29 4	10.36	8.34	10.46	8.42	10.66	8.58	10.86	8.74	11.06
29 5	10.43	8.40	10.52	8.47	10.73	8.64	10.93	8.80	11.13
29 6	10.50	8.45	10.59	8.53	10.79	8.69	10.99	8.85	11.20
29 7	10.56	8.50	10.66	8.58	10.86	8.74	11.06	8.91	11.27
29 8	10.63	8.56	10.72	8.63	10.93	8.80	11.12	8.96	11.33
29 9	10.69	8.61	10.79	8.69	10.99	8.85	11.19	9.02	11.39
30 0	10.76	8.66	10.86	8.74	11.06	8.91	11.26	9.07	11.46
30 1	10.83	8.72	10.93	8.80	11.12	8.96	11.32	9.12	11.52
30 2	10.89	8.77	10.99	8.85	11.18	9.02	11.38	9.18	11.59
30 3	10.95	8.82	11.05	8.90	11.25	9.07	11.45	9.23	11.66
30 4	11.02	8.88	11.12	8.96	11.31	9.12	11.51	9.28	11.72
30 5	11.08	8.93	11.18	9.01	11.38	9.18	11.58	9.34	11.79
30 6	11.15	8.98	11.25	9.06	11.44	9.23	11.64	9.39	11.85
30 7	11.21	9.04	11.31	9.12	11.51	9.28	11.71	9.44	11.92
30 8	11.28	9.09	11.38	9.17	11.58	9.34	11.78	9.50	11.99
30 9	11.34	9.14	11.44	9.22	11.64	9.39	11.84	9.55	12.05
31 0	11.41	9.19	11.51	9.28	11.71	9.44	11.91	9.60	12.12
31 1	11.47	9.25	11.57	9.33	11.77	9.49	11.97	9.66	12.18
31 2	11.54	9.30	11.64	9.38	11.84	9.55	12.04	9.71	12.25
31 3	11.60	9.35	11.70	9.43	11.90	9.60	12.11	9.76	12.32
31 4	11.66	9.40	11.77	9.49	11.97	9.65	12.17	9.82	12.38
31 5	11.73	9.46	11.83	9.54	12.03	9.71	12.24	9.87	12.45
31 6	11.79	9.51	11.90	9.59	12.10	9.76	12.30	9.92	12.51
31 7	11.86	9.56	11.96	9.65	12.16	9.81	12.37	9.98	12.58
31 8	11.92	9.62	12.03	9.70	12.23	9.87	12.43	10.03	12.64
31 9	11.99	9.67	12.09	9.75	12.29	9.92	12.50	10.09	12.71
32 0	12.05	9.72	12.15	9.80	12.36	9.97	12.57	10.14	12.78
32 1	12.12	9.77	12.21	9.86	12.42	10.03	12.63	10.19	12.84
32 2	12.18	9.83	12.28	9.91	12.49	10.08	12.70	10.25	12.91
32 3	12.24	9.88	12.34	9.96	12.55	10.13	12.76	10.30	12.97
32 4	12.31	9.93	12.40	10.02	12.62	10.19	12.83	10.35	13.04
32 5	12.37	9.98	12.47	10.07	12.68	10.24	12.89	10.41	13.10
32 6	12.43	10.04	12.54	10.12	12.75	10.29	12.96	10.46	13.17
32 7	12.50	10.09	12.60	10.17	12.81	10.34	13.03	10.52	13.24
32 8	12.56	10.14	12.67	10.23	12.88	10.39	13.09	10.57	13.30
32 9	12.62	10.19	12.72	10.28	12.94	10.45	13.15	10.62	13.37

TABLE.—Continued.

6

21° C.	22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
8.14	10.31	8.30	10.51	8.46	10.72	8.63	10.93	8.80	28.0
8.19	10.38	8.35	10.58	8.52	10.79	8.69	10.99	8.85	28.1
8.25	10.45	8.41	10.65	8.58	10.86	8.74	11.06	8.91	28.2
8.30	10.52	8.46	10.72	8.63	10.93	8.80	11.13	8.97	28.3
8.36	10.59	8.52	10.79	8.69	11.00	8.86	11.20	9.02	28.4
8.41	10.66	8.58	10.86	8.74	11.06	8.91	11.27	9.08	28.5
8.47	10.72	8.63	10.93	8.80	11.13	8.97	11.33	9.13	28.6
8.52	10.79	8.69	11.00	8.86	11.20	9.02	11.40	9.19	28.7
8.58	10.86	8.74	11.06	8.91	11.27	9.08	11.47	9.24	28.8
8.64	10.93	8.80	11.13	8.97	11.33	9.13	11.54	9.30	28.9
8.69	11.00	8.86	11.20	9.02	11.40	9.19	11.61	9.36	29.0
8.75	11.06	8.91	11.27	9.08	11.47	9.24	11.68	9.41	29.1
8.80	11.13	8.97	11.33	9.13	11.54	9.30	11.75	9.47	29.2
8.86	11.20	9.02	11.40	9.19	11.60	9.35	11.81	9.52	29.3
8.91	11.27	9.08	11.47	9.24	11.67	9.41	11.88	9.58	29.4
8.97	11.33	9.13	11.54	9.30	11.74	9.46	11.94	9.63	29.5
9.02	11.39	9.18	11.60	9.35	11.81	9.52	12.01	9.69	29.6
9.08	11.46	9.24	11.67	9.41	11.87	9.57	12.08	9.75	29.7
9.13	11.53	9.29	11.74	9.46	11.94	9.63	12.15	9.80	29.8
9.18	11.60	9.35	11.81	9.52	12.01	9.69	12.22	9.86	29.9
9.24	11.66	9.40	11.87	9.57	12.08	9.74	12.29	9.91	30.0
9.29	11.73	9.46	11.93	9.63	12.14	9.80	12.36	9.97	30.1
9.34	11.79	9.51	12.00	9.68	12.21	9.85	12.42	10.02	30.2
9.40	11.86	9.57	12.07	9.74	12.28	9.91	12.49	10.08	30.3
9.45	11.93	9.62	12.13	9.79	12.34	9.96	12.56	10.13	30.4
9.50	11.99	9.67	12.20	9.85	12.41	10.02	12.63	10.19	30.5
9.56	12.06	9.73	12.27	9.90	12.48	10.07	12.70	10.24	30.6
9.61	12.13	9.78	12.34	9.96	12.55	10.13	12.77	10.30	30.7
9.67	12.19	9.84	12.40	10.01	12.61	10.18	12.84	10.36	30.8
9.72	12.26	9.89	12.47	10.07	12.68	10.24	12.90	10.41	30.9
9.77	12.32	9.95	12.54	10.12	12.75	10.29	12.97	10.47	31.0
9.83	12.39	10.00	12.60	10.17	12.82	10.35	13.04	10.52	31.1
9.88	12.46	10.05	12.67	10.23	12.89	10.40	13.11	10.58	31.2
9.94	12.52	10.11	12.74	10.28	12.95	10.46	13.17	10.63	31.3
9.99	12.59	10.16	12.81	10.34	13.02	10.51	13.24	10.69	31.4
10.04	12.66	10.22	12.87	10.39	13.09	10.57	13.31	10.74	31.5
10.10	12.72	10.27	12.94	10.45	13.15	10.62	13.37	10.80	31.6
10.15	12.79	10.32	13.01	10.50	13.22	10.68	13.44	10.86	31.7
10.21	12.85	10.38	13.07	10.55	13.29	10.73	13.51	10.91	31.8
10.26	12.92	10.43	13.14	10.61	13.35	10.78	13.57	10.97	31.9
10.31	12.99	10.49	13.20	10.66	13.42	10.84	13.64	11.02	32.0
10.37	13.05	10.54	13.27	10.72	13.49	10.90	13.71	11.08	32.1
10.42	13.12	10.59	13.34	10.77	13.55	10.95	13.77	11.13	32.2
10.48	13.18	10.65	13.40	10.83	13.62	11.01	13.84	11.19	32.3
10.53	13.25	10.70	13.47	10.88	13.69	11.06	13.91	11.24	32.4
10.58	13.32	10.76	13.53	10.94	13.75	11.11	13.97	11.30	32.5
10.64	13.38	10.81	13.60	10.99	13.82	11.17	14.04	11.35	32.6
10.69	13.45	10.87	13.66	11.04	13.89	11.22	14.11	11.41	32.7
10.75	13.51	10.92	13.73	11.10	13.95	11.28	14.17	11.46	32.8
10.80	13.58	10.97	13.80	11.15	14.02	11.33	14.24	11.52	32.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
33.0	12.69	10.24	12.79	10.33	13.01	10.50	13.22	10.68	13.43
33.1	12.76	10.30	12.86	10.38	13.07	10.56	13.28	10.73	13.50
33.2	12.82	10.35	12.92	10.43	13.13	10.61	13.35	10.79	13.56
33.3	12.88	10.40	12.99	10.49	13.20	10.66	13.41	10.84	13.63
33.4	12.95	10.45	13.05	10.54	13.26	10.71	13.48	10.89	13.69
33.5	13.01	10.50	13.11	10.59	13.32	10.77	13.54	10.95	13.76
33.6	13.08	10.56	13.18	10.64	13.39	10.82	13.61	11.00	13.82
33.7	13.14	10.61	13.24	10.70	13.45	10.87	13.67	11.05	13.89
33.8	13.20	10.66	13.30	10.75	13.52	10.93	13.74	11.10	13.95
33.9	13.26	10.71	13.37	10.80	13.58	10.98	13.80	11.16	14.02
34.0	13.33	10.77	13.43	10.85	13.64	11.03	13.86	11.21	14.08
34.1	13.39	10.82	13.49	10.91	13.71	11.08	13.93	11.26	14.15
34.2	13.45	10.87	13.56	10.96	13.77	11.13	13.99	11.31	14.21
34.3	13.52	10.92	13.62	11.01	13.83	11.19	14.06	11.36	14.27
34.4	13.58	10.97	13.68	11.06	13.90	11.24	14.12	11.41	14.34
34.5	13.64	11.03	13.75	11.11	13.96	11.29	14.18	11.47	14.40
34.6	13.70	11.08	13.81	11.16	14.02	11.34	14.25	11.52	14.47
34.7	13.77	11.13	13.87	11.22	14.08	11.39	14.31	11.57	14.53
34.8	13.83	11.18	13.94	11.27	14.14	11.44	14.37	11.62	14.59
34.9	13.89	11.23	14.00	11.32	14.20	11.49	14.43	11.67	14.66
35.0	13.96	11.28	14.06	11.37	14.27	11.55	14.50	11.73	14.72
35.1	14.02	11.33	14.13	11.42	14.33	11.60	14.56	11.78	14.78
35.2	14.08	11.38	14.19	11.47	14.39	11.65	14.62	11.83	14.85
35.3	14.14	11.44	14.25	11.52	14.46	11.70	14.69	11.88	14.91
35.4	14.21	11.49	14.31	11.57	14.52	11.75	14.75	11.93	14.97
35.5	14.27	11.54	14.38	11.63	14.59	11.81	14.81	11.99	15.04
35.6	14.33	11.59	14.44	11.68	14.65	11.86	14.87	12.04	15.10
35.7	14.39	11.64	14.50	11.73	14.71	11.91	14.94	12.09	15.16
35.8	14.46	11.69	14.56	11.78	14.78	11.96	15.00	12.14	15.23
35.9	14.52	11.74	14.63	11.83	14.84	12.01	15.06	12.19	15.29
36.0	14.58	11.79	14.69	11.88	14.90	12.06	15.13	12.24	15.35
36.1	14.64	11.85	14.75	11.94	14.97	12.11	15.19	12.30	15.42
36.2	14.71	11.90	14.81	11.99	15.03	12.16	15.25	12.35	15.48
36.3	14.77	11.95	14.88	12.04	15.09	12.22	15.32	12.40	15.54
36.4	14.83	12.00	14.94	12.09	15.16	12.27	15.38	12.45	15.61
36.5	14.89	12.05	15.00	12.14	15.22	12.32	15.44	12.50	15.67
36.6	14.96	12.10	15.06	12.19	15.28	12.37	15.51	12.56	15.73
36.7	15.02	12.15	15.13	12.24	15.35	12.42	15.57	12.61	15.80
36.8	15.08	12.20	15.19	12.29	15.41	12.47	15.63	12.66	15.86
36.9	15.14	12.25	15.25	12.34	15.47	12.53	15.70	12.71	15.92
37.0	15.20	12.30	15.31	12.40	15.53	12.58	15.76	12.77	15.99
37.1	15.27	12.36	15.38	12.45	15.60	12.63	15.82	12.82	16.05
37.2	15.33	12.41	15.44	12.50	15.66	12.68	15.89	12.87	16.11
37.3	15.39	12.46	15.50	12.55	15.72	12.73	15.95	12.92	16.18
37.4	15.45	12.51	15.56	12.60	15.79	12.78	16.01	12.97	16.24
37.5	15.51	12.56	15.63	12.65	15.85	12.84	16.08	13.03	16.30
37.6	15.57	12.61	15.69	12.70	15.91	12.89	16.14	13.08	16.37
37.7	15.64	12.66	15.75	12.75	15.97	12.94	16.20	13.13	16.43
37.8	15.70	12.71	15.81	12.81	16.04	12.99	16.26	13.18	16.49
37.9	15.76	12.76	15.88	12.86	16.10	13.04	16.33	13.23	16.56

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight		
10.85	13.64	11.03	13.86	11.21	14.09	11.39	14.31	11.57	33.0	
10.91	13.71	11.08	13.93	11.26	14.15	11.44	14.38	11.63	33.1	
10.96	13.78	11.13	13.99	11.31	14.22	11.49	14.44	11.68	33.2	
11.02	13.84	11.19	14.06	11.37	14.28	11.55	14.51	11.74	33.3	
11.07	13.91	11.24	14.13	11.42	14.35	11.60	14.58	11.79	33.4	
11.12	13.97	11.29	14.19	11.48	14.42	11.66	14.64	11.85	33.5	
11.17	14.04	11.35	14.26	11.53	14.48	11.71	14.71	11.90	33.6	
11.23	14.11	11.40	14.32	11.58	14.55	11.77	14.78	11.96	33.7	
11.28	14.17	11.45	14.39	11.64	14.62	11.82	14.85	12.01	33.8	
11.33	14.24	11.51	14.45	11.69	14.68	11.88	14.91	12.07	33.9	
11.38	14.30	11.56	14.52	11.75	14.75	11.93	14.98	12.12	34.0	
11.44	14.37	11.61	14.59	11.80	14.81	11.98	15.05	12.18	34.1	
11.49	14.43	11.67	14.65	11.85	14.88	12.04	15.11	12.23	34.2	
11.54	14.50	11.72	14.72	11.91	14.95	12.09	15.18	12.29	34.3	
11.59	14.57	11.78	14.78	11.96	15.01	12.15	15.25	12.34	34.4	
11.65	14.63	11.83	14.85	12.02	15.08	12.20	15.31	12.40	34.5	
11.70	14.70	11.88	14.91	12.07	15.14	12.26	15.38	12.45	34.6	
11.75	14.76	11.94	14.98	12.12	15.21	12.31	15.45	12.51	34.7	
11.81	14.83	11.99	15.05	12.18	15.28	12.36	15.51	12.56	34.8	
11.86	14.89	12.04	15.11	12.23	15.34	12.42	15.58	12.62	34.9	
11.91	14.96	12.10	15.18	12.28	15.41	12.47	15.65	12.67	35.0	
11.96	15.03	12.15	15.24	12.34	15.47	12.53	15.71	12.73	35.1	
12.02	15.09	12.20	15.31	12.39	15.54	12.58	15.78	12.78	35.2	
12.07	15.15	12.25	15.37	12.44	15.61	12.64	15.85	12.84	35.3	
12.12	15.22	12.31	15.44	12.50	15.67	12.69	15.91	12.89	35.4	
12.17	15.28	12.36	15.50	12.55	15.74	12.75	15.98	12.95	35.5	
12.23	15.34	12.41	15.56	12.60	15.80	12.80	16.05	13.00	35.6	
12.28	15.41	12.47	15.63	12.66	15.87	12.85	16.11	13.05	35.7	
12.33	15.47	12.52	15.69	12.71	15.93	12.91	16.18	13.11	35.8	
12.38	15.53	12.57	15.76	12.76	16.00	12.96	16.24	13.16	35.9	
12.43	15.59	12.62	15.82	12.82	16.06	13.02	16.31	13.21	36.0	
12.49	15.66	12.68	15.89	12.87	16.13	13.07	16.37	13.27	36.1	
12.54	15.72	12.73	15.95	12.92	16.19	13.12	16.44	13.32	36.2	
12.59	15.78	12.78	16.02	12.98	16.26	13.18	16.50	13.37	36.3	
12.64	15.85	12.84	16.08	13.03	16.32	13.23	16.56	13.43	36.4	
12.70	15.91	12.89	16.15	13.08	16.39	13.28	16.63	13.48	36.5	
12.75	15.97	12.94	16.21	13.14	16.45	13.34	16.69	13.53	36.6	
12.80	16.04	12.99	16.28	13.19	16.52	13.39	16.76	13.59	36.7	
12.85	16.10	13.05	16.34	13.24	16.58	13.44	16.82	13.64	36.8	
12.91	16.16	13.10	16.40	13.29	16.65	13.49	16.89	13.70	36.9	
12.96	16.23	13.15	16.47	13.35	16.71	13.55	16.95	13.75	37.0	
13.01	16.29	13.20	16.53	13.40	16.77	13.60	17.02	13.80	37.1	
13.06	16.35	13.26	16.60	13.45	16.84	13.65	17.08	13.86	37.2	
13.11	16.42	13.31	16.66	13.50	16.90	13.71	17.15	13.91	37.3	
13.16	16.48	13.36	16.72	13.56	16.97	13.76	17.21	13.96	37.4	
13.21	16.54	13.41	16.79	13.61	17.03	13.81	17.27	14.02	37.5	
13.27	16.61	13.46	16.85	13.66	17.09	13.87	17.34	14.07	37.6	
13.32	16.67	13.52	16.92	13.72	17.16	13.92	17.40	14.12	37.7	
13.37	16.73	13.57	16.98	13.77	17.22	13.97	17.46	14.17	37.8	
13.42	16.80	13.62	17.04	13.82	17.28	14.03	17.53	14.23	37.9	

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
38.0	15.82	12.81	15.94	12.91	16.16	13.09	16.39	13.28	16.62
38.1	15.88	12.86	16.00	12.96	16.22	13.14	16.45	13.33	16.68
38.2	15.94	12.91	16.06	13.01	16.29	13.19	16.51	13.38	16.75
38.3	16.01	12.96	16.12	13.06	16.35	13.25	16.57	13.44	16.81
38.4	16.07	13.02	16.18	13.11	16.41	13.30	16.64	13.49	16.87
38.5	16.13	13.07	16.25	13.16	16.47	13.35	16.70	13.54	16.94
38.6	16.19	13.12	16.31	13.21	16.53	13.40	16.76	13.59	17.00
38.7	16.25	13.17	16.37	13.26	16.60	13.45	16.83	13.64	17.06
38.8	16.31	13.22	16.43	13.31	16.66	13.50	16.89	13.69	17.13
38.9	16.37	13.27	16.49	13.37	16.72	13.55	16.95	13.74	17.19
39.0	16.44	13.32	16.55	13.42	16.78	13.61	17.01	13.79	17.25
39.1	16.50	13.37	16.61	13.47	16.84	13.66	17.07	13.85	17.31
39.2	16.56	13.42	16.67	13.52	16.91	13.71	17.14	13.90	17.38
39.3	16.62	13.47	16.74	13.57	16.97	13.76	17.20	13.95	17.44
39.4	16.68	13.52	16.80	13.62	17.03	13.81	17.26	14.00	17.50
39.5	16.74	13.57	16.86	13.67	17.09	13.86	17.32	14.05	17.56
39.6	16.80	13.62	16.92	13.72	17.15	13.91	17.39	14.10	17.63
39.7	16.87	13.68	16.98	13.77	17.21	13.96	17.45	14.15	17.69
39.8	16.93	13.73	17.04	13.82	17.28	14.02	17.51	14.21	17.75
39.9	16.99	13.78	17.10	13.87	17.34	14.07	17.57	14.26	17.81
40.0	17.05	13.83	17.16	13.92	17.40	14.12	17.63	14.31	17.88
40.1	17.11	13.88	17.23	13.98	17.46	14.17	17.70	14.36	17.94
40.2	17.17	13.93	17.29	14.03	17.52	14.22	17.76	14.41	18.00
40.3	17.23	13.98	17.35	14.08	17.58	14.27	17.82	14.46	18.06
40.4	17.29	14.03	17.41	14.13	17.64	14.32	17.88	14.51	18.12
40.5	17.35	14.08	17.47	14.18	17.71	14.37	17.94	14.56	18.19
40.6	17.41	14.13	17.53	14.23	17.77	14.42	18.01	14.62	18.25
40.7	17.48	14.18	17.59	14.28	17.83	14.47	18.07	14.67	18.31
40.8	17.54	14.23	17.65	14.33	17.89	14.52	18.13	14.72	18.37
40.9	17.60	14.28	17.71	14.38	17.95	14.57	18.19	14.77	18.43
41.0	17.66	14.33	17.77	14.43	18.01	14.62	18.25	14.82	18.49
41.1	17.72	14.38	17.84	14.48	18.07	14.68	18.31	14.87	18.56
41.2	17.78	14.43	17.90	14.53	18.13	14.73	18.37	14.92	18.62
41.3	17.84	14.48	17.96	14.58	18.20	14.78	18.44	14.97	18.68
41.4	17.90	14.53	18.03	14.63	18.26	14.83	18.50	15.03	18.74
41.5	17.96	14.58	18.08	14.68	18.32	14.88	18.56	15.08	18.80
41.6	18.02	14.63	18.14	14.73	18.38	14.93	18.62	15.13	18.86
41.7	18.08	14.68	18.20	14.78	18.44	14.98	18.68	15.18	18.93
41.8	18.14	14.73	18.26	14.83	18.50	15.03	18.74	15.23	18.99
41.9	18.20	14.78	18.32	14.88	18.56	15.08	18.81	15.28	19.05
42.0	18.27	14.83	18.38	14.93	18.62	15.13	18.87	15.33	19.11
42.1	18.33	14.88	18.44	14.98	18.68	15.18	18.93	15.38	19.17
42.2	18.39	14.93	18.50	15.03	18.74	15.23	18.99	15.43	19.23
42.3	18.45	14.98	18.56	15.08	18.80	15.28	19.05	15.48	19.29
42.4	18.51	15.03	18.62	15.13	18.87	15.33	19.11	15.53	19.36
42.5	18.57	15.08	18.68	15.18	18.93	15.38	19.17	15.58	19.42
42.6	18.63	15.13	18.75	15.23	18.99	15.43	19.23	15.63	19.48
42.7	18.69	15.18	18.81	15.28	19.05	15.48	19.29	15.69	19.54
42.8	18.75	15.23	18.87	15.33	19.11	15.53	19.36	15.74	19.60
42.9	18.81	15.28	18.93	15.38	19.17	15.58	19.42	15.79	19.66

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight		Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
13.47	16.86	13.67		17.11	13.87	17.35	14.08	17.59	14.28	38.0
13.53	16.92	13.72		17.17	13.93	17.41	14.13	17.66	14.33	38.1
13.58	16.99	13.78		17.23	13.98	17.47	14.18	17.72	14.38	38.2
13.63	17.05	13.83		17.30	14.03	17.54	14.23	17.78	14.43	38.3
13.68	17.11	13.88		17.36	14.08	17.60	14.29	17.85	14.49	38.4
13.73	17.18	13.93		17.42	14.14	17.66	14.34	17.91	14.54	38.5
13.79	17.24	13.98		17.48	14.19	17.73	14.39	17.97	14.59	38.6
13.84	17.30	14.04		17.55	14.24	17.79	14.44	18.04	14.64	38.7
13.89	17.36	14.09		17.61	14.29	17.85	14.49	18.10	14.70	38.8
13.94	17.43	14.14		17.67	14.34	17.92	14.55	18.16	14.75	38.9
13.99	17.49	14.19		17.74	14.40	17.98	14.60	18.23	14.80	39.0
14.05	17.55	14.24		17.80	14.45	18.04	14.65	18.29	14.85	39.1
14.10	17.62	14.30		17.86	14.50	18.11	14.70	18.35	14.91	39.2
14.15	17.68	14.35		17.92	14.55	18.17	14.76	18.42	14.96	39.3
14.20	17.74	14.40		17.99	14.60	18.23	14.81	18.48	15.01	39.4
14.25	17.81	14.45		18.05	14.66	18.30	14.86	18.54	15.06	39.5
14.30	17.87	14.50		18.11	14.71	18.36	14.91	18.61	15.12	39.6
14.36	17.93	14.56		18.18	14.76	18.42	14.96	18.67	15.17	39.7
14.41	17.99	14.61		18.24	14.81	18.48	15.02	18.73	15.22	39.8
14.46	18.06	14.66		18.30	14.87	18.55	15.07	18.80	15.27	39.9
14.51	18.12	14.71		18.36	14.92	18.61	15.12	18.86	15.32	40.0
14.56	18.18	14.77		18.43	14.97	18.67	15.17	18.92	15.38	40.1
14.61	18.24	14.82		18.49	15.02	18.74	15.22	18.99	15.43	40.2
14.67	18.30	14.87		18.55	15.07	18.80	15.27	19.05	15.48	40.3
14.72	18.37	14.92		18.61	15.12	18.86	15.33	19.11	15.53	40.4
14.77	18.43	14.97		18.68	15.17	18.92	15.38	19.18	15.59	40.5
14.82	18.49	15.03		18.74	15.23	18.99	15.43	19.24	15.64	40.6
14.87	18.55	15.08		18.80	15.28	19.05	15.48	19.30	15.69	40.7
14.92	18.61	15.13		18.86	15.33	19.11	15.53	19.37	15.74	40.8
14.97	18.68	15.18		18.93	15.38	19.18	15.59	19.43	15.80	40.9
15.03	18.74	15.23		18.99	15.43	19.24	15.64	19.49	15.85	41.0
15.08	18.80	15.28		19.05	15.48	19.30	15.69	19.56	15.90	41.1
15.13	18.86	15.33		19.11	15.53	19.36	15.74	19.62	15.95	41.2
15.18	18.93	15.38		19.17	15.58	19.43	15.79	19.68	16.01	41.3
15.23	18.99	15.43		19.24	15.64	19.49	15.84	19.75	16.06	41.4
15.28	19.05	15.48		19.30	15.69	19.55	15.90	19.81	16.11	41.5
15.33	19.11	15.53		19.36	15.74	19.61	15.95	19.87	16.16	41.6
15.38	19.17	15.58		19.42	15.79	19.68	16.00	19.94	16.21	41.7
15.43	19.23	15.63		19.48	15.84	19.74	16.05	20.00	16.27	41.8
15.48	19.29	15.69		19.55	15.89	19.80	16.10	20.06	16.32	41.9
15.53	19.36	15.74		19.61	15.94	19.86	16.16	20.13	16.37	42.0
15.58	19.42	15.79		19.67	15.99	19.93	16.21	20.19	16.42	42.1
15.63	19.48	15.84		19.73	16.05	19.99	16.26	20.25	16.48	42.2
15.69	19.54	15.89		19.80	16.10	20.05	16.31	20.31	16.53	42.3
15.74	19.60	15.94		19.86	16.15	20.11	16.36	20.38	16.58	42.4
15.79	19.66	15.99		19.92	16.20	20.18	16.41	20.44	16.63	42.5
15.84	19.72	16.04		19.98	16.25	20.24	16.47	20.50	16.69	42.6
15.89	19.79	16.09		20.04	16.30	20.30	16.52	20.57	16.74	42.7
15.94	19.85	16.14		20.10	16.35	20.36	16.57	20.63	16.79	42.8
15.99	19.91	16.19		20.17	16.41	20.43	16.62	20.69	16.84	42.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
43.0	18.87	15.33	18.99	15.43	19.23	15.63	19.48	15.84	19.72
43.1	18.93	15.38	19.05	15.48	19.29	15.68	19.54	15.89	19.79
43.2	18.99	15.43	19.11	15.53	19.35	15.73	19.60	15.94	19.85
43.3	19.05	15.48	19.17	15.58	19.41	15.78	19.66	15.99	19.91
43.4	19.11	15.53	19.23	15.63	19.47	15.83	19.72	16.04	19.97
43.5	19.17	15.58	19.29	15.68	19.53	15.88	19.79	16.09	20.03
43.6	19.23	15.63	19.35	15.73	19.59	15.93	19.85	16.14	20.09
43.7	19.29	15.68	19.41	15.78	19.65	15.98	19.91	16.19	20.15
43.8	19.35	15.73	19.47	15.83	19.72	16.03	19.97	16.24	20.21
43.9	19.40	15.78	19.53	15.88	19.78	16.08	20.03	16.29	20.27
44.0	19.46	15.83	19.59	15.93	19.84	16.13	20.09	16.34	20.34
44.1	19.52	15.88	19.65	15.98	19.90	16.18	20.15	16.39	20.40
44.2	19.58	15.93	19.71	16.03	19.96	16.23	20.21	16.44	20.46
44.3	19.64	15.98	19.77	16.08	20.02	16.28	20.27	16.49	20.52
44.4	19.70	16.02	19.83	16.13	20.08	16.33	20.33	16.55	20.58
44.5	19.76	16.07	19.89	16.18	20.14	16.38	20.39	16.60	20.64
44.6	19.82	16.12	19.95	16.23	20.20	16.43	20.45	16.65	20.70
44.7	19.88	16.17	20.01	16.27	20.26	16.48	20.52	16.70	20.76
44.8	19.94	16.22	20.07	16.32	20.32	16.53	20.58	16.75	20.82
44.9	20.00	16.27	20.12	16.37	20.38	16.58	20.64	16.80	20.88
45.0	20.06	16.32	20.18	16.42	20.44	16.63	20.70	16.85	20.95
45.1	20.12	16.37	20.24	16.47	20.50	16.68	20.76	16.90	21.01
45.2	20.18	16.41	20.30	16.52	20.56	16.73	20.82	16.95	21.07
45.3	20.24	16.46	20.36	16.57	20.62	16.78	20.88	17.00	21.13
45.4	20.29	16.51	20.42	16.62	20.68	16.83	20.94	17.05	21.19
45.5	20.35	16.56	20.48	16.67	20.74	16.88	21.00	17.10	21.25
45.6	20.41	16.61	20.54	16.72	20.80	16.93	21.06	17.15	21.31
45.7	20.47	16.66	20.60	16.76	20.86	16.98	21.12	17.20	21.37
45.8	20.53	16.71	20.66	16.81	20.92	17.03	21.18	17.25	21.43
45.9	20.59	16.76	20.72	16.86	20.98	17.08	21.24	17.30	21.49
46.0	20.65	16.80	20.78	16.91	21.04	17.13	21.30	17.35	21.54
46.1	20.71	16.85	20.83	16.96	21.10	17.18	21.36	17.40	21.60
46.2	20.76	16.90	20.89	17.01	21.16	17.23	21.42	17.45	21.66
46.3	20.82	16.95	20.95	17.06	21.22	17.28	21.48	17.50	21.72
46.4	20.88	17.00	21.01	17.11	21.28	17.33	21.54	17.55	21.78
46.5	20.94	17.05	21.07	17.16	21.34	17.38	21.60	17.60	21.84
46.6	21.00	17.10	21.13	17.21	21.40	17.43	21.66	17.65	21.90
46.7	21.06	17.15	21.19	17.26	21.46	17.48	21.72	17.70	21.96
46.8	21.12	17.20	21.25	17.31	21.52	17.53	21.78	17.75	22.02
46.9	21.18	17.25	21.31	17.36	21.58	17.58	21.84	17.80	22.09
47.0	21.24	17.30	21.37	17.41	21.64	17.63	21.90	17.85	22.15
47.1	21.30	17.35	21.43	17.46	21.70	17.68	21.96	17.90	22.21
47.2	21.36	17.40	21.49	17.51	21.76	17.73	22.02	17.95	22.27
47.3	21.42	17.45	21.55	17.56	21.82	17.78	22.08	18.01	22.33
47.4	21.48	17.50	21.61	17.61	21.88	17.83	22.15	18.06	22.39
47.5	21.54	17.55	21.67	17.66	21.94	17.88	22.21	18.11	22.45
47.6	21.60	17.60	21.73	17.71	22.00	17.94	22.27	18.16	22.51
47.7	21.66	17.65	21.79	17.76	22.06	17.99	22.33	18.21	22.58
47.8	21.72	17.70	21.85	17.81	22.12	18.04	22.39	18.26	22.64
47.9	21.78	17.75	21.91	17.86	22.18	18.09	22.45	18.31	22.70

TABLE.—Continued.

6

21° C.	22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
16.04	19.97	16.24	20.23	16.46	20.49	16.67	20.75	16.90	43.0
16.09	20.03	16.30	20.29	16.51	20.55	16.73	20.82	16.95	43.1
16.14	20.09	16.35	20.35	16.56	20.61	16.78	20.88	17.00	43.2
16.19	20.15	16.40	20.41	16.61	20.68	16.83	20.94	17.05	43.3
16.24	20.21	16.45	20.47	16.66	20.74	16.88	21.01	17.10	43.4
16.29	20.28	16.50	20.53	16.71	20.80	16.93	21.07	17.16	43.5
16.34	20.34	16.55	20.60	16.76	20.86	16.98	21.13	17.21	43.6
16.40	20.40	16.60	20.66	16.82	20.93	17.04	21.19	17.26	43.7
16.45	20.46	16.65	20.72	16.87	20.99	17.09	21.25	17.31	43.8
16.50	20.52	16.70	20.78	16.92	21.05	17.14	21.32	17.36	43.9
16.55	20.58	16.76	20.84	16.97	21.11	17.19	21.38	17.41	44.0
16.60	20.64	16.81	20.90	17.02	21.17	17.24	21.44	17.47	44.1
16.65	20.71	16.86	20.96	17.07	21.23	17.29	21.50	17.52	44.2
16.70	20.77	16.91	21.02	17.12	21.30	17.35	21.57	17.57	44.3
16.75	20.83	16.96	21.09	17.17	21.36	17.40	21.63	17.62	44.4
16.80	20.89	17.01	21.15	17.22	21.42	17.45	21.69	17.67	44.5
16.85	20.95	17.06	21.21	17.28	21.48	17.50	21.75	17.73	44.6
16.90	21.01	17.11	21.27	17.33	21.54	17.55	21.81	17.78	44.7
16.95	21.07	17.16	21.33	17.38	21.60	17.60	21.88	17.83	44.8
17.01	21.13	17.21	21.39	17.43	21.67	17.65	21.94	17.88	44.9
17.06	21.19	17.26	21.45	17.48	21.73	17.71	22.00	17.93	45.0
17.11	21.25	17.31	21.52	17.53	21.79	17.76	22.06	17.98	45.1
17.16	21.31	17.36	21.58	17.58	21.85	17.81	22.13	18.04	45.2
17.21	21.37	17.41	21.64	17.63	21.91	17.86	22.19	18.09	45.3
17.26	21.43	17.46	21.70	17.68	21.98	17.91	22.25	18.14	45.4
17.31	21.49	17.51	21.76	17.73	22.04	17.96	22.32	18.20	45.5
17.36	21.55	17.56	21.82	17.79	22.10	18.02	22.38	18.25	45.6
17.41	21.61	17.61	21.88	17.84	22.16	18.07	22.45	18.30	45.7
17.46	21.67	17.66	21.94	17.89	22.23	18.12	22.51	18.36	45.8
17.51	21.73	17.71	22.01	17.94	22.29	18.17	22.57	18.41	45.9
17.56	21.79	17.76	22.07	17.99	22.35	18.23	22.64	18.47	46.0
17.61	21.85	17.81	22.13	18.04	22.42	18.28	22.70	18.52	46.1
17.66	21.91	17.86	22.19	18.09	22.48	18.33	22.76	18.57	46.2
17.71	21.97	17.91	22.26	18.15	22.54	18.39	22.83	18.63	46.3
17.76	22.03	17.96	22.32	18.20	22.61	18.44	22.89	18.68	46.4
17.81	22.09	18.01	22.38	18.25	22.67	18.49	22.96	18.73	46.5
17.86	22.16	18.06	22.44	18.30	22.73	18.55	23.02	18.79	46.6
17.91	22.22	18.11	22.51	18.36	22.80	18.60	23.08	18.84	46.7
17.96	22.28	18.17	22.57	18.41	22.86	18.65	23.15	18.90	46.8
18.01	22.34	18.22	22.63	18.46	22.92	18.70	23.21	18.95	46.9
18.06	22.41	18.27	22.69	18.51	22.99	18.76	23.28	19.00	47.0
18.11	22.47	18.32	22.76	18.57	23.05	18.81	23.34	19.05	47.1
18.16	22.53	18.38	22.82	18.62	23.12	18.86	23.41	19.11	47.2
18.21	22.59	18.43	22.88	18.67	23.18	18.92	23.47	19.16	47.3
18.26	22.66	18.48	22.94	18.72	23.24	18.97	23.54	19.22	47.4
18.31	22.72	18.53	23.01	18.78	23.31	19.02	23.60	19.27	47.5
18.36	22.78	18.58	23.07	18.83	23.37	19.08	23.67	19.33	47.6
18.42	22.84	18.64	23.13	18.88	23.44	19.13	23.73	19.38	47.7
18.47	22.91	18.69	23.20	18.93	23.50	19.18	23.80	19.43	47.8
18.52	22.97	18.74	23.26	18.99	23.56	19.24	23.86	19.49	47.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
48.0	21.84	17.80	21.97	17.91	22.24	18.14	22.51	18.36	22.76
48.1	21.90	17.85	22.03	17.96	22.30	18.19	22.57	18.41	22.82
48.2	21.96	17.90	22.09	18.01	22.36	18.24	22.63	18.46	22.88
48.3	22.02	17.95	22.15	18.06	22.42	18.29	22.69	18.51	22.94
48.4	22.08	18.00	22.21	18.11	22.48	18.34	22.75	18.56	23.01
48.5	22.14	18.05	22.27	18.16	22.54	18.39	22.81	18.61	23.07
48.6	22.20	18.10	22.33	18.21	22.60	18.44	22.87	18.66	23.13
48.7	22.26	18.15	22.39	18.26	22.66	18.49	22.93	18.71	23.19
48.8	22.32	18.20	22.45	18.31	22.72	18.54	22.99	18.76	23.26
48.9	22.38	18.25	22.51	18.36	22.78	18.59	23.06	18.81	23.32
49.0	22.44	18.30	22.57	18.41	22.84	18.64	23.12	18.86	23.38
49.1	22.50	18.35	22.63	18.46	22.90	18.69	23.18	18.91	23.44
49.2	22.56	18.40	22.69	18.51	22.96	18.74	23.24	18.96	23.51
49.3	22.62	18.45	22.75	18.56	23.02	18.79	23.30	19.02	23.57
49.4	22.68	18.50	22.81	18.61	23.08	18.84	23.36	19.07	23.63
49.5	22.74	18.55	22.87	18.66	23.15	18.89	23.42	19.12	23.69
49.6	22.80	18.60	22.93	18.71	23.21	18.94	23.48	19.17	23.76
49.7	22.86	18.65	22.99	18.76	23.27	18.99	23.55	19.22	23.82
49.8	22.92	18.70	23.05	18.81	23.33	19.04	23.61	19.27	23.88
49.9	22.98	18.75	23.11	18.86	23.39	19.09	23.67	19.32	23.94
50.0	23.04	18.80	23.17	18.91	23.45	19.14	23.73	19.38	24.01
50.1	23.10	18.85	23.23	18.96	23.51	19.19	23.79	19.43	24.07
50.2	23.16	18.90	23.30	19.02	23.57	19.24	23.85	19.48	24.13
50.3	23.22	18.95	23.36	19.07	23.63	19.29	23.91	19.53	24.19
50.4	23.28	19.00	23.42	19.12	23.69	19.35	23.98	19.58	24.26
50.5	23.34	19.05	23.48	19.17	23.75	19.40	24.04	19.63	24.32
50.6	23.40	19.10	23.54	19.22	23.81	19.45	24.10	19.69	24.38
50.7	23.46	19.15	23.60	19.27	23.87	19.50	24.16	19.74	24.45
50.8	23.51	19.20	23.66	19.32	23.93	19.55	24.22	19.79	24.51
50.9	23.57	19.25	23.72	19.37	23.99	19.60	24.28	19.84	24.57
51.0	23.63	19.30	23.78	19.42	24.05	19.65	24.35	19.89	24.64
51.1	23.69	19.35	23.84	19.47	24.12	19.70	24.41	19.94	24.70
51.2	23.75	19.40	23.90	19.52	24.18	19.75	24.47	20.00	24.76
51.3	23.81	19.45	23.96	19.57	24.24	19.80	24.53	20.05	24.82
51.4	23.87	19.50	24.02	19.62	24.30	19.85	24.59	20.10	24.89
51.5	23.93	19.55	24.08	19.67	24.36	19.90	24.65	20.15	24.95
51.6	23.99	19.60	24.14	19.72	24.42	19.95	24.72	20.20	25.01
51.7	24.05	19.65	24.20	19.77	24.48	20.01	24.78	20.26	25.07
51.8	24.11	19.70	24.26	19.82	24.54	20.06	24.84	20.31	25.14
51.9	24.17	19.75	24.32	19.87	24.60	20.11	24.90	20.36	25.20
52.0	24.23	19.80	24.38	19.92	24.66	20.16	24.96	20.41	25.27
52.1	24.30	19.85	24.44	19.97	24.73	20.21	25.03	20.46	25.33
52.2	24.36	19.90	24.50	20.02	24.79	20.26	25.09	20.52	25.39
52.3	24.42	19.95	24.56	20.07	24.85	20.31	25.15	20.57	25.46
52.4	24.48	20.00	24.62	20.12	24.91	20.37	25.21	20.62	25.52
52.5	24.54	20.05	24.68	20.17	24.97	20.42	25.28	20.67	25.58
52.6	24.60	20.10	24.74	20.22	25.03	20.47	25.34	20.72	25.65
52.7	24.66	20.15	24.80	20.28	25.09	20.52	25.40	20.78	25.71
52.8	24.72	20.20	24.86	20.33	25.15	20.57	25.46	20.83	25.77
52.9	24.78	20.25	24.92	20.38	25.22	20.62	25.53	20.88	25.84

TABLE.—Continued.

6

21° C.	22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
18.57	23.03	18.79	23.32	19.04	23.63	19.29	23.93	19.54	48.0
18.62	23.09	18.84	23.39	19.09	23.69	19.34	23.99	19.60	48.1
18.67	23.16	18.90	23.45	19.14	23.76	19.40	24.06	19.65	48.2
18.72	23.22	18.95	23.52	19.20	23.83	19.45	24.13	19.71	48.3
18.77	23.28	19.00	23.58	19.25	23.89	19.50	24.19	19.76	48.4
18.83	23.35	19.06	23.64	19.31	23.96	19.56	24.26	19.82	48.5
18.88	23.41	19.11	23.71	19.36	24.02	19.61	24.32	19.87	48.6
18.93	23.47	19.16	23.77	19.41	24.08	19.67	24.39	19.92	48.7
18.98	23.54	19.21	23.83	19.47	24.14	19.72	24.45	19.98	48.8
19.03	23.60	19.27	23.90	19.52	24.21	19.77	24.52	20.03	48.9
19.08	23.66	19.32	23.96	19.57	24.27	19.83	24.59	20.09	49.0
19.14	23.73	19.37	24.03	19.63	24.34	19.88	24.65	20.15	49.1
19.19	23.79	19.43	24.09	19.68	24.40	19.94	24.72	20.20	49.2
19.24	23.85	19.48	24.15	19.73	24.47	19.99	24.78	20.26	49.3
19.29	23.92	19.54	24.22	19.79	24.53	20.04	24.85	20.31	49.4
19.35	23.98	19.59	24.28	19.84	24.60	20.10	24.91	20.37	49.5
19.40	24.04	19.64	24.35	19.89	24.66	20.15	24.98	20.42	49.6
19.45	24.11	19.70	24.41	19.95	24.73	20.21	25.05	20.48	49.7
19.51	24.17	19.75	24.48	20.00	24.79	20.27	25.11	20.54	49.8
19.56	24.24	19.80	24.54	20.05	24.86	20.32	25.18	20.59	49.9
19.61	24.30	19.86	24.61	20.11	24.92	20.38	25.25	20.65	50.0
19.66	24.37	19.91	24.67	20.16	24.99	20.43	25.31	20.70	50.1
19.72	24.43	19.96	24.74	20.22	25.05	20.49	25.38	20.76	50.2
19.77	24.49	20.02	24.80	20.27	25.12	20.54	25.45	20.82	50.3
19.82	24.56	20.07	24.86	20.33	25.18	20.60	25.51	20.87	50.4
19.87	24.62	20.12	24.93	20.38	25.25	20.65	25.58	20.93	50.5
19.93	24.69	20.18	24.99	20.44	25.32	20.71	25.65	20.98	50.6
19.98	24.75	20.23	25.06	20.49	25.38	20.76	25.71	21.04	50.7
20.03	24.81	20.29	25.12	20.55	25.45	20.82	25.78	21.10	50.8
20.08	24.88	20.34	25.19	20.60	25.51	20.87	25.85	21.15	50.9
20.14	24.94	20.39	25.25	20.66	25.58	20.93	25.91	21.21	51.0
20.19	25.01	20.45	25.32	20.71	25.64	20.98	25.98	21.27	51.1
20.24	25.07	20.50	25.38	20.77	25.71	21.04	26.05	21.32	51.2
20.30	25.13	20.55	25.45	20.82	25.78	21.09	26.11	21.38	51.3
20.35	25.20	20.61	25.51	20.87	25.84	21.15	26.18	21.44	51.4
20.40	25.26	20.66	25.58	20.93	25.91	21.21	26.25	21.49	51.5
20.46	25.33	20.72	25.64	20.98	25.97	21.26	26.32	21.55	51.6
20.51	25.39	20.77	25.71	21.04	26.04	21.32	26.39	21.61	51.7
20.56	25.46	20.82	25.77	21.09	26.11	21.37	26.45	21.66	51.8
20.61	25.52	20.88	25.84	21.15	26.17	21.43	26.52	21.72	51.9
20.67	25.58	20.93	25.90	21.20	26.24	21.49	26.59	21.78	52.0
20.72	25.65	20.98	25.97	21.26	26.31	21.54	26.66	21.83	52.1
20.77	25.71	21.04	26.03	21.31	26.37	21.60	26.72	21.89	52.2
20.83	25.78	21.09	26.10	21.37	26.44	21.65	26.79	21.95	52.3
20.88	25.84	21.15	26.16	21.42	26.51	21.71	26.86	22.01	52.4
20.93	25.90	21.20	26.23	21.48	26.57	21.77	26.93	22.06	52.5
20.98	25.97	21.26	26.29	21.53	26.64	21.82	26.99	22.12	52.6
21.04	26.03	21.31	26.36	21.59	26.71	21.88	27.06	22.18	52.7
21.09	26.10	21.36	26.42	21.64	26.77	21.93	27.13	22.24	52.8
21.15	26.16	21.42	26.49	21.70	26.84	21.99	27.20	22.29	52.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
53.0	24.84	20.30	24.98	20.43	25.28	20.68	25.59	20.93	25.90
53.1	24.90	20.36	25.04	20.48	25.34	20.73	25.65	20.98	25.96
53.2	24.96	20.41	25.10	20.53	25.40	20.78	25.71	21.04	26.03
53.3	25.02	20.46	25.16	20.58	25.46	20.83	25.77	21.09	26.09
53.4	25.08	20.51	25.23	20.63	25.52	20.88	25.84	21.14	26.15
53.5	25.14	20.56	25.29	20.68	25.59	20.93	25.90	21.20	26.22
53.6	25.20	20.61	25.35	20.74	25.65	20.98	25.96	21.25	26.28
53.7	25.26	20.66	25.41	20.79	25.71	21.04	26.03	21.30	26.35
53.8	25.32	20.71	25.47	20.84	25.77	21.09	26.09	21.36	26.41
53.9	25.38	20.76	25.53	20.89	25.83	21.14	26.15	21.41	26.47
54.0	25.44	20.81	25.59	20.94	25.90	21.19	26.22	21.47	26.54
54.1	25.50	20.86	25.65	20.99	25.96	21.25	26.28	21.52	26.60
54.2	25.56	20.91	25.71	21.04	26.02	21.30	26.34	21.57	26.67
54.3	25.62	20.96	25.77	21.09	26.08	21.35	26.41	21.63	26.73
54.4	25.68	21.02	25.84	21.14	26.14	21.40	26.47	21.68	26.79
54.5	25.75	21.07	25.90	21.20	26.20	21.46	26.53	21.73	26.86
54.6	25.81	21.12	25.96	21.25	26.27	21.51	26.59	21.79	26.92
54.7	25.87	21.17	26.02	21.30	26.33	21.56	26.66	21.84	26.99
54.8	25.93	21.22	26.08	21.35	26.39	21.62	26.72	21.90	27.05
54.9	25.99	21.27	26.14	21.40	26.45	21.67	26.78	21.95	27.11
55.0	26.05	21.32	26.20	21.45	26.52	21.72	26.85	22.00	27.18
55.1	26.11	21.37	26.26	21.51	26.58	21.77	26.91	22.05	27.24
55.2	26.17	21.43	26.32	21.56	26.64	21.83	26.97	22.11	27.31
55.3	26.23	21.48	26.38	21.61	26.70	21.88	27.04	22.16	27.37
55.4	26.29	21.53	26.45	21.66	26.76	21.93	27.10	22.21	27.43
55.5	26.35	21.58	26.51	21.71	26.83	21.98	27.16	22.26	27.49
55.6	26.41	21.63	26.57	21.76	26.89	22.04	27.23	22.32	27.55
55.7	26.47	21.68	26.63	21.81	26.95	22.09	27.29	22.37	27.62
55.8	26.53	21.73	26.69	21.87	27.01	22.14	27.35	22.42	27.69
55.9	26.59	21.79	26.75	21.92	27.07	22.19	27.41	22.48	27.75
56.0	26.65	21.84	26.81	21.97	27.14	22.24	27.48	22.53	27.82
56.1	26.72	21.89	26.87	22.02	27.20	22.30	27.54	22.58	27.88
56.2	26.78	21.94	26.93	22.07	27.26	22.35	27.60	22.64	27.94
56.3	26.84	21.99	26.99	22.12	27.32	22.40	27.66	22.69	28.01
56.4	26.90	22.04	27.05	22.18	27.38	22.45	27.73	22.74	28.07
56.5	26.96	22.09	27.12	22.23	27.44	22.50	27.79	22.79	28.14
56.6	27.02	22.14	27.18	22.28	27.51	22.56	27.85	22.85	28.20
56.7	27.08	22.19	27.24	22.33	27.57	22.61	27.91	22.90	28.26
56.8	27.14	22.25	27.30	22.38	27.63	22.66	27.98	22.95	28.33
56.9	27.20	22.30	27.36	22.43	27.69	22.71	28.04	23.01	28.39
57.0	27.26	22.35	27.42	22.48	27.75	22.77	28.10	23.06	28.46
57.1	27.32	22.40	27.48	22.54	27.81	22.82	28.16	23.11	28.52
57.2	27.38	22.45	27.54	22.59	27.88	22.87	28.23	23.17	28.59
57.3	27.44	22.50	27.60	22.64	27.94	22.92	28.29	23.22	28.65
57.4	27.50	22.55	27.66	22.69	28.00	22.97	28.35	23.27	28.72
57.5	27.56	22.61	27.73	22.74	28.06	23.03	28.42	23.33	28.78
57.6	27.62	22.66	27.79	22.79	28.13	23.08	28.48	23.38	28.85
57.7	27.68	22.71	27.85	22.85	28.19	23.13	28.54	23.43	28.91
57.8	27.75	22.76	27.91	22.90	28.25	23.19	28.60	23.49	28.97
57.9	27.81	22.81	27.97	22.95	28.31	23.24	28.67	23.54	29.04

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight		
21.20	26.23	21.47	26.56	21.75	26.91	22.05	27.27	22.35	53.0	
21.25	26.29	21.53	26.62	21.81	26.97	22.10	27.33	22.41	53.1	
21.31	26.35	21.58	26.69	21.86	27.04	22.16	27.40	22.47	53.2	
21.36	26.42	21.64	26.75	21.92	27.11	22.22	27.47	22.52	53.3	
21.42	26.48	21.69	26.82	21.97	27.17	22.27	27.54	22.58	53.4	
21.47	26.55	21.74	26.88	22.03	27.24	22.33	27.61	22.64	53.5	
21.52	26.61	21.80	26.95	22.08	27.31	22.39	27.67	22.70	53.6	
21.58	26.68	21.85	27.01	22.14	27.38	22.44	27.74	22.75	53.7	
21.63	26.74	21.91	27.08	22.20	27.44	22.50	27.81	22.81	53.8	
21.69	26.81	21.96	27.15	22.25	27.51	22.56	27.88	22.87	53.9	
21.74	26.87	22.02	27.21	22.31	27.58	22.61	27.95	22.93	54.0	
21.79	26.94	22.07	27.28	22.37	27.64	22.67	28.01	22.98	54.1	
21.85	27.00	22.13	27.35	22.42	27.71	22.73	28.08	23.04	54.2	
21.90	27.07	22.18	27.41	22.48	27.78	22.78	28.15	23.10	54.3	
21.96	27.13	22.24	27.48	22.53	27.85	22.84	28.22	23.16	54.4	
22.01	27.20	22.29	27.55	22.59	27.91	22.90	28.29	23.22	54.5	
22.06	27.26	22.35	27.61	22.65	27.98	22.95	28.36	23.27	54.6	
22.12	27.33	22.40	27.68	22.70	28.05	23.01	28.43	23.33	54.7	
22.17	27.39	22.46	27.75	22.76	28.11	23.07	28.49	23.39	54.8	
22.23	27.46	22.51	27.81	22.81	28.18	23.13	28.56	23.45	54.9	
22.28	27.52	22.57	27.88	22.87	28.25	23.18	28.63	23.51	55.0	
22.33	27.59	22.63	27.95	22.93	28.32	23.24	28.70	23.56	55.1	
22.39	27.65	22.68	28.01	22.98	28.38	23.30	28.77	23.62	55.2	
22.44	27.72	22.74	28.08	23.04	28.45	23.35	28.84	23.68	55.3	
22.49	27.78	22.79	28.15	23.10	28.52	23.41	28.90	23.74	55.4	
22.55	27.85	22.85	28.21	23.15	28.58	23.47	28.97	23.80	55.5	
22.60	27.92	22.90	28.28	23.21	28.65	23.53	29.04	23.86	55.6	
22.66	27.98	22.96	28.34	23.26	28.72	23.58	29.11	23.91	55.7	
22.71	28.05	23.01	28.41	23.32	28.78	23.64	29.18	23.97	55.8	
22.76	28.11	23.07	28.48	23.38	28.85	23.70	29.24	24.03	55.9	
22.82	28.18	23.12	28.54	23.43	28.92	23.75	29.31	24.09	56.0	
22.87	28.24	23.18	28.61	23.49	28.99	23.81	29.38	24.14	56.1	
22.92	28.31	23.23	28.68	23.54	29.05	23.87	29.45	24.20	56.2	
22.98	28.37	23.29	28.74	23.60	29.12	23.93	29.52	24.26	56.3	
23.03	28.44	23.34	28.81	23.66	29.19	23.98	29.58	24.32	56.4	
23.09	28.50	23.40	28.87	23.71	29.26	24.04	29.65	24.38	56.5	
23.14	28.56	23.45	28.94	23.77	29.32	24.10	29.72	24.43	56.6	
23.20	28.63	23.51	29.01	23.83	29.39	24.15	29.79	24.49	56.7	
23.25	28.69	23.56	29.07	23.88	29.46	24.21	29.86	24.55	56.8	
23.31	28.76	23.62	29.14	23.94	29.53	24.27	29.93	24.61	56.9	
23.36	28.82	23.67	29.20	23.99	29.59	24.32	29.99	24.66	57.0	
23.42	28.89	23.73	29.27	24.05	29.66	24.38	30.06	24.72	57.1	
23.47	28.95	23.78	29.34	24.11	29.73	24.44	30.13	24.78	55.2	
23.52	29.02	23.84	29.40	24.16	29.80	24.49	30.20	24.84	55.3	
23.58	29.08	23.90	29.47	24.22	29.86	24.55	30.27	24.90	57.4	
23.63	29.15	23.95	29.53	24.27	29.93	24.61	30.34	24.95	57.5	
23.69	29.21	24.01	29.60	24.33	30.00	24.66	30.41	25.01	57.6	
23.74	29.28	24.06	29.66	24.39	30.07	24.72	30.48	25.07	57.7	
23.80	29.34	24.12	29.73	24.44	30.14	24.78	30.55	25.13	57.8	
23.85	29.41	24.17	29.80	24.50	30.20	24.83	30.62	25.19	57.9	

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
58.0	27.87	22.86	28.03	23.00	28.38	23.29	28.73	23.59	29.10
58.1	27.93	22.91	28.09	23.05	28.44	23.34	28.79	23.65	29.17
58.2	27.99	22.96	28.15	23.10	28.50	23.40	28.86	23.70	29.23
58.3	28.05	23.02	28.21	23.16	28.56	23.45	28.92	23.75	29.29
58.4	28.11	23.07	28.28	23.21	28.62	23.50	28.98	23.81	29.36
58.5	28.17	23.12	28.34	23.26	28.69	23.56	29.04	23.86	29.42
58.6	28.23	23.17	28.40	23.31	28.75	23.61	29.11	23.91	29.48
58.7	28.29	23.22	28.46	23.36	28.81	23.66	29.17	23.97	29.55
58.8	28.35	23.27	28.52	23.41	28.88	23.71	29.23	24.02	29.61
58.9	28.41	23.32	28.58	23.47	28.94	23.77	29.30	24.08	29.68
59.0	28.47	23.37	28.64	23.52	29.00	23.82	29.36	24.13	29.74
59.1	28.53	23.43	28.71	23.57	29.06	23.87	29.42	24.18	29.80
59.2	28.59	23.48	28.77	23.62	29.12	23.93	29.49	24.24	29.87
59.3	28.65	23.53	28.83	23.67	29.19	23.98	29.55	24.29	29.93
59.4	28.71	23.58	28.89	23.73	29.25	24.03	29.61	24.34	29.99
59.5	28.78	23.63	28.95	23.78	29.31	24.08	29.68	24.40	30.06
59.6	28.84	23.68	29.01	23.83	29.37	24.14	29.74	24.45	30.13
59.7	28.90	23.73	29.07	23.88	29.43	24.19	29.80	24.50	30.19
59.8	28.96	23.79	29.13	23.93	29.50	24.24	29.87	24.56	30.26
59.9	29.02	23.84	29.20	23.98	29.56	24.30	29.93	24.61	30.32
60.0	29.08	23.89	29.26	24.04	29.62	24.35	29.99	24.67	30.39
60.1	29.14	23.94	29.32	24.09	29.68	24.40	30.06	24.72	30.45
60.2	29.20	23.99	29.38	24.14	29.74	24.46	30.12	24.77	30.52
60.3	29.26	24.04	29.44	24.19	29.81	24.51	30.19	24.83	30.59
60.4	29.32	24.10	29.50	24.25	29.87	24.56	30.25	24.88	30.65
60.5	29.38	24.15	29.56	24.30	29.93	24.61	30.32	24.94	30.72
60.6	29.45	24.20	29.63	24.35	29.99	24.67	30.38	24.99	30.78
60.7	29.51	24.25	29.69	24.40	30.06	24.72	30.45	25.04	30.85
60.8	29.57	24.30	29.75	24.46	30.12	24.77	30.51	25.10	30.91
60.9	29.63	24.35	29.81	24.51	30.18	24.83	30.57	25.16	30.98
61.0	29.69	24.41	29.87	24.56	30.25	24.88	30.64	25.21	31.05
61.1	29.75	24.46	29.93	24.61	30.31	24.93	30.70	25.27	31.11
61.2	29.81	24.51	29.99	24.66	30.38	24.98	30.77	25.32	31.18
61.3	29.87	24.56	30.06	24.72	30.44	25.04	30.83	25.38	31.25
61.4	29.93	24.61	30.12	24.77	30.50	25.09	30.90	25.44	31.32
61.5	29.99	24.67	30.18	24.82	30.57	25.15	30.96	25.49	31.39
61.6	30.06	24.72	30.25	24.87	30.63	25.20	31.03	25.55	31.45
61.7	30.12	24.77	30.31	24.93	30.69	25.26	31.09	25.60	31.52
61.8	30.18	24.82	30.37	24.98	30.76	25.31	31.16	25.66	31.59
61.9	30.25	24.88	30.44	25.03	30.82	25.37	31.23	25.71	31.66
62.0	30.31	24.93	30.50	25.09	30.89	25.43	31.29	25.77	31.72
62.1	30.37	24.98	30.56	25.14	30.95	25.48	31.36	25.83	31.79
62.2	30.43	25.03	30.63	25.20	31.01	25.54	31.43	25.88	31.86
62.3	30.50	25.09	30.69	25.25	31.08	25.59	31.49	25.94	31.93
62.4	30.56	25.14	30.75	25.31	31.14	25.65	31.56	25.99	31.99
62.5	30.62	25.20	30.82	25.36	31.21	25.70	31.63	26.05	32.06
62.6	30.69	25.25	30.88	25.42	31.28	25.76	31.69	26.11	32.13
62.7	30.75	25.31	30.94	25.47	31.34	25.81	31.76	26.17	32.20
62.8	30.81	25.36	31.01	25.53	31.41	25.87	31.83	26.23	32.27
62.9	30.87	25.42	31.07	25.58	31.47	25.92	31.89	26.29	32.34

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight		Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
23.91		29.47	24.23	29.87	24.55	30.27	24.89	30.69	25.25	58.0
23.96		29.54	24.28	29.93	24.61	30.34	24.95	30.76	25.31	58.1
24.02		29.60	24.34	29.99	24.66	30.41	25.01	30.83	25.37	58.2
24.07		29.67	24.39	30.06	24.72	30.48	25.07	30.90	25.43	58.3
24.13		29.73	24.45	30.13	24.78	30.54	25.13	30.97	25.49	58.4
24.18		29.80	24.50	30.20	24.83	30.61	25.18	31.04	25.55	58.5
24.23		29.86	24.56	30.26	24.89	30.68	25.23	31.11	25.61	58.6
24.29		29.93	24.61	30.33	24.94	30.75	25.29	31.18	25.67	58.7
24.34		29.99	24.67	30.40	25.00	30.82	25.35	31.25	25.73	58.8
24.40		30.06	24.72	30.47	25.06	30.88	25.41	31.32	25.80	58.9
24.45		30.13	24.78	30.53	25.12	30.95	25.47	31.40	25.86	59.0
24.51		30.19	24.83	30.60	25.18	31.02	25.53	31.47	25.92	59.1
24.56		30.26	24.89	30.67	25.24	31.09	25.59	31.54	25.98	59.2
24.61		30.33	24.95	30.74	25.30	31.16	25.65	31.61	26.04	59.3
24.67		30.39	25.00	30.81	25.36	31.23	25.71	31.68	26.10	59.4
24.72		30.46	25.06	30.87	25.41	31.30	25.77	31.76	26.16	59.5
24.78		30.53	25.11	30.94	25.47	31.38	25.83	31.83	26.23	59.6
24.83		30.59	25.17	31.01	25.53	31.45	25.89	31.90	26.29	59.7
24.89		30.66	25.23	31.08	25.59	31.52	25.95	31.97	26.35	59.8
24.94		30.73	25.29	31.15	25.65	31.59	26.02	32.04	26.42	59.9
24.99		30.79	25.34	31.22	25.71	31.66	26.08	32.12	26.48	60.0
25.05		30.86	25.40	31.29	25.77	31.73	26.14	32.19	26.54	60.1
25.11		30.93	25.46	31.36	25.83	31.80	26.20	32.27	26.61	60.2
25.16		30.99	25.52	31.43	25.89	31.87	26.27	32.34	26.67	60.3
25.22		31.06	25.57	31.50	25.95	31.94	26.33	32.41	26.73	60.4
25.28		31.13	25.63	31.57	26.01	32.02	26.39	32.49	26.80	60.5
25.34		31.20	25.69	31.64	26.07	32.09	26.45	32.56	26.86	60.6
25.39		31.27	25.75	31.71	26.13	32.16	26.52	32.64	26.92	60.7
25.45		31.33	25.80	31.78	26.19	32.23	26.58	32.71	26.99	60.8
25.51		31.40	25.86	31.85	26.25	32.30	26.64	32.78	27.05	60.9
25.56		31.47	25.92	31.92	26.31	32.38	26.70	32.86	27.12	61.0
25.62		31.54	25.98	31.99	26.37	32.45	26.76	32.93	27.18	61.1
25.68		31.61	26.04	32.06	26.43	32.52	26.83	33.01	27.24	61.2
25.73		31.67	26.10	32.13	26.49	32.59	26.89	33.08	27.31	61.3
25.79		31.74	26.16	32.20	26.55	32.67	26.95	33.16	27.37	61.4
25.85		31.81	26.22	32.27	26.61	32.74	27.01	33.23	27.44	61.5
25.90		31.88	26.28	32.34	26.67	32.81	27.08	33.31	27.50	61.6
25.96		31.95	26.34	32.41	26.73	32.88	27.14	33.38	27.56	61.7
26.02		32.01	26.40	32.49	26.79	32.96	27.20	33.46	27.63	61.8
26.08		32.09	26.46	32.56	26.85	33.03	27.27	33.53	27.69	61.9
26.14		32.16	26.51	32.63	26.92	33.10	27.33	33.60	27.76	62.0
26.20		32.23	26.57	32.70	26.98	33.18	27.39	33.68	27.82	62.1
26.25		32.30	26.63	32.77	27.04	33.25	27.46	33.75	27.88	62.2
26.31		32.37	26.69	32.84	27.10	33.33	27.52	33.83	27.95	62.3
26.37		32.44	26.75	32.91	27.16	33.40	27.58	33.90	28.01	62.4
26.43		32.51	26.81	32.99	27.23	33.47	27.65	33.98	28.08	62.5
26.49		32.58	26.87	33.06	27.29	33.55	27.71	34.05	28.15	62.6
26.55		32.65	26.93	33.13	27.35	33.62	27.77	34.13	28.22	62.7
26.61		32.72	26.99	33.20	27.41	33.70	27.84	34.21	28.28	62.8
26.67		32.79	27.06	33.28	27.48	33.77	27.90	34.29	28.35	62.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
63.0	30.94	25.47	31.14	25.64	31.54	25.98	31.96	26.35	32.41
63.1	31.00	25.52	31.20	25.69	31.61	26.03	32.03	26.41	32.48
63.2	31.06	25.58	31.26	25.75	31.67	26.09	32.10	26.46	32.55
63.3	31.13	25.63	31.33	25.80	31.74	26.15	32.17	26.52	32.62
63.4	31.19	25.69	31.39	25.86	31.80	26.21	32.23	26.58	32.69
63.5	31.26	25.74	31.46	25.91	31.87	26.26	32.30	26.64	32.76
63.6	31.32	25.80	31.52	25.97	31.93	26.32	32.37	26.70	32.83
63.7	31.38	25.85	31.59	26.02	32.00	26.38	32.44	26.76	32.90
63.8	31.45	25.91	31.65	26.08	32.07	26.44	32.51	26.82	32.97
63.9	31.51	25.96	31.72	26.13	32.13	26.49	32.58	26.88	33.04
64.0	31.58	26.02	31.78	26.19	32.20	26.55	32.65	26.94	33.11
64.1	31.64	26.07	31.85	26.25	32.27	26.61	32.72	26.99	33.18
64.2	31.70	26.13	31.91	26.30	32.34	26.67	32.79	27.05	33.25
64.3	31.77	26.18	31.97	26.36	32.40	26.72	32.86	27.11	33.32
64.4	31.83	26.24	32.04	26.41	32.47	26.78	32.92	27.17	33.39
64.5	31.90	26.29	32.11	26.47	32.54	26.84	32.99	27.23	33.46
64.6	31.96	26.35	32.17	26.53	32.60	26.90	33.06	27.29	33.53
64.7	32.03	26.40	32.24	26.58	32.67	26.95	33.13	27.35	33.60
64.8	32.09	26.46	32.30	26.64	32.74	27.01	33.20	27.41	33.67
64.9	32.16	26.51	32.37	26.69	32.81	27.07	33.27	27.47	33.74
65.0	32.22	26.57	32.43	26.75	32.87	27.13	33.34	27.53	33.82
65.1	32.29	26.63	32.50	26.80	32.94	27.19	33.41	27.59	33.89
65.2	32.35	26.68	32.57	26.86	33.01	27.25	33.48	27.65	33.96
65.3	32.42	26.74	32.63	26.92	33.08	27.31	33.55	27.71	34.03
65.4	32.48	26.79	32.70	26.97	33.15	27.37	33.62	27.77	34.10
65.5	32.55	26.85	32.76	27.03	33.22	27.43	33.69	27.83	34.18
65.6	32.61	26.90	32.83	27.09	33.28	27.49	33.76	27.89	34.25
65.7	32.68	26.96	32.89	27.15	33.35	27.54	33.83	27.95	34.32
65.8	32.75	27.01	32.96	27.21	33.42	27.60	33.90	28.01	34.40
65.9	32.81	27.07	33.03	27.26	33.49	27.66	33.97	28.07	34.47
66.0	32.88	27.13	33.10	27.32	33.56	27.72	34.04	28.13	34.54
66.1	32.94	27.19	33.17	27.38	33.63	27.78	34.11	28.19	34.62
66.2	33.01	27.25	33.23	27.44	33.70	27.84	34.18	28.26	34.69
66.3	33.07	27.30	33.30	27.50	33.77	27.90	34.25	28.32	34.76
66.4	33.14	27.36	33.37	27.56	33.84	27.96	34.33	28.38	34.84
66.5	33.21	27.42	33.44	27.62	33.91	28.02	34.40	28.45	34.91
66.6	33.28	27.48	33.51	27.68	33.98	28.08	34.47	28.51	34.99
66.7	33.35	27.54	33.58	27.73	34.05	28.14	34.54	28.57	35.06
66.8	33.41	27.60	33.65	27.79	34.12	28.20	34.62	28.64	35.14
66.9	33.48	27.65	33.72	27.85	34.19	28.27	34.69	28.70	35.21
67.0	33.55	27.71	33.79	27.91	34.26	28.33	34.76	28.76	35.29
67.1	33.62	27.77	33.86	27.97	34.34	28.39	34.83	28.82	35.37
67.2	33.69	27.83	33.92	28.03	34.41	28.45	34.91	28.89	35.44
67.3	33.76	27.89	33.99	28.09	34.48	28.52	34.98	28.95	35.52
67.4	33.82	27.95	34.06	28.15	34.55	28.58	35.05	29.01	35.60
67.5	33.89	28.01	34.13	28.21	34.62	28.64	35.13	29.08	35.67
67.6	33.96	28.06	34.20	28.27	34.69	28.70	35.20	29.14	35.75
67.7	34.03	28.12	34.27	28.34	34.76	28.76	35.28	29.21	35.82
67.8	34.09	28.18	34.34	28.40	34.84	28.83	35.35	29.27	35.90
67.9	34.16	28.24	34.41	28.46	34.91	28.89	35.43	29.34	35.98

TABLE.—Continued.

6

21° C.			22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
26.73	32.87	27.12	33.35	27.54	33.84	27.96	34.36	28.42	63.0		
26.79	32.94	27.18	33.42	27.60	33.92	28.03	34.44	28.49	63.1		
26.85	33.01	27.24	33.50	27.66	33.99	28.09	34.52	28.55	63.2		
26.91	33.08	27.31	33.57	27.72	34.07	28.16	34.60	28.62	63.3		
26.96	33.15	27.37	33.64	27.79	34.15	28.23	34.67	28.69	63.4		
27.02	33.22	27.43	33.72	27.85	34.22	28.29	34.75	28.76	63.5		
27.09	33.30	27.49	33.79	27.91	34.30	28.36	34.83	28.82	63.6		
27.15	33.37	27.56	33.86	27.98	34.38	28.43	34.91	28.89	63.7		
27.21	33.44	27.62	33.93	28.04	34.45	28.49	34.98	28.96	63.8		
27.27	33.51	27.68	34.01	28.11	34.53	28.56	35.07	29.03	63.9		
27.33	33.59	27.74	34.08	28.17	34.61	28.63	35.15	29.10	64.0		
27.39	33.66	27.81	34.16	28.24	34.68	28.69	35.23	29.17	64.1		
27.45	33.73	27.87	34.23	28.30	34.76	28.76	35.31	29.24	64.2		
27.51	33.80	27.93	34.31	28.37	34.84	28.83	35.39	29.31	64.3		
27.57	33.88	27.99	34.39	28.43	34.92	28.89	35.48	29.38	64.4		
27.63	33.95	28.06	34.46	28.50	34.99	28.96	35.56	29.45	64.5		
27.70	34.02	28.12	34.54	28.57	35.07	29.03	35.64	29.52	64.6		
27.76	34.10	28.19	34.61	28.63	35.15	29.10	35.72	29.60	64.7		
27.82	34.17	28.25	34.69	28.70	35.23	29.17	35.80	29.67	64.8		
27.88	34.24	28.31	34.76	28.76	35.31	29.24	35.89	29.74	64.9		
27.94	34.32	28.38	34.84	28.83	35.39	29.31	35.97	29.81	65.0		
28.00	34.39	28.44	34.92	28.89	35.47	29.38	36.05	29.88	65.1		
28.06	34.47	28.51	34.99	28.96	35.55	29.44	36.13	29.95	65.2		
28.13	34.54	28.57	35.07	29.03	35.63	29.51	36.21	30.02	65.3		
28.19	34.61	28.63	35.15	29.10	35.71	29.58	36.30	30.10	65.4		
28.25	34.69	28.70	35.23	29.16	35.79	29.65	36.38	30.17	65.5		
28.32	34.76	28.76	35.30	29.23	35.87	29.72	36.46	30.24	65.6		
28.38	34.84	28.83	35.38	29.30	35.94	29.79	36.55	30.32	65.7		
28.45	34.91	28.89	35.46	29.37	36.02	29.86	36.63	30.39	65.8		
28.51	34.99	28.96	35.54	29.44	36.11	29.93	36.71	30.46	65.9		
28.57	35.06	29.02	35.62	29.51	36.19	30.00	36.79	30.54	66.0		
28.64	35.14	29.09	35.70	29.58	36.27	30.07	36.88	30.61	66.1		
28.70	35.22	29.16	35.77	29.64	36.35	30.15	36.96	30.68	66.2		
28.76	35.30	29.23	35.85	29.71	36.43	30.22	37.04	30.76	66.3		
28.83	35.38	29.29	35.93	29.78	36.52	30.29	37.13	30.83	66.4		
28.89	35.45	29.36	36.01	29.85	36.60	30.36	37.22	30.90	66.5		
28.96	35.53	29.43	36.09	29.92	36.68	30.43	37.30	30.98	66.6		
29.03	35.61	29.50	36.17	29.99	36.76	30.51	37.39	31.05	66.7		
29.09	35.69	29.57	36.25	30.06	36.84	30.58	37.48	31.13	66.8		
29.15	35.77	29.64	36.33	30.13	36.93	30.65	37.57	31.21	66.9		
29.22	35.84	29.71	36.41	30.20	37.01	30.72	37.65	31.28	67.0		
29.29	35.92	29.77	36.49	30.27	37.09	30.80	37.74	31.36	67.1		
29.35	36.00	29.84	36.57	30.34	37.18	30.87	37.83	31.44	67.2		
29.42	36.08	29.91	36.65	30.41	37.26	30.94	37.91	31.51	67.3		
29.49	36.16	29.98	36.73	30.49	37.35	31.02	38.00	31.59	67.4		
29.55	36.24	30.05	36.81	30.56	37.44	31.09	38.09	31.65	67.5		
29.62	36.32	30.12	36.90	30.63	37.52	31.17	38.18	31.74	67.6		
29.69	36.40	30.19	36.98	30.70	37.61	31.24	38.26	31.82	67.7		
29.75	36.48	30.26	37.06	30.77	37.69	31.32	38.35	31.89	67.8		
29.82	36.56	30.33	37.14	30.84	37.78	31.39	38.44	31.97	67.9		

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
68.0	34.23	28.30	34.48	28.52	34.98	28.95	35.50	29.41	36.05
68.1	34.30	28.36	34.55	28.58	35.05	29.01	35.57	29.47	36.13
68.2	34.36	28.42	34.62	28.64	35.13	29.08	35.65	29.54	36.21
68.3	34.43	28.48	34.69	28.70	35.20	29.14	35.72	29.60	36.29
68.4	34.50	28.54	34.76	28.76	35.27	29.21	35.80	29.67	36.37
68.5	34.57	28.59	34.83	28.82	35.35	29.27	35.87	29.73	36.45
68.6	34.64	28.65	34.90	28.88	35.42	29.33	35.95	29.80	36.52
68.7	34.70	28.71	34.97	28.95	35.49	29.40	36.02	29.86	36.60
68.8	34.77	28.77	35.04	29.01	35.57	29.46	36.10	29.93	36.68
68.9	34.84	28.83	35.12	29.07	35.64	29.53	36.18	29.99	36.76
69.0	34.91	28.89	35.19	29.13	35.71	29.59	36.25	30.06	36.84
69.1	34.97	28.95	35.26	29.19	35.79	29.65	36.33	30.13	36.91
69.2	35.04	29.01	35.33	29.26	35.86	29.72	36.41	30.20	36.99
69.3	35.12	29.07	35.40	29.32	35.93	29.78	36.48	30.27	37.07
69.4	35.19	29.14	35.47	29.38	36.01	29.85	36.56	30.33	37.15
69.5	35.27	29.20	35.55	29.45	36.08	29.91	36.64	30.40	37.24
69.6	35.34	29.26	35.62	29.51	36.16	29.97	36.72	30.47	37.32
69.7	35.41	29.33	35.69	29.57	36.23	30.04	36.79	30.54	37.40
69.8	35.49	29.39	35.76	29.64	36.31	30.11	36.87	30.61	37.48
69.9	35.56	29.46	35.83	29.70	36.39	30.17	36.95	30.67	37.56
70.0	35.64	29.52	35.91	29.76	36.46	30.24	37.02	30.74	37.64
70.1	35.71	29.59	35.98	29.82	36.54	30.31	37.10	30.81	37.72
70.2	35.78	29.65	36.05	29.89	36.61	30.38	37.19	30.88	37.80
70.3	35.86	29.72	36.13	29.95	36.69	30.44	37.27	30.95	37.89
70.4	35.93	29.78	36.20	30.01	36.76	30.51	37.35	31.01	37.97
70.5	36.01	29.85	36.28	30.08	36.84	30.58	37.43	31.09	38.05
70.6	36.08	29.91	36.35	30.15	36.92	30.64	37.51	31.16	38.13
70.7	36.16	29.97	36.43	30.21	36.99	30.71	37.59	31.23	38.22
70.8	36.23	30.04	36.50	30.28	37.07	30.78	37.67	31.30	38.30
70.9	36.31	30.11	36.58	30.35	37.15	30.85	37.75	31.37	38.38
71.0	36.38	30.17	36.65	30.41	37.23	30.91	37.83	31.44	38.47
71.1	36.46	30.24	36.73	30.48	37.31	30.98	37.91	31.51	38.55
71.2	36.53	30.30	36.80	30.55	37.39	31.05	37.99	31.59	38.63
71.3	36.60	30.37	36.88	30.61	37.47	31.12	38.07	31.66	38.72
71.4	36.68	30.44	36.95	30.68	37.55	31.19	38.16	31.73	38.80
71.5	36.75	30.50	37.03	30.75	37.63	31.26	38.24	31.80	38.88
71.6	36.83	30.57	37.11	30.81	37.71	31.33	38.32	31.87	38.97
71.7	36.90	30.64	37.19	30.88	37.79	31.40	38.40	31.94	39.05
71.8	36.98	30.70	37.27	30.95	37.87	31.47	38.49	32.01	39.14
71.9	37.05	30.77	37.34	31.01	37.94	31.54	38.57	32.09	39.23
72.0	37.13	30.84	37.42	31.08	38.02	31.61	38.65	32.17	39.31
72.1	37.21	30.90	37.50	31.15	38.11	31.68	38.74	32.24	39.40
72.2	37.29	30.97	37.58	31.22	38.19	31.75	38.82	32.32	39.49
72.3	37.36	31.03	37.66	31.29	38.27	31.82	38.90	32.39	39.57
72.4	37.44	31.10	37.73	31.36	38.35	31.89	38.98	32.47	39.66
72.5	37.52	31.17	37.81	31.42	38.43	31.96	39.07	32.54	39.75
72.6	37.60	31.24	37.89	31.49	38.51	32.04	39.16	32.62	39.83
72.7	37.67	31.31	37.97	31.56	38.59	32.11	39.24	32.69	39.92
72.8	37.75	31.37	38.05	31.63	38.67	32.18	39.33	32.77	40.01
72.9	37.83	31.44	38.13	31.70	38.76	32.26	39.41	32.84	40.10

TABLE.—Continued.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight		Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
29.89		36.63	30.40	37.23	30.91	37.86	31.47	38.53	32.05	68.0
29.95		36.71	30.47	37.31	30.99	37.95	31.54	38.61	32.13	68.1
30.02		36.79	30.54	37.39	31.06	38.03	31.62	38.70	32.21	68.2
30.09		36.87	30.61	37.48	31.13	38.12	31.69	38.79	32.29	68.3
30.16		36.95	30.68	37.56	31.21	38.21	31.77	38.88	32.37	68.4
30.23		37.03	30.75	37.65	31.28	38.30	31.84	38.96	32.45	68.5
30.30		37.12	30.82	37.73	31.35	38.38	31.92	39.06	32.53	68.6
30.37		37.20	30.89	37.82	31.43	38.47	31.99	39.15	32.61	68.7
30.43		37.28	30.96	37.90	31.50	38.56	32.07	39.24	32.69	68.8
30.50		37.36	31.03	37.98	31.57	38.64	32.15	39.33	32.77	68.9
30.57		37.45	31.10	38.07	31.65	38.73	32.23	39.43	32.86	69.0
30.64		37.53	31.17	38.15	31.72	38.82	32.31	39.52	32.94	69.1
30.71		37.61	31.25	38.24	31.79	38.90	32.39	39.61	33.02	69.2
30.78		37.69	31.32	38.32	31.87	38.99	32.47	39.70	33.10	69.3
30.85		37.78	31.39	38.41	31.94	39.08	32.55	39.80	33.18	69.4
30.92		37.86	31.46	38.50	32.02	39.17	32.63	39.89	33.26	69.5
30.99		37.94	31.54	38.58	32.09	39.26	32.71	39.98	33.34	69.6
31.06		38.03	31.61	38.67	32.17	39.35	32.78	40.07	33.43	69.7
31.13		38.11	31.68	38.75	32.25	39.45	32.86	40.17	33.51	69.8
31.20		38.19	31.75	38.84	32.33	39.54	32.95	40.26	33.59	69.9
31.27		38.28	31.83	38.92	32.41	39.63	33.02	40.35	33.67	70.0
31.35		38.36	31.90	39.01	32.49	39.72	33.11	40.44	33.75	70.1
31.42		38.45	31.97	39.10	32.57	39.81	33.19	40.53	33.84	70.2
31.49		38.53	32.05	39.19	32.65	39.90	33.27	40.62	33.92	70.3
31.56		38.61	32.12	39.28	32.72	39.99	33.35	40.72	34.00	70.4
31.63		38.70	32.20	39.37	32.80	40.08	33.43	40.81	34.08	70.5
31.70		38.78	32.28	39.46	32.88	40.17	33.51	40.90	34.17	70.6
31.78		38.87	32.36	39.55	32.96	40.26	33.59	40.99	34.25	70.7
31.85		38.95	32.43	39.64	33.04	40.35	33.68	41.08	34.33	70.8
31.92		39.04	32.51	39.73	33.12	40.45	33.76	41.18	34.42	70.9
31.99		39.12	32.59	39.82	33.20	40.54	33.84	41.27	34.50	71.0
32.07		39.21	32.67	39.91	33.28	40.63	33.92	41.36	34.58	71.1
32.15		39.30	32.74	40.00	33.36	40.72	34.00	41.46	34.67	71.2
32.22		39.39	32.82	40.09	33.44	40.81	34.08	41.55	34.75	71.3
32.30		39.48	32.90	40.18	33.52	40.90	34.17	41.64	34.83	71.4
32.37		39.57	32.98	40.27	33.60	40.99	34.25	41.74	34.92	71.5
32.45		39.65	33.05	40.36	33.68	41.08	34.33	41.83	35.00	71.6
32.53		39.74	33.13	40.45	33.76	41.18	34.41	41.93	35.08	71.7
32.60		39.83	33.21	40.54	33.84	41.27	34.50	42.02	35.17	71.8
32.68		39.92	33.29	40.63	33.92	41.36	34.58	42.11	35.25	71.9
32.76		40.01	33.37	40.72	34.00	41.45	34.66	42.21	35.34	72.0
32.83		40.10	33.45	40.81	34.08	41.55	34.74	42.30	35.42	72.1
32.91		40.18	33.52	40.90	34.16	41.64	34.83	42.40	35.51	72.2
32.98		40.27	33.60	40.99	34.24	41.73	34.91	42.49	35.59	72.3
33.06		40.36	33.68	41.08	34.33	41.82	34.99	42.58	35.68	72.4
33.14		40.45	33.76	41.17	34.41	41.92	35.08	42.68	35.76	72.5
33.22		40.54	33.84	41.26	34.49	42.01	35.16	42.77	35.85	72.6
33.29		40.62	33.91	41.35	34.57	42.10	35.24	42.87	35.93	72.7
33.37		40.71	33.99	41.45	34.65	42.19	35.33	42.96	36.02	72.8
33.45		40.80	34.07	41.54	34.73	42.29	35.41	43.06	36.10	72.9

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
73.0	37.91	31.51	38.21	31.77	38.84	32.33	39.50	32.92	40.18
73.1	37.98	31.58	38.29	31.84	38.92	32.40	39.59	32.99	40.27
73.2	38.06	31.65	38.37	31.90	39.00	32.48	39.67	33.07	40.36
73.3	38.14	31.71	38.45	31.97	39.08	32.55	39.76	33.15	40.44
73.4	38.22	31.78	38.53	32.04	39.17	32.62	39.84	33.22	40.53
73.5	38.30	31.85	38.61	32.12	39.25	32.70	39.93	33.30	40.62
73.6	38.38	31.92	38.69	32.19	39.34	32.77	40.02	33.37	40.70
73.7	38.46	31.99	38.77	32.26	39.42	32.85	40.10	33.45	40.79
73.8	38.54	32.06	38.85	32.34	39.50	32.92	40.19	33.53	40.88
73.9	38.62	32.13	38.93	32.41	39.59	32.99	40.28	33.60	40.97
74.0	38.70	32.20	39.01	32.48	39.67	33.07	40.36	33.68	41.05
74.1	38.78	32.27	39.09	32.55	39.76	33.15	40.45	33.76	41.14
74.2	38.86	32.35	39.18	32.63	39.84	33.22	40.53	33.83	41.23
74.3	38.94	32.42	39.26	32.70	39.92	33.30	40.62	33.91	41.32
74.4	39.02	32.49	39.34	32.77	40.01	33.37	40.71	33.98	41.41
74.5	39.10	32.56	39.43	32.85	40.09	33.45	40.79	34.06	41.50
74.6	39.18	32.63	39.51	32.92	40.18	33.53	40.88	34.14	41.59
74.7	39.26	32.70	39.59	32.99	40.27	33.60	40.97	34.22	41.68
74.8	39.35	32.78	39.68	33.07	40.35	33.68	41.05	34.30	41.77
74.9	39.43	32.85	39.76	33.15	40.44	33.76	41.14	34.38	41.86
75.0	39.51	32.92	39.84	33.22	40.53	33.83	41.23	34.46	41.95
75.1	39.60	32.99	39.93	33.30	40.61	33.91	41.32	34.54	42.04
75.2	39.68	33.07	40.01	33.37	40.70	33.98	41.41	34.61	42.13
75.3	39.76	33.15	40.09	33.45	40.78	34.06	41.50	34.69	42.22
75.4	39.84	33.22	40.18	33.53	40.87	34.14	41.58	34.77	42.31
75.5	39.93	33.30	40.27	33.60	40.96	34.22	41.67	34.85	42.40
75.6	40.01	33.37	40.35	33.68	41.04	34.30	41.76	34.93	42.49
75.7	40.09	33.45	40.44	33.76	41.13	34.38	41.85	35.01	42.58
75.8	40.18	33.53	40.53	33.83	41.22	34.45	41.94	35.09	42.67
75.9	40.27	33.60	40.61	33.91	41.31	34.53	42.03	35.17	42.76
76.0	40.35	33.68	40.70	33.98	41.40	34.61	42.12	35.25	42.85
76.1	40.44	33.76	40.78	34.06	41.48	34.68	42.21	35.33	42.95
76.2	40.53	33.83	40.87	34.14	41.57	34.77	42.30	35.41	43.04
76.3	40.61	33.91	40.96	34.22	41.66	34.84	42.39	35.50	43.13
76.4	40.70	33.98	41.04	34.29	41.75	34.92	42.48	35.58	43.22
76.5	40.78	34.06	41.13	34.37	41.83	35.00	42.57	35.66	43.32
76.6	40.87	34.14	41.22	34.45	41.92	35.08	42.66	35.74	43.41
76.7	40.96	34.22	41.30	34.53	42.01	35.16	42.75	35.82	43.50
76.8	41.04	34.29	41.39	34.60	42.10	35.24	42.84	35.90	43.60
76.9	41.13	34.37	41.48	34.68	42.19	35.32	42.93	35.98	43.69
77.0	41.22	34.45	41.57	34.76	42.28	35.40	43.02	36.07	43.79
77.1	41.31	34.52	41.65	34.84	42.37	35.48	43.11	36.15	43.88
77.2	41.39	34.60	41.74	34.91	42.46	35.56	43.20	36.24	43.97
77.3	41.48	34.68	41.83	34.99	42.54	35.64	43.30	36.32	44.07
77.4	41.57	34.75	41.91	35.07	42.63	35.72	43.39	36.40	44.16
77.5	41.66	34.83	42.00	35.15	42.72	35.80	43.48	36.49	44.26
77.6	41.75	34.91	42.09	35.23	42.81	35.88	43.57	36.57	44.35
77.7	41.83	34.98	42.17	35.30	42.90	35.96	43.67	36.66	44.45
77.8	41.92	35.06	42.26	35.38	42.99	36.04	43.76	36.74	44.54
77.9	42.01	35.14	42.35	35.46	43.08	36.13	43.85	36.82	44.64

TABLE.—Continued.

6

21° C.	22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	
33.52	40.88	34.15	41.63	34.81	42.38	35.49	43.15	36.18	73.0
33.60	40.97	34.23	41.72	34.89	42.47	35.58	43.24	36.27	73.1
33.68	41.06	34.31	41.81	34.98	42.56	35.66	43.33	36.35	73.2
33.75	41.15	34.39	41.90	35.06	42.66	35.74	43.43	36.43	73.3
33.83	41.24	34.47	41.99	35.14	42.75	35.83	43.52	36.52	73.4
33.91	41.33	34.55	42.08	35.22	42.84	35.91	43.61	36.60	73.5
33.98	41.42	34.63	42.17	35.31	42.93	35.99	43.70	36.68	73.6
34.06	41.51	34.71	42.27	35.39	43.03	36.08	43.80	36.77	73.7
34.14	41.60	34.79	42.36	35.47	43.12	36.16	43.89	36.85	73.8
34.22	41.69	34.87	42.45	35.55	43.21	36.24	43.98	36.93	73.9
34.30	41.78	34.95	42.54	35.64	43.31	36.33	44.08	37.02	74.0
34.38	41.87	35.03	42.63	35.72	43.40	36.41	44.18	37.11	74.1
34.46	41.96	35.12	42.72	35.80	43.49	36.49	44.28	37.20	74.2
34.54	42.06	35.20	42.82	35.88	43.58	36.58	44.38	37.29	74.3
34.62	42.15	35.28	42.91	35.97	43.68	36.66	44.48	37.38	74.4
34.70	42.24	35.36	43.00	36.05	43.77	36.74	44.57	37.47	74.5
34.78	42.33	35.45	43.09	36.13	43.86	36.83	44.67	37.56	74.6
34.86	42.42	35.53	43.19	36.22	43.95	36.91	44.77	37.65	74.7
34.94	42.51	35.61	43.28	36.30	44.05	36.99	44.87	37.75	74.8
35.02	42.61	35.69	43.37	36.39	44.15	37.08	44.97	37.84	74.9
35.10	42.70	35.78	43.46	36.47	44.25	37.17	45.07	37.93	75.0
35.18	42.79	35.86	43.56	36.55	44.34	37.26	45.18	38.02	75.1
35.26	42.88	35.95	43.65	36.64	44.44	37.35	45.29	38.12	75.2
35.34	42.97	36.03	43.74	36.72	44.53	37.44	45.39	38.21	75.3
35.43	43.07	36.11	43.83	36.81	44.63	37.53	45.50	38.31	75.4
35.51	43.16	36.20	43.92	36.89	44.73	37.62	45.61	38.40	75.5
35.59	43.25	36.28	44.02	36.97	44.83	37.71	45.71	38.50	75.6
35.67	43.35	36.36	44.12	37.06	44.93	37.80	45.82	38.60	75.7
35.75	43.44	36.45	44.21	37.15	45.03	37.89	45.92	38.69	75.8
35.84	43.53	36.53	44.31	37.24	45.13	37.98	46.02	38.79	75.9
35.92	43.63	36.62	44.41	37.33	45.24	38.08	46.12	38.88	76.0
36.00	43.72	36.70	44.50	37.42	45.34	38.17	46.23	38.98	76.1
36.08	43.81	36.79	44.60	37.50	45.44	38.27	46.34	39.08	76.2
36.17	43.91	36.87	44.70	37.59	45.55	38.36	46.45	39.18	76.3
36.25	44.00	36.96	44.80	37.68	45.65	38.46	46.56	39.29	76.4
36.34	44.10	37.04	44.89	37.77	45.75	38.55	46.67	39.39	76.5
36.42	44.19	37.13	44.99	37.86	45.86	38.65	46.78	39.49	76.6
36.51	44.29	37.22	45.09	37.95	45.96	38.74	46.89	39.59	76.7
36.59	44.38	37.30	45.19	38.04	46.07	38.84	47.00	39.69	76.8
36.68	44.48	37.39	45.30	38.13	46.18	38.93	47.11	39.80	76.9
36.76	44.57	37.47	45.40	38.23	46.29	39.03	47.23	39.90	77.0
36.85	44.67	37.56	45.50	38.32	46.40	39.13	47.34	40.00	77.1
36.93	44.76	37.65	45.60	38.42	46.51	39.23	47.45	40.11	77.2
37.02	44.86	37.73	45.70	38.51	46.62	39.34	47.57	40.22	77.3
37.10	44.95	37.82	45.81	38.60	46.73	39.44	47.68	40.32	77.4
37.19	45.05	37.91	45.91	38.70	46.84	39.54	47.80	40.43	77.5
37.28	45.15	37.99	46.01	38.79	46.95	39.64	47.91	40.54	77.6
37.36	45.25	38.08	46.12	38.89	47.06	39.74	48.02	40.65	77.7
37.45	45.35	38.18	46.23	38.98	47.17	39.85	48.14	40.75	77.8
37.53	45.45	38.27	46.34	39.08	47.28	39.95	48.26	40.86	77.9

6

TABLE 17.—ALCOHOL

SCALE READING	17.5° C.		18° C.		19° C.		20° C.		21° C.
	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume
78.0	42.09	35.22	42.43	35.54	43.17	36.21	43.94	36.91	44.73
78.1	42.18	35.30	42.52	35.62	43.27	36.29	44.04	36.99	44.83
78.2	42.26	35.38	42.61	35.70	43.36	36.38	44.13	37.08	44.92
78.3	42.35	35.45	42.70	35.77	43.45	36.46	44.23	37.16	45.02
78.4	42.44	35.53	42.78	35.85	43.54	36.54	44.32	37.25	45.12
78.5	42.52	35.61	42.87	35.93	43.63	36.63	44.42	37.33	45.22
78.6	42.61	35.69	42.96	36.01	43.72	36.71	44.51	37.42	45.32
78.7	42.69	35.77	43.05	36.09	43.82	36.79	44.60	37.50	45.42
78.8	42.78	35.84	43.14	36.17	43.91	36.88	44.70	37.59	45.52
78.9	42.86	35.92	43.23	36.26	44.00	36.96	44.79	37.68	45.62
79.0	42.95	36.00	43.32	36.34	44.09	37.04	44.89	37.76	45.72
79.1	43.04	36.08	43.41	36.42	44.19	37.13	44.98	37.85	45.82
79.2	43.13	36.16	43.50	36.50	44.28	37.21	45.08	37.94	45.92
79.3	43.22	36.25	43.59	36.59	44.38	37.30	45.18	38.02	46.02
79.4	43.31	36.33	43.68	36.67	44.47	37.38	45.28	38.11	46.13
79.5	43.40	36.41	43.77	36.75	44.56	37.47	45.38	38.20	46.24
79.6	43.49	36.49	43.86	36.83	44.65	37.56	45.48	38.30	46.34
79.7	43.58	36.57	43.95	36.92	44.75	37.64	45.58	38.39	46.45
79.8	43.67	36.66	44.05	37.00	44.84	37.73	45.68	38.48	46.56
79.9	43.76	36.74	44.14	37.09	44.93	37.81	45.78	38.57	46.67
80.0	43.85	36.82	44.24	37.17	45.04	37.90	45.88	38.67	46.77

TABLE—Concluded.

6

21° C.		22° C.		23° C.		24° C.		25° C.		SCALE READING
Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight	Per cent by volume	Per cent by weight		
37.62	45.56	38.37	46.45	39.18	47.40	40.05	48.37	40.97	78.0	
37.71	45.66	38.46	46.56	39.29	47.51	40.16	48.49	41.08	78.1	
37.79	45.76	38.56	46.67	39.39	47.63	40.27	48.60	41.18	78.2	
37.88	45.86	38.65	46.78	39.49	47.74	40.37	48.72	41.29	78.3	
37.97	45.96	38.75	46.89	39.59	47.85	40.48	48.84	41.40	78.4	
38.06	46.07	38.84	47.00	39.69	47.97	40.59	48.95	41.51	78.5	
38.15	46.17	38.93	47.11	39.80	48.08	40.69	49.07	41.62	78.6	
38.24	46.28	39.03	47.22	39.90	48.19	40.80	49.19	41.73	78.7	
38.33	46.39	39.13	47.34	40.00	48.31	40.90	49.31	41.84	78.8	
38.43	46.50	39.23	47.45	40.11	48.42	41.01	49.42	41.95	78.9	
38.52	46.61	39.33	47.56	40.21	48.53	41.12	49.54	42.05	79.0	
38.61	46.72	39.43	47.67	40.32	48.65	41.22	49.66	42.16	79.1	
38.70	46.83	39.54	47.79	40.42	48.76	41.33	49.77	42.27	79.2	
38.80	46.93	39.64	47.90	40.53	48.88	41.44	49.89	42.38	79.3	
38.89	47.04	39.74	48.01	40.63	48.99	41.54	50.01	42.49	79.4	
38.98	47.15	39.84	48.12	40.74	49.10	41.65	50.13	42.60	79.5	
39.08	47.26	39.94	48.23	40.84	49.22	41.76	50.24	42.71	79.6	
39.18	47.37	40.04	48.34	40.95	49.33	41.86	50.36	42.82	79.7	
39.28	47.48	40.14	48.46	41.05	49.45	41.97	50.48	42.93	79.8	
39.38	47.59	40.24	48.57	41.16	49.56	42.08	50.59	43.04	79.9	
39.48	47.70	40.35	48.68	41.26	49.68	42.18	50.71	43.15	80.0	

GLYCEROL IN DRY WINES.

7

Method I. (By Direct Weighing)—Tentative.

Evaporate 100 cc. of the wine in a porcelain dish on a water bath to a volume of about 10 cc. and treat the residue with about 5 grams of fine sand and 4-5 cc. of milk of lime (containing about 15% of calcium oxid) for each gram of extract present and evaporate almost to dryness. Treat the moist residue with 50 cc. of 90% alcohol by volume, remove the substance adhering to the sides of the dish with a spatula and rub the whole mass to a paste. Heat the mixture on a water bath, with constant stirring, to incipient boiling and decant the liquid through a filter into a small flask. Wash the residue repeatedly by decantation with 10 cc. portions of hot 90% alcohol until the filtrate amounts to about 150 cc. Evaporate the filtrate to a sirupy consistency in a porcelain dish on a hot, but not boiling, water bath; transfer the residue to a small, glass-stoppered, graduated cylinder with 20 cc. of absolute alcohol and add 3 portions of 10 cc. each of anhydrous ether, shaking thoroughly after each addition. Let stand until clear, then pour off through a filter, and wash the cylinder and filter with a mixture of 1 part of absolute alcohol to 1½ parts of anhydrous ether, also pouring the wash liquor through the filter. Evaporate the filtrate to a sirupy consistency, dry for an hour at the temperature of boiling water, weigh, ignite and weigh again. The loss on ignition gives the weight of glycerol.

8

Method II. (By Ozidation with Dichromate)—Tentative.

Evaporate 100 cc. of the wine in a porcelain dish on a water bath, the temperature of which is maintained at 85°-90°C., to a volume of 10 cc. and treat the residue with about 5 grams of fine sand and 5 cc. of milk of lime (containing the equivalent of 15 grams of calcium oxid). Proceed from this point as directed under XIX, 6, beginning with the clause "evaporate almost to dryness, with frequent stirring". Observe the precautions given concerning the temperature at which all evaporations are to be made.

9

GLYCEROL IN SWEET WINES.—TENTATIVE.

With wines whose extract exceeds 5 grams per 100 cc., heat 100 cc. to boiling in a flask and treat with successive small portions of milk of lime until the wine becomes first darker and then lighter in color. Cool, add 200 cc. of 95% alcohol by volume, allow the precipitate to subside, filter and wash with 95% alcohol. Treat the combined filtrate and washings as directed in 7 or 8.

10

GLYCEROL-ALCOHOL RATIO.—TENTATIVE.

Express this ratio as $x : 100$, in which x is obtained by multiplying the percentage by weight of glycerol by 100 and dividing the result by the percentage of alcohol by weight.

EXTRACT.

11

From the Specific Gravity of the Dealccoholized Wine.—Tentative.

Calculate the specific gravity of the dealcoholized wine by the following formula:

$$S = G + 1 - A \text{ in which}$$

S = specific gravity of the dealcoholized wine;

G = specific gravity of the wine, 3; and

A = specific gravity of the distillate obtained in the determination of alcohol, 4 (a).

From **IX, 9**, ascertain the per cent by weight of extract in the dealcoholized wine corresponding to the value of *S*. Multiply the figure thus obtained by the value of *S* to obtain the grams of extract per 100 cc. of wine.

12**By Evaporation.—Tentative.**

(a) *In dry wines, having an extract content of less than 3 grams per 100 cc.*—Evaporate 50 cc. of the sample on a water bath to a sirupy consistency in a 75 cc. flat-bottomed platinum dish, approximately 85 mm. in diameter. Heat the residue for 2½ hours in a drying oven at the temperature of boiling water, cool in a desiccator and weigh as soon as the dish and contents reach room temperature.

(b) *In sweet wines.*—When the extract content is between 3 and 6 grams per 100 cc., treat 25 cc. of the sample as directed under (a).

When the extract exceeds 6 grams per 100 cc., however, the result, obtained as directed under **11**, is accepted and no gravimetric determination is attempted. This is because of the serious error connected with drying levulose at high temperature.

13**NON-SUGAR SOLIDS.—TENTATIVE.**

Determine the non-sugar solids (sugar-free extract) by subtracting the amount of reducing sugars before inversion, **14**, from the extract, **11** or **12**. If sucrose is present in the wine, determine the non-sugar solids by subtracting the sum of reducing sugars before inversion and the sucrose from the extract.

14**REDUCING SUGARS.—TENTATIVE.**

(a) *Dry wines.*—Place 200 cc. of the wine in a porcelain dish, exactly neutralize with N/1 sodium hydroxid, calculating the amount required from the determination of acidity, **25**, and evaporate to about one fourth the original volume. Transfer to a 200 cc. flask, add sufficient neutral lead acetate solution to clarify, dilute to the mark with water, shake and filter through a folded filter. Remove the lead with dry potassium oxalate and determine reducing sugars as directed under **VIII, 25**.

(b) *Sweet wines.*—In the case of sweet wines approximate the sugar content by subtracting 2 from the result in the determination of the extract and employ such a quantity of the sample that the aliquot taken for the copper reduction shall not exceed 245 mg. of invert sugar. Proceed as directed in (a) except that this smaller quantity of the sample is taken for the determination.

SUCROSE.**15****By Reducing Sugars Before and After Inversion.—Tentative.**

Proceed as directed under **VIII, 18**, using the method given under **VIII, 25**, for the determination of reducing sugars.

16**By Polarization.—Tentative.**

Polarize part of the filtrate, obtained in **14**, before and after inversion in a 200 mm. tube as directed under **VIII, 14** or **16**. In calculating the percentage of sucrose the relation of the amount of sample contained in 100 cc. to the normal weight must be taken into consideration.

17**COMMERCIAL GLUCOSE.—TENTATIVE.**

Polarize a portion of the filtrate, obtained in **14**, after inversion in a 200 mm. jacketed tube at 87°C. as directed under **IX, 25**. In calculating the per-

centage of glucose the relation of the amount of sample contained in 100 cc. to the normal weight for the instrument must be taken into consideration.

18 **ASH.—TENTATIVE.**

Proceed as directed under **VIII, 4**, employing the residue from 50 cc. of the wine.

19 **ASH-EXTRACT RATIO.—TENTATIVE.**

Express results as 1 : x, in which x is the quotient obtained by dividing the grams of extract per 100 cc. by the grams of ash per 100 cc.

20 **ALKALINITY OF THE WATER-SOLUBLE ASH.—TENTATIVE.**

Extract the ash, obtained as directed under **18**, with successive small portions of hot water until the filtrate amounts to about 60 cc. and proceed as directed under **IX, 18**. Express the alkalinity in terms of the number of cc. of N/10 acid required to neutralize the water-soluble ash from 100 cc. of the wine.

21 **ALKALINITY OF THE WATER-INSOLUBLE ASH.—TENTATIVE.**

Ignite the filter and residue from **20** in the platinum dish in which the wine was ashed, and proceed as directed under **IX, 19**. Express the alkalinity in terms of the number of cc. of N/10 acid required to neutralize the water-insoluble ash from 100 cc. of the wine.

22 **PHOSPHORIC ACID.—TENTATIVE.**

Dissolve the ash, obtained as directed under **18**, in 50 cc. of boiling nitric acid (1 to 9), filter, wash the filter and determine phosphoric acid in the combined filtrate and washings, as directed in **I, 6** or **9**. If the ash ignites without difficulty, no free phosphoric acid need be suspected. Should there be any free acid, the ash remains black even after repeated leaching. In such cases calcium acetate or a mixture containing 3 parts of sodium carbonate and 1 of sodium nitrate should be added to avoid loss of phosphoric acid before attempting to ash.

23 **SULPHURIC ACID.—TENTATIVE.**

Precipitate directly the sulphuric acid in 50 cc. of the wine by means of barium chlorid solution, after acidifying with a small excess of hydrochloric acid, and determine the resulting barium sulphate as directed under **II, 20**. Allow the precipitate to stand for at least 6 hours before filtering. Report as sulphur trioxid (SO_3).

24 **CHLORIN.—TENTATIVE.**

To 100 cc. of dry wine or 50 cc. of sweet wine add sufficient sodium carbonate to make distinctly alkaline. Evaporate to dryness, ignite at a heat not above low redness, cool, extract the residue with hot water, acidify the water extract with nitric acid and determine chlorin as directed under **III, 15**.

25 **TOTAL ACIDS.—TENTATIVE.**

Measure 20 cc. of the wine into a 250 cc. beaker, heat rapidly to incipient boiling and immediately titrate with N/10 sodium hydroxid. Determine the end point with neutral 0.05% azolitmin solution as an outside indicator. Place the indicator in the cavities of a spot plate and spot the wine into the azolitmin solution. The end point is reached when the color of the indicator remains unchanged by the addition of a few drops of N/10 alkali to the wine.

In the case of wines which are artificially colored and which cannot be satisfactorily titrated in the above manner, it will be found helpful to use phenolphthalein powder (1 part of phenolphthalein mixed with 100 parts of dry, powdered potassium sulphate) as an indicator. Place this indicator in the cavities of a spot plate and spot the wine into the powder. The end of the titration is indicated when the powder acquires a pink tint.

Express the result in terms of tartaric acid. One cc. of N/10 sodium hydroxid is equivalent to 0.0075 gram of tartaric acid.

VOLATILE ACIDS.

26

Method I.—Tentative.

Heat rapidly to incipient boiling 50 cc. of the wine in a 500 cc. distillation flask and pass steam through until 15 cc. of the distillate require only 2 drops of N/10 sodium hydroxid for neutralization. The water used to generate the steam should be boiled several minutes before connecting the steam generator with the distillation flask in order to expel carbon dioxide. Titrate rapidly with N/10 sodium hydroxid, using phenolphthalein as an indicator. The color should remain about 10 seconds. Express the result as acetic acid. One cc. of N/10 sodium hydroxid is equivalent to 0.0060 gram of acetic acid.

27

Method II. (Hortvet Method³)—Tentative.

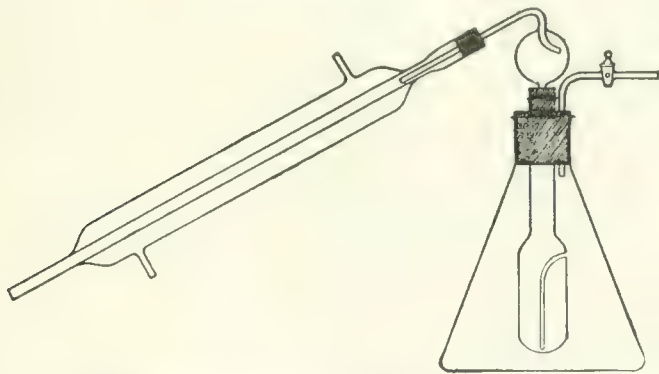


FIG. 8. APPARATUS FOR THE DETERMINATION OF VOLATILE ACIDS.

Introduce 10 cc. of the wine, previously freed from carbon dioxide, into the inner tube of a modified Sellier distillation apparatus (Fig. 8), add a small piece of paraffin to prevent foaming, and adjust the tube and its contents in place within the larger flask containing 100 cc. of recently boiled water. Connect with a condenser as illustrated in Fig. 8 and distil by heating the outer flask. When 50 cc. of the distillate have been collected, empty the receiver into a beaker and titrate with N/10 sodium hydroxid, using phenolphthalein as an indicator. Continue the distillation and titrate each succeeding 10 cc. of distillate until not more than 1 drop of standard alkali is required to reach the neutral point. Usually 80 cc. of distillate will contain all the volatile acid.

28

FIXED ACIDS.—TENTATIVE.

Multiply the amount of volatile acids by 1.25 and subtract this from the total acids, to obtain the amount of fixed acids, expressed as tartaric acid.

29

TOTAL TARTARIC ACID⁴.—TENTATIVE.

Neutralize 100 cc. of the wine with N/1 sodium hydroxid, calculating from the acidity, 25, the number of cc. of N/1 alkali necessary for the neutralization. If the volume of the solution is increased more than 10% by the addition of the alkali, evaporate to approximately 100 cc. Add to the neutralized solution 0.075 gram of tartaric acid for each cc. of N/1 alkali added and, after the tartaric acid has dissolved, add 2 cc. of glacial acetic acid and 15 grams of potassium chlorid. After the potassium chlorid has dissolved, add 15 cc. of 95% alcohol by volume, stir vigorously until the potassium bitartrate starts to precipitate, and then let stand in an ice box for at least 15 hours. Decant the liquid from the separated potassium bitartrate on a Gooch, prepared with a very thin film of asbestos, or on filter paper in a Büchner funnel. Wash the precipitate and filter 3 times with a few cc. of a mixture of 15 grams of potassium chlorid, 20 cc. of 95% alcohol by volume and 100 cc. of water, using not more than 20 cc. of the wash solution in all. Transfer the asbestos or paper and precipitate to the beaker in which the precipitation was made, wash out the Gooch or Büchner funnel with hot water, using about 50 cc. in all, heat to boiling and titrate the hot solution with N/10 sodium hydroxid, using phenolphthalein as an indicator. Increase the number of cc. of N/10 alkali required by 1.5 cc. to allow for the solubility of the precipitate. One cc. of N/10 alkali is equivalent, under these conditions, to 0.015 gram of tartaric acid. Subtract the amount of tartaric acid added from this result to obtain the grams of total tartaric acid per 100 cc. of the wine.

30

FREE TARTARIC ACID AND CREAM OF TARTAR⁵.—TENTATIVE.

Calculate the free tartaric acid and cream of tartar in the following manner:

- Let A = total tartaric acid in 100 cc. of wine divided by 0.015;
 B = total alkalinity of the ash (sum of C and D);
 C = alkalinity of water-soluble ash;
 D = alkalinity of water-insoluble ash.

Then

- (1) If A is greater than B,
 Cream of tartar = $0.0188 \times C$; and
 Free tartaric acid = $0.015 \times (A - B)$.
- (2) If A equals B or is smaller than B but greater than C,
 Cream of tartar = $0.0188 \times C$; and
 Free tartaric acid = 0.
- (3) If A is smaller than C,
 Cream of tartar = $0.0188 \times A$; and
 Free tartaric acid = 0.

TANNIN AND COLORING MATTER.—OFFICIAL.

31

REAGENTS.

- (a) *N/10 oxalic acid*.—One cc. is equivalent to 0.004157 gram of tannin.
 (b) *Standard potassium permanganate solution*.—Dissolve 1.333 grams of potassium permanganate in 1 liter of water and standardize the solution against (a).

(c) *Indigo solution*.—Dissolve 6 grams of sodium sulphindigotate in 500 cc. of water by heating; cool, add 50 cc. of concentrated sulphuric acid, make up to 1 liter and filter.

(d) *Purified boneblack*⁶.—Boil 100 grams of finely powdered boneblack with successive portions of hydrochloric acid (1 to 3), filter and wash with boiling water until free from chlorin. Keep covered with water.

32

DETERMINATION⁷.

Dealcoholize 100 cc. of the wine by evaporation and dilute with water to the original volume. Transfer 10 cc. to a 2 liter porcelain dish; add about a liter of water and exactly 20 cc. of the indigo solution. Add the standard potassium permanganate solution, 1 cc. at a time, until the blue color changes to green; then add a few drops at a time until the color becomes golden yellow. Designate the number of cc. of permanganate solution used as "a".

Treat 10 cc. of the dealcoholized wine, prepared as above, with boneblack for 15 minutes; filter and wash the boneblack thoroughly with water. Add a liter of water and 20 cc. of the indigo solution and titrate with permanganate as above. Designate the number of cc. of permanganate used as "b".

Then $a-b=c$, the number of cc. of the permanganate solution required for the oxidation of the tannin and coloring matter in 10 cc. of the wine.

33

CRUDE PROTEIN.—TENTATIVE.

Determine nitrogen in 50 cc. of the wine, as directed under **I, 18, 21** or **23**, and multiply the result by 6.25.

34

PENTOSANS.—TENTATIVE.

Proceed as directed in **VIII, 64**, except that 100 cc. of the wine and 43 cc. of hydrochloric acid (sp. gr. 1.19) are used in beginning the distillation. Owing to the interference of sugars this determination can be made in dry wines only.

35

GUM AND DEXTRIN.—TENTATIVE.

Evaporate 100 cc. of the wine to about 10 cc. and add 10 cc. of 96% alcohol by volume. If gum or dextrin be present (indicated by the formation of a voluminous precipitate), continue the addition of alcohol, slowly and with stirring, until 100 cc. have been added. Let stand overnight, filter, and wash with 80% alcohol by volume. Dissolve the precipitate on the paper with hot water, hydrolyze the filtrate and washings with hydrochloric acid and proceed as directed under **VIII, 60**.

36

NITRATES.—TENTATIVE.

(a) *White wine*.—Treat a few drops of the wine in a porcelain dish with 2-3 cc. of concentrated sulphuric acid, which contains about 0.1 gram of diphenylamin⁸ per 100 cc. The deep blue color formed in the presence of nitrates appears so quickly that it is not obscured, even in sweet wine, by the blackening produced by the action of sulphuric acid on the sugar.

(b) *Red wine*.—Clarify with basic lead acetate, filter, remove the excess of lead from the filtrate with sodium sulphate, filter again and treat a few drops of this filtrate as directed under (a).

37

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI**.

38

PRESERVATIVES.—TENTATIVE.

Proceed as directed under X.

The detection of added boric acid is somewhat difficult because a small amount of it is normally present in certain wines. Therefore, a quantitative determination should be made. The determination of sulphurous acid must also be quantitative. A small amount of salicylic acid is also normal in wine, and for that reason not more than 50 cc. of the sample should be used in testing for that preservative.

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XVII. DISTILLED LIQUORS.

1 SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer or a small, accurately graduated hydrometer.

2 ALCOHOL BY WEIGHT.—OFFICIAL.

Weigh 20–25 grams of the sample into a distillation flask, dilute with 100 cc. of water, distil nearly 100 cc. and weigh the distillate or make to volume at 20°C. and in either case, determine the specific gravity as directed under 1. Obtain the corresponding percentage of alcohol by weight from XVI, 5, multiply this figure by the weight of the distillate, and divide by the weight of the sample taken to obtain the per cent of alcohol by weight.

The alcohol content of the distillate may be checked by determining the immersion refractometer reading and obtaining, from XVI, 6, the percentage of alcohol.

ALCOHOL BY VOLUME.

3 Method I.—Official.

From the specific gravity of the distillate, obtained under 2, ascertain the corresponding percentage of alcohol by volume from XVI, 5. Multiply this figure by the volume of distillate and divide by the volume of the sample (calculated from the specific gravity) to obtain the percentage of alcohol by volume in the original sample.

4 Method II.—Tentative.

Measure 25 cc. of the sample at 20°C. into a distillation flask, dilute with 100 cc. of water, distil nearly 100 cc., make to volume at 20°C. and determine the specific gravity as directed in 1. Obtain, from XVI, 5, the corresponding percentage of alcohol by volume in the distillate. Multiply by 4 to obtain the percentage of alcohol by volume in the original substance.

The alcohol content of the distillate may be checked by determining the immersion refractometer reading and obtaining the percentage of alcohol from XVI, 6.

5 EXTRACT.—OFFICIAL.

Weigh, or measure at 20°C., 100 cc. of the sample, evaporate nearly to dryness on the water bath, then transfer to a water oven, and dry at the temperature of boiling water for 2½ hours.

6 ASH.—OFFICIAL.

Proceed as directed under VIII, 4, employing the residue from the determination of the extract, 5.

7 ACIDITY.—TENTATIVE.

Titrate 100 cc. of the sample (or 50 cc. diluted to 100 cc. if the sample is dark) with N/10 alkali, using phenolphthalein as an indicator. Express the result as acetic acid; 1 cc. of N/10 alkali is equivalent to 0.0060 gram of acetic acid.

8

ESTERS.—TENTATIVE.

Measure 200 cc. of the sample into a distillation flask, add 25 cc. of water and distil slowly 200 cc., using a mercury valve to prevent loss of alcohol. Exactly neutralize the free acid in 50 cc. of the distillate with N/10 alkali, add a measured excess of 25–50 cc. of N/10 alkali, and either boil for an hour under a reflux condenser, cool and titrate with N/10 acid, or allow the solution to stand overnight in a stoppered flask with the excess of alkali, heat with a tube condenser for 30 minutes at a temperature below the boiling point, cool and titrate. Calculate the number of cc. of N/10 alkali used in the saponification of the esters as ethyl acetate; 1 cc. of N/10 alkali is equivalent to 0.0088 gram of ethyl acetate. Run a blank using water in place of the distillate and make any necessary correction.

ALDEHYDES.—TENTATIVE.

9

REAGENTS.

(a) *Aldehyde-free alcohol*.—Redistil 95% alcohol over sodium or potassium hydroxid, then add 2–3 grams per liter of meta-phenyldiamin hydrochlorid, digest at ordinary temperature for several days (or reflux on a steam bath for several hours) and then distil slowly, rejecting the first 100 cc. and the last 200 cc. of the distillate.

(b) *Sulphite-fuchsin solution*.—Dissolve 0.50 gram of pure fuchsin in 500 cc. of water, then add 5 grams of sulphur dioxid dissolved in water, make up to 1 liter and allow to stand until colorless. This solution does not keep indefinitely; therefore, prepare in small quantities and keep at a low temperature.

(c) *Standard acetaldehyde solution*.—Prepare according to the directions of Vasey¹ as follows: Grind aldehyde ammonia in a mortar with anhydrous ether and decant the ether. Repeat this operation several times, then dry the purified salt in a current of air and finally in vacuo over sulphuric acid. Dissolve 1.386 grams of this purified aldehyde ammonia in 50 cc. of 95% alcohol by volume, add 22.7 cc. of N/1 alcoholic sulphuric acid, then make up to 100 cc. and add 0.8 cc. of alcohol for the volume of the ammonium sulphate precipitate. Allow the mixture to stand overnight and filter. This solution contains 1 gram of acetaldehyde in 100 cc. and will retain its strength.

The standard found most convenient for use is 2 cc. of this strong aldehyde solution diluted to 100 cc. with 50% alcohol by volume. One cc. of this solution is equivalent to 0.0002 gram of acetaldehyde. This solution should be made up fresh every day or so, as it loses strength.

10

DETERMINATION.

Determine the aldehyde in the distillate, prepared as directed under 8. Dilute 5–10 cc. of the distillate to 50 cc. with aldehyde-free alcohol 50% by volume, add 25 cc. of the sulphite-fuchsin solution and allow to stand for 15 minutes at 15°C. The solutions and reagents should be at 15°C. when they are mixed. Prepare standards of known strength and blanks in the same way. The comparison standards found most convenient for use contain 0.0001, 0.0002, 0.0004, 0.0005 and 0.0006 gram of acetaldehyde.

FURFURAL.—TENTATIVE.

11

REAGENTS.

(a) *Standard furfural solution*.—Dissolve 1 gram of redistilled furfural in 100 cc. of 95% alcohol by volume. Standards are made by diluting 1 cc. of this solu-

tion to 100 cc. with 50% alcohol by volume. One cc. of this solution contains 0.0001 gram of furfural.

(b) *Furfural-free alcohol*.—Prepare as directed in 9 (a).

12

DETERMINATION.

Dilute 10–20 cc. of the distillate, as prepared under 8, to 50 cc. with furfural-free alcohol, 50% by volume. Add 2 cc. of colorless anilin and 0.5 cc. of hydrochloric acid (sp. gr. 1.125) and keep for 15 minutes in a water bath at about 15°C. Prepare standards of known strength and blanks in the same way. The comparison standards found most convenient for use contain 0.00005, 0.0001, 0.00015, 0.0002, 0.00025 and 0.0003 gram of furfural.

FUSEL OIL.—TENTATIVE.

13

REAGENTS.

(a) *Purified carbon tetrachlorid*.—Mix crude carbon tetrachlorid with one tenth its volume of concentrated sulphuric acid, shake thoroughly at frequent intervals and allow to stand overnight. Wash free of acid and impurities with tap water. Remove the water, add an excess of sodium hydroxid solution and distil the carbon tetrachlorid from it.

(b) *Oxidizing solution*.—Dissolve 100 grams of potassium dichromate in 900 cc. of water and add 100 cc. of concentrated sulphuric acid.

14

DETERMINATION.

(1) To 100 cc. of the sample add 20 cc. of N/2 sodium hydroxid and saponify the mixture by boiling for an hour under a reflux condenser; or, (2) Mix 100 cc. of the liquor with 20 cc. of N/2 sodium hydroxid, allow to stand overnight at room temperature and distil directly. Connect the flask with a distillation apparatus, distil 90 cc., add 25 cc. of water and continue the distillation until an additional 25 cc. are collected.

Whenever aldehydes are present in excess of 15 parts per 100,000, add to the distillate 0.5 gram of meta-phenylendiamin hydrochlorid, reflux for an hour, distil 100 cc., add 25 cc. of water and continue the distillation until an additional 25 cc. are collected.

Approximately saturate the distillate with finely ground sodium chlorid and add saturated sodium chlorid solution until the specific gravity is 1.10.

Extract this salt solution 4 times with the purified carbon tetrachlorid, using 40, 30, 20 and 10 cc., respectively, and wash the carbon tetrachlorid 3 times with 50 cc. portions of saturated sodium chlorid solution, and twice with saturated sodium sulphate solution. Then transfer the carbon tetrachlorid to a flask containing 50 cc. of the oxidizing solution and boil for 8 hours under a reflux condenser.

Add 30 cc. of water and distil until only about 20 cc. remain; add 80 cc. of water and again distil until 15–20 cc. are left. Neutralize the distillate to methyl orange, and titrate with N/10 sodium hydroxid, using phenolphthalein as an indicator. If the distillations have been properly conducted, the distillate will not show a marked acid reaction to methyl orange. Should considerably more than 1 cc. of N/10 alkali be consumed at this point, the result will be unreliable and the determination should be repeated. One cc. of N/10 sodium hydroxid is equivalent to 0.0088 gram of amyl alcohol.

Rubber stoppers can be used in the saponification and first distillation, but corks covered with tinfoil must be used in the oxidation and second distillation. Corks and tinfoil must be renewed frequently.

Conduct a blank determination upon 100 cc. of carbon tetrachlorid beginning the blank at that point of the procedure immediately after the extraction and just before the washings with sodium chlorid and sodium sulphate solutions.

15**SUGARS.—TENTATIVE.**

Proceed as directed under **XVI, 15, 16** or **17**.

METHYL ALCOHOL.**16***Trillat Method².—Tentative.*

To 50 cc. of the sample add 50 cc. of water and 8 grams of lime and fractionate by the aid of Glinsky bulb tubes. Dilute the first 15 cc. of the distillate to 150 cc., mix with 15 grams of potassium dichromate and 70 cc. of sulphuric acid (1 to 5), and allow to stand for an hour with occasional shaking.

Distil, reject the first 25 cc. and collect 100 cc. Mix 50 cc. of the distillate with 1 cc. of redistilled dimethylanilin, transfer to a stout, tightly stoppered flask, and keep on a bath at 70°–80°C. for 3 hours with occasional shaking. Make distinctly alkaline with sodium hydroxid solution, and distil off the excess of dimethylanilin, stopping the distillation when 25 cc. have passed over.

Acidify the residue in the flask with acetic acid, shake and test a few cc. by adding 4 or 5 drops of 1% suspension of lead dioxid. If methyl alcohol is present, a blue coloration occurs which is increased by boiling.

Ethyl alcohol thus treated yields a blue coloration changing immediately to green, later to yellow, and becoming colorless when boiled.

17*Riche and Bardy Method².—Tentative.*

The following method for the detection of methyl alcohol in commercial spirit of wine depends on the formation of methylanilin violet:

Place 10 cc. of the sample, previously redistilled over potassium carbonate if necessary, in a small flask with 15 grams of iodine and 2 grams of red phosphorus. Keep in ice water for 10–15 minutes until action has ceased. Distil off, on a water bath, the methyl and ethyl iodids formed into about 30 cc. of water. Wash with dilute alkali to eliminate free iodine. Separate the heavy, oily liquid which settles and transfer to a flask containing 5 cc. of anilin. If the action be too violent, place the flask in cold water; if too slow, stimulate by gently warming the flask. After an hour boil the product with water and add about 20 cc. of 15% sodium hydroxid solution; when the bases rise to the top as an oily layer, fill the flask up to the neck with water and draw them off with a pipette. Oxidize 1 cc. of the oily liquid by adding 10 grams of a mixture of 100 parts of clean sand, 2 of common salt, and 3 of cupric nitrate; mix thoroughly, transfer to a glass tube, and heat to 90°C. for 8–10 hours. Exhaust the product with warm alcohol, filter and make up to 100 cc. with alcohol. If the sample of spirits is pure, the liquid has a red tint, but, in the presence of 1% of methyl alcohol, it has a distinct violet shade; with 2.5% the shade is very distinct, and still more so with 5%. To detect more minute quantities of methyl alcohol, dilute 5 cc. of the colored liquid to 100 cc. with water, and dilute 5 cc. of this again to 400 cc. Heat the liquid thus obtained in a porcelain dish and immerse in it a fragment of white merino (free from sulphur) for 30 minutes. If the alcohol is pure, the wool will remain white, but, if methyl alcohol is present, the fiber will become violet, the depth of tint giving a fairly approximate indication of the proportion of methyl alcohol present.

18 *Immersion Refractometer Method. (Leach and Lythgoe⁴)—Tentative.*

Determine by the immersion refractometer at 20°C. the refraction of the distillate obtained in the determination of alcohol. If, on reference to the table under **19**, the refraction shows the percentage of alcohol agreeing with that obtained from the specific gravity, it may safely be assumed that no methyl alcohol is present. If, however, there is an appreciable amount of methyl alcohol, the low refractometer reading will at once indicate the fact. If the absence from the solution of refractive substances other than water and the alcohols is assured, this difference in refraction is conclusive of the presence of methyl alcohol.

The addition of methyl alcohol to ethyl alcohol decreases the refraction in direct proportion to the amount present; hence the quantitative calculation is readily made by interpolation in the table under **19**, using the figures for pure ethyl and methyl alcohol of the same alcoholic strength as the sample.

Example.—The distillate has a specific gravity of 0.97080, corresponding to 18.38% alcohol by weight, and has a refraction of 35.8 at 20°C. by the immersion refractometer; by interpolation in the refractometer table the readings of ethyl and methyl alcohol corresponding to 18.38% alcohol are 47.3 and 25.4, respectively, the difference being 21.9; $47.3 - 35.8 = 11.5$; $(11.5 \div 21.9) 100 = 52.5$, showing that 52.5% of the total alcohol present is methyl alcohol.

19

TABLE 18.

Scale readings of the Zeiss immersion refractometer at 20°C., corresponding to each per cent by weight of methyl and ethyl alcohols.

PER CENT ALCO- HOL BY WEIGHT	SCALE READINGS		PER CENT ALCO- HOL BY WEIGHT	SCALE READINGS		PER CENT ALCO- HOL BY WEIGHT	SCALE READINGS		PER CENT ALCO- HOL BY WEIGHT	SCALE READINGS	
	Methyl alco- hol	Ethyl alco- hol		Methyl alco- hol	Ethyl alco- hol		Methyl alco- hol	Ethyl alco- hol		Methyl alco- hol	Ethyl alco- hol
0	14.5	14.5	25	29.7	60.1	50	39.8	90.3	75	29.7	101.0
1	14.8	16.0	26	30.3	61.9	51	39.7	91.1	76	29.0	101.0
2	15.4	17.6	27	30.9	63.7	52	39.6	91.8	77	28.3	100.9
3	16.0	19.1	28	31.6	65.5	53	39.6	92.4	78	27.6	100.9
4	16.6	20.7	29	32.2	67.2	54	39.5	93.0	79	26.8	100.8
5	17.2	22.3	30	32.8	69.0	55	39.4	93.6	80	26.0	100.7
6	17.8	24.1	31	33.5	70.4	56	39.2	94.1	81	25.1	100.6
7	18.4	25.9	32	34.1	71.7	57	39.0	94.7	82	24.3	100.5
8	19.0	27.8	33	34.7	73.1	58	38.6	95.2	83	23.6	100.4
9	19.6	29.6	34	35.2	74.4	59	38.3	95.7	84	22.8	100.3
10	20.2	31.4	35	35.8	75.8	60	37.9	96.2	85	21.8	100.1
11	20.8	33.2	36	36.3	76.9	61	37.5	96.7	86	20.8	99.8
12	21.4	35.0	37	36.8	78.0	62	37.0	97.1	87	19.7	99.5
13	22.0	36.9	38	37.3	79.1	63	36.5	97.5	88	18.6	99.2
14	22.6	38.7	39	37.7	80.2	64	36.0	98.0	89	17.3	98.9
15	23.2	40.5	40	38.1	81.3	65	35.5	98.3	90	16.1	98.6
16	23.9	42.5	41	38.4	82.3	66	35.0	98.7	91	14.9	98.3
17	24.5	44.5	42	38.8	83.3	67	34.5	99.1	92	13.7	97.8
18	25.2	46.5	43	39.2	84.2	68	34.0	99.4	93	12.4	97.2
19	25.8	48.5	44	39.3	85.2	69	33.5	99.7	94	11.0	96.4
20	26.5	50.5	45	39.4	86.2	70	33.0	100.0	95	9.6	95.7
21	27.1	52.4	46	39.5	87.0	71	32.3	100.2	96	8.2	94.9
22	27.8	54.3	47	39.6	87.8	72	31.7	100.4	97	6.7	94.0
23	28.4	56.3	48	39.7	88.7	73	31.1	100.6	98	3.5	93.0
24	29.1	58.2	49	39.8	89.5	74	30.4	100.8	99	3.5	92.0
.....	100	2.0	91.0

20

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI**.

21

WATER-INSOLUBLE COLOR IN WHISKIES.—TENTATIVE.

Evaporate 50 cc. of the sample just to dryness on a steam bath. Take up with cold water, using approximately 15 cc., filter and wash until the filtrate amounts to nearly 25 cc. To this filtrate add 25 cc. of absolute alcohol, or 26.3 cc. of 95% alcohol by volume, and make up to 50 cc. by the addition of water. Mix thoroughly and compare in a colorimeter with the original material. Calculate from these readings the per cent of color insoluble in water.

COLORS INSOLUBLE IN AMYL ALCOHOL.

22

Modified Marsh Method.—Tentative.

Evaporate 50 cc. of whisky just to dryness on a steam bath. Dissolve the residue in water and 95% alcohol by volume and make to a volume of 50 cc., using a total volume of 26.3 cc. of 95% alcohol. Place 25 cc. of this solution in a separatory funnel and add 20 cc. of freshly shaken Marsh reagent (100 cc. of pure amyl alcohol, 3 cc. of sirupy phosphoric acid and 3 cc. of water), shaking lightly so as not to form an emulsion. Allow the layers to separate and repeat this shaking and standing twice again. After the layers have separated completely draw off the lower or aqueous layer, which contains the caramel, into a 25 cc. cylinder and make up to volume with 50% alcohol by volume. Compare this solution in a colorimeter with the untreated 25 cc. Calculate the result of this reading to the per cent of color insoluble in amyl alcohol.

CARAMEL.

23

Amthor Test Modified by Lasche⁵.—Tentative.

Add 10 cc. of paraldehyde to 5 cc. of the sample in a test tube and shake. Add absolute alcohol, a few drops at a time, shaking after each addition until the mixture becomes clear. Allow to stand. Turbidity after 10 minutes is an indication of caramel.

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⁴ J. Am. Chem. Soc., 1905, **27**: 964.

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XVIII. BEERS.

1 PREPARATION OF SAMPLE.—TENTATIVE.

Remove carbon dioxid by transferring the contents of the bottle to a large flask and shaking vigorously or by pouring back and forth between beakers, care being taken that the temperature of the beer is not below 20°C.

2 COLOR.—TENTATIVE.

Determine the depth of color of the sample in a $\frac{1}{4}$ inch cell with a Lovibond tintometer, using the beer scale. Express the result in terms of a $\frac{1}{4}$ inch cell.

3 SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer.

4 ALCOHOL.—TENTATIVE.

Determine as directed under **XVI, 4**.

EXTRACT.

5 Method I.—Official.

Measure 25 cc. of the carbon dioxid-free beer at 20°C. into a tared, flat-bottomed platinum dish, approximately 85 mm. in diameter, and evaporate just to dryness on a steam bath and heat to constant weight in a vacuum oven at 70°C.

6 Method II.—Tentative.

The immersion refractometer reading of the beer at 20°C. minus the immersion refractometer reading of the distillate at 20°C. times 0.2571 equals the grams of extract in 100 cc. of beer.

7 Method III.—Tentative.

Calculate the specific gravity of the dealcoholized beer by the following formula:

$$S = G \div 1 - A \text{ in which}$$

S = the specific gravity of the dealcoholized beer;

G = the specific gravity of the beer; and

A = the specific gravity of the distillate obtained in the determination of alcohol.

From **IX, 9**, ascertain the per cent by weight of extract in the dealcoholized beer corresponding to the value of S. Multiply the figure thus obtained by S to obtain the grams of extract per 100 cc. of beer.

8 EXTRACT OF ORIGINAL WORT (APPROXIMATE).—TENTATIVE.

Calculate the grams of extract per 100 cc. in the original wort by the following formula:

$$O = 2A + E \text{ in which}$$

O = extract of the original wort;

A = alcohol (grams per 100 cc.); and

E = extract of the dealcoholized beer (grams per 100 cc.).

9

DEGREE OF FERMENTATION.—TENTATIVE.

Calculate the degree of fermentation by the following formula:

$$D = \frac{100 \times 2A}{O} \text{ in which}$$

D = degree of fermentation;

A = alcohol (grams per 100 cc.); and

O = extract of original wort.

10

TOTAL ACIDS.—TENTATIVE.

Proceed as directed under **XVI, 25**. Express the result as lactic acid, grams per 100 cc. One cc. of N/10 sodium hydroxid is equivalent to 0.0090 gram of lactic acid.

11

VOLATILE ACIDS.—TENTATIVE.

Proceed as directed under **XVI, 27**. Express the result as acetic acid, grams per 100 cc.

12

REDUCING SUGARS.—TENTATIVE.

Dilute 25 cc. of the carbon dioxid-free beer, measured at 20°C., with water to 100 cc. of the same temperature. Determine the reducing sugars in 25 cc. of this solution, as directed under **VIII, 42**. Express the result as grams of anhydrous maltose per 100 cc. of beer.

13

DEXTRIN.—TENTATIVE.

To 50 cc. of the carbon dioxid-free beer measured at 20°C., add 15 cc. of hydrochloric acid (sp. gr. 1.125), dilute to 200 cc., attach to a reflux condenser and keep in a boiling water bath for 2 hours. Cool, nearly neutralize with sodium hydroxid solution, complete to a volume of 250 cc., filter and determine dextrose as directed under **VIII, 52** or **54**. From the number of grams of dextrose per 100 cc. of beer, subtract 1.053 times the amount of maltose as found in **12** and multiply the remainder by 0.9 to obtain the number of grams of dextrin per 100 cc. of beer.

14

DIRECT POLARIZATION.—TENTATIVE.

Read the polarization of the original sample in degrees Ventzke in a 200 mm. tube at 20°C. If the beer is turbid, clarify by shaking with alumina cream, filter and correct the reading for dilution.

15

GLYCEROL.—TENTATIVE.

Proceed as directed under **XVI, 8**.

16

ASH.—OFFICIAL.

Evaporate to dryness 25 cc. of the carbon dioxid-free sample, measured at 20°C., and proceed as directed under **VIII, 4**.

17

PHOSPHORIC ACID.—TENTATIVE.

To 25 cc. of the carbon dioxid-free beer, measured at 20°C., add 20 cc. of 2% calcium acetate solution, evaporate to dryness and ignite at low redness to a white ash. Add 10–15 cc. of boiling nitric acid (1 to 9) and determine phosphoric acid (P_2O_5) as directed under **I, 9**.

18

PROTEIN.—OFFICIAL.

Measure, at 20°C., 25 cc. of the carbon dioxid-free beer into a Kjeldahl digestion flask, add a small amount of tannin to prevent frothing, evaporate to dryness, determine nitrogen as directed under I, 18, 21 or 23, multiply the result by 6.25 and calculate the percentage of protein.

19

PRESERVATIVES.—TENTATIVE.

Proceed as directed under X.

20

COLORING MATTERS.—TENTATIVE.

Proceed as directed under XI.

21

METALS.—TENTATIVE.

Proceed as directed under XII.



XIX. VINEGARS.

(Unless otherwise noted, express results as grams per 100 cc.)

1 PHYSICAL EXAMINATION.—TENTATIVE.

Note the appearance, color, odor and taste.

2 PREPARATION OF SAMPLE.—TENTATIVE.

If the sample is turbid, filter before proceeding with the analysis.

3 SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer.

4 ALCOHOL.—TENTATIVE.

Measure 100 cc. of the sample into a round-bottomed, distillation flask. Make faintly alkaline with saturated sodium hydroxid solution, add a small piece of paraffin, distil almost 50 cc., make up to 50 cc. at the temperature of the sample and determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer. Obtain from XVI, 5, the per cent by volume, or grams per 100 cc., noting that the alcoholic strength of the distillate is twice that of the original vinegar.

GLYCEROL.—TENTATIVE.

5 REAGENTS.

(a) *Strong potassium dichromate solution.*—Dissolve 74.56 grams of dry, recrystallized potassium dichromate in water, add 150 cc. of concentrated sulphuric acid, cool and make up to 1 liter at 20°C. One cc. of this solution is equivalent to 0.01 gram of glycerol. The high coefficient of expansion of this strong solution necessitates its preparation at exactly 20°C. and the measurement of any definite volume at the same temperature. If desired, the measurements may be made at room temperature by means of a weighing burette, the volume used in this case being ascertained by dividing the weight of the solution used by its specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$.

(b) *Dilute potassium dichromate solution.*—Measure 25 cc. of the strong potassium dichromate solution at exactly 20°C. into a 500 cc. volumetric flask, dilute with water and make up to the mark at room temperature. Twenty cc. of this solution are equivalent to 1 cc. of (a).

(c) *Ferrous ammonium sulphate solution.*—Dissolve 30 grams of crystallized ferrous ammonium sulphate in water, add 50 cc. of concentrated sulphuric acid, cool and dilute at room temperature. One cc. of this solution is approximately equivalent to 1 cc. of (b). Its value changes slightly from day to day and it must be standardized against (b) whenever used.

(d) *Potassium ferricyanid indicator.*—Dissolve 1 gram of crystallized potassium ferricyanid in 50 cc. of water. This solution must be freshly prepared.

(e) *Milk of lime.*—Introduce 150 grams of calcium oxid, selected from clean, hard lumps, prepared preferably from marble, into a large porcelain or iron dish, slake with water, cool and add sufficient water to make 1 liter.

(f) *Silver carbonate*.—Dissolve 0.1 gram of silver sulphate in about 50 cc. of water, add an excess of sodium carbonate solution, allow the precipitate to settle and wash with water several times by decantation until the washings are practically neutral. This reagent must be freshly prepared immediately before use.

6

DETERMINATION.

All evaporations should be made on a water bath, the temperature of which is maintained at 85°–90°C.

Evaporate 100 cc. of the vinegar to 5 cc., add 20 cc. of water and again evaporate to 5 cc. to expel acetic acid. Treat the residue with about 5 grams of fine sand and 15 cc. of the milk of lime and evaporate almost to dryness, with frequent stirring, avoiding the formation of a dry crust or evaporation to complete dryness. Treat the moist residue with 5 cc. of water, rub into a homogeneous paste, then add slowly 45 cc. of absolute alcohol, washing down the sides of the dish to remove adhering paste, and stir thoroughly. Heat the mixture on a water bath, with constant stirring, to incipient boiling, transfer to a suitable vessel and centrifugalize. Decant the clear liquid into a porcelain dish and wash the residue with several small portions of hot 90% alcohol by volume by aid of the centrifuge. (If a centrifuge is not available, decant the liquid through a fluted filter into a porcelain dish. Wash the residue repeatedly with small portions of hot 90% alcohol, twice by decantation, and then by transferring all the material to the filter. Continue the washing until the filtrate amounts to 150 cc.) Evaporate to a sirupy consistency, add 10 cc. of absolute alcohol to dissolve this residue and transfer to a 50 cc. glass-stoppered cylinder, washing the dish with successive small portions of absolute alcohol until the volume of the solution amounts to 20 cc. Then add 3 portions of 10 cc. each of anhydrous ether, shaking thoroughly after each addition. Let stand until clear, then pour off through a filter, and wash the cylinder and filter with a mixture of 2 volumes of absolute alcohol and 3 of anhydrous ether. If a heavy precipitate has formed in the cylinder, centrifugalize at low speed, decant the clear liquid and wash 3 times with 20 cc. portions of the alcohol-ether mixture, shaking the mixture thoroughly each time and separating the precipitate by means of the centrifuge. Wash the paper with the alcohol-ether mixture, and evaporate the filtrate and washings on the water bath to about 5 cc., add 20 cc. of water and again evaporate to 5 cc.; again add 20 cc. of water and evaporate to 5 cc.; finally add 10 cc. of water and evaporate to 5 cc.

These evaporations are necessary to remove all the ether and alcohol, and, when conducted at 85°–90°C., result in no loss of glycerol if the concentration of the latter is less than 50%.

Transfer the residue with hot water to a 50 cc. graduated flask, cool, add the silver carbonate, prepared from 0.1 gram of silver sulphate, shake and allow to stand 10 minutes; then add 0.5 cc. of basic lead acetate solution [VIII, 13 (a)], shake occasionally and allow to stand 10 minutes; make up to the mark, shake well, filter, rejecting the first portion of the filtrate, and pipette 25 cc. of the clear filtrate into a 250 cc. volumetric flask.

Add 1 cc. of concentrated sulphuric acid to precipitate the excess of lead and then 30 cc. of the strong potassium dichromate solution. Add carefully 24 cc. of concentrated sulphuric acid, rotating the flask gently to mix the contents and avoid violent ebullition, and then place in a *boiling* water bath for exactly 20 minutes. Remove the flask from the bath, dilute, cool and make up to the mark at room temperature. The amount of strong dichromate solution used must be sufficient to

leave an excess of about 12.5 cc. at the end of the oxidation, the amount given above (30 cc.) being sufficient for ordinary vinegar containing about 0.35 gram or less of glycerol per 100 cc.

Standardize the ferrous ammonium sulphate solution against the dilute potassium dichromate solution by introducing from the respective burettes approximately 20 cc. of each of the 2 solutions into a beaker containing 100 cc. of water. Complete the titration using the potassium ferrieyanid solution as an outside indicator. From this titration calculate the volume (F) of the ferrous ammonium sulphate solution equivalent to 20 cc. of the dilute and also, therefore, to 1 cc. of the strong dichromate solution.

In place of the dilute dichromate solution, substitute a burette containing the oxidized glycerol with an excess of the strong dichromate solution and ascertain how many cc. are equivalent to (F) cc. of the ferrous ammonium sulphate solution and also, therefore, to 1 cc. of the strong dichromate solution. Then 250 divided by this last equivalent equals the number of cc. of the strong dichromate solution present in excess in the 250 cc. flask after oxidation of the glycerol.

The number of cc. of the strong dichromate solution added, minus the excess found after oxidation, multiplied by 0.02 gives the grams of glycerol per 100 cc. of vinegar.

7

SOLIDS.—TENTATIVE.

Measure 10 cc. of the sample into a tared, flat-bottomed platinum dish of 50 mm. bottom diameter, evaporate on a boiling water bath for 30 minutes, and dry for exactly $2\frac{1}{2}$ hours in a water oven at the temperature of boiling water. Cool in a desiccator and weigh. It is essential that the size and shape of the dish and the time of drying be followed strictly.

8

TOTAL REDUCING SUBSTANCES BEFORE INVERSION.—TENTATIVE.

Proceed as directed under VIII, 25, using 10 cc. of the sample. In the case of malt vinegar, express the results as dextrose; in all other cases as invert sugar.

9

REDUCING SUGARS BEFORE INVERSION AFTER EVAPORATION.—TENTATIVE.

Evaporate 50 cc. of the sample on the water bath to a volume of 5 cc. Add 25 cc. of water and again evaporate to 5 cc. Transfer to a 100 cc. volumetric flask, make up to the mark, and proceed as directed under 8, using a quantity equivalent to 10 or 20 cc. of the sample.

10

REDUCING SUGARS AFTER INVERSION.—TENTATIVE.

Proceed as directed under 9. After the last evaporation to 5 cc. transfer to a 100 cc. volumetric flask with 70 cc. of water, and invert as directed under VIII, 14. Nearly neutralize with sodium hydroxid solution, make up to the mark and proceed as directed under VIII, 25, using a quantity equivalent to 10 or 20 cc. of the sample.

11

LEAD PRECIPITATE.—TENTATIVE.

To 10 cc. of the sample in a test tube, add 2 cc. of 20% lead acetate solution, shake and let stand 30 minutes. Describe the precipitate as turbid, light, normal, heavy or very heavy.

12

POLARIZATION.—TENTATIVE.

If the lead precipitate is normal, add to 50 cc. of the sample 5 cc. of basic lead acetate solution [VIII, 13 (a)], shake, let stand 30 minutes, filter and polarize,

preferably in a 200 mm. tube, correcting for dilution. If basic lead acetate gives only a turbidity, add to the sample, already treated with basic lead acetate, 10 cc. of alumina cream [VIII. 13 (b)], shake, let stand 30 minutes, filter and polarize, correcting for dilution. In the case of malt vinegar, treat 100 cc. of the sample with 5 cc. of 10% phosphotungstic acid solution and filter. To 50 cc. of the filtrate add 5 cc. of the basic lead acetate solution, filter and polarize, correcting the reading obtained for dilution.

13**ASH.—TENTATIVE.**

(a) Measure 25 cc. of the vinegar into a tared platinum dish, evaporate to dryness on the steam bath and proceed as directed under VIII, 4.

(b) Evaporate 25 cc. of the sample to dryness as directed under (a), heat in a muffle at low heat to expel inflammable gases, treat the charred portion with a few cc. of water, and evaporate to dryness on a water bath; replace in the muffle at low redness for 15 minutes, and continue the alternate evaporation and heating until a white or gray ash is obtained, at no time exceeding a dull red heat; cool in a desiccator and weigh.

Useful information may often be obtained by noting the odor given off by the solids during charring.

14**SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.**

Treat the ash, obtained in 13, as directed under IX, 17.

15**ALKALINITY OF THE SOLUBLE ASH.—TENTATIVE.**

Proceed as directed under IX, 18, expressing the result as the number of cc. of N, 10 hydrochloric acid required to neutralize the soluble ash from 100 cc. of the vinegar.

16**SOLUBLE AND INSOLUBLE PHOSPHORIC ACID.—TENTATIVE.**

Determine phosphoric acid in the water-soluble and water-insoluble portions of the ash as directed under I, 9, dissolving the water-insoluble portion in about 50 cc. of boiling nitric acid (1 to 9). Express the result as mg. of phosphorus pentoxid (P_2O_5) in 100 cc. of the vinegar.

17**TOTAL ACIDS.—TENTATIVE.**

Dilute 10 cc. of the sample with recently boiled and cooled water until it appears very slightly colored, and titrate with N/2 alkali, using phenolphthalein as an indicator. One cc. of N/2 alkali is equivalent to 0.030 gram of acetic acid.

18**FIXED ACIDS.—TENTATIVE.**

Measure 10 cc. of the vinegar into a 200 cc. porcelain casserole, evaporate just to dryness, add 5–10 cc. of water, and again evaporate; repeat until at least 5 evaporations have taken place. Add about 200 cc. of recently boiled and cooled water and titrate with N/10 alkali, using phenolphthalein as an indicator. One cc. of N/10 alkali is equivalent to 0.0067 gram of malic acid.

19**VOLATILE ACIDS.—TENTATIVE.**

To obtain the volatile acids subtract the fixed acids, calculated as acetic acid, from the total acids.

20

COLOR.—TENTATIVE.

Determine the depth of color in a Lovibond tintometer by good, reflected daylight, using a $\frac{1}{2}$ inch cell and the brewer's scale. Express the result in terms of a $\frac{1}{2}$ inch cell.

FORMIC ACID.

21

Fincke Method¹.—Tentative.

Employ the apparatus described under **X, 39**, Fig. 6. Introduce 100 cc. of the sample into flask (A), add 0.4–0.5 gram of tartaric acid, and place in position as shown in Fig. 6, the flask (B) having previously been charged with a suspension of 15 grams of calcium carbonate in 100 cc. of water. Heat the contents of flasks (A) and (B) to boiling and distil with steam from the generator (S), the vapor passing first through the sample in flask (A), then through the boiling suspension of calcium carbonate in flask (B), after which it is condensed and measured in the receiver (C). Maintain the volume of liquid in flask (B) as nearly constant as possible and reduce the volume of the sample in flask (A) to 30–40 cc. by heating with small Bunsen flames, the distillation being continued until 1 liter of distillate is collected. Disconnect the apparatus, filter the calcium carbonate suspension, and wash the calcium carbonate that remains on the paper with a little hot water. Render the filtrate faintly acid with hydrochloric acid, add 10–15 cc. of mercuric chlorid reagent [**X, 38 (b)**], mix and heat on a boiling water bath for 2 hours. Filter on a tared Gooch, wash the precipitate thoroughly with cold water and finally with a little alcohol. Dry in a boiling water oven for 30 minutes, cool in a desiccator, weigh, and calculate the weight of formic acid present by multiplying the weight of the precipitate by 0.0975.

22

ALCOHOL PRECIPITATE.—TENTATIVE.

Evaporate 100 cc. of the vinegar to about 15 cc. When there is considerable sugar in the vinegar, if the sample is evaporated to too low a volume, a gummy or stringy precipitate is formed on adding the alcohol instead of a flocculent one. When the sugar content is high, therefore, the evaporation should not be carried beyond 20 cc. To this residue add slowly and with constant stirring 200 cc. of 95% alcohol by volume and allow the mixture to stand overnight. From this point proceed as directed under **XIII, 18**, beginning with the sentence, "Filter and wash with 80% alcohol by volume".

23

PENTOSANS.—TENTATIVE.

Proceed as directed in **VIII, 64**, except that 100 cc. of the vinegar and 43 cc. of hydrochloric acid (sp. gr. 1.19) are used in beginning the distillation.

TARTARIC ACID AND TARTRATES.

24

Qualitative Test.—Tentative.

Evaporate 50 cc. of the vinegar in a porcelain dish to a volume of about 10 cc., filter into a test tube, add 1 cc. of 25% calcium chlorid solution and 2 cc. of 50% ammonium acetate solution and allow to stand overnight. In the presence of tartaric acid a deposit of calcium tartrate is formed, the crystals of which may be identified under the microscope by their characteristic form.

25

TOTAL TARTARIC ACID.—TENTATIVE.

Evaporate 200 cc. of the sample to a sirupy consistency to remove excess of acetic acid, dilute to the original volume with water in a volumetric flask, determine the

acidity as directed in 17, and determine total tartaric acid in a 100 cc. aliquot as directed under XVI, 29, except that 20 cc. of alcohol are used in the precipitation instead of 15 cc.

FREE MINERAL ACIDS.

26

Logwood Method².—Tentative.

Prepare an extract of logwood as follows: Pour 100 cc. of boiling water upon 2 grams of fresh logwood chips, allow the infusion to stand for a few hours and filter. Place drops of the liquid on a porcelain surface and dry on a water bath. Add to one of the spots a drop of the vinegar to be tested (after concentration if desirable) and evaporate to dryness. A yellow tint remains if free mineral acids are absent, a red tint if they are present.

27

Methyl Violet Method.—Tentative.

Add 5–10 cc. of water to 5 cc. of vinegar and, after mixing well, add 4 or 5 drops of methyl violet solution (1 part of methyl violet 2B in 10,000 parts of water). A blue or green coloration indicates the presence of a free mineral acid.

28

Quantitative Method. (Hehner Method).—Tentative.

To a measured amount of the sample add a measured excess of standard alkali, evaporate to dryness, incinerate and titrate the ash with standard acid, using methyl orange as an indicator. The difference between the number of cc. of alkali first added and the number of cc. of acid needed to titrate the ash represents the free mineral acid present.

29

METALS.—TENTATIVE.

Proceed as directed under XII.

DEXTRIN.

30

Qualitative Test.—Tentative.

Evaporate 100 cc. of the vinegar to a volume of about 15 cc. Add slowly and with constant stirring 200 cc. of 95% alcohol by volume and allow to stand overnight. The precipitate formed should be tested for dextrin by the optical rotation and color reaction with iodine.

SPICES AND ADDED PUNGENT MATERIALS.

31

Qualitative Test.—Tentative.

Neutralize exactly a portion of the vinegar and test by taste and smell. Agitate the liquid with ether in a separatory funnel, remove and evaporate the ethereal layer, and note the odor and taste of the residue.

32

COLORING MATTERS.—TENTATIVE.

Proceed as directed under XI.

33

PRESERVATIVES.—TENTATIVE.

Proceed as directed under X.

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¹ Z. Nahr. Genussm., 1911, 21: 1; 22: 83.

² Allen, Commercial Organic Analysis. 4th ed., 1909–14, 1: 503.

XX. FLAVORING EXTRACTS.

VANILLA EXTRACT AND ITS SUBSTITUTES.

1 SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer

2 ALCOHOL.—OFFICIAL.

Proceed as directed under **XVII, 2** or **3**.

3 GLYCEROL.—TENTATIVE.

Proceed as directed under **XVI, 7, 8** or **9**, the method selected depending upon the amount of sugar present, employing an amount of the sample containing 0.10-0.40 gram of glycerol.

VANILLIN AND COUMARIN.

Modified Hess and Prescott Method¹.—Tentative.

(This method is not applicable to concentrated vanillin and coumarin preparations in which the amount of vanillin and coumarin present exceeds the quantity dissolved by 100 cc. of water at 20°C.

In such cases employ a smaller amount of the sample and dilute to 50 cc.)

4 PREPARATION OF SOLUTION.

Measure 50 cc. of the extract at 20°C. into a 250 cc. beaker with marks showing volumes of 80 and 50 cc., dilute to 80 cc. and evaporate to 50 cc. on a water bath kept at 70°C. Dilute again with water to 80 cc. and evaporate to 50 cc. Transfer to a 100 cc. flask, rinsing the beaker with hot water; add 25 cc. of 8% lead acetate solution; make up to the mark with water, shake and allow to stand 18 hours (overnight) at 37°-40°C. Decant into a small, dry filter, reserving the filtrate for the determination of vanillin and coumarin, the normal lead number, **6**, and the residual color, **14**.

5 DETERMINATION.

Transfer a 50 cc. aliquot of the filtrate to a separatory funnel and extract with 4 successive 15 cc. portions of ether (previously washed twice with an equal volume of water to remove alcohol). Wash the combined ether solutions 4 or 5 times with 2% ammonium hydroxid solution (2% NH_3 by weight), using 10 cc. the first time and 5 cc. thereafter, and reserve the ether solution for the determination of coumarin. Slightly acidify the combined ammoniacal solutions with hydrochloric acid; cool and extract in a separatory funnel with 4 portions of washed ether, using about 40 cc. altogether. Evaporate the ethereal solutions at room temperature, dry over sulphuric acid and weigh. If the residue is considerably discolored or gummy, re-extract in the dry state with boiling petroleum ether (b. p. 40°C. or below) not less than 15 times; evaporate the solvent, dry and weigh. The residue should now be white, crystalline vanillin, with a melting point of approximately

63.54
21.6/21

80°C. A small amount of this residue, dissolved in 2 drops of concentrated hydrochloric acid, should develop a pink color upon the addition of a crystal of resorcin.

Evaporate at room temperature the original ether extract of the sample, from which the vanillin has been removed by means of ammonium hydroxid, and dry over sulphuric acid. The residue, if pure coumarin, should melt at approximately 67°C. and should respond to Leach's test for coumarin as follows: A small portion of the residue, dissolved in not more than 0.5 cc. of hot water, should yield a brown precipitate upon the addition of a few drops of N/10 iodine. This precipitate finally gathers in green flecks, leaving a clear, brown solution. The reaction is especially marked if the reagent is applied with a glass rod to a few drops of the solution on a white plate or tile.

6**NORMAL LEAD NUMBER².—TENTATIVE.**

To a 10 cc. aliquot of the filtrate from the lead acetate precipitate, as obtained in 4, add 25 cc. of water, 0.5–1.0 cc. of sulphuric acid, and 100 cc. of 95% alcohol by volume. Let stand overnight, filter on a Gooch crucible, wash with 95% alcohol, dry at a moderate heat, ignite at low redness for 3 minutes, taking care to avoid the reducing flame, and weigh. Conduct a blank determination employing water containing 4 or 5 drops of glacial acetic acid in place of the sample. The normal lead number is calculated by the following formula:

$$P = \frac{100 \times 0.6831 (S - W)}{5} = 13.662 (S - W) \text{ in which}$$

P = normal lead number (grams of metallic lead in the precipitate obtained from 100 cc. of the sample);

S = grams of lead sulphate corresponding to 2.5 cc. of the lead acetate solution as determined in a blank analysis; and

W = grams of lead sulphate obtained in 10 cc. of the filtrate from the lead acetate precipitate, as obtained in 4.

7**TOTAL SOLIDS.—TENTATIVE.**

Proceed as directed under IX, 4, employing 10 grams of the sample.

8**ASH.—OFFICIAL.**

Evaporate 10 grams of the extract and determine the ash as directed under VIII, 4.

9**ASH CONSTITUENTS.—TENTATIVE.**

Proceed as directed under III or XXVIII, 21–26, inclusive.

10**SUCROSE.—TENTATIVE.**

Determine as directed under VIII, 14 or 18.

VANILLA RESINS.**11****Qualitative Test.—Tentative.**

Place 50 cc. of the extract in a glass dish and evaporate the alcohol on a water bath. When the alcohol is removed, make up to about the original volume with hot water. If alkali has not been used in the manufacture of the extract, the resins will appear as a flocculent red to brown residue. Acidify with acetic acid to free the resins from the bases, separating the resins completely and leaving a partly decolorized, clear, supernatant liquid after standing a short time. Collect the resins on a filter, wash with water and reserve the filtrate for further tests.

Place a portion of the filter with the attached resins in a few cc. of dilute potassium hydroxid solution. The resins are dissolved, giving a deep red solution; acidify, and the resins are precipitated.

Dissolve a portion of the resins in alcohol. To one portion add a few drops of ferric chlorid solution; to another portion hydrochloric acid; neither produces any marked change in color. Most resins, however, in alcoholic solution give color reactions with ferric chlorid or hydrochloric acid.

To a portion of the filtrate obtained above add a few drops of basic lead acetate solution. The precipitate is so bulky as to almost solidify, due to the excessive amount of organic acids, gums and other extractive matter. The filtrate from this precipitate is almost colorless.

Test another portion of the filtrate from the resin for tannin with a solution of gelatin. Tannin is present in varying but small quantities, but should not be present in great excess.

12 METHYL ALCOHOL.—TENTATIVE.

Proceed as directed under XVII, 16, 17 or 18, using the distillate from the determination of alcohol, 2.

13 COLOR VALUE.—TENTATIVE.

Pipette 2 cc. of the extract into a 50 cc. graduated flask and make up to the mark with a mixture of equal parts of 95% alcohol by volume and water. Determine the color value of this diluted extract in terms of red and yellow by means of a Lovibond tintometer, using a 1 inch cell. To obtain the color value of the original extract, multiply the figures for each color by 25.

14 RESIDUAL COLOR AFTER PRECIPITATION WITH LEAD ACETATE³.—TENTATIVE.

Determine the color value, in terms of red and yellow, of the filtrate from the lead acetate precipitate as obtained in 4, using a 1 inch Lovibond cell. Multiply the reading by 2 to reduce the results to the basis of the original extract. If the actual reading of the solution is greater than 5 red and 15 yellow, as may happen if the extract is highly colored with caramel, a $\frac{1}{2}$ or $\frac{1}{4}$ inch cell should be employed, and the readings multiplied, respectively, by 4 or 8. Divide the figures for red and yellow, respectively, by the corresponding figures of the original extract and multiply the quotients by 100, to obtain the percentages of the 2 colors remaining in the lead acetate filtrate.

Calculate also the ratio of red to yellow in both extract and lead acetate filtrate.

COLORS INSOLUBLE IN AMYL ALCOHOL.

15 Modified Marsh Method.—Tentative.

Proceed as directed under XVII, 22, using 25 cc. of the extract and shaking with 25 cc. of the Marsh reagent instead of 20 cc.

16 COLORING MATTERS OTHER THAN CARAMEL.—TENTATIVE.

Proceed as directed under XI.

LEMON AND ORANGE EXTRACTS.

17 SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer.

18

ALCOHOL.—TENTATIVE.

Dilute 50 cc. of the extract, measured at 20°C., with water to about 200 cc., allow the mixture to stand until the oil separates in a clear layer at the top, or centrifugalize, then make up to the mark, using the lower meniscus of the oil. Pour the mixture into a dry Erlenmeyer flask containing 5 grams of light magnesium carbonate, stopper, shake well and filter quickly through a large, dry, folded filter. Introduce a 150 cc. aliquot of the filtrate, measured at 20°C., into a 300–500 cc. distillation flask, attach the flask to a vertical condenser and distil almost 100 cc. Complete the volume of the distillate to 100 cc. at 20°C., mix well and determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$. Ascertain the corresponding per cent of alcohol by volume from **XVI, 5** and multiply the result thus obtained by $\frac{23}{25}$ to obtain the percentage of alcohol by volume in the original sample.

19

GLYCEROL.—TENTATIVE.

Proceed as directed under **3**.

LEMON AND ORANGE OILS.

20

By Polarization. (Mitchell Method)—Tentative.

Without diluting polarize the extract at 20°C. in a 200 mm. tube. Divide the reading in degrees Ventzke by 3.2 in the case of lemon extract and by 5.2 in the case of orange extract; in the absence of other optically active substances, the result will be the percentage of oil by volume. A small amount of cane sugar is occasionally present; if so, determine as directed under **28** and correct the reading accordingly.

21

By Precipitation. (Mitchell Method)—Tentative.

Pipette 20 cc. of the extract into a Babcock milk bottle, add 1 cc. of hydrochloric acid (1 to 1), then 25–28 cc. of water previously warmed to 60°C., mix, let stand in water at 60°C. for 5 minutes, centrifugalize for 5 minutes, fill with warm water to bring the oil into the graduated neck of the flask, again centrifugalize for 2 minutes, place the flask in water at 60°C. for a few minutes and note the per cent of oil by volume. If oil of lemon is present in amounts over 2%, add 0.4% to the percentage of oil noted to correct for the solubility of the oil. If less than 2% and more than 1% is present, add 0.3% for this correction.

When the extract is made in accordance with the U. S. P., the results by the methods given under **20** and **21** should agree within 0.2%.

To obtain the per cent by weight from the per cent by volume, as found by either of these methods, multiply the volume percentage by 0.86 in the case of lemon extracts, and by 0.85 in the case of orange extracts, and divide the result by the specific gravity of the original extract.

TOTAL ALDEHYDES.

Chace Method⁴.—Tentative.

22

REAGENTS.

(a) *Aldehyde-free alcohol.*—Allow 95% alcohol by volume, containing 5 grams of meta-phenyldiamin hydrochlorid per liter, to stand for 24 hours with frequent shaking. (Nothing is gained by previous treatment with potassium hydrid.) Boil under a reflux condenser for at least 8 hours, longer if necessary, allow to stand overnight and distil, rejecting the first 10 and the last 5 per cent

which come over. Store in a dark, cool place in well filled bottles. Twenty-five cc. of this alcohol, on standing 20 minutes at 14°–16°C. with 20 cc. of the sulphite fuchsin solution, should develop only a faint pink coloration. If a stronger color is developed, repeat the treatment with meta-phenylindiamin hydrochlorid as above.

(b) *Sulphite-fuchsin solution*.—Dissolve 0.5 gram of fuchsin in 250 cc. of water, add an aqueous solution of sulphur dioxide containing 16 grams of the gas, allow to stand until colorless, or nearly so, and make up to 1 liter with water. Let stand 12 hours before using and keep in a refrigerator. This solution is liable to deteriorate and should be reasonably fresh when used.

(c) *Standard citral solution*.—Use 0.5 or 1 mg. of C.P. citral per cc. in 50% aldehyde-free alcohol.

23

DETERMINATION.

Weigh approximately 25 grams of the extract in a stoppered weighing flask, transfer to a 50 cc. flask and make up to the mark at room temperature with aldehyde-free alcohol. Measure, at room temperature, 2 cc. of this solution into a comparison tube. Add 25 cc. of the aldehyde-free alcohol (previously cooled to 14°–16°C.), then 20 cc. of the sulphite-fuchsin solution (also cooled) and finally make up to the 50 cc. mark with more aldehyde-free alcohol. Mix thoroughly, stopper and keep at 14°–16°C. for 15 minutes. Prepare a standard for comparison at the same time and in the same manner, using 2 cc. of the standard citral solution, and compare the colors developed. Calculate the amount of citral present and repeat the determination, using a quantity sufficient to give the sample approximately the strength of the standard. From this result calculate the amount of citral in the sample. If the comparisons are made in Nessler tubes, standards containing 1, 1.5, 2, 2.5, 3, 3.5, and 4 mg. of citral, may be prepared and the trial comparison made against these, the final comparison being made with standards lying between 1.5 and 2.5 mg. with 0.25 mg. increments.

It is absolutely essential to keep the reagents and comparison tubes at the required temperature, 14°–16°C. Where the comparisons are made in a bath (this being possible only where the bath is of glass), the standards should be discarded within 25 minutes after adding the sulphite-fuchsin solution. Give samples and standards identical treatment.

CITRAL.

Hiltner Method⁵.—Tentative.

24

REAGENTS.

(a) *Meta-phenylendiamin hydrochlorid solution*.—Prepare a 1% solution of meta-phenylendiamin hydrochlorid in 95% alcohol by volume. Decolorize, if necessary, by shaking with fuller's earth and filter through a double filter. The solution should be bright and clear, free from suspended matter, and practically colorless. Prepare this solution only for immediate use.

(b) *Alcohol*.—For the analysis of lemon extracts, 90–95% alcohol by volume should be used, but for terpeneless extracts, 40–50% alcohol by volume is sufficient. Filter to remove any suspended matter. The alcohol need not be purified from aldehyde. If not practically colorless, render slightly alkaline with sodium hydroxid and distil.

25

DETERMINATION.

All of the operations may be carried on at room temperature. Weigh 25 grams of the extract into a 50 cc. graduated flask and make up to the mark with alcohol.

Stopper the flask and mix the contents thoroughly. Pipette 2 cc. of this solution into a colorimeter tube; add 10 cc. of the meta-phenyldiamin hydrochlorid solution and complete the volume to 50 cc. (or other standard volume) with alcohol. Compare at once the color with that of the standard, prepared at the same time, using 2 cc. of standard citral solution and 10 cc. of the meta-phenyldiamin hydrochlorid solution, and making up to standard volume with alcohol. From the result of this first determination, calculate the amount of standard citral solution that should be used in order to give approximately the same citral strength as the sample under examination; then repeat the determination.

26

TOTAL SOLIDS.—OFFICIAL.

Proceed as directed under XVII, 5, employing 10 cc. of the sample measured at 20°C.

27

ASH.—OFFICIAL.

Ignite the residue from 10 cc. of the extract as directed under VIII, 4.

28

SUCROSE.—TENTATIVE.

Neutralize the normal weight of the extract, evaporate to dryness, wash several times with ether, dissolve in water and determine as directed under VIII, 14 or 18.

29

METHYL ALCOHOL.—TENTATIVE.

Proceed as directed under XVII, 16, 17 or 18, using the distillate from the determination of alcohol, 18.

COLORING MATTERS.

30

GENERAL.—TENTATIVE.

Proceed as directed under XI.

31

LEMON AND ORANGE PEEL COLOR.

Albrech Method.—Tentative.

Place a few cc. of the extract in each of 2 test tubes; to one, add slowly 3–4 volumes of concentrated hydrochloric acid; to the other, several drops of concentrated ammonium hydroxid. If the color is due to lemon or orange peel only it is materially deepened by such treatment.

LEMON AND ORANGE OILS.

32

SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer.

33

INDEX OF REFRACTION.—TENTATIVE.

Determine the index of refraction with any standard instrument, making the reading at 20°C.

34

OPTICAL ROTATION.—TENTATIVE.

Determine the rotation at 20°C. with any standard instrument, using a 50 mm. tube and sodium light. The results should be stated in angular degrees on a 100 mm. basis. If instruments having the sugar scale are used, the reading on orange oils is above the range of the scale, but readings may be obtained by the use of

standard laevo-rotatory quartz plates, or by the use of a 25 mm. tube. The true rotation cannot be obtained by diluting the oil with alcohol and correcting the rotation in proportion to the dilution.

CITRAL.

Kleber Method⁶.—Tentative.

35

REAGENTS.

(a) *Phenylhydrazin solution*.—Prepare a 10% solution in absolute alcohol. A sufficiently pure product can be obtained by distilling the commercial article, rejecting the first portions coming over which contain ammonia.

(b) *N/2 hydrochloric acid*.

36

DETERMINATION.

Weigh 15 grams of the sample into a small, glass-stoppered flask; add 10 cc. of the phenylhydrazin solution. Allow to stand 30 minutes at room temperature, titrate with N/2 hydrochloric acid, using either methyl or ethyl orange as an indicator. Titrate similarly 10 cc. of the phenylhydrazin solution. The difference in the number of cc. of N/2 acid used in these 2 titrations, multiplied by the factor 0.076, gives the weight of citral in the sample. If difficulty is experienced in detecting the end point of the reaction, titrate until the solution is distinctly acid, transfer to a separatory funnel, and draw off the alcoholic portion. Wash the oil with water, adding the washings to the alcoholic solution, titrate back with N/2 alkali and make the necessary corrections.

37

Hiltner Method⁶.—Tentative.

Weigh 2 grams of lemon oil or 8 grams of orange oil into a 100 cc. graduated flask, dilute to the mark with 95% alcohol by volume and proceed as under 25, using 2 cc. of the dilute solution for the comparison.

TOTAL ALDEHYDES.

38

Chace Method⁸.—Tentative.

Weigh a small quantity of the sample into a small, stoppered flask and dilute with aldehyde-free alcohol in the proportion of 2 grams of lemon oil or 4 grams of orange oil to 100 cc. of solution. Determine the total aldehydes as directed under 23, expressing the result as citral.

39

PHYSICAL CONSTANTS OF THE 10 PER CENT DISTILLATE⁷.—TENTATIVE.

Place 50 cc. of the sample in a 3-bulb Ladenburg flask having the main bulb 6 cm. in diameter and of 120 cc. capacity and the condensing bulbs of the following dimensions: 3.5 cm., 3 cm., 2.5 cm.; the distance from the bottom of the flask to the opening of the side arm should be 20 cm. Distil the oil at the rate of 2 cc. per minute until 5 cc. have been distilled. Determine the refractive index and rotation of this distillate as directed in 33 and 34.

40

PINENE.

Chace Method⁸.—Tentative.

Mix the 10% distillate, obtained in 39, with 5 cc. of glacial acetic acid; cool the mixture thoroughly in a freezing bath and add 10 cc. of ethyl nitrite. Then add

slowly, with constant stirring, 2 cc. of hydrochloric acid (2 to 1). Keep the mixture in the freezing bath 15 minutes. Filter off the crystals formed, using suction, and wash with 95% alcohol by volume. Return the combined filtrate and washings to the freezing bath for 15 minutes. Filter off the crystals formed, using the original filter paper. Wash the combined crops of crystals thoroughly with alcohol. Dry at room temperature and dissolve in a minimum amount of chloroform. Add methyl alcohol to the chloroform solution, a little at a time, until the nitroso-chlorids crystallize out, mount the separated and dried crystals in olive oil and examine under the microscope. Pinene nitroso-chlorid crystals have irregular pyramidal ends while limonene nitroso-chlorid crystallizes in needles.

ALMOND EXTRACT.

41

ALCOHOL.—TENTATIVE.

Inasmuch as almond extract usually contains only about 1% of almond oil the alcohol can, in most cases, be calculated from the specific gravity of the extract. If the extract is high in solids, determine the alcohol as follows: Add 25 cc. of the extract, measured at 20°C., to 75 cc. of saturated sodium chlorid solution in a separatory funnel and extract twice with 50 cc. portions of petroleum ether (b. p. 40°–60°C.). Collect the petroleum ether extract in a second separatory funnel and wash twice with 2 portions (25 cc.) of saturated brine. Combine the original salt solution with the washings; add a little powdered pumice and distil into a 100 cc. flask. When almost 100 cc. have been distilled, make up to the mark with water at 20°C. and determine alcohol from the specific gravity, as directed under XVII, 4.

BENZALDEHYDE.—TENTATIVE.

42

REAGENT.

Phenylhydrazin solution.—Add 1.5 cc. of glacial acetic acid to 20 cc. of water and mix with 1 cc. of phenylhydrazin.

43

DETERMINATION.

Measure out 2 portions of 10 cc. each of the extract into 300 cc. Erlenmeyer flasks and add 10 cc. of the phenylhydrazin solution to 1 flask and 15 cc. to the other. Allow to stand overnight in a dark place, add 200 cc. of water and filter on a tared Gooch crucible, provided with a thin layer of asbestos. Wash first with cold water, finally with 10 cc. of 10% alcohol, and dry for 3 hours in a vacuum oven at 70°C., or to constant weight over sulphuric acid. The weight of the precipitate multiplied by the factor 5.408 gives the weight of benzaldehyde in 100 cc. of the sample. If duplicate determinations do not agree, repeat the operation using a larger quantity of the phenylhydrazin solution.

HYDROCYANIC ACID.

44

Qualitative Test.—Tentative.

Add several drops of ferrous sulphate solution and a single drop of ferric chlorid solution to several cc. of the extract. Mix thoroughly, add sodium hydroxid solution, drop by drop, until no further precipitate forms and then dilute hydrochloric acid to dissolve the precipitated hydroxids. In the presence even of small amounts of hydrocyanic acid, a Prussian blue coloration or suspension will develop.

45

Quantitative Method.—Tentative.

(In the absence of chlorids.)

Measure 25 cc. of the extract into a small flask and add 5 cc. of freshly precipitated magnesium hydroxid (chlorin-free). Titrate with N/10 silver nitrate solution, using potassium chromate as an indicator; 1 cc. of N/10 silver nitrate is equivalent to 0.00268 gram of hydrocyanic acid.

NITROBENZOL.

46

Qualitative Test.—Tentative.

Boil a few cc. of the extract with some zinc dust and acetic acid and filter. Add to the filtrate a drop of chloroform, make strongly alkaline with sodium hydroxid solution and heat. The presence of nitrobenzol in the original extract is indicated by the development of the characteristic odor of phenylisonitrile.

CASSIA, CINNAMON AND CLOVE EXTRACTS.

47

ALCOHOL.—TENTATIVE.

Determine as directed under 41.

OIL.

48

Hortvet and West Method⁹.—Tentative.

Transfer 10 cc. of the extract to a separatory funnel, add 30 cc. of water, acidify with 1 cc. of hydrochloric acid (1 to 1) and extract 3 times with ether, using not less than 100 cc. altogether. Wash the combined ether solutions twice with water and, in the case of cinnamon extract, dry by shaking with a small amount of granulated calcium chlorid. Transfer to a tared, wide-mouthed weighing bottle and evaporate the ether as rapidly as possible on a boiling water bath, rotating the liquid upon the sides of the bottle in order to rid the residual oil of traces of ether. Weigh the residue and divide the weight by the specific gravity of the oil in order to obtain the per cent of oil by volume. In the case of clove oil, allow the weighing bottle to remain in the balance case until the usual film of moisture has evaporated. The time of weighing, however, should not be delayed over 3 minutes.

Determine the refractive index of the residual oils at 20°C.

Dissolve a drop of the oil in several drops of alcohol and add a drop of ferric chlorid solution. The following tabulation gives the specific gravity, refractive index at 20°C. and color reaction with ferric chlorid solution:

OIL	SPECIFIC GRAVITY	REFRACTIVE INDEX AT 20°C.	COLOR REACTION WITH FERRIC CHLORID SOLUTION
Cassia.....	1.05	1.585-1.600	Brown
Cinnamon.....	1.03	1.590-1.599	Green
Cloves.....	1.055	1.560-1.565	Deep blue

GINGER EXTRACT.

49

ALCOHOL.—TENTATIVE.

Determine as directed under XVII, 4.

50

SOLIDS.—TENTATIVE.

Evaporate 10 cc. of the extract nearly to dryness on a water bath, dry for 2 hours in a water oven and weigh.

GINGER.

51

Seeker Method.—Tentative.

Dilute 10 cc. of the extract to 30 cc., evaporate to 20 cc., decant into a separatory funnel and extract with an equal volume of ether. Allow the ether to evaporate spontaneously in a porcelain dish, and to the residue add 5 cc. of 75% sulphuric acid and about 5 mg. of vanillin. Allow to stand 15 minutes and add an equal volume of water; in the presence of ginger extract an azure blue color develops.

CAPSICUM.

52

La Wall Method Modified by Doyle.—Tentative.

To 10 cc. of the extract add cautiously dilute sodium hydroxid solution until the solution reacts very slightly alkaline with litmus paper. Evaporate at about 70°C. to approximately one fourth the original volume, render slightly acid with dilute sulphuric acid, testing with litmus paper. Transfer to a separatory funnel, rinsing the dish with water, and extract with an equal volume of ether, avoiding emulsification by shaking the funnel gently 1–2 minutes. Draw off the lower layer and wash the ether extract once with about 10 cc. of water. Transfer the washed ether extract to a small evaporating dish, render decidedly alkaline with N/2 alcoholic potassium hydroxid and evaporate at about 70°C. until the residue is pasty; then add about 20 cc. more of N/2 alcoholic potash and allow to stand on a steam bath until the gingerol is completely saponified (about 30 minutes). Dissolve the residue in a little water and transfer with water to a small separatory funnel. The volume should not exceed 50 cc. Extract the alkaline solution with an equal volume of ether. Wash the ether extract repeatedly with small amounts of water until no longer alkaline to litmus. Transfer the washed extract to a small evaporating dish, and allow the ether to evaporate spontaneously. Finally test the residue for capsicum by moistening the tip of the finger, rubbing it on the bottom and sides of the dish, and then applying the finger to the end of the tongue. A hot, stinging or prickly sensation, which persists for several minutes, indicates capsicum or other foreign pungent substances.

PEPPERMINT, SPEARMINT AND WINTERGREEN EXTRACTS.

53

ALCOHOL.—TENTATIVE.

Proceed as directed under 41.

OIL.

54

Howard Method¹⁰ Modified.—Tentative.

Pipette 10 cc. of the extract into a Babcock milk bottle, add 1 cc. of carbon disulphid, mix thoroughly, then add 25 cc. of cold water and 1 cc. of concentrated hydrochloric acid. Close the mouth of the bottle and shake vigorously; centrifugalize for 6 minutes and remove all but 3–4 cc. of the supernatant liquid, which should be practically clear, by aspirating through a glass tube of small bore. Connect the stem of the bottle with a filter pump, immerse the bottle in water kept at approximately 70°C. for 3 minutes, remove from the bath every 15 seconds and shake vigorously. Continue in the same manner for 45 seconds, using a boiling water bath. Remove from the bath and shake while cooling. Disconnect from the suction and fill the bottle to the neck with saturated salt solution at room temperature, centrifugalize for 2 minutes and read the volume of the separated oil from the top of the meniscus. Multiply the reading by 2 to obtain the per cent of oil by volume.

In the case of wintergreen, use as a floating medium a mixture of 1 volume of concentrated sulphuric acid and 3 of saturated sodium sulphate solution.

METHYL SALICYLATE IN WINTERGREEN EXTRACT.

55

Hortvet and West Method⁹ Modified.—Tentative.

Mix 10 cc. of the extract with 10 cc. of 10% potassium hydroxid solution. Heat on the steam bath until the volume is reduced about one half, add a distinct excess of hydrochloric acid (1 to 1), cool and extract with 3 portions of ether, 40, 30 and 20 cc., respectively. Filter the extract through a dry filter into a weighed dish, wash the paper with 10 cc. of ether and allow the filtrate and washings to evaporate spontaneously. Dry in a desiccator containing sulphuric acid and weigh. Multiply the weight of salicylic acid so found by 9.33 to obtain the per cent by volume of wintergreen oil in the sample.

ANISE AND NUTMEG EXTRACTS.

OIL.

56

Hortvet and West Method⁹.—Tentative.

To 10 cc. of the extract in a Babcock milk bottle add 1 cc. of hydrochloric acid (1 to 1), then sufficient half saturated salt solution, previously heated to 60°C., to fill the flask nearly to the neck. Cork and let stand in water at 60°C. for about 15 minutes, rotate occasionally and centrifugalize for 10 minutes at about 800 revolutions per minute. Add brine till the oil rises into the neck of the bottle, and again centrifugalize for 10 minutes. If the separation is not satisfactory, or the liquid is not clear, cool to about 10°C. and centrifugalize for an additional 10 minutes. Multiply the reading by 2 to obtain the percentage of oil by volume.

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XXI. MEAT AND MEAT PRODUCTS.

MEAT.

1

PREPARATION OF SAMPLE.—TENTATIVE.

In the case of fresh meat, separate the sample as completely as possible from the bones and pass through a sausage mill rapidly and repeatedly until thoroughly mixed and macerated. Chill the sample to prevent decomposition and begin all determinations as soon as practicable after the sample is prepared.

In the case of canned meats, pass the entire contents of a can through a sausage mill as directed above. Remove sausage from the casings and mix by repeated grinding in a sausage mill. Dry the portion of the sample, which is not needed for analysis, either in vacuo or by evaporating with alcohol, extract the fat with gasoline (b. p. below 60°C.), allow the gasoline to evaporate spontaneously and expel the last traces by heating for a short time on the steam bath. Do not heat the meat or separated fat longer than necessary, owing to the tendency of the latter to decompose. Reserve the fat for examination according to the methods given under XXIII. Keep the fat in a cool place and complete the examination before the fat becomes rancid.

2

MOISTURE.—TENTATIVE.

Proceed as directed under VIII, 2 or 3, using the latter method in cases in which it is desired to employ the dried sample for further determinations.

3

ASH.—OFFICIAL.

Proceed as directed under VIII, 4.

4

CRUDE FAT OR ETHER EXTRACT.—OFFICIAL.

Proceed as directed under VIII, 10.

5

TOTAL PHOSPHORUS.—TENTATIVE.

Destroy the organic matter as directed under I, 5 (a), (b), (c) or (d) and proceed as directed under I, 6 or 9.

WATER-SOLUBLE PHOSPHORUS¹.—TENTATIVE.

6

PREPARATION OF SOLUTIONS.

(a) *Cold water extract of flesh*.—Weigh out 10–12 grams of fresh muscle and divide equally between 2 small beakers. Moisten the sample with a few cc. of water, and break up the lumps with a glass rod. Add 50 cc. of water to each beaker and stir the contents for 15 minutes. Allow the insoluble residue to settle for 3–5 minutes, decant the liquid through filters into beakers and add 25 cc. of water to each residue. Stir 7–8 minutes and, after allowing to settle, decant onto the same filter. Continue this treatment, using 25 cc. of water each time, until the filtrates measure about 230 cc. each. Allow the filters to drain completely between extractions. Whenever the major portion of the residue has been mechanically transferred to the filter, return it to the beaker. After the last extraction, transfer the entire con-

tents of each beaker onto the filter and, when drained, wash twice with small quantities of water. Combine the 2 extracts.

(b) *Hot water-ammonium sulphate extract of blood*.—Weigh out 30–35 grams of fresh whole blood as caught from the animal into a porcelain mortar. Grind and transfer to a 400 cc. beaker with hot water. Make up to about 150 cc. with boiling water. Place over a flame, gradually bring to boiling, with constant stirring, then add 20 cc. of 20% ammonium sulphate solution and continue boiling, with constant stirring, for about 10 minutes. Decant onto an 18 cm. filter paper, receiving the filtrate in an 800 cc. beaker. Transfer the coagulum from the filter, along with that remaining in the beaker, to a mortar. Grind to a smooth paste and transfer to a beaker with boiling 3½% ammonium sulphate solution. Make up to about 50 cc. with the latter, stir for 8 minutes and again filter. Return the coagulum to the mortar and grind again, transferring to the beaker as before with boiling 3½% ammonium sulphate solution. Repeat this process of 8 minute extractions of the coagulum in 3½% ammonium sulphate solution and filtration as directed above, without further grinding, until the filtrate measures about 450 cc. Wash out each beaker twice with 8–10 cc. of hot 3½% ammonium sulphate solution, transferring the coagulum and extract to the filter. Wash the coagulum twice with boiling 3½% ammonium sulphate solution from a wash bottle. Always allow the filter to drain well between additions of extract or wash solutions.

(c) *Hot water-ammonium sulphate extract of liver*.—Weigh by difference from a closed weighing bottle 15–20 grams of finely ground liver into a 400 cc. beaker. Add a few cc. of cold water and beat up with a stirring rod to separate the particles of tissue. Add enough boiling water to make the volume about 150 cc., place over a flame and bring to boiling. Add 10 cc. of 20% ammonium sulphate solution and continue to boil for 10 minutes. Allow to settle for a moment and decant the boiling hot liquid onto an 18 cm. filter. Add 50 cc. of boiling water and stir for 8 minutes, without further heating, and decant onto the filter again. Repeat this addition of 50 cc. of hot water, stirring and decanting 8 times, returning the coagulum to the beaker as soon as any considerable amount collects upon the filter. With the eighth portion of water transfer the entire contents of the beaker onto the filter and wash twice with hot water. Always allow the filter to drain well between additions of extract or wash water.

(d) *Hot water-ammonium sulphate extract of brain*.—Weigh out about 10 grams of brain into a 250 cc. beaker. Add a few cc. of water and work up the brain and water with a glass rod. Make up to about 100 cc. with boiling water, place over a flame, and gradually bring to boiling, with constant stirring. While boiling vigorously (not before) add 20 cc. of 20% ammonium sulphate solution, boil gently for about 10 minutes, allow to settle for a moment and decant slowly onto a linen filter containing acid-washed, glassmaker's sand, receiving the extract in an 800 cc. beaker. Add to the beaker containing the coagulum 50 cc. of 3½% ammonium sulphate solution, stir 1 minute, keep boiling and decant the liquid onto the filter. Repeat this process of 1 minute extractions of the coagulum in 3½% ammonium sulphate solution, and filtration as directed above, until the filtrate measures about 450 cc. Wash out the beaker twice with 8–10 cc. of hot 3½% ammonium sulphate solution, completing the transfer of the coagulum and extract to the sand, and wash the coagulum twice with the wash solution. Always allow the filter to drain well between additions of extract or wash solution.

In making extracts of brain, give careful attention to the handling of the sample. The coagulum is very soft. It should be stirred only enough to keep it in motion. If handled roughly in returning from the sand filter to the beaker, it becomes too

finely divided and retains a great deal of liquid. To prevent the extract or the coagulum from coming into contact with the linen before passing through the sand, pour the extract slowly into a slight depression in the center of the sand or, better yet, into a thin film of absorbent cotton, $1\frac{1}{2}$ inches in diameter, laid over a depression in the sand. The coagulum remains on the cotton and its return to the beaker is facilitated thereby. If the cotton is not broken up by needless stirring, it can be taken out of the beaker with a glass rod and returned to the sand each time a partial extract is to be filtered. Care is necessary to prevent loss through bumping, on account of sand in the beakers during the final extractions. Each partial extract should be boiling hot at the time filtration begins.

7

DETERMINATION.

To the extracts, prepared as directed in **6**, add 50 cc. of magnesia mixture [**I, 4 (c)**] and stir thoroughly. Allow to stand 15 minutes, add 25 cc. of ammonium hydroxid (sp. gr. 0.90), cover and allow to stand 3 days. Filter and wash the precipitate with 2½% ammonium hydroxid. Dissolve the precipitate on the filter paper and that remaining in the beaker in nitric acid (1 to 1) and hot water, receiving the solution in a 400 cc. beaker. Neutralize with ammonium hydroxid, make slightly acid with nitric acid, add 5 grams of ammonium nitrate and determine phosphorus as directed under **I, 6**.

8

TOTAL NITROGEN.—OFFICIAL.

Proceed as directed under **I, 18, 21** or **23**, using about 2 grams of the fresh sample. In the Kjeldahl and Gunning methods digest with sulphuric acid for at least 4 hours; in the Kjeldahl-Gunning-Arnold method for 2 hours after the mixture has become clear.

9

SOLUBLE AND INSOLUBLE NITROGEN.—TENTATIVE.

Exhaust 7–25 grams of the sample depending upon the water content in the following manner: Weigh into a 150 cc. beaker, add 5–10 cc. of cold (15°C.) ammonia-free water and stir to a homogeneous paste. Then add 50 cc. of cold water, stir every 3 minutes for 15 minutes, let stand for 2–3 minutes and decant the liquid upon a quantitative filter, collecting the filtrate in a 500 cc. graduated flask. Drain the beaker, pressing out the liquid from the meat residue by the aid of a glass rod. Add to the residue in the beaker 50 cc. of cold water, stir for 5 minutes and, after standing 2–3 minutes, decant as before. If a considerable portion of the meat is carried over onto the filter, transfer it back to the beaker by means of a glass rod. Repeat the extractions, using the following additional amounts of cold water: 50, 50, 25, 25, 25 and 25 cc. After the last extraction transfer the entire insoluble portion to the filter and wash with three 10 cc. portions of water, allowing the material to drain thoroughly after each addition of water. Dilute to the mark, mix thoroughly and determine the total soluble nitrogen in a 50 cc. aliquot as directed under **I, 18, 21** or **23**. Subtract the percentage of soluble nitrogen from the percentage of total nitrogen, **8**, to obtain the percentage of insoluble nitrogen. To obtain the percentage of insoluble protein multiply the percentage of insoluble nitrogen by 6.25.

10

CONNECTIVE TISSUE NITROGEN.—TENTATIVE.

Exhaust 10 grams of the sample with cold water as directed under **9**, and boil the exhausted residue repeatedly with successive portions of about 100 cc. of water until the total hot water extract amounts to approximately 1 liter. Filter, concentrate and determine nitrogen in the concentrated extract. Multiply the percentage of nitrogen so obtained by 5.55 to obtain the percentage of nitrogenous substances of the connective tissue.

11

COAGULABLE PROTEINS.—TENTATIVE.

(For uncooked meat only.)

Measure 150 cc. of the extract, from **9**, into a 250 cc. beaker and evaporate to 40 cc. on a steam bath, with occasional stirring. Neutralize to phenolphthalein, then add 1 cc. of N/1 acetic acid and boil gently for 5 minutes. The coagulum should separate out at once, leaving a clear liquid. Filter on quantitative paper, wash the beaker thoroughly 4 times with hot water, taking special care to clean the sides. Finally wash the coagulum on the filter 3 times, dilute the combined filtrate and washings to a definite volume and reserve for the determination of proteoses, peptones and gelatin, **12**, and creatin, **16**. Transfer the coagulum with the paper to a Kjeldahl flask and remove, with concentrated sulphuric acid, any of the material adhering to the beaker, taking the usual 25 cc. of acid in 5 cc. portions for this purpose, heating the acid in the beaker on a hot plate and rubbing with a glass rod. Proceed as directed under **8**. Multiply the percentage of nitrogen obtained by 6.25 to obtain the percentage of coagulable proteins.

PROTEOSES, PEPTONES AND GELATIN.

12

Modified Tannin-Salt Method².—Tentative.

Transfer a 50 cc. aliquot of the filtrate, obtained in **11**, to a 100 cc. graduated flask, add 15 grams of sodium chlorid and 10 cc. of cold water, shake until the sodium chlorid has dissolved and cool to 12°C. Add 30 cc. of 24% tannin solution, cooled to 12°C., fill to the mark with water previously cooled to 12°C., shake and allow the mixture to stand at a temperature of 12°C. for 12 hours or overnight. Filter at 12°C., transfer 50 cc. of the filtrate to a Kjeldahl flask and add a few drops of sulphuric acid. Place the flask in a steam bath, connect with a vacuum pump and evaporate to dryness. Determine nitrogen in the residue as directed in **1**, **18**, using 30 cc. of sulphuric acid for the digestion. Conduct a blank determination, using the same amount of reagents, and correct the result accordingly. Multiply the corrected result by 2 and deduct the amount of nitrogen as found from the nitrogen determined in another 50 cc. aliquot of the filtrate from the coagulable proteins without the tannin-salt treatment; the difference multiplied by 6.25 gives the percentage of proteoses, peptones and gelatin.

13

MEAT BASES.—TENTATIVE.

Deduct from the percentage of total nitrogen, **8**, the sum of the percentages of nitrogen, obtained in the determination of insoluble proteins, **9**, coagulable proteins, **11**, and proteoses, peptones and gelatin, **12**, to obtain the percentage of nitrogen of the meat bases. Multiply the result by 3.12 to obtain the percentage of meat bases.

AMMONIA.

Folin Aeration Method³.—Tentative.

14

APPARATUS.

Employ the apparatus illustrated in Fig. 9; *A* is a wash bottle one fourth full of 10% sulphuric acid; *B* is a tube containing the sample; *C* is a rubber disc and *D* is a 5 cc. bulb to prevent spray from being carried over into the tube (*E*) which contains the standard acid; *F* is a safety bottle.

15

DETERMINATION.

Introduce 2–4 grams of the finely divided meat into the tube (*B*) and add 20 cc. of ammonia-free water. Place a measured amount of N/25 or N/50 sulphuric or

hydrochloric acid in tube (E). Then add 1 cc. of saturated potassium oxalate solution to the sample in tube (B), introduce a few drops of kerosene and finally add just sufficient saturated potassium carbonate solution to render the mixture alkaline. Place the tubes in position at once, pass air through the apparatus and titrate the standard acid in tube (E) at hourly intervals, until ammonia ceases to be given off, using methyl red, cochineal or congo red as an indicator. If preferred, the ammonia, collected in tube (E), may also be determined by nesslerizing as directed under IV, 11.

16

CREATIN.—OFFICIAL.

Evaporate an aliquot or the remaining portion of the filtrate and washings from the coagulable proteins, 11, (a portion having been used in 12), to 5–10 cc., transfer with a minimum amount of hot water to a 50 cc. measuring flask, keeping the volume below 30 cc., add 10 cc. of 2N hydrochloric acid and mix. Hydrolyze in an auto-

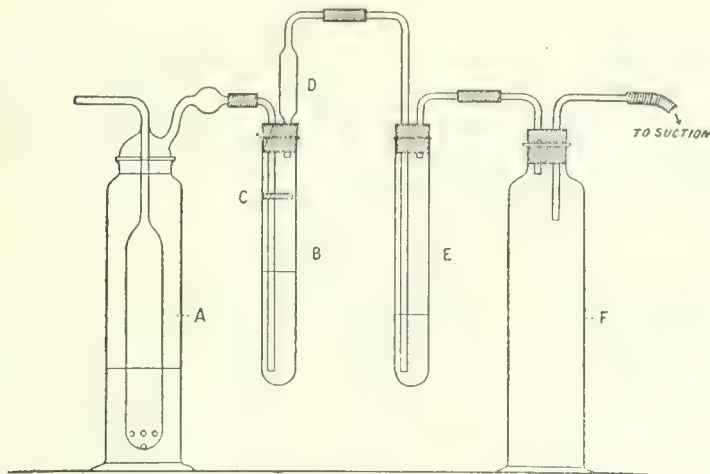


FIG. 9. APPARATUS FOR THE FOLIN AMMONIA DETERMINATION.

clave at 117°–120°C. for 20 minutes, allow the flask to cool somewhat, remove and chill under running water. Partially neutralize the excess of acid by adding 7.5 cc. of 10% sodium hydroxid solution, free from carbonates, dilute to the mark and mix. Make a preliminary reading on 20 cc. to ascertain the volume to use to obtain a reading of approximately 8 mm. and transfer to a 500 cc. graduated flask. Add 10 cc. of 10% sodium hydroxid solution and 30 cc. of saturated picric acid solution (1.2%). Mix and rotate for 30 seconds and let stand exactly 4½ minutes. Dilute to the mark at once with water, shake thoroughly and read in a Duboscq colorimeter, comparing the color with N/2 potassium dichromate solution, set at 8 mm.

If the reading is too high or too low (above 9.5 or below 7 mm.), calculate the quantity necessary to obtain a reading of about 8 mm. The strength of the dichromate solution used must be checked against a standard creatin solution. To obtain the values, divide 81 by the reading and multiply by the volume factor to obtain mg. of creatinin; this value multiplied by 1.16 gives creatin, which, divided by the weight of the sample and multiplied by 100 gives the per cent of creatin.

The use of Kober's shade and the painting of the plunger, as suggested for this nephelometer, assists in getting a sharper end point, relieves the eye strain and may be employed if desired.

Example.—20 grams of meat are extracted with water as in 9, and the extract diluted to 500 cc.; 150 cc. of this latter solution (equivalent to 6 grams of meat) are treated as in 11. The filtrate thus obtained is then evaporated and hydrolyzed as above and then diluted to 50 cc.; 25 cc. of this last solution are treated with sodium hydroxid solution and picric acid solution as directed above and diluted to 500 cc. This latter solution gives a reading of 9 mm.

$$\frac{81}{9} \times \frac{50}{25} = 18 \text{ mg. creatinin;}$$

$$\frac{0.018 \times 1.16 \times 100}{6} = 0.35\% \text{ creatin.}$$

STARCH.

(In chopped meat, sausage, deviled meat, etc.)

17

Qualitative Test.—Tentative.

Treat 5–6 grams of the sample with boiling water for 2–3 minutes, cool the mixture and test the supernatant liquid with iodine solution. In using this test, a small amount of starch may be present as the result of the use of spices. If a marked reaction is given, however, it may be concluded that starch or flour has been added, and a quantitative determination may be made. The qualitative method may be replaced by a microscopic examination, which discloses not only the presence of added starch, but also the variety employed.

18

Mayrhofer Method, Price Modification⁴.—Tentative.

Treat in a 200 cc. beaker 10 grams of the finely divided sample with 75 cc. of an 8% solution of potassium hydroxid in 95% alcohol by volume and heat on a steam bath until all the meat is dissolved (30–45 minutes). Add an equal volume of 95% alcohol, cool and allow to stand for at least an hour. Filter by suction through a thin layer of asbestos in a Gooch crucible. Wash twice with warm 4% potassium hydroxid in 50% alcohol by volume and then twice with warm 50% alcohol. Discard the washings. Retain as much of the precipitate in the beaker as possible until the last washing. Place the crucible with contents in the original beaker, add 40 cc. of water and 25 cc. of concentrated sulphuric acid. Stir during the addition of the acid and make sure that the acid comes in contact with all the precipitate. Allow to stand about 5 minutes, add 40 cc. of water and heat just to boiling, stirring constantly. Transfer the solution to a 250 cc. graduated flask, add 2 cc. of 20% phosphotungstic acid solution, allow to cool to room temperature and make up to the mark with water. Filter through a starch-free filter paper, pipette 100 cc. of the filtrate into a 200 cc. graduated flask, neutralize with sodium hydroxid solution, make up to volume and determine the dextrose present in a 50 cc. portion of the filtrate, as directed under VIII, 25, titrating the cuprous oxide precipitate as directed under VIII, 29. The weight of the dextrose multiplied by 0.9 gives the weight of the starch.

GLYCOGEN.

19

Qualitative Test⁵.—Tentative.

Boil 50 grams of the macerated sample with 50 cc. of water for 15–30 minutes. Filter the broth through moistened filter paper or fine linen. To a portion of the filtrate in a test tube add a few drops of a mixture of 2 parts of iodine, 4 of potassium

iodid and 100 of water. In the presence of a considerable amount of glycogen the latter produces a dark brown color, which is destroyed by heating and reappears on cooling. When starch is present, it may be precipitated by treating the water extract with 2 volumes of glacial acetic acid, filtering and applying the test for glycogen to the filtrate.

Trowbridge and Francis Method⁶.—Tentative.

20

PREPARATION OF SOLUTION.

Weigh out by difference about 25 grams of the finely ground and thoroughly mixed sample. Place in a 400 cc. beaker and mix with 50 cc. of potassium hydroxid solution ($1\frac{1}{2}$ to 1), free from carbonate. Cover the beaker with a watch glass and digest on the water bath for 2 hours, with occasional stirring. At the end of the 2 hours, dilute to approximately 200 cc. with cold water.

Add to the solution an equal volume of 95% alcohol by volume, cover with a watch glass and set aside for 10–12 hours. Decant the supernatant liquid through a folded 18.5 cm. filter, allowing the glycogen to remain in the beaker and wash by decantation with 66% alcohol (2 volumes of 95% alcohol to 1 of water) until the glycogen is white, or nearly so. Usually about 4 washings are required. Transfer the washed precipitate from the beaker to the filter and wash 2–3 times with 66% alcohol. The solution filters slowly and the funnel should be covered with a watch glass to prevent excessive evaporation. The albuminous substance present retards the filtration if permitted to dry on the paper. If the washing by decantation is not made nearly complete, it will be difficult to obtain the glycogen free from the coloring matter.

After the washing is completed, close the bottom of the funnel by a piece of rubber tubing and a pinch-cock. Fill the funnel with warm water, cover with the watch glass and let stand 2–3 hours, or overnight. Open the pinch-cock and allow all of the solution to pass through the filter into a beaker. Close the funnel with the pinch-cock and fill with warm water as before. Allow this water to remain in the funnel for an hour and then filter as before. At first the glycogen solution appears quite turbid. This washing with warm water should be continued until the filtrate becomes perfectly clear. To the solution of glycogen in water, add double its volume of 95% alcohol by volume and let stand overnight to complete the reprecipitation of the glycogen. Filter and wash as before with 66% alcohol.

21

DETERMINATION.

If desired, the last filtration may be made through a tared Gooch crucible and the weight of glycogen determined after drying to constant weight. This gives results that are approximately correct. More satisfactory results are obtained by hydrolyzing the glycogen with dilute hydrochloric acid and determining the resultant dextrose. Dissolve the glycogen on the filter in warm water as directed above, collecting the filtrate and washings in a 300 cc. graduated flask and keeping the volume within 225 cc. Add 12.5 cc. of hydrochloric acid (sp. gr. 1.19) to the combined filtrate and washings, mix and place in a boiling water bath for 3 hours. Cool, neutralize with sodium hydroxid solution, cool again, make up to volume with water and determine dextrose in an aliquot of the solution as directed under VIII, 54, determining the reduced copper as directed under VIII, 29. Multiply the corresponding weight of dextrose by 0.9 to obtain its equivalent of glycogen and correct this result for dilution to obtain the per cent of glycogen in the sample.

SUGAR⁷.—TENTATIVE.

22

REAGENTS.

(a) *Mercuric nitrate solution*.—Warm 220 grams of yellow mercuric oxid with 300 cc. of water and treat with small portions of nitric acid, stirring until dissolved. Make up to 1 liter and filter.

(b) *Phosphotungstic acid solution*.—Prepare a 20% solution of phosphotungstic acid in 2.5% hydrochloric acid.

23

DETERMINATION.

Boil 100 grams of the sample with about 350 cc. of water for about 20 minutes, cool, add an excess (10–30 cc.) of the mercuric nitrate solution, nearly neutralize with sodium hydroxid solution and make up to 500 cc., exclusive of fat. Mix thoroughly, allow the mixture to settle and decant the clear liquid through a large, dry, folded filter. To an aliquot of the filtrate add 1–2 cc. of concentrated hydrochloric acid for each 100 cc., heat to boiling and saturate thoroughly with a rapid current of hydrogen sulphid. Remove the excess of hydrogen sulphid by means of a current of air, cool, make up to a definite volume with water and filter. To an aliquot of the filtrate add an excess of the phosphotungstic acid solution, noting the increase in volume of the solution caused by this addition and place in an ice box for several hours or overnight. Filter, introduce 50 cc. of the filtrate into a 400 cc. beaker, neutralize with concentrated sodium hydroxid solution, add 50 cc. of Soxhlet's solution [VIII, 19], heat so that boiling begins in 4 minutes, boil 2 minutes and filter through an alundum crucible of suitable porosity, using very gentle suction. If the filtrate is green or yellow, refilter through the same crucible until the filtrate is clear blue. Wash the precipitate with a very small amount of 5% sodium hydroxid solution, refiltering the washings if they are turbid. Dissolve the precipitated cuprous oxid in nitric acid (1 to 1) and determine copper as directed under VIII, 29. Find the corresponding amount of dextrose or invert sugar from VIII, 27, and calculate the per cent in the original sample by proper correction for the various aliquots taken in the determination. To convert invert sugar to sucrose multiply by the factor 0.95.

NITRATES.

Schlösing-Wagner Method⁸.—Tentative.

24

REAGENT.

Ferrous chlorid solution.—Dissolve nails or other small pieces of iron in concentrated hydrochloric acid, keeping an excess of iron present until the evolution of gas ceases. Keep the solution in 50 cc. glass-stoppered bottles entirely filled. Employ only freshly opened bottles of the reagent for the determination.

25

APPARATUS.

Provide a 250 cc. flask with a 2-holed rubber stopper. Through one of the holes pass the stem of a funnel having a glass stop-cock, and into the other fit a delivery tube leading downward at an angle from the flask to a trough containing water. Terminate the upper end of the delivery tube just below the rubber stopper in the flask and place the lower end under the surface of the water in the trough, the exit being immediately beneath the mouth of an inverted measuring tube, filled with 40% potassium hydroxid solution. Cover the trough end of the delivery tube with a piece of rubber tubing. Midway on the delivery tube between the flask and the measuring tube place a short length of rubber tubing and a pinch-cock.

26

DETERMINATION.

Extract 100 grams of finely ground meat by boiling repeatedly with successive small portions of water, decanting the extracts through a muslin or paper filter into a casserole, and concentrate the combined extracts to a volume of about 50 cc.

Introduce 50 cc. of the ferrous chlorid solution and 50 cc. of 10% hydrochloric acid into the flask, close the stop-cock of the funnel, open the pinch-cock on the delivery tube, move the end of the latter so that escaping air will not pass into the measuring tube, and boil the contents of the flask until the air is expelled, as indicated by a slight pressure against the fingers when the rubber tubing is compressed after momentary removal of the flame. Now close the delivery tube with the pinch-cock and place the exit end beneath the measuring tube. Introduce the concentrated extract of the sample into the flask, a little at a time, through the funnel tube, opening the pinch-cock on the delivery tube and boiling the contents of the flask at intervals to force the nitric oxid gas into the measuring tube. Finally rinse the casserole and the funnel tube with a little boiled water, add the rinsings to the contents of the evolution flask in the manner just described and boil until nitric oxid no longer passes over into the measuring tube. Calculate the volume of nitric oxid at 0°C. and 760 mm. pressure. One cc. of nitric oxid at 0°C. and 760 mm. pressure is equivalent to 0.004512 gram of potassium nitrate.

Phenoldisulphonic Acid Method⁹.—Tentative.

27

REAGENTS.

(a) *Phenoldisulphonic acid solution.*—Heat 6 grams of phenol with 37 cc. of concentrated sulphuric acid on a water bath, cool and add 3 cc. of water.

(b) *Standard comparison solution.*—Dissolve 1 gram of pure, dry potassium nitrate in water and dilute to 1 liter. Evaporate 10 cc. of this solution to dryness on a steam bath, add 2 cc. of the phenoldisulphonic acid solution, mix quickly and thoroughly by means of a glass rod, heat for about a minute in a steam bath and dilute to 100 cc. One cc. of this solution is equivalent to 0.1 mg. of potassium nitrate. Prepare a series of standard comparison tubes by introducing amounts ranging from 1–20 cc. of this solution (0.1–2.0 mg. of potassium nitrate) into 50 cc. Nessler tubes, adding 5 cc. of strong ammonium hydroxid to each and diluting to 50 cc. These standard tubes are permanent for several weeks if kept tightly stoppered.

28

DETERMINATION.

Weigh 1 gram of the sample into a 100 cc. flask, add 20–30 cc. of water and heat on a steam bath for 15 minutes, shaking occasionally. Add 3 cc. of saturated silver sulphate solution for each per cent of sodium chlorid present, then 10 cc. of basic lead acetate solution and 5 cc. of alumina cream, shaking after each addition. Make up to the mark with water, shake and filter through a folded filter, returning the filtrate to the filter until it runs through clear. Evaporate 25 cc. of the filtrate to dryness, add 1 cc. of the phenoldisulphonic acid solution, mix quickly and thoroughly by means of a glass rod, add 1 cc. of water and 3 or 4 drops of concentrated sulphuric acid and heat on a steam bath for 2–3 minutes, being careful not to char the material. Then add about 25 cc. of water and an excess of ammonium hydroxid, transfer to a 100 cc. graduated flask, add 1–2 cc. of alumina cream if not perfectly clear, dilute to the mark with water and filter. Fill a 50 cc. Nessler tube to the mark with the filtrate and determine the amount of potassium nitrate present in

the sample by comparison with the standard comparison tubes. If the solution is too dark for comparison with the standards, dilute with water and correct the result accordingly.

29

PRESERVATIVES.—TENTATIVE.

Proceed as directed under X.

30

METALS.—TENTATIVE.

Proceed as directed under XII.

31

COLORING MATTERS.—TENTATIVE.

Proceed as directed under XI.

MEAT EXTRACTS AND SIMILAR PRODUCTS.

32

PREPARATION OF SAMPLE.—TENTATIVE.

Remove liquid and semi-liquid meat extracts and similar preparations from the container and mix thoroughly before sampling. A little heating expedites the mixing of pasty extracts. In many liquid preparations a sediment forms which should be carefully removed from the bottom of the container and included in the sample. If the sample is in the form of cubes, grind 10-12 of the cubes in a mortar.

33

MOISTURE.—TENTATIVE.

Proceed as directed under VIII, 2, employing about 2 grams of powdered preparations, about 3 grams of pasty preparations, or 5-10 grams of liquid extracts, according to the solid content. Dry the powdered preparations directly without admixture. Dissolve the pasty preparations in water and dry with sufficient ignited sand, asbestos or pumice stone to absorb the solution. When glycerol is present, proceed as directed under VIII, 3.

34

ASH.—OFFICIAL.

Proceed as directed under VIII, 4. Add sufficient water to pasty preparations to effect solution and evaporate to dryness in order that the solids may be distributed evenly over the bottom of the dish.

35

TOTAL PHOSPHORUS.—TENTATIVE.

Proceed as directed under 5.

36

CHLORIN.—TENTATIVE.

Dissolve about 1 gram of the sample, prepared as directed in 32, in 20 cc. of 5% sodium carbonate solution and proceed as directed under III, 18.

37

FAT.—TENTATIVE.

Transfer the residue from the determination of moisture to a continuous extraction apparatus and proceed as directed under VIII, 10.

38

TOTAL NITROGEN.—OFFICIAL.

Proceed as directed under I, 18, 21 or 23.

39

INSOLUBLE PROTEIN¹⁰.—TENTATIVE.

Dissolve 5 grams of powdered preparations, 8–10 grams of pasty extracts, or 20–25 grams of fluid extracts, in cold water. Filter and wash with cold water. Transfer the filter paper and contents to a Kjeldahl flask and determine nitrogen as directed under **I, 18, 21 or 23**. However, if a large amount of insoluble matter is present, transfer the weighed sample to a graduated flask, make up to a definite volume, shake thoroughly, filter through a folded filter and determine nitrogen in an aliquot of the filtrate. Deduct the percentage of nitrogen in the total filtrate from the percentage of total nitrogen, **38**, to obtain the percentage of nitrogen in the insoluble protein. Multiply this percentage of nitrogen by 6.25 to obtain the percentage of insoluble protein.

40

COAGULABLE PROTEIN.—TENTATIVE.

Prepare a solution of the sample as directed in **39**. Employ as large an aliquot of the filtrate as practicable or an aliquot of the filtrate from the insoluble protein, **39**, and neutralize to phenolphthalein by the addition of acetic acid or sodium hydroxide, whichever may be necessary, add 1 cc. of N/1 acetic acid, boil for 2–3 minutes, cool to room temperature, dilute to 500 cc. and pass through a folded filter.

Determine nitrogen in 50 cc. of the filtrate as directed under **I, 18, 21 or 23**. Ten times the percentage of nitrogen so obtained subtracted from the percentage of soluble nitrogen (total nitrogen minus the percentage of nitrogen occurring as insoluble protein) gives the percentage of nitrogen present as coagulable protein. Multiply this figure by 6.25 to obtain the percentage of coagulable protein in the sample.

41

AMMONIA.—TENTATIVE.

Mix 1 gram of meat extract with 2 cc. of N/1 hydrochloric acid, wash into a Folin apparatus with about 5 cc. of water and proceed as directed under **15**.

42

PROTEOSES AND GELATIN¹¹.—TENTATIVE.

Evaporate the filtrate from **40** to a small volume and saturate with zinc sulphate (about 85 grams to 50 cc., avoiding such an excess as would later cause bumping). Let stand several hours, filter and wash the precipitate with saturated zinc sulphate solution, place the filter and precipitate in a Kjeldahl flask and determine nitrogen as directed under **I, 18, 21 or 23**. Or, if the precipitate is voluminous, which rarely happens, make up to a definite volume with saturated zinc sulphate solution, filter and determine the nitrogen in an aliquot of the filtrate, as directed under **I, 18, 21 or 23**, and subtract the nitrogen thus obtained from the nitrogen in the filtrate from the coagulable protein to obtain the nitrogen of the precipitated protein (proteoses and gelatin).

43

GELATIN.—TENTATIVE.

Prepare a 50% solution of the sample, using hot water. Allow to cool and place in an ice box for 2 hours. If gelatin is present, the solution will set.

The ratio of total creatinin to total nitrogen in a normal meat extract (1 : 1.5) assists in determining the presence of gelatin or gelatin derivatives. The ratio is decreased when gelatin or gelatin derivatives are present in any considerable amount.

AMINO NITROGEN.

*Van Slyke Method*¹².—*Tentative.*

44

REAGENTS.

(a) *Alkaline permanganate solution.*—Dissolve 50 grams of potassium permanganate and 25 grams of potassium hydroxid in sufficient water to make 1 liter.

(b) *Sodium nitrite solution.*—Dissolve 30 grams of sodium nitrite in sufficient water to make 100 cc.

(c) *Glacial acetic acid.*

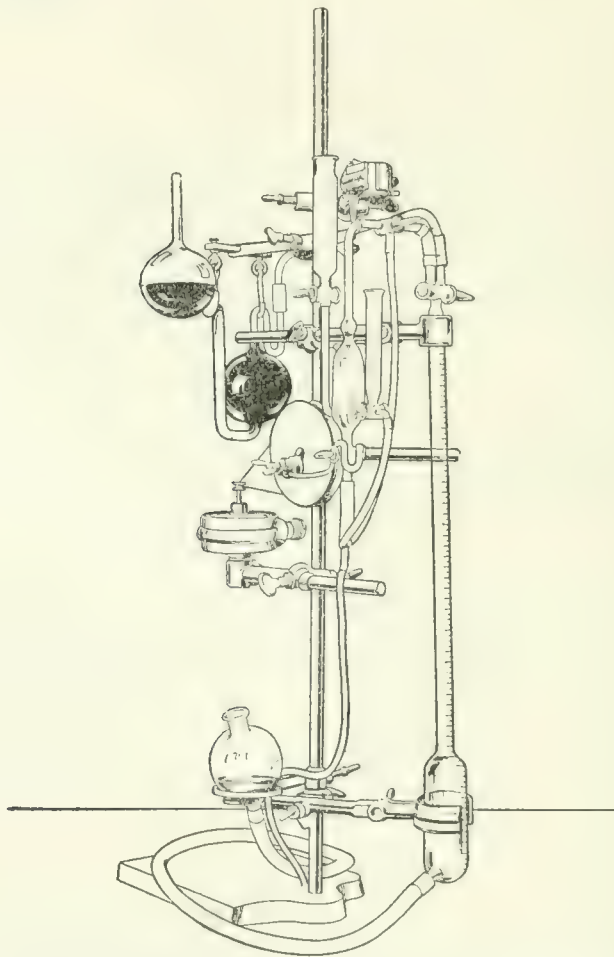


FIG. 10. VAN SLYKE APPARATUS FOR THE DETERMINATION OF AMINO NITROGEN.
(By courtesy of the *Journal of Biological Chemistry*.)

45

APPARATUS.

Employ the apparatus shown in Figs. 10 and 11, the former illustrating the manner in which the entire apparatus is arranged and the latter showing the details

of the deaminizing bulb and connections. The Hempel gas pipette is filled with the alkaline permanganate solution.

46

DETERMINATION.

Fill with water the burette (*F*), the capillary tube leading to the Hempel pipette and also the other capillary as far as *c*. Introduce into *A* sufficient glacial acetic acid to fill one fifth of *D*, the tube (*A*) being etched with a mark to measure this

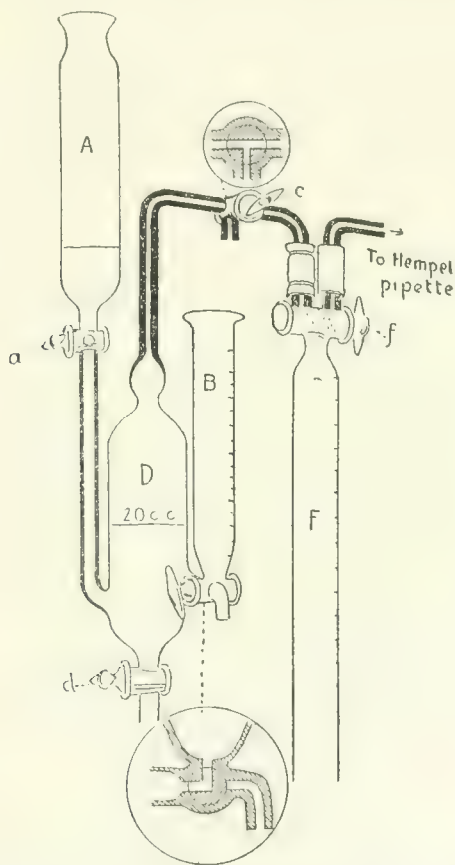


FIG. 11. DETAILS OF THE DEAMINIZING BULB AND CONNECTION.

(By courtesy of the *Journal of Biological Chemistry*.)

amount. Allow the acid to run into *D*, the cock *c* being turned so as to allow the air to escape from *D*. Pour the sodium nitrite solution into *A* until *D* is full of solution and enough excess is present to rise a little above the cock into *A*. *A* is also marked for measuring off this amount. Then close the gas exit from *D* at *c*, and, *a* being open, shake *D* for a few seconds until the liquid is forced down to the 20 cc. mark in *D*. Then close *a*, open *c* and shake the apparatus rapidly with the motor for 2 minutes, these operations being for the purpose of expelling all the air from *D*. Then turn *c* and *f* so that *D* and *F* are connected.

Measure off in *B* 10 cc. or less, as the case may be, of the solution of the sample containing not more than 20 mg. of amino nitrogen (about 1-2 grams of the sample) and allow it to run into *D*. Connect *D* with the motor as shown in Fig. 10 and shake for 5 minutes.

If the solution of the sample is viscous and threatens to foam over, rinse out *B*, and then through it introduce a little caprylic alcohol into *D*, or, if it is known beforehand that the sample will cause excessive foaming, introduce a little caprylic alcohol into *D* through *B*, rinsing *B* with alcohol and ether or drying with a roll of filter paper before adding the solution of the sample.

During the shaking there is an evolution of nitrogen mixed with nitric oxid, the gases being collected in *F*. Force all the gas in *D* into *F* by opening *a* and filling *D* with liquid from *A*. Connect *F* with the Hempel pipette and force the gas into the latter by means of the leveling bulb, allowing the cock *a* to remain open during this and the succeeding operation in order to permit displacement of the liquid in *D* by the nitric oxid. formed in the interval. Connect the driving rod with the pipette by lifting the hook from the shoulder of *D* and placing the other hook, on the opposite side of the driving rod, over the horizontal lower tube of the pipette. Shake the pipette rather slowly for a minute which, with any but almost completely exhausted permanganate solutions, completes the absorption of nitric oxid. Then return the gas to the burette, adjust the level with the leveling bulb and note the volume of nitrogen, the temperature and barometric pressure, and calculate the volume of nitrogen under standard conditions of temperature and pressure. Obtain the corresponding weight of nitrogen, divide the latter by 2, and from the quotient calculate the apparent per cent of amino nitrogen in the sample. Correct the result for a blank test performed as above, using 10 cc. of water instead of the solution of the sample. The amount of gas obtained in the blank is usually 0.3-0.4 cc., and nitrite solutions giving a much larger correction should be rejected.

In the case of beef extracts and similar preparations 5 minutes is sufficient time to allow for the completion of the reaction in *D*. In general the same time serves for the decomposition of alpha-amino acids but with ammonia, methylamin and most amines other than alpha-amines 1-1½ hours should be allowed. For determinations on such substances mix the solution of the sample with the reagents as described above, allow the mixture to stand in the apparatus till the end of the required time, and conclude the reaction by shaking the apparatus with the motor for 2-3 minutes. Continue the determination from this point as directed above.

47

ACID ALCOHOL-SOLUBLE NITROGEN¹³.—TENTATIVE.

Transfer 10 cc. of an aqueous solution of the sample (10 grams of the sample dissolved in sufficient water to make 100 cc.) or, if the sample is insoluble in water, 1 gram of the sample and 10 cc. of water, to a 200 cc. glass-stoppered measuring cylinder, add 1.2 cc. of 12% hydrochloric acid, mix and add absolute alcohol to the 200 cc. mark. Mix thoroughly and set aside for several hours. If necessary make up to volume, filter, transfer 100 cc. of the filtrate to a Kjeldahl flask, evaporate the alcohol on a water bath and determine nitrogen in the residue as directed under I, 18, 21 or 23.

48

CREATIN.—OFFICIAL.

Dissolve about 7 grams of the sample in cold (20°C.) ammonia-free water in a 150 cc. beaker, transfer the solution to a 250 cc. measuring flask, dilute to the mark

and mix thoroughly. Transfer a 20 cc. aliquot of this solution to a 50 cc. measuring flask and proceed as directed under **16**. Subtract from the combined creatinin value the equivalent of the pre-formed creatinin, **49**, and multiply the difference by 1.16 to convert into creatin. Express the result as per cent of creatin.

49

CREATININ.—OFFICIAL.

For creatinin in beef extract measure about 5 cc. of the solution employed in **48** into a 500 cc. measuring flask, add 10 cc. of 10% sodium hydroxid solution and 30 cc. of the saturated picric acid solution (1.2%), mix and rotate for 30 seconds. Allow to stand exactly $4\frac{1}{2}$ minutes, then dilute to the mark at once with water. Shake thoroughly and read the depth of color after standing. If the reading is less than 7 or over 9.5 mm., repeat, calculating the quantity of solution necessary to obtain a reading of about 8 mm. Express the result as per cent of creatinin, making the calculations as indicated under **16**.

GLYCEROL.

50*Cook Method*¹⁴.—*Tentative.*

Weigh 2 grams of a solid or 5 grams of a liquid preparation in a small lead dish or Hofmeister Schälchen containing 20 grams of ignited sand. Transfer the dish and its contents to a mortar containing more ignited sand and several grams of anhydrous sodium sulphate and mix thoroughly. Transfer the mixture, including the dish, to a Soxhlet apparatus which has a piece of cotton placed in the side arm to prevent solid particles from being siphoned over. Extract the entire mass with redistilled anhydrous acetone for 10 hours. Distil the acetone from the extract, carefully removing the last trace by means of a vacuum pump. Take up the residue in water, add 5 cc. of 10% silver nitrate solution, make up to a volume of 100 cc., shake, allow to stand overnight, filter and determine glycerol in an aliquot of the filtrate as directed under **XIX, 6**, beginning at the point "Add * * * 30 cc. of the strong potassium dichromate solution". With solid meat and yeast extracts a blank of 0.5–1.0 % is obtained in most cases.

51

SUGAR.—TENTATIVE.

Heat 20 grams of the sample with about 200 cc. of water on a steam bath until all soluble substances have gone into solution, cool and proceed from this point as directed under **23**. Reducing sugar up to 0.5% may be present as a natural constituent of meat extracts.

52

PRESERVATIVES.—TENTATIVE.

Proceed as directed under **X**.

53

METALS.—TENTATIVE.

Proceed as directed under **XII**.

54

NITRATES.—TENTATIVE.

Proceed as directed under **26** or **28**.

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XXII. DAIRY PRODUCTS.

MILK.

1 SOLIDS.—OFFICIAL.

Heat 3-5 grams of the milk at the temperature of boiling water until it ceases to lose weight, using a tared, flat-bottomed dish of not less than 5 cm. diameter. If desired, previously place 15-20 grams of pure, dry sand in the dish. Cool in a desiccator and weigh rapidly to avoid absorption of hygroscopic moisture.

2 ASH.—OFFICIAL.

Weigh about 20 grams of the milk in a tared dish, add 6 cc. of nitric acid, evaporate to dryness and ignite at a temperature just below redness until the ash is free from carbon.

3 TOTAL NITROGEN.—OFFICIAL.

Place about 5 grams of the milk in a Kjeldahl digestion flask and proceed, without evaporation, as directed under **I, 18, 21** or **23**. Multiply the percentage of nitrogen by 6.38 to obtain the equivalent percentage of nitrogen compounds.

CASEIN.

(This determination should be made while the milk is fresh, or nearly so.

When it is not practicable to make this determination within

24 hours, add 1 part of formaldehyde to 2500

parts of milk and keep in a cool place.)

4 *Method I.—Official.*

Place 10 grams of the milk in a beaker with 90 cc. of water at 40°-42°C. and add at once 1.5 cc. of 10% acetic acid. Stir and let stand 3-5 minutes. Then decant on a filter, wash by decantation 2-3 times with cold water and transfer the precipitate to the filter. Wash once or twice on the filter. The filtrate should be clear, or very nearly so. If the first portions of the filtrate are not clear, repeat the filtration, after which complete the washing of the precipitate. Determine nitrogen in the washed precipitate and filter paper as directed under **I, 18, 21** or **23**, multiply by 6.38 and calculate the percentage of casein.

In samples of milk which have been preserved, the acetic acid should be added in small portions, a few drops at a time, with stirring, and the addition continued until the liquid above the precipitate becomes clear, or very nearly so.

5 *Method II.—Official.*

To 10 grams of the milk add 50 cc. of water at 40°C., then 2 cc. of alum solution saturated at 40°C., or higher. Allow the precipitate to settle, transfer to a filter and wash with cold water. Determine nitrogen in the precipitate and filter paper as directed under **I, 18, 21** or **23**, multiply by 6.38 and calculate the percentage of casein.

ALBUMIN.

6

Method I.—Tentative.

Exactly neutralize the filtrate, obtained in **4**, with sodium hydroxid solution, add 0.3 cc. of 10% acetic acid and heat on a steam bath until the albumin is completely precipitated. Collect the precipitate on a filter, wash with cold water and determine the nitrogen as directed under **I**, **18**, **21** or **23**, multiply by 6.38 and calculate the percentage of albumin.

7

Method II.—Tentative.

To the filtrate obtained from the casein determination, **5**, add 0.3 cc. of 10% acetic acid, boil until the albumin is completely precipitated and proceed as directed in **6**.

LACTOSE.

Optical Method.—Tentative.

8

REAGENTS.

(a) *Acid mercuric nitrate solution.*—Dissolve mercury in double its weight of nitric acid (sp. gr. 1.42) and dilute with an equal volume of water.

(b) *Mercuric iodid solution.*—Dissolve 33.2 grams of potassium iodid and 13.5 grams of mercuric chlorid in 20 cc. of glacial acetic acid and 640 cc. of water.

9

DETERMINATION.

Determine the specific gravity of the milk by means of a delicate hydrometer or, if preferred, a pycnometer. The quantity of sample to be taken for the determination varies with the specific gravity and is to be measured at the same temperature at which the specific gravity is taken. The volume to be measured will be found in **10** which is based upon twice the normal weight of lactose (32.9 grams per 100 metric cc.) for the Ventzke sugar scale.

Place the quantity of milk indicated in **10** in a flask graduated at 102.6 cc. Add 1 cc. of the acid mercuric nitrate solution or 30 cc. of the mercuric iodid solution (an excess of these reagents does no harm), fill to the mark, shake, filter through a dry filter and polarize. It is not necessary to heat before polarizing. If a 200 mm. tube is used, divide the polariscope reading by 2 (or, if a 400 mm. tube is used, by 4) to obtain the per cent of lactose in the sample.

TABLE 19.

10 *Volumes of milk corresponding to a lactose double normal weight¹.*

SPECIFIC GRAVITY OF MILK	VOLUME OF MILK FOR A LACTOSE DOUBLE NORMAL WEIGHT (VENTZKE SCALE)	SPECIFIC GRAVITY OF MILK	VOLUME OF MILK FOR A LACTOSE DOUBLE NORMAL WEIGHT (VENTZKE SCALE)
	cc.		cc.
1.024	64.25	1.030	63.90
1.025	64.20	1.031	63.80
1.026	64.15	1.032	63.75
1.027	64.05	1.033	63.70
1.028	64.00	1.034	63.65
1.029	63.95	1.035	63.55
		1.036	63.50

11

Gravimetric Method.—Official.

Dilute 25 grams of the milk with 400 cc. of water in a 500 cc. graduated flask, add 10 cc. of copper sulphate solution [VIII, 19 (a)] and about 7.5 cc. of a potassium hydroxid solution of such strength that 1 volume is just sufficient to precipitate completely the copper as hydroxid from 1 volume of the copper sulphate solution. Instead of potassium hydroxid solution of this strength, 8.8 cc. of N/2 sodium hydroxid solution may be used. After the addition of the alkali solution the mixture must still have an acid reaction and contain copper in solution. Fill the flask to the 500 cc. mark, mix, filter through a dry filter and determine lactose in an aliquot of the filtrate as directed under VIII, 46 or 48.

FAT.

12

Roese-Gottlieb Method².—Official.

Weigh 10–11 grams of the milk into a Röhrig tube or some similar apparatus, add 1.25 cc. of concentrated ammonium hydroxid (2 cc. if the sample is sour) and mix thoroughly. Add 10 cc. of 95% alcohol by volume and mix well. Then add 25 cc. of washed ether and shake vigorously for 30 seconds, then 25 cc. of petroleum ether (redistilled slowly at a temperature below 60°C.) and shake again for 30 seconds. Let stand 20 minutes, or until the upper liquid is practically clear. Draw off as much as possible of the ether-fat solution (usually 0.5–0.8 cc. will be left) into a weighed flask through a small, quick-acting filter. The flask should always be weighed with a similar one as a counterpoise. Re-extract the liquid remaining in the tube, this time with only 15 cc. of each ether, shake vigorously 30 seconds with each and allow to settle. Draw off the clear solution through the small filter into the same flask as before and wash the tip of spigot, the funnel and the filter with a few cc. of a mixture of the 2 ethers in equal parts. For absolutely exact results the re-extraction must be repeated. This third extraction yields usually not more than about 1 mg. of fat (about 0.02% on a 4 gram charge) if the previous ether-fat solutions have been drawn off closely. Evaporate the ethers slowly on a steam bath, then dry the fat in a boiling water oven to constant weight.

Confirm the purity of the fat by dissolving in a little petroleum ether. Should a residue remain, remove the fat completely with petroleum ether, dry the residue, weigh and deduct the weight. Finally correct this weight by a blank determination on the reagents used.

Babcock Method.—Official.

13

APPARATUS.

(a) *Standard Babcock test bottles.*—The standard Babcock test bottles for milk and cream shall be as follows:

(1) *8%, 18 gram, 6 inch milk test bottle.*—The total per cent graduation shall be 8. The total height of the bottle shall be 150–165 mm. ($5\frac{3}{8}$ – $6\frac{1}{2}$ inches). The capacity of the bulb up to the junction with the neck shall be not less than 45 cc. The graduated portion of the neck shall have a length of not less than 63.5 mm. ($2\frac{1}{2}$ inches) and the neck shall be cylindrical for at least 9 mm. below the lowest and above the highest graduation marks. The graduations shall represent whole per cents, halves and tenths of a per cent.

(2) *50%, 9 gram, 6 inch cream test bottle.*—The total per cent graduation shall be 50. The total height of the bottle shall be 150–165 mm. ($5\frac{3}{8}$ – $6\frac{1}{2}$ inches). The capacity of the bulb up to the junction with the neck shall be not less than 45 cc. The graduated portion of the neck shall have a length of not less than 63.5 mm. ($2\frac{1}{2}$

inches) and the neck shall be cylindrical for at least 9 mm. below the lowest and above the highest graduation marks. The graduations shall represent five per cents, whole and halves of a per cent.

(3) *50%, 9 gram, 9 inch, cream test bottle.*—Same as (2) except that the total height of the bottle shall be 210–225 mm. ($8\frac{1}{4}$ – $8\frac{3}{4}$ inches).

(b) *Centrifuge.*

(c) *Pipettes.*—Graduated to deliver 17.6 cc. of water at 20°C. in 5–8 seconds.

(d) *Graduates.*—Capacity 17.5 cc. or a Swedish acid bottle delivering that amount.

14

CALIBRATION OF APPARATUS.

(a) *Graduation.*—The unit of graduation for all Babcock glassware shall be the true cc. (0.998877 gram of water at 4°C.).

With bottles, the capacity of each per cent on the scale shall be 0.20 cc.

With pipettes and graduates, the delivery shall be the intent of the graduation; and the graduation shall be read with the bottom of the meniscus in line with the mark.

(b) *Testing.*—The method for testing Babcock bottles shall be calibration with mercury (13.5471 grams of clean, dry mercury at 20°C., to be equal to 5% on the scale), the bottle being previously filled to zero with mercury.

The mercury and cork, alcohol and burette, and alcohol and brass plunger methods may be employed for the rapid testing of Babcock bottles, but the accuracy of all questionable bottles shall be determined by calibration with mercury.

The method for testing pipettes and graduates shall be calibration by measuring in a burette the quantity of water (at 20°C.) delivered.

(c) *Limit of error.*—For standard Babcock milk bottles the error at any point of the scale shall not exceed 0.1%.

For standard Babcock cream bottles the error at any point of the scale shall not exceed 0.5 %.

For standard milk pipettes the error shall not exceed 0.05 cc.

For acid measures the error shall not exceed 0.2 cc.

15

DETERMINATION.

Pipette 17.6 cc. of the carefully mixed sample into a test bottle and add 17.5 cc. of commercial sulphuric acid (sp. gr. 1.82–1.83). Mix and, when the curd is dissolved, centrifugalize for 4 minutes at the required speed for the machine used. Add boiling water, filling to the neck of the bottle, and whirl for 1 minute; again add boiling water so as to bring the fat within the scale on the neck of the bottle, and, after whirling for 1 minute more, read the length of the fat column, making the reading at 57°–60°C. at which temperature the fat is wholly liquid. The reading gives directly the per cent of fat in the milk.

Details of the manipulation of the Babcock test and its application in the analysis of dairy products other than milk are described by Farrington and Woll³, and Van Slyke⁴.

ADDED WATER.

(In conjunction with the copper, acetic or sour serum refraction method, the determination of the ash of the sour serum or of the acetic serum should be made in all cases where the indices of refraction fall below the minimum limit so as to eliminate all possibility of abnormal milk.)

16

ACETIC SERUM.—TENTATIVE.

(a) *Zeiss immersion refractometer reading*.—To 100 cc. of milk at a temperature of about 20°C. add 2 cc. of 25% acetic acid (sp. gr. 1.035) in a beaker and heat the mixture, covered with a watch glass, in a water bath for 20 minutes at a temperature of 70°C. Place the beaker on ice water for 10 minutes and separate the curd from the serum by filtering through a 12.5 cm. folded filter. Transfer about 35 cc. of the serum to 1 of the beakers that accompanies the control-temperature bath used in connection with the Zeiss immersion refractometer, and take the refractometer reading at exactly 20°C., using a thermometer graduated to tenths of a degree. A reading below 39 indicates added water; between 39 and 40, the addition of water is suspected.

(b) *Ash*.—Transfer 25 cc. of the serum to a flat-bottomed platinum dish and evaporate to dryness on a water bath. Then heat over a low flame (to avoid spattering) until the contents are thoroughly charred, place the dish in an electric muffle, preferably with pyrometer attached, and ignite to a white ash at a temperature not greater than 500°C. (900°F.). Cool and weigh. Express the result as grams per 100 cc. Results below 0.715 gram per 100 cc. indicate added water. Multiply by the factor 1.021 (dilution of the acetic serum being 2%) to obtain the result on the sour serum ash.

17

SOUR SERUM.—TENTATIVE.

(a) *Zeiss immersion refractometer reading*.—Allow the milk to sour spontaneously, filter and determine the immersion refractometer reading of the clear serum at 20°C. A reading below 38.3 indicates added water.

(b) *Ash*^s.—Determine the ash in 25 cc. of the serum, obtained in (a), as directed in 16 (b). Results below 0.730 gram per 100 cc. indicate added water.

18

ZEISS REFRACTOMETER READING OF COPPER SERUM.—TENTATIVE.

To 1 volume of copper sulphate solution (72.5 grams of copper sulphate per liter, adjusted if necessary to read 36 at 20°C. on the scale of the Zeiss immersion refractometer, or, to a specific gravity of 1.0443 at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$) add 4 volumes of milk. Shake well and filter. Determine the Zeiss refractometer reading of the clear serum at 20°C. A reading below 36 indicates added water.

GELATIN.

19

Qualitative Test.—Tentative.

To 10 cc. of the milk add an equal volume of acid mercuric nitrate solution (mercury dissolved in twice its weight of nitric acid (sp. gr. 1.42) and this solution diluted to 25 times its volume with water), shake the mixture, add 20 cc. of water, shake again, allow to stand 5 minutes and filter. If much gelatin is present, the filtrate will be opalescent and cannot be obtained quite clear. To a portion of the filtrate contained in a test tube, add an equal volume of saturated aqueous picric acid solution. A yellow precipitate will be produced in the presence of any considerable amount of gelatin, while smaller amounts will be indicated by a cloudiness. In the absence of gelatin the filtrate will remain perfectly clear.

20

PRESERVATIVES.—TENTATIVE.

Proceed as directed under X. To test for salicylic or benzoic acid acidify 100 cc. of the milk with 5 cc. of hydrochloric acid (1 to 3), shake until curdled, filter and treat the clear filtrate as directed under X, 2, 3 or 8.

To test for formaldehyde proceed as directed under **X, 17, 18, 19, 20, 21, 22, or 23**, applying the test directly to the milk.

COLORING MATTERS.

21*Leach Method.—Tentative.*

Warm about 150 cc. of milk in a casserole over a flame and add about 5 cc. of acetic acid, then slowly continue the heating nearly to the boiling point while stirring. Gather the curd, when possible, into one mass with a stirring rod and pour off the whey. If the curd breaks up into small flecks, separate from the whey by straining through a sieve or colander. Press the curd free from adhering liquid, transfer to a small flask and macerate for several hours, preferably overnight, in about 50 cc. of ether, the flask being tightly corked and shaken at intervals. Decant the ether extract into an evaporating dish, remove the ether by evaporation and test the fatty residue for annatto as directed in **XI, 24**.

The curd of an uncolored milk is perfectly white after complete extraction with ether, as is also that of a milk colored with annatto. If the extracted fat-free curd is distinctly colored an orange or yellowish color, a coal tar dye is indicated. In many cases upon treating a lump of a fat-free curd in a test tube with a little strong hydrochloric acid the color changes to pink, indicating the presence of a dye similar to aniline yellow or butter yellow or perhaps one of the acid azo yellows or oranges. In such cases, separate and identify the coloring matter present in the curd as directed under **XI**. If aniline yellow, butter yellow, or other oil-soluble dye is present, the greater part will be found in the ether extract containing the fat. In such cases proceed as directed under **XI, 3**.

In some cases the presence of coal tar dyes can be detected by treating about 10 cc. of the milk directly with an equal volume of hydrochloric acid (sp. gr. 1.20) in a porcelain casserole, giving the dish a slight rotary motion. In the presence of some dyes the separated curd acquires a pink coloration.

CREAM.

22**SOLIDS.—OFFICIAL.**

Proceed as directed in **1**, employing 2-3 grams of the sample.

23**ASH.—OFFICIAL.**

Proceed as directed under **2**.

24**TOTAL NITROGEN.—OFFICIAL.**

Proceed as directed under **3**.

LACTOSE.

25*Gravimetric Method.—Official.*

Proceed as directed under **11**.

FAT.

26*Extraction Method.—Official.*

Weigh 4-5 grams of the homogeneous sample into a Röhrig tube or similar apparatus, dilute with water to about 10.5 cc. and proceed as directed under **12**.

27*Babcock Method.—Official.*

Weigh 9 or 18 grams, depending upon the fat content of the sample, into a standard Babcock cream bottle and proceed as directed under **15**.

28

GELATIN.—TENTATIVE.

Proceed as directed under 19.

29

COLORING MATTERS.—TENTATIVE.

Proceed as directed under XI, particularly 3 and 24 for the detection of oil-soluble coal tar dyes and annatto.

30

PRESERVATIVES.—TENTATIVE.

Proceed as directed under X.

31

CONDENSED MILK (UNSWEETENED).

Dilute 40 grams of the homogeneous sample with 60 grams of water and proceed as directed under 1 to 15, inclusive, 19, 20 and 21, correcting the results for the dilution.

CONDENSED MILK (SWEETENED).

32

PREPARATION OF SAMPLE.—OFFICIAL.

If cold, place the can in water at 30°–35°C. until warm. Open, scrape out all milk adhering to the interior of the can and mix by transferring the contents to a dish sufficiently large to stir thoroughly and make the whole mass homogeneous. Weigh 100 grams into a 500 cc. flask and make up to the mark with water. If the milk will not dissolve completely, weigh out each portion for analysis separately.

33

TOTAL SOLIDS.—OFFICIAL.

Use 10 cc. of the solution, prepared as directed in 32, and proceed as directed in 1, drying either on sand or asbestos fiber.

34

ASH.—OFFICIAL.

Evaporate 10 cc. of the solution, prepared as directed in 32, to dryness on a water bath and ignite the residue as directed under VIII, 4.

35

PROTEIN.—OFFICIAL.

Determine nitrogen as directed under I, 18, 21 or 23, using 10 cc. of the solution, prepared as directed in 32, without evaporation and multiply by 6.38.

36

LACTOSE.—OFFICIAL.

Dilute 100 cc. of the solution, prepared as directed in 32, in a 250 cc. flask to about 200 cc., add 6 cc. of Fehling's copper sulphate solution [VIII, 19 (a)] and make up to the mark. Filter through a dry filter and determine lactose as directed in VIII, 46 or 48.

37

FAT OR ETHER EXTRACT.

Röse-Gottlieb Method.—Official.

Weigh 4–5 grams of the homogeneous sample into a Röhrig tube or some similar apparatus, dilute with water to about 10.5 cc. and proceed as directed under 12.

38

SUCROSE.—TENTATIVE.

Determine sucrose by difference, deducting the milk solids (lactose, protein, fat and ash) from the total solids.

BUTTER AND ITS SUBSTITUTES.**39****PREPARATION OF SAMPLE.—OFFICIAL.**

If large quantities of butter are to be sampled, use a butter trier or sampler. Melt completely the portions thus drawn, 100–500 grams, in a closed vessel at as low a temperature as possible. When softened, cool and, at the same time, shake the mass violently until it is homogeneous and solidified sufficiently to prevent the separation of the water and fat. Then pour a portion into the vessel from which it is to be weighed. The sample should completely or nearly fill the vessel and should be kept in a cool place until analyzed.

40**MOISTURE.—OFFICIAL.**

Weigh 1.5–2.5 grams of the sample into a flat-bottomed dish, having a surface of at least 20 sq. cm., dry at the temperature of boiling water and weigh at hourly intervals until the weight becomes constant. The use of clean, dry sand or asbestos is admissible.

ETHER EXTRACT.**41***Indirect Method.—Official.*

Dissolve the dry butter, obtained in the moisture determination in which no absorbent was used, in absolute ether or petroleum ether, transfer to a weighed Gooch, with the aid of a wash bottle filled with the solvent and wash until free from fat. Dry the Gooch and contents at the temperature of boiling water until the weight is constant and determine the fat.

42*Direct Method.—Official.*

From the dry butter, obtained in the determination of moisture, either with or without the use of an absorbent, extract the fat with anhydrous, alcohol-free ether, receiving the solution in a weighed flask. Evaporate the ether, dry the extract at the temperature of boiling water and weigh at hourly intervals until the weight is constant.

43**CASEIN, ASH AND CHLORIN.—OFFICIAL.**

Cover the crucible, containing the residue from the fat determination by the indirect method, **41**, and heat gently at first, then raise the temperature gradually to just below redness. The cover may then be removed and heating continued until the contents of the crucible are white. The loss in weight represents casein, and the residue in the crucible, mineral matter. Dissolve this mineral matter in water slightly acidified with nitric acid and determine chlorin, either gravimetrically as directed under **I, 16 (a)**, or volumetrically as directed under **III, 15**.

44**SALT.—OFFICIAL.**

Weigh in a counterpoised beaker 5–10 grams of butter, using portions of about 1 gram from different parts of the sample. Add about 20 cc. of hot water and, after the butter is melted, transfer the whole to a separatory funnel. Insert the stopper and shake for a few moments. Let stand until all the fat has collected on the top of the water, then draw off the latter into a flask, being careful to let none of the fat globules pass. Again add hot water, rinsing the beaker, and repeat the extraction 10–15 times, using 10–20 cc. of water each time. The washings will contain all but a mere trace of the sodium chlorid originally present in the butter. Determine the amount in the whole or an aliquot of the liquid by titration with standard silver nitrate, using potassium chromate as an indicator.

FAT.

45

PREPARATION OF SAMPLE.—OFFICIAL.

Melt the butter and keep in a dry place at about 60°C. for 2-3 hours or until the water and curd have entirely separated. Filter the clear, supernatant fat through a dry filter paper in a hot water funnel or in an oven at about 60°C. If the filtered liquid fat is not perfectly clear, refilter.

46

EXAMINATION.

Proceed as directed under **XXIII**.

47

Microscopic Method.—Official.

Place on a slide a small portion of the fresh, unmelted sample taken from the inside of the mass, add a drop of pure olive oil, apply a cover-glass with gentle pressure, and examine with a magnification of 120-150 diameters for crystals of lard, etc. Examine the same specimen with polarized light and a selenite plate without the use of oil. Pure fresh butter will show neither crystals nor a parti-colored field with selenite. Renovated butter or other fats melted and cooled and mixed with butter will usually present crystals and variegated colors with the selenite plate.

For further microscopic study dissolve in a test tube 3-4 cc. of the fat in 15 cc. of ether. Close the tube with a loose plug of cotton wool and allow to stand 12-24 hours at 20°-25°C. When crystals form at the bottom of the tube, remove with a pipette, glass rod or tube, place on a slide, cover and examine under a microscope. The crystals formed by later deposits may be examined in a similar way. Compare with crystals obtained in the same way from samples of known purity.

48

PRESERVATIVES.—TENTATIVE.

Proceed as directed under **X**.

49

COLORING MATTERS.—TENTATIVE.

Pour about 2 grams of the filtered fat, dissolved in ether, into each of 2 test tubes. Into one of the tubes pour 1-2 cc. of hydrochloric acid and into the other about the same volume of dilute potassium hydroxid solution. Shake the tubes well and allow to stand. In the presence of azo dyes the test tube to which the acid has been added will show a pink to wine-red coloration, while the potash solution in the other tube will show no color. If, on the other hand, annatto or other vegetable color has been used, the potash solution will be colored yellow, while no color will be apparent in the acid solution.

General test.—Proceed as directed under **XI**, particularly **3** and **24**, for the detection of oil-soluble coloring matters and annatto.

RENOVATED BUTTER AND OLEOMARGARINE.

50

Foam Test.—Tentative.

Heat 2-3 grams of the sample, either in a spoon or dish, over a free flame. True butter will foam abundantly, whereas process butter will bump and sputter, like hot grease, without foaming. Oleomargarine behaves like process butter, but chemical tests will determine whether the sample is oleomargarine or butter.

51

Melted Fat Test.—Tentative.

Melt 50–100 grams of butter or renovated butter at 50°C. The curd from butter will settle, leaving a clear supernatant fat; in the case of renovated butter, the supernatant fat remains more or less turbid.

CHEESE.

52

SELECTION AND PREPARATION OF THE SAMPLE.—OFFICIAL.

When the cheese can be cut, take a narrow, wedge-shaped segment reaching from the outer edge to the center of the cheese. Cut this into strips and pass 3 times through a sausage machine. When the cheese cannot be cut, take the sample with a cheese trier. If only 1 plug can be obtained, take it perpendicular to the surface of the cheese at a point one third the distance from the edge to the center and extending either entirely or half way through it. When possible, draw 3 plugs, 1 from the center, 1 from a point near the outer edge, and 1 from a point half way between the other 2. For inspection purposes reject the rind but for investigations requiring the absolute amount of fat in the cheese include the rind in the sample. Either grind the plugs in a sausage machine or cut them very finely and mix carefully, preferably the former.

53

MOISTURE.—TENTATIVE.

Place 2–3 grams of very short fiber asbestos (the long fiber may be made suitable by rubbing it through a fine sieve) in a flat-bottomed platinum dish, 6–7 cm. in diameter, and press the asbestos down firmly. Place in the dish a glass rod, about 5 mm. in diameter and slightly longer than the diameter of the dish. Ignite, cool and weigh the dish and contents. Then weigh into the dish 4–5 grams of the sample, prepared as directed under 52, and mix the cheese and asbestos intimately with the glass rod until the mass is homogeneous. Leave the mass in as loose a condition as possible to facilitate the drying. Dry the mixture in an oven at 100°C. and weigh at 1–1½ hour intervals until the weight becomes constant (3 weighings are usually sufficient).

54

ASH AND SALT.—OFFICIAL.

The dry residue from the moisture determination may be used for the determination of ash. If the cheese be rich in fat, the asbestos will be saturated with it. Ignite cautiously to avoid spattering and remove the lamp while the fat is burning off. When the flame has died out, complete the burning in a muffle at low redness. When desired, the salt may be determined in the ash, as directed in 43.

55

NITROGEN.—OFFICIAL.

Determine nitrogen as directed under I, 18, 21 or 23, using about 2 grams of cheese, and multiply the percentage of nitrogen by 6.38 to obtain the per cent of nitrogen compounds.

56

ACIDITY.—TENTATIVE.

To 10 grams of finely divided cheese add water at a temperature of 40°C. until the volume equals 105 cc., shake vigorously and filter. Titrate 25 cc. portions of the filtrate, representing 2.5 grams of the sample, with standard sodium hydroxid, preferably N/10, using phenolphthalein as an indicator. Express the result in terms of lactic acid.

57

COLORING MATTERS.—TENTATIVE.

Proceed as directed under XI.

FAT.

58

PREPARATION OF SAMPLE.—TENTATIVE.

(a) *Alkaline extraction*.—Treat about 300 grams of the cheese, cut into fragments the size of a pea, with 700 cc. of 5% potassium hydroxid solution at 20°C. in a large, wide-necked flask, shaking vigorously to dissolve the casein. In 5–10 minutes the casein will be dissolved and the fat will rise to the surface in lumps. Collect the lumps of fat into as large a mass as possible by shaking gently. Pour cold water into the flask until the fat is driven up into the neck and remove it by suitable means. Wash the fat thus obtained with just sufficient water to remove the residue of the alkali which it may contain. The fat is not perceptibly attacked by the alkali in this treatment, is practically all separated in a short time and is then easily prepared for chemical analysis by filtering and drying as directed in 45.

(b) *Acid extraction*.—Pass the cheese through a grinding machine, transfer to a large flask and cover with warm water, using 1 cc. for every gram of cheese. Shake thoroughly and add sulphuric acid (sp. gr. 1.82–1.825) slowly and in small quantities, shaking after each addition of acid. The total amount of acid used should be the same as the amount of water employed. Remove the fat, which separates after standing a few minutes, by means of a separatory funnel, wash free from acid, filter and dry as directed in 45.

59

EXAMINATION.—TENTATIVE.

Proceed as directed under XXIII.

ESTIMATION.

60

Gravimetric Method.—Official.

Cover the perforations in the bottom of an extraction tube with dry asbestos felt, and place on this a mixture containing equal parts of anhydrous copper sulphate and pure, dry sand to a depth of about 5 cm., packing loosely. Cover the upper surface of this material with a layer of asbestos. Place on this 2–5 grams of the sample and extract with anhydrous ether for 5 hours in a continuous extraction apparatus. Remove the cheese and grind it with pure sand in a mortar to a fine powder, return the mixed cheese and sand to the extraction tube, wash the mortar with ether, add the washings to the tube and continue the extraction for at least 10 hours.

61

Schmidt-Bondzynski Method, Modified.—Tentative.

Rub up, by means of a glass rod, 1 gram of the homogeneous sample with 9 cc. of water and 1 cc. of concentrated ammonium hydroxid in a narrow 100–125 cc. beaker. Digest the mixture at a low heat until the casein is well softened; neutralize with concentrated hydrochloric acid, using litmus as an indicator and add 10 cc. more of concentrated hydrochloric acid. Add a pinch of sand to prevent bumping and boil gently for 5 minutes, keeping the beaker covered with a watch glass. Cool the solution, transfer to a Röhrig tube or some similar apparatus, rinse the beaker with 25 cc. of washed ethyl ether and shake well. Add 25 cc. of redistilled petroleum ether (b. p. below 65°C.), let the solutions separate and proceed from this point as directed in 12.

Weigh about 6 grams of the cheese in a tared dish. Add 10 cc. of boiling water and stir with a rod until the cheese softens and an even emulsion is formed, preferably adding a few drops of strong ammonium hydroxid, and keep the beaker in hot water until the emulsion is nearly completed and the mass free from lumps. If the sample is a whole milk cheese, employ a Babcock cream bottle. After cooling, transfer the contents of the beaker to the test bottle by adding to the beaker about half of the 17.6 cc. of sulphuric acid usually employed in this test, stirring with a rod, and pouring carefully into the bottle, using the remainder of the acid in 2 portions for washing out the beaker. Then proceed as directed in 15. Multiply the fat reading by 18 and divide by the weight of the sample taken to obtain the per cent of fat.

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XXIII. FATS AND OILS.

1

PREPARATION OF SAMPLE.—OFFICIAL.

Melt solid fats and filter by means of a hot water funnel or similar apparatus. Make the different determinations on samples of this melted, homogeneous mass. Filter oils that are not clear. Keep oils and fats in a cool place and protected from light and air, otherwise they will soon become rancid. Weigh out at one time as many portions as are needed for the various determinations, using a small beaker or weighing burette.

SPECIFIC GRAVITY.

2

At $\frac{20^{\circ}\text{C.}}{4^{\circ}}$.—Tentative.

Determine the specific gravity of the oil at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer.

If the specific gravity of the oil is determined at other than standard temperature, the approximate specific gravity at 20°C. may be calculated by means of the following formula:

$G = G' + 0.0007 (T - 20^{\circ}\text{C.})$ in which

G = specific gravity at 20°C. ;

G' = specific gravity at $\frac{T^{\circ}\text{C.}}{4^{\circ}}$;

T = temperature at which the specific gravity was determined;

0.0007 = mean correction¹ for 1°C.

At the Temperature of Boiling Water.—Official.

3

STANDARDIZATION OF FLASKS.

(a) Fill a tared, 25–30 cc. specific gravity flask with freshly boiled, hot water. Place in a briskly boiling water bath for 30 minutes, replacing any evaporation from the flask by the addition of boiling water. Then insert the stopper, previously heated to 100°C. , remove the flask, cool and weigh.

(b) The following formula may be used for calculating the weight of water (W^T) which a given flask will hold at T° (weighed in air with brass weights at the temperature of the room) from the weight of water (W^t) (weighed in air with brass weights at the temperature of the room) contained therein at t° :

$$W^T = W^t \frac{d^T}{d^t} [1 + 0.000026 (T - t)] \text{ in which}$$

d^T = the density of water at T° ;

d^t = the density of water at t° .

4

DETERMINATION.

Fill the flask, dried at the temperature of boiling water, with the dry, hot, freshly filtered fat, which should be entirely free from air bubbles; keep in the water bath 30 minutes at the temperature of boiling water. Insert the stopper, previously heated to 100°C. , cool and weigh. Divide the weight of contained fat by the weight of contained water previously found to obtain the specific gravity.

The weight of water at boiling temperature must be determined under the barometric conditions prevailing at the time the determination is made.

INDEX OF REFRACTION.

5

General Directions.—Tentative.

Place the instrument in such a position that diffuse daylight or any form of artificial light can readily be obtained for illumination. Circulate through the prisms a stream of water of constant temperature.

Determine the index of refraction with any standard instrument, reading oils at 20°C. and fats at 40°C.

The readings of the Zeiss butyro-refractometer can be reduced to standard temperature by the following formula²:

$$R = R' + 0.55 (T' - T) \text{ in which}$$

R = the reading reduced to temperature T ;

R' = the reading at T' °C.

T' = the temperature at which reading R' is made;

T = the standard temperature;

0.55 = correction in scale divisions for 1°C.

With oils the factor 0.58 is substituted in the formula for 0.55, since they have a higher index of refraction.

The readings of instruments, which give the index of refraction directly, can be reduced to standard temperature by substituting the factor 0.000365 for 0.55 in the formula. As the temperature rises the refractive index falls.

The instrument used may be standardized with water at 20°C., the theoretical refractive index of water at that temperature being 1.3330. Any correction found should be made on all readings.

The index of refraction varies directly with the specific gravity. If the results appear abnormal, compare the specific refractive power³ with the normal. Calculate the specific refractive power from the formula $\frac{N-1}{D}$, in which N equals the refractive index and D the specific gravity. According to Procter⁴ the Lorenz formula $\frac{N^2-1}{(N^2+2)D}$ gives much more satisfactory results than $\frac{N-1}{D}$.

6

By Means of the Abbé Refractometer.—Official.

To charge the instrument, open the double prism by means of the screw head and place a few drops of the sample on the prism or, if preferred, open the prisms slightly by turning the screw head and pour a few drops of the sample into the funnel-shaped aperture between the prisms. Then close the prisms firmly by tightening the screw head. Allow the instrument to stand for a few minutes before the reading is made, so that the temperature of the sample and the instrument will be the same.

The method of measurement is based upon the observation of the position of the *border line of total reflection* in relation to the faces of a prism of flint glass. Bring this border line into the field of vision of the telescope by rotating the double prism by means of the alidade in the following manner: Hold the sector firmly, move the alidade backward or forward until the field of vision is divided into a light and a dark portion. The line dividing these portions is the "border line". This, as a rule, will not be a sharp line but a band of color which is eliminated by rotating the screw head of the compensator until a sharp, colorless line is obtained. The border line should now be adjusted so that it falls on the point of intersection of the 2 cross hairs. Read the refractive index of the substance directly on the scale of the sector. Check the correctness of the instrument, as directed under 5, or by means of the quartz plate which accompanies it, using monobromnaphthalene, and make the necessary correction in the reading.

7

By Means of the Zeiss Butyro-Refractometer.—Official.

Place 2 or 3 drops of the filtered fat on the surface of the lower prism. Close the prisms and adjust the mirror until it gives the sharpest reading. If the reading be indistinct after running water of a constant temperature through the instrument for some time, the fat is unevenly distributed on the surfaces of the prism. As the index of refraction is greatly affected by temperature, care must be used to keep the latter constant. The instrument should be carefully adjusted by means of the standard fluid which is supplied with it. Convert the degrees of the instrument into refractive indices from 8.

8

TABLE 20.

Butyro-refractometer readings and indices of refraction.

READING	INDEX OF REFRACTION	READING	INDEX OF REFRACTION	READING	INDEX OF REFRACTION	READING	INDEX OF REFRACTION
40.0	1.4524	50.0	1.4593	60.0	1.4659	70.0	1.4723
40.5	1.4527	50.5	1.4596	60.5	1.4662	70.5	1.4726
41.0	1.4531	51.0	1.4600	61.0	1.4665	71.0	1.4729
41.5	1.4534	51.5	1.4603	61.5	1.4668	71.5	1.4732
42.0	1.4538	52.0	1.4607	62.0	1.4672	72.0	1.4735
42.5	1.4541	52.5	1.4610	62.5	1.4675	72.5	1.4738
43.0	1.4545	53.0	1.4613	63.0	1.4678	73.0	1.4741
43.5	1.4548	53.5	1.4616	63.5	1.4681	73.5	1.4744
44.0	1.4552	54.0	1.4619	64.0	1.4685	74.0	1.4747
44.5	1.4555	54.5	1.4623	64.5	1.4688	74.5	1.4750
45.0	1.4558	55.0	1.4626	65.0	1.4691	75.0	1.4753
45.5	1.4562	55.5	1.4629	65.5	1.4694	75.5	1.4756
46.0	1.4565	56.0	1.4633	66.0	1.4697	76.0	1.4759
46.5	1.4569	56.5	1.4636	66.5	1.4700	76.5	1.4762
47.0	1.4572	57.0	1.4639	67.0	1.4704	77.0	1.4765
47.5	1.4576	57.5	1.4642	67.5	1.4707	77.5	1.4768
48.0	1.4579	58.0	1.4646	68.0	1.4710	78.0	1.4771
48.5	1.4583	58.5	1.4649	68.5	1.4713	78.5	1.4774
49.0	1.4586	59.0	1.4652	69.0	1.4717	79.0	1.4777
49.5	1.4590	59.5	1.4656	69.5	1.4720	79.5	1.4780

MELTING POINT OF FATS AND FATTY ACIDS.

Wiley Method.—Official.

9

REAGENT.

Alcohol-water mixture.—Specific gravity same as that of the fat to be examined. Prepare by boiling, separately, water and 95% alcohol by volume for 10 minutes to remove the gases which may be held in solution. While still hot pour the water into the test tube until it is almost half full. Nearly fill the test tube with the hot alcohol, poured down the side of the inclined tube to avoid too much mixing. If the alcohol be added after the water has cooled, the mixture will contain so many air bubbles as to be unfit for use.

10

DETERMINATION.

Prepare disks of fat as follows: Allow the melted and filtered fat to fall a distance of 15-20 cm. from a dropping tube upon a piece of ice or upon the surface of cold mercury. The disks thus formed should be 1-1.5 cm. in diameter and weigh about

200 mg. When solid remove the disk and allow to stand 2-3 hours in order to obtain the normal melting point.

Place a test tube, 30 by 3.5 cm., containing the alcohol-water mixture, in a tall beaker, 35 by 10 cm., containing ice and water, until cold. Then drop the disk of fat into the tube and it will at once sink to a point where the density of the alcohol-water mixture is exactly equivalent to its own. Lower an accurate thermometer, which can be read to $0.1^{\circ}\text{C}.$, into the test tube until the bulb is just above the disk. In order to secure an even temperature in all parts of the alcohol-water mixture around the disk, stir gently with the thermometer. Slowly heat the water in the beaker, constantly stirring it by means of an air blast or other suitable device.

When the temperature of the alcohol-water mixture rises to about $6^{\circ}\text{C}.$ below the melting point of the fat, the disk of fat begins to shrivel and gradually rolls up into an irregular mass. Lower the thermometer until the fat particle is even with the center of the bulb. Rotate the thermometer bulb gently and regulate the temperature so that about 10 minutes for the increment of the last $2^{\circ}\text{C}.$ are required. As soon as the fat mass becomes spherical, read the thermometer. Remove the tube from the bath and again cool. Place in the bath a second tube containing the alcohol-water mixture. The test tube is of sufficiently low temperature to cool the bath to the desired point, ice water having been used for cooling. After the first or preliminary determination, regulate the temperature of the bath so as to reach a maximum of about $1.5^{\circ}\text{C}.$ above the melting point of the fat under examination.

Do not allow the edge of the disk to touch the sides of the tube. If so, make a new determination. Run triplicate determinations of which the second and third results should agree closely.

11

Capillary Tube Method⁵.—Tentative.

Draw the melted fat or fatty acids into a thin-walled capillary tube. Use a column of fat 1-2 cm. long, according to the length of the thermometer bulb. Seal 1 end of the tube and cool on ice 12-15 hours. Attach the capillary tube to the bulb of an accurate thermometer, graduated to $0.2^{\circ}\text{C}.$, immerse in a large test tube of water surrounded by a beaker of water and heat very slowly. An apparatus similar to that indicated in Fig. 12 may be used. The temperature at which the substance becomes transparent is taken as the melting point.

TITER TEST.

Alcoholic or Aqueous Sodium Hydroxid Method.—Tentative.

12

APPARATUS.

Standard thermometer.—The thermometer must have a zero mark, 0.1° graduations between 10° - $60^{\circ}\text{C}.$, and auxiliary reservoirs at the upper end and between the 0° and the 10° marks. The cavity in the capillary tube between the 0° and the 10° marks must be at least 1 cm. below the 10° mark, which must be about 3-4 cm. above the bulb, the total length of the thermometer being about 38 cm. The bulb should be about 3 cm. long and 6 mm. in diameter. The stem of the thermometer should be 6 mm. in diameter and made of the best thermometer tubing, with scale etched on the stem, the graduation to be clear cut and distinct. The thermometer should have been annealed for 75 hours at $450^{\circ}\text{C}.$, and the bulb should be of Jena normal 16^{III} glass, moderately thin, so that the thermometer will be quick-acting.

13

DETERMINATION.

Saponify 75 grams of the sample in a metal dish with 60 cc. of 30% sodium hydroxid solution (36° Baumé) and 75 cc. of 95% alcohol by volume or 120 cc. of water. Evaporate to dryness over a very low flame or on an iron or asbestos plate, stirring constantly. Dissolve the dry soap in a liter of boiling water and, if alcohol has been used, boil for 40 minutes to remove it, adding sufficient water to replace that lost in boiling. Liberate the fatty acids by adding 100 cc. of 30% sulphuric

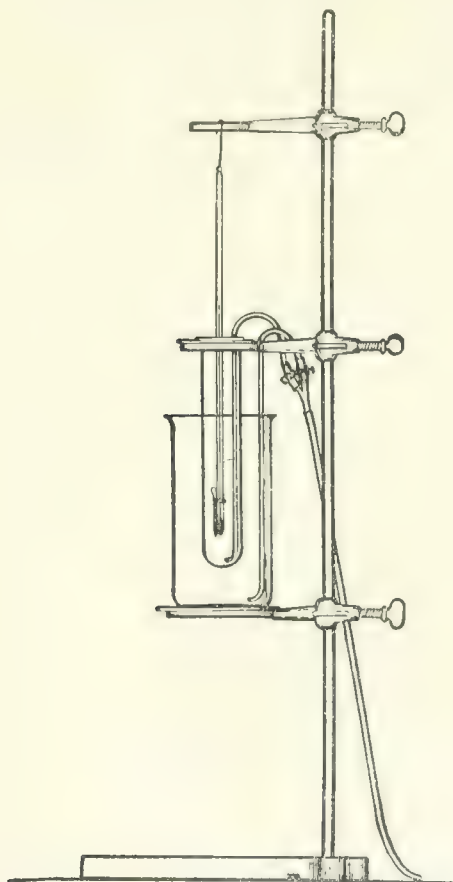


FIG. 12. APPARATUS FOR THE MELTING POINT DETERMINATION.

acid (25° Baumé) and boil until they form a clear, transparent layer. Wash with boiling water until free from sulphuric acid, collect in a small beaker and place on a steam bath until the water has settled and the fatty acids are clear; then decant into a dry beaker, filter while hot and dry 20 minutes at 100°C. When dried, cool the fatty acids to 15°–20°C. above the expected titer and transfer to the titer tube, 25 by 100 mm. (1 by 4 inches) and made of glass about 1 mm. in thickness. Place in a 16 ounce wide-mouthed bottle of clear glass, 70 by 150 mm. (2.8 by 6

inches), fitted with a perforated cork, so as to hold the tube rigidly when in position. Suspend the standard thermometer so that it can be used as a stirrer, and stir the mass slowly until the mercury remains stationary for 30 seconds. Then allow the thermometer to hang quietly, with the bulb in the center of the mass, and observe the rise of the mercury column. The highest point to which it rises is regarded as the titer of the fatty acids.

Test the fatty acids for complete saponification as follows:

Place 3 cc. in a test tube and add 15 cc. of 95% alcohol by volume. Bring the mixture to a boil and add an equal volume of ammonium hydroxid (sp. gr. 0.96). A clear solution should result. The titer must be made at about 20°C. for all fats having a titer above 30°C., and at 10°C. below the titer for all other fats.

14

Glycerol-Potassium Hydroxid Method.—Tentative.

Heat 75 cc. of glycerol-potassium hydroxid solution (25 grams of potassium hydroxid in 100 cc. of high-test glycerol) to 150°C. in an 800 cc. beaker; then add 50 cc. of the oil or melted fat, previously filtered if necessary to remove foreign substances. Saponification often takes place almost immediately, but heating, with frequent stirring, should be continued for 15 minutes, avoiding a temperature much above 150°C. When the saponification is complete, as indicated by the perfectly homogeneous solution, pour the soap into an 800 cc. casserole containing about 500 cc. of nearly boiling water, add carefully 50 cc. of 30% sulphuric acid and heat the solution, with frequent stirring, until the layer of fatty acids separates out perfectly clear. Transfer the fatty acids to a tall separatory funnel, wash 3–4 times with boiling water to remove all mineral acids, draw the fatty acids off into a small beaker, and allow to stand on a steam bath until the water has settled out and the acids are clear. Filter into a dry beaker and heat to 150°C. on a thin asbestos plate, stirring continually with the thermometer, transfer to a titer tube, fill it to within 2.5 cm. of the top and take the titer as directed in 13.

IODIN ABSORPTION NUMBER.

Hübl Method.—Official.

15

REAGENTS.

(a) *Hübl's iodine solution.*—Dissolve 26 grams of pure iodine in 500 cc. of 95% alcohol by volume. Dissolve 30 grams of mercuric chlorid in 500 cc. of 95% alcohol by volume. Filter the latter solution, if necessary, and mix the 2 solutions. Let the mixed solution stand 12 hours before using. The solution loses strength with age, but can be used so long as 35 cc. of N/10 thiosulphate are equivalent to 25 cc. of the iodine solution.

(b) *N/10 sodium thiosulphate.*—Standardize this solution as follows: Place in a glass-stoppered flask 20 cc. of the N/10 potassium dichromate and 10 cc. of the 15% potassium iodide solution. Add 5 cc. of strong hydrochloric acid. Dilute with 100 cc. of water and allow the N/10 sodium thiosulphate to flow slowly into the flask until the yellow color of the liquid has almost disappeared, add a few drops of the starch indicator and, with constant shaking, continue to add the N/10 sodium thiosulphate until the blue color just disappears.

(c) *Starch indicator.*—Prepare as directed under VII, 3 (a).

(d) *15% potassium iodide solution.*

(e) *N/10 potassium dichromate.*—The dichromate solution should be checked against pure iron.

16

DETERMINATION.

Weigh about 0.500 gram of fat, or 0.250 gram of oil (0.100–0.200 gram in the case of drying oils which have a very high absorbent power), into a 500 cc. glass-stoppered flask or bottle. Dissolve the fat or oil in 10 cc. of chloroform. Add 30 cc. of the Hübl iodine solution in the case of fats, or 40–50 cc. in the case of oils. Place the bottle in a dark place and allow to stand for 3 hours, shaking occasionally.

This time must be adhered to closely in order to obtain good results. The time allowed does not give the complete iodine absorption power of an oil or fat and can not be compared with determinations in which 6–12 hours have been used. It gives very satisfactory comparative results, but the time factor must be closely observed.

The excess of iodine should be at least as much as is absorbed. Add 20 cc. of the 15% potassium iodide solution, shake thoroughly and then add 100 cc. of water, washing down any free iodine that may be found on the stopper. Titrate the iodine with the N/10 sodium thiosulphate, adding the latter gradually, with constant shaking, until the yellow color of the solution has almost disappeared. Add a few drops of the starch indicator and continue the titration until the blue color has entirely disappeared. Toward the end of the titration, stopper the bottle and shake violently, so that any iodine remaining in solution in the chloroform may be taken up by the potassium iodide solution. Conduct 2 blank determinations along with that on the sample. The number of cc. of the N/10 sodium thiosulphate required by the blank less the amount used in the determination gives the thiosulphate equivalent of the iodine absorbed by the fat or oil. Ascertain the iodine number by calculating the per cent by weight of iodine absorbed.

Hanus Method.—Official.

17

REAGENTS.

Hanus' iodine solution.—Dissolve 13.2 grams of iodine in 1 liter of glacial acetic acid (99.5%) which shows no reduction with dichromate and sulphuric acid. Add enough bromine to double the halogen content as determined by titration (3 cc. of bromine are about the proper amount). The iodine may be dissolved by heating but the solution should be cold when the bromine is added.

A convenient way to prepare the Hanus solution is as follows: Measure 825 cc. of acetic acid which has shown no reduction by the dichromate test and dissolve in it 13.615 grams of iodine with the aid of heat. Cool and titrate 25 cc. of this solution against the N/10 sodium thiosulphate. Add 3 cc. of bromine to 200 cc. of acetic acid and titrate 5 cc. of the solution against the N/10 sodium thiosulphate. Calculate the quantity of bromine solution required exactly to double the halogen content of the remaining 800 cc. of iodine solution as follows:

$$A = \frac{B}{C} \text{ in which}$$

A = cc. of bromine solution required;

B = 800 × the thiosulphate equivalent of 1 cc. of iodine solution;

C = the thiosulphate equivalent of 1 cc. of bromine solution.

Example: 136.15 grams of iodine are dissolved in 8250 cc. of acetic acid. 30 cc. of bromine are dissolved in 2000 cc. of acetic acid. Titrating 50 cc. of the iodine solution against the standard thiosulphate shows that 1 cc. of the iodine solution equals 1.1 cc. of the thiosulphate (0.0165 gram of iodine). Titrating 5 cc. of the bromine solution shows that 1 cc. of the bromine solution equals 4.6 cc. of the thiosulphate. Then the quantity of bromine solution required to double the halogen content of the remaining 8200 cc. of iodine solution is equivalent to $\frac{8200 \times 1.1}{4.6}$ or 1961 cc. Upon

mixing the 2 solutions in this proportion, a total volume of 10161 cc. is obtained, containing 135.3 grams of iodine. In order to reduce this solution to the proper strength (13.2 grams iodine per liter), $10.161 \times 13.2 = 134.1$; $135.3 - 134.1 = 1.2$ grams of iodine present in excess, or $\frac{1.2 \times 1000}{13.2} = 91$ cc. of acetic acid which must be added.

The other reagents used are described under 15.

18**DETERMINATION.**

Weigh about 0.500 gram of fat, or 0.250 gram of oil (0.100–0.200 gram in the case of drying oils which have a very high absorbent power), into a 500 cc. glass-stoppered flask or bottle. Dissolve the fat, or oil, in 10 cc. of chloroform, add 25 cc. of the Hanus iodine solution and allow to stand for 30 minutes, shaking occasionally. The excess of iodine should be at least 60% of the amount added. Add 10 cc. of the 15% potassium iodide solution and continue as directed under 16.

SAPONIFICATION NUMBER (KOETTSTORFER NUMBER).—OFFICIAL.**1****REAGENTS.**

(a) *N/2 hydrochloric acid.*—Prepare as directed under I, 16 (a).

(b) *Alcoholic potassium hydroxide solution.*—Dissolve 40 grams of the purest potassium hydroxide in 1 liter of 95% redistilled alcohol by volume. The alcohol should be redistilled from potassium hydroxide over which it has been standing for some time, or with which it has been boiled for some time using a reflux condenser. The solution must be clear and the potassium hydroxide free from carbonates.

20**DETERMINATION.**

Weigh accurately about 5 grams of the filtered sample into a 250–300 cc. Erlenmeyer flask. Pipette 50 cc. of the alcoholic potassium hydroxide solution into the flask, allowing the pipette to drain for a definite time. Connect the flask with an air condenser and boil until the fat is completely saponified (about 30 minutes). Cool and titrate with the *N/2* hydrochloric acid, using phenolphthalein as an indicator. Calculate the Koettstorfer number (mg. of potassium hydroxide required to saponify 1 gram of fat). Conduct 2 or 3 blank determinations, using the same pipette and draining for the same length of time as above.

21**SOLUBLE ACIDS.—OFFICIAL.**

Place the flask, used in 20, on a water bath and evaporate the alcohol. Add such an amount of *N/2* hydrochloric acid that its volume plus the amount used in titrating for the saponification number will be 1 cc. in excess of the amount required to neutralize the 50 cc. of the alcoholic potassium hydroxide solution added, and place on the steam bath until the separated fatty acids form a clear layer on the upper surface of the liquid. Fill to the neck with hot water and cool in ice water until the cake of fatty acids is thoroughly hardened. Pour the liquid contents of the flask through a filter into a liter flask. Fill the flask again with hot water, set on the steam bath until the fatty acids collect at the surface, cool by immersing in ice water, and again filter the liquid into the liter flask. Repeat this treatment with hot water 3 times, cooling and collecting the washings in the liter flask after each treatment. Titrate the combined washings with *N/10* alkali, using phenolphthalein as an indicator. Subtract 5 (corresponding to the excess of 1 cc. of *N/2* acid) from the number of cc. of *N/10* alkali used and multiply by 0.0088 to obtain the weight of soluble acids as butyric acid. Calculate the percentage of soluble acids.

22

INSOLUBLE ACIDS (HEHNER NUMBER).—OFFICIAL.

Allow the flask, containing the cake of insoluble fatty acids from **21** and the paper through which the soluble fatty acids have been filtered, to drain and dry for 12 hours. Transfer the cake, together with as much of the fatty acids as can be removed from the filter paper, to a weighed, wide-mouthed beaker flask. Then place the funnel, containing the filter, in the neck of the flask and wash the paper thoroughly with hot absolute alcohol. Remove the funnel, evaporate off the alcohol, dry for 2 hours at 100°C., cool in a desiccator and weigh. Again dry for 2 hours, cool and weigh. If there is any considerable decrease in weight, re-heat for 2 hours and weigh again. Calculate the percentage of insoluble fatty acids.

SOLUBLE VOLATILE ACIDS (REICHERT-MEISSEL NUMBER).

(As these determinations are entirely empirical, the directions given must be followed exactly. In reporting results the method used should always be stated.)

Reichert-Meissl Method.—Official.

23

REAGENTS.

(a) *Sodium hydroxid solution (1 to 1).*—The sodium hydroxid should be as free as possible from carbonates. Protect the solution from contact with carbon dioxide. Allow to settle and use only the clear liquid.

(b) *Potassium hydroxid solution.*—Dissolve 100 grams of the purest potassium hydroxid in 58 cc. of hot water. Cool in a stoppered vessel, decant the clear solution and protect from contact with carbon dioxide.

(c) *95% alcohol by volume.*—Distilled over sodium hydroxid.

(d) *Dilute sulphuric acid.*—Dilute 200 cc. of the strongest acid to 1 liter with water.

(e) *Barium (or sodium) hydroxid solution.*—Standardize an approximately N/10 solution.

(f) *Indicator.*—Dissolve 1 gram of phenolphthalein in 100 cc. of 95% alcohol.

(g) *Pumice stone.*—Heat small pieces to a white heat, plunge in water, and keep under water until used.

24

SAPONIFICATION.

Weigh 5.75 cc., about 5 grams, of the filtered sample, into a saponification flask and proceed by 1 of the following 3 methods.

(1) *Under pressure with alcohol.*—Place 10 cc. of the 95% alcohol in the flask containing the fat (the flask must be made of strong, well-annealed glass, capable of resisting the tension of alcoholic vapor at 100°C.) and add 2 cc. of the sodium hydroxid solution. Insert a soft, cork stopper into the neck of the flask, tie down and place the stoppered flask on a water or steam bath for at least an hour, rotating the flask gently from time to time. Cool before opening.

(2) *Under pressure without the use of alcohol.*—Place 2 cc. of the potassium hydroxid solution in the flask containing the fat (the flask being round-bottomed and made of well-annealed glass to resist pressure), cork and heat as directed under (1). This method avoids the formation of esters and the removal of the alcohol after saponification.

(3) *With a reflux condenser and the use of alcohol.*—Place 10 cc. of the 95% alcohol in the flask containing the fat, add 2 cc. of the sodium hydroxid solution and heat on a steam bath until the saponification is complete, using a reflux condenser.

After the saponification, if alcohol was used, remove it by evaporation on a steam bath.

25

DISTILLATION AND TITRATION.

Dissolve the soap, obtained as directed under 24, by adding 135 cc. of recently boiled water (132 cc. if potassium hydroxid was used in the saponification) and warm on the water bath, with occasional shaking, until the solution is clear. Cool to 60°–70°C., add 5 cc. of the dilute sulphuric acid (8 cc. if potassium hydroxid was used in the saponification), stopper loosely and heat on the water bath until the fatty acids form a clear, transparent layer. Cool to room temperature, add a few pieces of the pumice stone and connect with a glass condenser by means of a bulb tube. Heat slowly with a free flame until ebullition begins and distil, regulating the flame so as to collect 110 cc. of distillate in as nearly 30 minutes as possible. Mix this distillate, filter through a dry filter, and titrate 100 cc. with the standard barium or sodium hydroxid solution, using phenolphthalein as an indicator. The red color should remain unchanged for 2–3 minutes.

Multiply the number of cc. of N/10 alkali used by 1.1, divide by the weight of fat taken and multiply by 5 to obtain the Reichert-Meissl number. Correct the result by the figure obtained in a blank determination.

Leffman and Beam Method.—Official.

26

REAGENTS.

Glycerol-soda solution.—Add 20 cc. of the sodium hydroxid solution, prepared as directed under 23 (a), to 180 cc. of pure, concentrated glycerol.

The other reagents used are described under 23.

27

DETERMINATION.

Add 20 cc. of the glycerol-soda solution to about 5 grams of the fat in a flask, weighed as directed under 24, and heat over a free flame, or on an asbestos plate, until complete saponification takes place, as shown by the mixture becoming perfectly clear. If foaming occurs, shake the flask gently.

Add 135 cc. of recently boiled water, drop by drop at first, to prevent foaming, and 5 cc. of the dilute sulphuric acid, distil without previous melting of the fatty acids, using an apparatus similar to that illustrated in 28, Fig. 13, regulating the flame so as to collect 110 cc. of distillate in as nearly 30 minutes as possible. Filter, titrate the volatile acids and calculate the Reichert-Meissl number, as directed under 25. Conduct a blank determination and correct the result accordingly.

INSOLUBLE VOLATILE ACIDS (POLENSKE NUMBER).

28

Polenske Method^s.—Tentative.

Proceed as directed under 27 up to the point at which 110 cc. of distillate have been collected, except that only 20 minutes are allowed for the distillation, employing an apparatus of the *exact* dimensions illustrated in Fig. 13. Substitute a 25 cc. cylinder for the receiving flask to collect any drops that may fall after the flame has been removed. Immerse the flask containing the distillate almost completely in water at 15°C. for 15 minutes, filter the 110 cc. of distillate and determine the approximate Reichert-Meissl number, if desired, as in 27, avoiding too violent shaking of the distillate and consequent emulsification of the insoluble acids previous to filtration. Remove the remainder of the soluble acids from the insoluble acids upon the filter paper by washing with 3 successive portions of 15 cc. of water, previously passed through the condenser, the 25 cc. cylinder and the 110 cc. receiving flask. Then dissolve the insoluble acids by passing 3 successive 15 cc. por-

tions of neutral 90% alcohol by volume through the filter, each portion previously passed through the condenser, the 25 cc. cylinder and the 110 cc. receiving flask. Titrate the combined alcoholic washings with N/10 sodium hydroxid, using phenolphthalein as an indicator.

Run a blank in the same manner and subtract the quantity of the standard alkali required to neutralize the 45 cc. of alcohol, used in washing the apparatus and filter paper of the blank, from that required in each Polenske determination. Report the Polenske number as the number of cc. of N/10 alkali required to neutralize the insoluble volatile acids from the 110 cc. of distillate as obtained above. Since the entire distillate is filtered it is not necessary to multiply the burette reading by 1.1, as in 25, but a calculation must be made, as directed in 25, to reduce the actual number of cc. found in the titration to the number which would have been required had exactly 5 grams of fat been used.

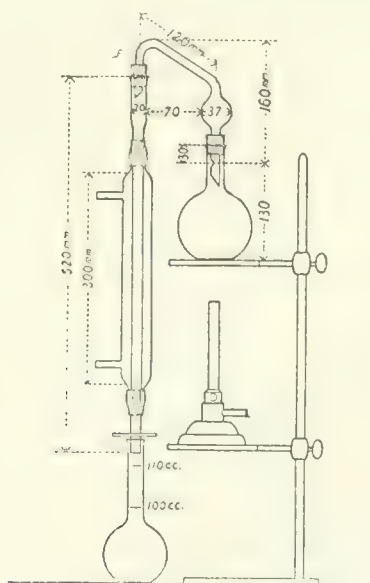


FIG. 13. APPARATUS⁷ FOR THE DETERMINATION OF THE POLENKE NUMBER.

LIQUID AND SOLID FATTY ACIDS.

29

Muter Method, Modified by Lane⁸.—Tentative.

Weigh 5 grams of the oil or fat into an Erlenmeyer flask, saponify, precipitate with lead acetate solution and treat the precipitated lead soap with ether, as directed under 37. Filter the ether solution of the soluble lead soap into a Muter tube or separatory funnel and decompose the soap by shaking with 40 cc. of hydrochloric acid (1 to 5). The soap is completely decomposed when the ether becomes clear and colorless.

Draw off the lead chlorid from the ether solution and wash the ether free from acid. Evaporate, until free from ether, an aliquot of this solution in an atmosphere of carbon dioxide, in order to prevent the oxidation of the oleic acid, and weigh to determine the per cent of liquid acids; determine the iodine number as directed in 16 or 18, using 0.2–0.3 gram of this residue.

As it is very difficult to dry the unsaturated acids without very serious oxidation, it is just as satisfactory to determine the weight of insoluble acids by the following method:

Wash the insoluble soap left on the filter into a flask, decompose with hydrochloric acid and heat until the fatty acids are melted. Fill the flask with hot water, cool, pour off the water and again wash the solidified fatty acids. Dissolve in hot 95% alcohol by volume, transfer to a weighed dish, remove the alcohol by evaporation, dry, weigh and calculate the per cent of solid fatty acids.

30

FREE FATTY ACIDS.—OFFICIAL.

Weigh 20 grams of fat, or oil, into a flask, add 50 cc. of 95% alcohol by volume which has been neutralized with dilute sodium hydroxid solution, using phenolphthalein as an indicator, and heat to boiling. Shake the flask thoroughly in order to dissolve the free fatty acids as completely as possible. Titrate with N/10 alkali, shaking thoroughly until the pink color persists after vigorous shaking.

Express the results either as percentage of oleic acid, as acid degree (cc. of N/1 alkali required to neutralize the free acids in 100 grams of oil or fat), or as acid value (mg. of potassium hydroxid required to saturate the free acids in 1 gram of fat or oil).

One cc. of N/10 alkali is equivalent to 0.0282 gram of oleic acid.

ACETYL VALUE.

31

Benedikt-Lewkowitsch Method².—Tentative.

Boil the oil or fat with an equal volume of acetic anhydrid for 2 hours, pour the mixture into a large beaker containing 500 cc. of water and boil for 30 minutes. To prevent bumping, pass a slow current of carbon dioxid into the liquid through a finely drawn out tube reaching nearly to the bottom. Allow the mixture to separate into 2 layers, siphon off the water, and boil the oily layer with fresh water until it is no longer acid to litmus paper. Separate the acetylated fat from the water and dry and filter in a drying oven.

Weigh 2–4 grams of the acetylated fats into a flask and saponify with alcoholic potash as in 20. If the distillation process is to be adopted, it is not necessary to work with a standardized alcoholic potassium hydroxid solution, but in the filtration method, which is much shorter, the alcoholic potassium hydroxid solution must be measured exactly. In either case evaporate the alcohol after saponification and dissolve the soap in water. Then either distil or filter as follows:

(1) *Distillation*.—Acidify with sulphuric acid (1 to 10) and distil as in 25. As several hundred cc. must be distilled, either run a current of steam through or add portions of water from time to time (500–700 cc. of distillate will be sufficient). Filter the distillates to remove any insoluble acids carried over by the steam and titrate with N/10 potassium hydroxid, using phenolphthalein as an indicator. Multiply the number of cc. of alkali employed by 5.61 and divide by the weight of substance used to obtain the acetyl value.

(2) *Filtration*.—Add to the soap solution a quantity of standard sulphuric acid exactly corresponding to the amount of alcoholic potassium hydroxid solution added, warm gently, filter off the free fatty acids which collect on top, wash with boiling water until the washings are no longer acid and titrate the filtrate with N/10 potassium hydroxid, using phenolphthalein as an indicator. Calculate the acetyl value as directed under (1).

CHOLESTEROL AND PHYTOSTEROL IN MIXTURES OF ANIMAL AND VEGETABLE FATS.

32

Alcohol Extraction Method¹⁰.—Tentative.

Introduce 200–300 grams of the melted fat into a flat-bottomed liter flask. Close the neck of the flask with a 3-holed stopper and insert through these holes: (1) a reflux condenser; (2) a right-angled glass tube, one arm of which reaches to a point 6 mm. above the surface of the melted fat, the other being closed a short distance from the flask by means of a short piece of rubber tubing and a pinch-cock; (3) a glass tube bent so that one arm reaches down to the bottom of the flask and the other serves as a delivery tube for a 700 cc. round-bottomed flask containing 500 cc. of 95% alcohol by volume.

Place the flasks, containing the melted fat and the alcohol, on a steam bath and heat so that the alcohol vapor passes through the melted fat in the liter flask and is condensed in the reflux condenser, finally collecting in a layer over the melted fat. After all the alcohol has passed in this manner into the flask containing the fat, disconnect the flask from which the alcohol has been distilled and attach a tube to the short piece of rubber tubing attached to the right-angled glass tube [see (2) above] and siphon the alcohol layer back into the alcohol distillation flask. Reconnect as at first and again distil the alcohol as in the first operation. When all the alcohol has been distilled, siphon it again into the distillation flask and extract in the same manner for a third time.

Discard the fat and retain the alcohol which now contains practically all of the cholesterol and phytosterol originally present in the fat. Concentrate the alcoholic solution to about 250 cc. and add 20 cc. of potassium hydroxid solution (1 to 1) to the boiling liquid. Boil for 10 minutes to insure complete saponification of the fat, cool to room temperature and pour into a large separatory funnel containing 500 cc. of warm ether. Shake to insure thorough mixing and add 500 cc. of water. Rotate the funnel gently to avoid the formation of extremely stubborn emulsions, but mix the water thoroughly with the alcohol-ether-soap solution. A clear, sharp separation takes place at once. Draw off the soap solution and wash the ether layer with 300 cc. of water, avoiding shaking. Repeat the washing of the ether solution with small quantities of water until all the soap is removed. Transfer the ether layer to a flask and distil the ether until the volume of liquid remaining in the flask measures about 25 cc. Transfer this residue to a tall 50 cc. beaker and continue the evaporation until all the ether is driven off and the residue is perfectly dry. If desired, a tared beaker may be used and the weight of the unsaponifiable matter determined at this point.

Add 3–5 cc. of acetic anhydrid to the residue in the beaker, cover the beaker with a watch glass and heat to boiling over a free flame. After boiling for a few seconds, remove the beaker from the flame, cool and add 35 cc. of 60% alcohol by volume. Mix the contents of the beaker thoroughly, filter off the alcoholic solution and wash the precipitate with 60% alcohol. Dissolve the precipitate on the filter with a stream of hot 80% alcohol by volume and wash the insoluble portion well with 80% alcohol. Acetates of cholesterol and phytosterol are dissolved while the greater portion of the impurities present (including paraffin and paraffin oil if present) remain behind on the filter. Cool the combined filtrate and washings to a temperature of 10°–12°C. and allow to stand at that temperature for 2–3 hours. During this time the acetates of cholesterol and phytosterol crystallize from the solution. Collect the crystals upon a filter, wash with cold 80% alcohol and then dissolve them in a minimum amount of hot absolute alcohol. Collect the alcoholic solution of the acetates in a small, glass evaporating dish, add 2 or 3 drops of water to the solution and heat if not perfectly clear. Allow the alcohol to evaporate

spontaneously, the contents of the dish being stirred occasionally to mix the deposit of crystals, which form upon the edges, with the main body of the liquid. As soon as a good deposit of crystals has formed, collect them upon a hardened filter, wash twice with cold 90% alcohol and dry by suction, drying finally at 100°C. for 30 minutes, and determine the melting point in the apparatus shown in 11, Fig. 12, using sulphuric acid in the outer beaker and glycerin in the inner tube.

The melting point of the first crop of crystals usually gives definite information as to the presence or absence of phytosterol but the conclusion indicated should be confirmed by recrystallizing the crystals from absolute alcohol and again determining the melting point. If the crystals are pure cholesteryl acetate, the melting point of the second crop should agree closely with that of the first. If phytosteryl acetate is present, however, a higher melting point will be noted, as phytosteryl acetate is less soluble in alcohol than cholesteryl acetate. The melting point of cholesteryl acetate is 114°C., that of phytosteryl acetate 125°-137°C.

33

Digitonin Method. (Marcusson and Schilling¹¹)—Tentative.

Shake vigorously 50 grams of the oil or fat for 15 minutes in a separatory funnel with 20 cc. of a 1% solution of digitonin in 95% alcohol by volume. Allow the mixture to stand for a time until the emulsion separates. The lower or fat layer should be quite clear while the alcohol layer contains a bulky, flocculent precipitate. Draw off as much as possible of the fat, avoiding any loss of the precipitate. Add 100 cc. of ether to the alcohol layer and filter the mixture. Wash the precipitate with ether until free from fat; after drying in the air, transfer it to a tall 50 cc. beaker, add 2-3 cc. of acetic anhydrid and cover the beaker with a watch glass. Then boil slowly over a low flame for 30 minutes. After cooling, add 30-35 cc. of 60% alcohol by volume and mix the contents of the beaker thoroughly. Filter off the alcohol solution and wash the precipitate with 60% alcohol, then dissolve it on the filter with a stream of hot 80% alcohol by volume from a wash bottle and set aside the filtrate in a cool place (10°C. or below). After the acetates have crystallized out of this solution filter them off, recrystallize from absolute alcohol, dry and determine the melting point of each crop of crystals, as directed under 32.

34

UNSAAPONIFIABLE RESIDUE¹².—TENTATIVE.

Saponify 5 grams of the oil or fat with alcoholic potassium hydroxid solution and remove the alcohol by evaporation. Wash into a separatory funnel with 70-100 cc. of water and extract with 50-60 cc. of ether. If the 2 liquids do not separate, add a few cc. of alcohol. Separate the water solution and wash the ether with water containing a few drops of sodium hydroxid solution. Again extract the soap solution and washings with ether and evaporate the combined extracts to dryness. In most cases it is advisable to add a little alcoholic potassium hydroxid solution to the residue and heat in order to saponify any traces of fats left unsaponified and extract again with ether. Transfer to a weighed dish and dry as quickly as possible in a water oven.

Many of the hydrocarbon oils are volatile at 100°C., so that the drying should not be carried any further than necessary. With resin oil, paraffin wax and the denser mineral oils there is little danger of loss at 100°C.

On account of the solubility of soap in ether and petroleum ether it is well to wash the residue with warm water containing a little phenolphthalein. If the reaction is alkaline, soap is present and the residue must be further purified.

RESIN OIL.

35

Qualitative Test.—*Tentative.*

Polarize the pure oil, or a definite dilution with petroleum ether, in a 200 mm. tube. Resin oil has a polarization in a 200 mm. tube of from $+30^{\circ}$ to $+40^{\circ}$ on the sugar scale (Schmidt and Haensch) while most oils¹³ read between $+1^{\circ}$ and -1° .

COTTONSEED OIL.

36

*Halphen Test*¹⁴.—*Official.*

Mix carbon disulphid, containing about 1% of sulphur in solution, with an equal volume of amyl alcohol. Mix equal volumes of this reagent and the oil under examination, and heat in a bath of boiling, saturated brine for 1–2 hours. In the presence of as little as 1% of cottonseed oil, a characteristic red or orange-red color is produced.

Lard and lard oil from animals fed on cottonseed meal will give a faint reaction; their fatty acids also give this reaction.

The depth of color is proportional, to a certain extent, to the amount of oil present, and by making comparative tests with cottonseed oil some idea as to the amount present can be obtained. Different oils react with different intensities, and oils which have been heated from 200° – 210°C .¹⁵ react with greatly diminished intensity. Heating 10 minutes at 250°C . renders cottonseed oil incapable of giving the reaction¹⁶.

PEANUT OIL.

37

*Modified Renard Test*¹⁷.—*Tentative.*

Weigh 20 grams of the oil into an Erlenmeyer flask. Saponify with alcoholic potash solution, neutralize exactly with dilute acetic acid, using phenolphthalein as an indicator, and wash into an 800–1000 cc. flask containing a boiling mixture of 100 cc. of water and 120 cc. of 20% lead acetate solution. Boil for a minute and then cool the precipitated soap by immersing the flask in water, occasionally giving it a whirling motion to cause the soap to stick to the sides of the flask. After the flask has cooled, decant the water and excess of lead acetate solution and wash the lead soap with cold water and 90% alcohol by volume. Add 200 cc. of ether, cork and allow to stand for some time until the soap is disintegrated; heat on a water bath, using a reflux condenser, and boil for about 5 minutes¹⁸. In the case of oils, most of the soap will be dissolved, while in lards, which contain much stearin, part of the soap will be left undissolved. Cool the ether solution of soap to 15° – 17°C . and allow to stand until all the insoluble soaps have separated out (about 12 hours).

Filter upon a Büchner funnel and thoroughly wash the insoluble lead soaps with ether. Wash the ether-insoluble lead soaps into a separatory funnel by means of a jet of ether, alternating at the end of the operation, if a little of the soap sticks to the paper, with hydrochloric acid (1 to 3). Add sufficient hydrochloric acid (1 to 3) so that the total volume of the latter amounts to about 200 cc. and enough ether to make the total volume of it 150–200 cc. and shake vigorously for several minutes. Allow the layers to separate, run off the acid layer, and wash the ether once with 100 cc. of dilute hydrochloric acid and then with several portions of water until the water washings are no longer acid to methyl orange. If a few undecomposed lumps of lead soap remain (indicated by solid particles remaining after the third washing with water), break these up by running off almost all the water layer and then add little concentrated hydrochloric acid, shake and then continue the washing with water as before.

Distil the ether from the solution of insoluble fatty acids and dry the latter in the flask by adding a little absolute alcohol and evaporating on a steam bath. Dissolve the dry fatty acids by warming with 100 cc. of 90% alcohol by volume and cool slowly to 15°C., shaking to aid crystallization. Allow to stand at 15°C. for 30 minutes. In the presence of peanut oil, crystals of arachidic acid will separate from the solution. Filter, wash the precipitate twice with 10 cc. of 90% alcohol by volume, and then with 70% alcohol by volume, care being taken to maintain the arachidic acid and the wash solutions at a definite temperature in order to apply the solubility corrections given below. Dissolve the arachidic acid upon the filter with boiling absolute alcohol, evaporate to dryness in a weighed dish, dry and weigh. Add to the weight 0.0025 gram for each 10 cc. of 90% alcohol used in the crystallization and washing, if conducted at 15°C.; if conducted at 20°C., add 0.0045 gram for each 10 cc. The melting point of arachidic acid thus obtained is 71°–72°C. Twenty times the weight of arachidic acid will give the approximate amount of peanut oil present. Arachidic acid has a characteristic appearance and may be identified by the microscope. As little as 5–10% of peanut oil can be detected by this method.

SESAME OIL.

38

Baudoin Test.—Official.

Dissolve 0.1 gram of finely powdered sugar in 10 cc. of hydrochloric acid (sp. gr. 1.20), add 20 cc. of the oil to be tested, shake thoroughly for a minute and allow to stand. The aqueous solution separates almost at once and, in the presence of even a very small admixture of sesame oil, is colored crimson. Some olive oils give a slight pink coloration with this reagent. Comparative tests with known samples containing sesame oil will differentiate them.

39

Villavecchia Test¹⁹.—Official.

Add 2 grams of furfural to 100 cc. of 95% alcohol by volume and mix thoroughly 0.1 cc. of this solution, 10 cc. of hydrochloric acid (sp. gr. 1.20), and 10 cc. of the oil by shaking them together in a test tube. A crimson color is developed as in the Baudoin test, **38**, where sugar is used.

Villavecchia explained this reaction on the basis that furfural is formed by the action of levulose and hydrochloric acid and therefore substituted furfural for sucrose. As furfural gives a violet tint with hydrochloric acid it is necessary to use the very dilute solution specified in the method.

BEEF FAT IN LARD.

40

Emery Method²⁰.—Tentative.

Weigh 5 grams of the melted fat into a glass-stoppered 25 cc. cylinder about 150–175 mm. in height. Add warm ether up to the 25 cc. mark, stopper securely and shake until the fat is completely dissolved. Allow the cylinder to stand for about 18 hours at a temperature of 16°–20°C. during which time some of the solid glycerides will crystallize out. Decant the clear solution carefully from the crystals, wash with three 5 cc. portions of cold ether, avoiding breaking up the deposit during the first 2 washings. Agitate the crystals with the third portion of ether and transfer to a small filter. Wash on the paper with successive small amounts of cold ether until 15–20 cc. have been used, then remove the last traces of ether by means of slight suction on the stem of the funnel. Break up any large lumps and allow the deposit to dry.

When thoroughly dry pulverize the glycerides and take their melting point in a closed 1 mm. tube, using an apparatus similar to that indicated in **11**, Fig. 12. Heat the water in the beaker rapidly to about 55°C. and maintain that temperature until the thermometer carrying the melting-point tube registers 50°–55°C., then heat again and carry the temperature of the outer bath somewhat rapidly to 67°C. when the lamp is removed. The melting point of the crystals is regarded as that point when the fused substance becomes perfectly clear and transparent. A dark background placed about 4 inches from the apparatus will prove of advantage. When the melting point of the glycerides obtained by this method is below 63.4°C. the presence of beef fat should be suspected, while a melting point of 63°C., or below, can be regarded as positive evidence that the sample is not pure lard. It is advisable to carry out this method with a control sample of pure lard in connection with each batch of samples analyzed.

FISH OIL AND MARINE ANIMAL OILS IN THE PRESENCE OF VEGETABLE OILS AND IN THE ABSENCE OF METALLIC SALTS.

41

Qualitative Test.—Tentative.

Dissolve in a test tube about 6 grams of the oil in 12 cc. of a mixture of equal parts of chloroform and glacial acetic acid. Add bromin, drop by drop, until a slight excess is indicated by the color, keeping the solution at about 20°C. Allow to stand 15 minutes or more and then place the test tube in boiling water. If vegetable oils only are present, the solution will become perfectly clear, while fish oils will remain cloudy or contain a precipitate due to the presence of insoluble bromids.

42

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI, 3**.

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XXIV. SPICES AND OTHER CONDIMENTS.

SPICES.

1 PREPARATION OF SAMPLE.—TENTATIVE.

Grind the sample so as to pass through a sieve having circular openings 1 mm. in diameter and mix thoroughly. Owing to the lack of uniformity of most spices and the peculiar tendency to stratify, extreme care is necessary in weighing out a portion for analysis. Stir the material with a spoon, having a capacity of approximately 2 grams, and dip a spoonful from the interior in order that only a very small amount needs to be added to or taken from the portion on the scale pan. In the determination of starch in spices by the diastase method, reduce a portion of the sample to an impalpable powder by grinding in a mortar.

2 MOISTURE.—TENTATIVE.

Dry 2 grams to constant weight at 110°C. From the resulting loss in weight subtract the amount of volatile ether extract as determined in 9.

3 ASH.—OFFICIAL.

Determine as directed under VIII, 4.

4 SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.

Proceed as directed under IX, 17, employing the ash obtained in 3.

5 ASH INSOLUBLE IN ACID.—TENTATIVE.

Boil the water-insoluble residue, obtained in 4, or the total ash obtained in 3, with 25 cc. of 10% hydrochloric acid (sp. gr. 1.050) for 5 minutes, collect the insoluble matter on a Gooch or an ashless filter, wash with hot water, ignite and weigh.

6 CALCIUM OXID IN ASH.—OFFICIAL.

Ignite 2-4 grams of the sample as directed under 3, digest with hot 10% hydrochloric acid, evaporate to dryness, moisten the dry residue with dilute hydrochloric acid and again evaporate to dryness to render the silica insoluble. Moisten the residue with 5-10 cc. of hydrochloric acid, add about 50 cc. of water, allow to stand on a water bath for a few minutes, filter and wash the insoluble residue with hot water. Determine calcium oxid in the combined filtrate and washings as directed under XXVIII, 23.

7 NITROGEN.—OFFICIAL.

Determine as directed under I, 18, 21 or 23, except in the case of black and white peppers in which use only the Kjeldahl-Gunning-Arnold method¹ [I, 23], employing 1 gram of the sample.

NITROGEN IN NON-VOLATILE ETHER EXTRACT.

8 Winton, Ogden and Mitchell Method.—Tentative.

(For black and white peppers.)

Extract 10 grams of the pepper for 20 hours in a continuous extraction apparatus with absolute ether, collecting the extract in a weighed 250 cc. flask. Evaporate

the ether, dry first at 100°C. and finally to constant weight at 110°C. Determine the nitrogen in the weighed extract, as directed in **I, 23**, digesting in the same flask used for the extraction. Calculate the parts of nitrogen per 100 parts of non-volatile ether extract. If desired, crude piperin may be calculated from the nitrogen by multiplying by 20.36.

9 VOLATILE AND NON-VOLATILE ETHER EXTRACT².—TENTATIVE.

Extract 2 grams of the ground material for 20 hours in a continuous extraction apparatus with anhydrous ether [**VIII, 9**]. Transfer the ethereal solution to a tared capsule and allow to evaporate at room temperature. Let stand for 18 hours over sulphuric acid and weigh the total ether extract. Heat the extract gradually and then to constant weight at 110°C. The loss is volatile ether extract; the residue, non-volatile ether extract.

10 ALCOHOL EXTRACT³.—TENTATIVE.

Place 2 grams of the sample in a 100 cc. flask and fill to the mark with 95% alcohol by volume. Stopper, shake for 8 hours at 30 minute intervals and allow to stand for 16 hours longer without shaking. Filter the extract through a dry filter, evaporate a 50 cc. aliquot of the filtrate to dryness in a flat-bottomed dish on a water bath and heat to constant weight at 110°C.

11 COLD-WATER EXTRACT.—TENTATIVE.

(For ginger.)

Place 4 grams of the sample in a 200 cc. graduated flask, add water to the mark, shake at 30 minute intervals during 8 hours and let stand 16 hours longer without shaking. Filter and evaporate a 50 cc. aliquot of the filtrate to dryness in a flat-bottomed, metal dish. Dry to constant weight at 100°C.

12 COPPER-REDUCING SUBSTANCES BY DIRECT INVERSION.—TENTATIVE.

Extract 4 grams of the sample with 5 successive portions of 10 cc. of ether on a filter that will retain completely the smallest starch granules. After the ether has evaporated, wash with 150 cc. of 10% alcohol by volume.

Owing to the formation of a glutinous mass which clogs the filter, it is not possible to wash samples of Batavia cassia with water or dilute alcohol. Therefore all preliminary washing is best omitted in determinations made on all varieties of cassia, as well as on cassia buds and cinnamon.

Carefully wash the residue from the paper into a 500 cc. flask with 200 cc. of water, using a small wash bottle, and gently rubbing the paper with the tip of the finger. Hydrolyze and determine the copper reducing material as directed under **VIII, 60**. Express the result in terms of starch.

13 STARCH.—TENTATIVE.

Extract 4 grams of the finely pulverized sample with ether and 10% alcohol by volume, as directed under **12**, and determine starch by the diastase method, as directed under **VIII, 62**.

14 CRUDE FIBER.—TENTATIVE.

Proceed as directed under **VIII, 68**, and remove all ether extractives by successive washings of the dry fiber with ether previous to weighing.

15

TANNIN.—TENTATIVE.

(For cloves and allspice.)

Extract 2 grams of the sample for 20 hours with anhydrous ether. Boil the residue for 2 hours with 300 cc. of water, cool, make up to 500 cc. and filter. Measure 25 cc. of this infusion into a 1200 cc. flask, add 20 cc. of indigo solution, 750 cc. of water and proceed as directed under XVI, 32. One cc. of N/10 oxalic acid is equivalent to 0.006232 gram of quercitannic acid, or 0.0008 gram of oxygen absorbed.

16

TOTAL SULPHUR.—OFFICIAL.

(For mustard.)

Proceed as directed under III, 17.

OLIVE OIL.

(For paprika.)

17

Qualitative Test.—Tentative.

Spread 5 grams of the paprika on a watch glass and dry over sulphuric acid for at least 12 hours. Measure 250 cc. of anhydrous, alcohol-free ether [VIII, 9] into a graduated flask on which the mark is situated near the lower end of the neck, and brush the paprika into it. Place a mark on the neck of the flask at the point where the meniscus is, and allow to stand for an hour, shaking at 20 minute intervals during that time. Bring the meniscus back to the mark placed upon the neck, either by cooling the flask and contents if the level has risen, or by adding absolute ether if it has fallen; let the solid particles settle and pipette off 100 cc. of the supernatant liquid, filter through an 11 cm. closely woven paper into a tared, air-dry, 250 cc., glass-stoppered Erlenmeyer flask that has been counterpoised against a similar flask; wash the paper with a little absolute ether. Then distil off the solvent and remove the flask from the bath as soon as the ether ceases to come over. Lay the flask on its side in a water oven and heat for 30 minutes; cool the open flask for at least 30 minutes in the air and weigh. Repeat this heating and weighing until the weight is constant to within 1 mg., 2 heatings usually being sufficient. Note the per cent of ether extract obtained. If more than 1½ hours of heating are required to obtain constant weight, or if the ether extract becomes colorless, reject it and start a new determination with freshly purified ether.

Dissolve the ether extract in the flask with 10 cc. of chloroform, add 30 cc. of Hanus' solution [XXIII, 17 (a)] and proceed as directed under XXIII, 18, allowing 30 minutes for the halogen absorption. Note the iodine number of the ether extract. The iodine number of pure paprika thus obtained should not be less than 125.

MICROSCOPIC EXAMINATION.—TENTATIVE.

18

GENERAL.

Adulterants of vegetable origin in spices are detected best by means of the microscope. A general knowledge of vegetable histology and the microscopic appearance of the spices and spice adulterants is essential. Some of the standard works⁴ on these subjects are listed in the bibliography.

19

REAGENTS.

Of the numerous reagents employed in histological work the following are the most useful in spice examinations:—

(a) *Glycerol solution* (1 to 1).

(b) *Absolute alcohol*.

(c) *Ether*.

(d) *Ammonium hydroxid*.—The concentrated solution, containing about 30% of ammonia gas, is used in making Schweitzer's reagent and for some other purposes. For the turmeric test the concentrated solution should be diluted with 10 parts of water.

(e) *5% potassium hydroxid solution*.

(f) *Chloral hydrate solution (8 to 5)*.

(g) *Schulze's mixture*.—Crystallized potassium chlorate mixed with nitric acid as needed.

(h) *Iodin-potassium iodid solution*.—A solution of 0.05 gram of iodine, 0.2 gram of potassium iodide in 15 cc. of water.

(i) *Chlor-zinc iodine solution*.—Dissolve 100 grams of zinc chloride in 60 cc. of water and to this add 20 grams of potassium iodide and 0.5 gram of iodine crystals. A few crystals of iodine should be left in the bottle to insure saturation and allowed to stand a few hours before using. The chlor-zinc iodine solution, prepared in this manner, will keep for months. If the color developed in the tissue is too deep a blue, a very slight dilution of the reagent is advisable.

(j) *Millon's reagent*.—Prepare as in XV, 9.

(k) *1% ferric acetate or chloride solution*.—Freshly prepared.

(l) *Alkanna tincture*.—Macerate 20 grams of alkanet root for several days with 100 cc. of alcohol.

(m) *Aqueous safranin solution*.

(n) *10% hydrochloric acid*.

(o) *Acetic acid*.—Glacial or 99% acetic acid diluted with 2 parts of water.

20

APPARATUS.

(a) *Dissecting microscope or hand lens*.

(b) *Compound microscope*.—Provided with $\frac{3}{4}$ and $\frac{1}{2}$ inch objectives, 1 and 2 inch oculars, double nosepiece, eyepiece micrometer and polarizing apparatus.

(c) *Sieves*.—A series of sieves with meshes ranging from 0.2–2 mm.

(d) *Slides, cover-glasses, needles, scalpels, forceps, etc.*

21

PREPARATION OF SAMPLE.

Reduce one portion to a fine powder in a mortar. Separate another portion into several grades of fineness by sieves of different mesh or by jarring on a sheet of paper. In the coarser grades, fragments of a suspicious nature may often be seen with the naked eye or under a simple microscope; these should be picked out for subsequent examination under the compound microscope.

22

EXAMINATION.

Mount a small quantity of the ground sample in water and examine under the compound microscope with both ordinary and polarized light. This gives general information as to the nature of the material and serves for the detection and identification of starch granules and various tissues. Draw a small drop of the iodine-potassium iodide solution into the same preparation by means of a piece of filter paper placed on the opposite edge of the cover-glass and examine. Starch granules will be colored blue or blue-black, cellulose yellow, and proteins either brown or yellow.

In the manner just described draw a little of the 5% potassium hydroxid solution under the cover-glass and again examine. This treatment gelatinizes the starch

granules, dissolves the proteins, saponifies the fats, and in other ways clears the preparation. It also imparts to tannins a reddish color. If this treatment does not clear the tissues satisfactorily, treat a fresh portion for some hours with the chloral hydrate solution.

Examine also the crude fiber obtained in the chemical analysis, as in this material the stone cells and other tissues are shown distinctly.

To isolate stone cells, bast fibers and other thick-walled cells macerate a portion of the sample in Schultze's mixture, using such proportion of potassium chlorate and nitric acid and heating for such a time as secures the desired results. Powdered charcoal and charred shells resist the bleaching action of potash, chloral hydrate and Schultze's mixture.

If it is desired to distinguish cellulose from infiltrated substances (lignin, suberin, etc.), add the freshly prepared chlor-zinc iodine solution to a water mount, whereby the former is colored blue and the latter yellow.

Test for proteins by cautiously warming on a slide with a drop of freshly prepared Millon's reagent. The proteins are partially decomposed, acquiring gradually a brick-red color. If it is desired to study the form of the aleurone (protein) granules, which in some plants are quite as characteristic as starch granules, prepare a mount in pure glycerol or oil.

To distinguish fats, oils, essential oils and resins from other cell contents, treat for an hour with the alkanna tincture, diluted with an equal bulk of water, which imparts to these substances a deep red color, or treat with ether, which dissolves them. Treat also with alcohol, which dissolves the essential oils and resins, but does not perceptibly affect the fats and oils.

In testing for tannins and tissues impregnated with these substances, add the 1% ferric acetate or chlorid solution. Both of these reagents give a green or blue color with tannins, but the former acts more slowly and is to be preferred.

Crystals of calcium oxalate are recognized by their characteristic forms and their behavior to polarized light. To distinguish calcium oxalate from calcium carbonate, treat with acetic acid, which does not affect the former, but dissolves the latter with effervescence. Both are soluble in hydrochloric acid.

PREPARED MUSTARD.

23

PREPARATION OF SAMPLE.—TENTATIVE.

Transfer the entire contents of the container to a dish sufficiently large to stir thoroughly and make the whole mass homogeneous. Preserve in a bottle having a tightly fitting glass stopper. Stir well each time before removing a portion for analysis.

24

SOLIDS.—TENTATIVE.

Weigh 5 grams of the sample into a flat-bottomed, platinum dish, distribute evenly over the bottom of the dish with a little water, place on a water bath until the mixture appears dry, and heat finally to constant weight at 100°C. in a water oven.

25

ASH.—OFFICIAL.

Ignite the dry residue, obtained in the determination of solids, 24, as directed under VIII, 4.

26

SALT.—TENTATIVE.

Determine chlorin in the ash as directed under III, 15.

27

ETHER EXTRACT.—TENTATIVE.

Weigh 10 grams of the sample into a capsule and mix with about 30 grams of sand. Heat on a water bath until the mixture appears dry and complete the drying in a water oven. Grind until all the lumps are broken up, and determine the ether extract as directed under **VIII, 10**.

28

PROTEIN.—OFFICIAL.

Determine the nitrogen as directed under **I, 18, 21** or **23**, using 5 grams of the sample. Multiply the result by 6.25 to obtain the amount of protein.

29

ACIDITY.—TENTATIVE.

Weigh 10 grams of the sample into a 200 cc. graduated flask, make up to the mark with water, shake, filter through a dry paper and determine the acidity in 100 cc. by titration with N/10 alkali, using phenolphthalein as an indicator. Express the result as acetic acid. One cc. of N/10 alkali is equivalent to 0.0060 gram of acetic acid.

30

COPPER-REDUCING SUBSTANCES.—TENTATIVE.

By Direct Inversion.

Proceed as directed under **VIII, 60**, except that 10 grams of the sample, without previous washing or extraction, are treated directly with 200 cc. of water and 20 cc. of 25% hydrochloric acid and the solution is made up to 250 cc. after neutralizing and before filtering and drawing off the aliquot. In analyses of samples containing starch, particular attention should be given that the amount of dextrose present in the aliquot taken for the reducing sugar determination does not exceed the maximum permitted for that determination. Calculate the result in terms of starch.

31

CRUDE FIBER.—TENTATIVE.

Transfer 8 grams of the sample (equivalent to about 2 grams of dry matter) to a porcelain or glass mortar. Treat with a little hot 1.25% sulphuric acid and rub into a uniform thin paste. It is absolutely essential that this paste be uniform in consistency and entirely free from lumps. Rinse the thin mixture into a 500 cc. Erlenmeyer flask, using a total volume of 200 cc. of the hot 1.25% sulphuric acid for the entire operation. Proceed as directed under **VIII, 68**, and remove all the fat, previous to weighing of the crude fiber, by repeated washings of the dry fiber with ether.

32

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI**.

33

PRESERVATIVES.—TENTATIVE.

Proceed as directed under **X**.

TOMATO PRODUCTS.

34

PREPARATION OF SAMPLE.—TENTATIVE.

Shake the package and contents thoroughly to incorporate any sediment, then transfer the entire contents of the container to a large glass or porcelain dish and mix thoroughly, continuing the stirring for at least 1 minute. Transfer the well mixed sample to a glass-stoppered container and shake or stir thoroughly each time before removing portions for analysis.

5

TOTAL SOLIDS.—TENTATIVE.

Weigh 10 grams of the sample into a flat-bottomed, platinum dish having a diameter of about 6 cm., spread the sample in a thin layer, evaporate to dryness on a steam bath and dry in a water oven for 4 hours.

36

INSOLUBLE SOLIDS.—TENTATIVE.

Wash 20 grams of the sample repeatedly with hot water, centrifugalizing after each addition of water and pouring the clear, supernatant liquid through a tared triple filter paper on a Büchner funnel. After 4-5 washings transfer the remaining insoluble matter to the filter, dry for 2 hours at 100°C., cool in a desiccator and weigh rapidly. The paper used should be dried for 2 hours at 100°C., cooled in a desiccator and weighed. A cylinder, 1-1½ inches in diameter and 5-6 inches long, is convenient for washing and centrifugalizing.

37

SOLUBLE SOLIDS.—TENTATIVE.

Subtract the percentage of insoluble solids from the percentage of total solids to obtain the percentage of soluble solids.

38

SAND.—TENTATIVE.

Weigh 100 grams of the well mixed sample into a 2-3 liter beaker, nearly fill the beaker with water, and mix the contents thoroughly. Allow to stand 5 minutes and decant the supernatant liquid into a second beaker. Refill the first with water and again mix the contents. After 5 minutes more decant the second beaker into a third, the first into the second, refill and again mix the first. Continue this operation, decanting from the third beaker into the sink until the lighter material is washed out from the ketchup. Then collect the sand from the 3 beakers on a tared Gooch crucible, dry, ignite and weigh. Attention is especially called to the fact that under "Sand" only the figure obtained by this method should be reported. The results obtained by the determination of ash insoluble in hydrochloric acid are not applicable to the determination of sand, since the sand is so unevenly distributed that reliable results can only be obtained by taking a larger sample than is possible in the determination of ash.

39

ASH.—OFFICIAL.

Evaporate 10 grams of the sample to dryness on a water bath and ignite as directed under VIII, 4.

40

ALKALINITY OF THE ASH.—TENTATIVE.

Proceed as directed under XIII, 7. Express the result as the number of cc. of N/10 acid required to neutralize the ash from 100 grams of the sample.

41

SODIUM CHLORID.—OFFICIAL.

Proceed as directed under III, 15, using either an aliquot of the solution obtained in 40 or a nitric acid solution of the whole ash.

42

REDUCING SUGARS BEFORE INVERSION.—TENTATIVE.

Weigh 20 grams of the sample into a 200 cc. flask, dilute with about 100 cc. of water, clarify with a slight excess of neutral lead acetate solution, dilute to the mark and filter. Remove the excess of lead with anhydrous sodium or potassium oxalate. Filter and determine reducing sugars as directed under VIII, 25. Express the result as per cent of invert sugar.

43 **REDUCING SUGARS AFTER INVERSION.—TENTATIVE.**

Transfer 50 cc. of the filtrate, obtained in **42**, to a 100 cc. flask, add 5 cc. of concentrated hydrochloric acid and let stand overnight. Nearly neutralize with sodium hydroxid solution, cool, dilute to the mark and determine reducing sugars in an aliquot as directed under **VIII, 25**. Express the result as per cent of invert sugar.

44 **SUCROSE.—TENTATIVE.**

Proceed as directed under **VIII, 18**.

45 **TOTAL ACIDS.—TENTATIVE.**

Proceed as directed under **XIX, 17**, employing 5 grams of the sample. Express the result as anhydrous citric acid. One cc. of N/10 alkali is equivalent to 0.0064 gram of anhydrous citric acid.

46 **VOLATILE ACIDS.—TENTATIVE.**

Proceed as directed under **XVI, 27**, employing 25 grams of the sample, increasing the amount of water used for the distillation and collecting a correspondingly larger amount of distillate. Express the result as acetic acid. One cc. of N/10 alkali is equivalent to 0.0060 gram of acetic acid. Reserve the neutralized distillate for the detection of butyric acid.

BUTYRIC ACID.**47** *Qualitative Test.—Tentative.*

Evaporate the neutralized distillate, obtained in **46**, to dryness on a steam bath. Decompose the residue with about 5 cc. of 10% sulphuric acid and note the odor.

48 **FIXED ACIDS.—TENTATIVE.**

Multiply the percentage of volatile acids, **46**, by 1.067 and subtract the product from the percentage of total acids, **45**, to obtain the per cent of fixed acids as citric acid.

MICRO-ANALYSIS OF TOMATO PULP, KETCHUP, PUREE AND SAUCE (PASTE).**49** **APPARATUS.**

(a) *Compound microscope*.—Giving magnifications of approximately 90, 180 and 500 diameters. These diameters can be obtained by a microscope equipped with 16 and 8 mm. apochromatic objectives and a X6 and a X18 compensating ocular (a X12 ocular may also be used if desired).

(b) *Thoma-Zeiss blood counting cell*.

(c) *Special Thoma-Zeiss cell*.—With the central disk of 19 mm. diameter for making the mold count⁶.

50 **MOLDS.—TENTATIVE.**

Clean the special Thoma-Zeiss cell so that Newton's rings are produced between the slide and the cover-glass. Remove the cover and place, by means of a knife blade or scalpel, a small drop of the sample upon the central disk; spread the drop evenly over the disk and cover with the cover-glass so as to give an even spread to the material.

It is of the utmost importance that the drop be mixed thoroughly and spread evenly, otherwise the insoluble matter, and consequently the molds, are most abundant at the center of the drop. Squeezing out of the more liquid portions around the margin must be avoided. In a satisfactory mount Newton's rings should be apparent when finally mounted and none of the liquid should be drawn across the mount and under the cover-glass.

Place the slide under the microscope and examine with about 90 diameters and with such adjustment that each field of view represents approximately 1.5 sq. mm. of area on the mount.

Observe each field as to the presence or absence of mold filaments and note the result as positive or negative. Examine at least 50 fields, prepared from 2 or more mounts. No field should be considered positive unless the aggregate length of the filaments present exceeds approximately one sixth the diameter of the field. Calculate the proportion of positive fields from the results of the examination of all the observed fields and report as percentage of fields containing mold filaments.

51

YEASTS AND SPORES.—TENTATIVE.

Fill a graduated cylinder with water to the 20 cc. mark, and then add the sample till the level of the mixture reaches the 30 cc. mark. Close the graduate, or pour the contents into an Erlenmeyer flask, and shake the mixture vigorously 15-20 seconds. To facilitate thorough mixing the mixture should not fill more than three fourths of the container in which the shaking is performed. For tomato sauce or pastes, or products running very high in the number of organisms, or of heavy consistency, 80 cc. of water should be used with 10 cc. or 10 grams of the sample. In the case of exceptionally thick or dry pastes, it may be necessary to make an even greater dilution.

Pour the mixture into a beaker. Thoroughly clean the Thoma-Zeiss counting cell so as to give good Newton's rings. Stir thoroughly the contents of the beaker with a scalpel or knife blade, and then, after allowing to stand 3-5 seconds, remove a small drop and place upon the central disk of the Thoma-Zeiss counting cell and cover immediately with the cover-glass, observing the same precautions in mounting the sample as given under 50. Allow the slide to stand not less than 10 minutes before beginning to make the count. Make the count with a magnification of about 180 (8 mm. apochromatic objective with the X6 compensating ocular).

Count the number of yeasts and spores on one half of the ruled squares on the disk (this amounts to counting the number in 8 of the blocks, each of which contains 25 of the small ruled squares). The total number thus obtained equals the number of organisms in 1/60 cmm. if a dilution of 1 part of the sample with 2 parts of water is used. If a dilution of 1 part of the sample with 8 parts of water is used, the number must be multiplied by 3. In making the counts, the analyst should avoid counting an organism twice when it rests on a boundary line between 2 adjacent squares.

52

BACTERIA.—TENTATIVE.

Estimate the bacteria from the mounted sample, used in 51, but allow the sample to stand not less than 15 minutes after mounting before counting. Use a magnification of about 500 (8 mm. apochromatic objective and X18 compensating ocular). Because of the somewhat clearer definition obtained with the X12 compensating ocular, some prefer it to the X18, though the magnification is correspondingly less, being about 375. Count and record the number of bacteria in a small area consisting of 5 of the small sized squares. Move the slide to another portion of the field

and count the number on another similar area. Count 5 such areas, preferably 1 from near each corner of the ruled portion of the slide and 1 from near the center. Determine the average number of bacteria per area and multiply by 2,400,000 which gives the number of bacteria per cc. If a dilution of 1 part of the sample with 8 parts of water instead of 1 part of the sample with 2 parts of water is used in making up the sample, then the total count obtained as above must be multiplied by 7,200,000. Omit the micrococci type of bacteria in making the count.

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- ¹ Z. anal. Chem., 1892, **31**: 525; Conn. Agr. Exp. Sta. Rept., 1898, (II), p. 190.
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- ³ Conn. Agr. Exp. Sta. Rept., 1898, (II), p. 187.
- ⁴ Winton. Microscopy of Vegetable Foods. 2nd ed., 1916; Vogl. Die wichtigsten vegetabilischen Nahrungs- und Genussmittel. 1899; Tschirch und Oesterle. Anatomischer Atlas der Pharmakognosie und Nahrungsmittelkunde. 1900; Greenish and Collin. Anatomical Atlas of Vegetable Powders. 1904; Greenish. Microscopical Examination of Foods and Drugs. 2nd ed., 1910; Koch. Die Mikroskopische Analyse der Drogenpulver. 1900-08.
- ⁵ U. S. Bur. Chem. Circ. 68, p. 4.

XXV. CACAO PRODUCTS.

1 PREPARATION OF SAMPLE.—TENTATIVE.

Mix powdered products thoroughly and preserve in tightly stoppered bottles. Chill sweet or bitter chocolate until it becomes hard and reduce to a finely granular condition by grating or shaving. Mix thoroughly and preserve in a tightly stoppered bottle in a cool place.

2 MOISTURE.—OFFICIAL.

Proceed as directed under IX, 2.

3 ASH.—OFFICIAL.

Proceed as directed under VIII, 4, employing sufficient sample to contain approximately 1 gram of water-, sugar- and fat-free material.

4 ASH INSOLUBLE IN ACID.—TENTATIVE.

Proceed as directed under XXIV, 5.

5 SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.

Proceed as directed under IX, 17, employing sufficient sample to contain approximately 1 gram of water-, sugar- and fat-free material.

6 ALKALINITY OF THE SOLUBLE ASH.—TENTATIVE.

Proceed as directed under IX, 18.

7 ALKALINITY OF THE INSOLUBLE ASH.—TENTATIVE.

Proceed as directed under IX, 19.

8 TOTAL NITROGEN.—OFFICIAL.

Determine total nitrogen as directed under I, 18, 21 or 23.

9 CRUDE FIBER.—TENTATIVE.

Proceed as directed under VIII, 68, employing sufficient sample to contain approximately 1 gram of water-, sugar- and fat-free material, except that both filtrations should be made upon paper, the washed fiber either being weighed upon a tared filter in the usual way or rinsed from the paper into a tared Gooch, dried and weighed.

The residue after fat extraction may be used directly for the crude fiber determination in the analysis of commercial cocoa and other finely ground or pulverized cacao products. If, however, the material is at all granular, it should be reduced to an impalpable powder; otherwise the results will be much too high. The pulverization may be satisfactorily performed by grinding with ether, as described under 10, treating the extracted residue with the hot 1.25% sulphuric acid and proceeding from this point as directed above.

STARCH.

10

Direct Acid Hydrolysis.—Tentative.

Weigh 4 grams of the sample, if unsweetened, or 10 grams if sweetened, into a small porcelain mortar, add 25 cc. of ether and grind. After the coarser material has settled, decant the ether, together with the fine suspended matter, onto an 11 cm. paper of sufficiently fine texture to retain the crude starch. Repeat this treatment until no more coarse material remains. After the ether has evaporated from the filter, transfer the fat-free residue to the mortar by means of a jet of cold water and rub to an even paste, filtering on the paper previously employed. Repeat this process until all the sugar is removed. In the case of sweetened products the filtrate should measure at least 500 cc. Determine crude starch in the extracted residue as directed under **VIII, 60**.

11

Diastase Method.—Tentative.

Remove fat and sugar from 4 grams of the sample, if unsweetened, or 10 grams if sweetened, as directed under **10**. Wash carefully the wet residue into a beaker with 100 cc. of water, heat to boiling over asbestos with constant stirring and continue the boiling and stirring for 30 minutes. Replace the water lost by evaporation and immerse the beaker in a water bath kept at 55°–60°C. When the liquid has cooled to the temperature of the bath, add 20 cc. of freshly prepared malt extract [**VIII, 61**] and digest the mixture for 2 hours with occasional stirring. Boil a second time for 30 minutes, dilute, cool and digest as before with another 20 cc. portion of the malt extract. Heat again to boiling, cool and transfer to a 250 cc. flask. Add 3 cc. of alumina cream, make up to the mark and filter through a dry paper. The residue on the paper should show no signs of starch when examined microscopically. Continue from this point as directed under **VIII, 62**, beginning with the words "Place 200 cc. of the filtrate in a flask with 20 cc. of hydrochloric acid".

12

FAT.—TENTATIVE.

Dry 2 grams of the material over sulphuric acid until all the moisture is practically removed. (Products rich in fat show a tendency to cake at the temperature of boiling water. Hence, drying by means of heat must be avoided.) Extract with anhydrous ether in a continuous extractor until no more fat is removed. Grind and repeat the extraction. Introduce the ether extract into a tared dish, allow the ether to evaporate and dry the residue to constant weight at 100°C.

The rapid centrifugal method¹, though useful and accurate under ordinary conditions, is unreliable during the summer months or in warm latitudes and has not been approved.

13

FAT CONSTANTS.—TENTATIVE.

Separate the fat in a manner similar to that described under **15** and determine the melting point, index of refraction, iodine absorption, saponification, Reichert-Meissl and Polenske numbers as directed under **XXIII**. Melting point determinations upon this material do not become normal until the fat has been kept for at least 24 hours in a cool place.

14

MILK FAT IN MILK CHOCOLATE.—TENTATIVE.

Estimate the amount of milk fat in milk chocolate from the following formula based on a Reichert-Meissl number of 0.5 for cocoa butter:

$$C = \frac{24A + 0.5B}{5} \text{ in which}$$

A = grams of butter fat in 5 grams of mixed fat;

B = 5 - A = grams of cocoa fat in 5 grams of mixed fat;

C = Reichert-Meissl number of extracted fat.

From which the

$$\text{Weight of butter fat in 5 grams of mixed fat} = \frac{C - 0.5}{4.7} \text{ and the}$$

$$\text{Per cent of butter fat} = \text{per cent of total fat} \times \frac{C - 0.5}{23.5}$$

15

SUCROSE AND LACTOSE.—TENTATIVE.

Prepare the sample by chilling well and shaving as finely as possible with a knife. Transfer 26 grams of this material to an 8 ounce nursing bottle, add about 100 cc. of petroleum ether and shake for 5 minutes. Centrifugalize until the solvent is clear. Draw off the same by suction and repeat the treatment with petroleum ether. Place the bottle containing the de-fatted residue in a warm place until the residual traces of petroleum ether are practically expelled. Add 100 cc. of water, shake until all the chocolate is loosened from the sides and bottom of the bottle and then shake for 3 minutes longer. Add basic lead acetate solution from a burette to complete precipitation, then sufficient water to make the total volume of liquid 110 cc. Mix thoroughly and filter through a folded filter. Make the direct polariscopic reading "a" in a 200 mm. tube. Precipitate the excess of lead by anhydrous potassium oxalate and invert the solution as directed under VIII, 14. Obtain the reading of the inverted solution. Multiply the invert reading by 2 to correct for dilution "b". From the figures obtained calculate the percentages of sucrose (S) and lactose (L) by the formulas

$$S = \frac{(a-b) (110 + x)}{142.66 - \frac{t}{2}}$$

$$L = \frac{a \left(1 + \frac{x}{100} \right)}{0.79} - S \text{ in which the value of } x \text{ is obtained from}$$

$$x = \frac{0.2244 (a - 21d)}{1 - 0.00204 (a - 21d)} \text{ in which the value of } d \text{ is obtained from}$$

$$d = \frac{a - b}{142.66 - \frac{t}{2}}$$

16

CASEIN IN MILK CHOCOLATE.—TENTATIVE.

It is unnecessary to de-fat the chocolate. Weigh 10 grams of the chocolate into a 500 cc. Erlenmeyer flask and add 250 cc. of 1% sodium oxalate solution. Heat to boiling and boil gently for a few minutes, then cool, add 5 grams of magnesium carbonate and filter. Determine nitrogen in 50 cc. of this filtrate. Pipette 100 cc. of the filtrate into a 200 cc. volumetric flask and dilute almost to the mark with water. Then precipitate the casein by the addition of 2 cc. of glacial acetic acid or 1 cc. of concentrated sulphuric acid. Make to volume, shake, filter and determine nitrogen in 100 cc. of the filtrate. The difference between the 2 nitrogen determinations gives the nitrogen derived from the casein which, multiplied by 6.38, gives the amount of casein present in 2 grams of the sample.

17

COLORING MATTERS.—TENTATIVE.

Proceed as directed under **XI**.

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¹ U. S. Bur. Chem. Bull. 137, p. 103.

XXVI. COFFEES.

GREEN COFFEE.

1 MACROSCOPIC EXAMINATION.—TENTATIVE.

A macroscopic examination is usually sufficient to show the presence of excessive amounts of black and blighted coffee beans, coffee hulls, stones and other foreign matter. These can be separated by hand picking and determined gravimetrically.

2 COLORING MATTERS.—TENTATIVE.

Shake vigorously 100 grams or more of the sample with cold water or 70% alcohol by volume. Strain through a coarse sieve and allow to settle. Identify soluble colors in the solution and insoluble pigments in the sediment as directed under **XI**.

ROASTED COFFEE.

3 MACROSCOPIC EXAMINATION.—TENTATIVE.

Artificial coffee beans are apparent from their exact regularity of form. Roasted legumes and lumps of chicory, when present in whole roasted coffee, can be picked out and identified microscopically. In the case of ground coffee sprinkle some of the sample on cold water and stir lightly. Fragments of pure coffee, if not over-roasted, will float, while fragments of chicory, legumes, cereals, etc., will sink immediately, chicory coloring the water a decided brown. In all cases identify the particles that sink, by microscopical examination.

4 PREPARATION OF SAMPLE.—TENTATIVE.

Grind the sample and pass through a sieve having holes 0.5 mm. in diameter and preserve in a tightly stoppered bottle.

5 MOISTURE.—TENTATIVE.

Dry 5 grams of the sample at 105°–110°C. for 5 hours and subsequent periods of an hour each until constant weight is obtained. The same procedure may be used, drying in vacuo at the temperature of boiling water. In the case of whole coffee, grind rapidly to a coarse powder and weigh at once portions for the determination without sifting and without unnecessary exposure to the air.

SOLUBLE SOLIDS.

6 *Winton Method.—Tentative.*

Place 4 grams of the sample in a 200 cc. flask, add water to the mark and allow the mass to infuse 8 hours, with occasional shaking; let stand 16 hours long r without shaking, filter, evaporate 50 cc. of the filtrate to dryness in a flat-bottomed dish, dry at 100°C. and weigh.

7 ASH.—OFFICIAL.

Proceed as directed under **VIII, 4**.

8 ASH INSOLUBLE IN ACID.—TENTATIVE.

Proceed as directed under **XXIV, 5**.

9 SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.

Proceed as directed under **IX, 17**.

10 ALKALINITY OF THE SOLUBLE ASH.—TENTATIVE.

Proceed as directed under **IX, 18**.

11 SOLUBLE PHOSPHORIC ACID IN THE ASH.—TENTATIVE.

Acidify the solution of soluble ash, obtained in **9**, with dilute nitric acid and determine phosphoric acid (P_2O_5) as directed under **I, 6** or **9**.

12 INSOLUBLE PHOSPHORIC ACID IN THE ASH.—TENTATIVE.

Determine phosphoric acid (P_2O_5) in the insoluble ash as directed under **I, 6** or **9**.

13 CHLORIN.—OFFICIAL.

Proceed as directed under **III, 18**.

CAFFEIN.

14 *Gortler Method¹.—Tentative.*

Moisten 11 grams of finely powdered coffee with 3 cc. of water, allow to stand 30 minutes and extract with chloroform for 3 hours in a Soxhlet extractor. Evaporate the extract, treat the residue of fat and caffeine with hot water, filter through a cotton plug and moistened filter paper and wash with hot water. Make up the filtrate and washings to 55 cc., pipette off 50 cc. and extract 4 times with chloroform. Evaporate the chloroform extract in a tared flask, dry the caffeine at 100°C. and weigh. Transfer the residue to a Kjeldahl flask with a small amount of hot water and determine nitrogen as directed in **I, 18, 21** or **23**. To obtain the weight of caffeine multiply the result by 3.464.

15 *Modified Stahlschmidt Method².—Tentative.*

Weigh 3.125 grams of the finely powdered sample into a 500 cc. flask, add 225 cc. of water (this volume will shrink to about 200 cc. by boiling), attach a reflux condenser and boil for 2 hours. Add 2 grams of dry basic lead acetate [**VIII, 13 (C)**] and boil 10 minutes more. Cool, transfer to a 250 cc. graduated flask, fill to the mark, filter through a dry filter, measure 200 cc. of the filtrate into a 250 cc. graduated flask and pass hydrogen sulphid through it to remove the excess of lead. Make the solution up to the mark and filter through a dry filter. Measure 200 cc. of this filtrate into an evaporating dish and concentrate to about 40 cc. Wash the concentrated solution with as little water as possible into a small separatory funnel and shake out 4 times with chloroform, using 25, 20, 15 and 10 cc., respectively. If any emulsion forms, break it up with a stirring rod and run the separated portions of chloroform through a 5 cm. filter paper into a small, tared Erlenmeyer flask. Evaporate off the chloroform on the steam bath, or recover the chloroform by attaching the flask to a condenser and distilling to a small volume. Dry the fine, white crystals of caffeine to constant weight at 75°C. Test the purity of this residue by determining nitrogen as directed in **I, 18, 21** or **23** and multiplying by the factor 3.464.

16

CRUDE FIBER.—TENTATIVE.

Proceed as directed under **VIII, 68.**

17

STARCH.—TENTATIVE.

Extract 5 grams of the finely pulverized sample on a hardened filter with 5 successive portions (10 cc. each) of ether; wash with small portions of 95% alcohol by volume until a total of 200 cc. have passed through, place the residue in a beaker and proceed as directed under **VIII, 62.**

18

SUGARS.—TENTATIVE.

Proceed as directed under **VIII, 58 and 59.**

19

PETROLEUM ETHER EXTRACT.—TENTATIVE.

Dry 2 grams of the coffee at 100°C., extract with petroleum ether (b. p. 35°–50°C.) for 16 hours, evaporate the solvent, dry the residue at 100°C. and weigh.

20

TOTAL ACIDITY.—TENTATIVE.

Treat 10 grams of the sample, prepared as directed under **4**, with 75 cc. of 80% alcohol by volume in an Erlenmeyer flask, stopper and allow to stand 16 hours, shaking occasionally. Filter and transfer an aliquot of the filtrate (25 cc. in the case of green coffee, 10 cc. in the case of roasted coffee) to a beaker, dilute to about 100 cc. with water and titrate with N/10 alkali, using phenolphthalein as an indicator. Express the result as the number of cc. of N/10 alkali required to neutralize the acidity of 100 grams of the sample.

21

VOLATILE ACIDITY.—TENTATIVE.

Into a volatile acid apparatus [**XVI, 27**; Fig. 8] introduce a few glass beads and over these place 20 grams of the unground sample. Add 100 cc. of recently boiled water to the sample, place a sufficient quantity of recently boiled water in the outer flask and distil until the distillate is no longer acid to litmus paper (usually 100 cc. of distillate will be collected). Titrate the distillate with N/10 alkali, using phenolphthalein as an indicator. Express the result as the number of cc. of N/10 alkali required to neutralize the acidity of 100 grams of the sample.

COATING AND GLAZING SUBSTANCES.

22

SUGAR AND DEXTRIN.—TENTATIVE.

Introduce 100 grams of the whole coffee into a beaker, add exactly 300 cc. of water, stir and allow to stand 5 minutes, with frequent stirring. Filter through a dry paper, add carefully to the filtrate sufficient dry lead acetate to precipitate all the caffetannic acid, avoiding an excess. Filter through dry paper and remove the lead from the filtrate by the addition of a slight excess of anhydrous potassium oxalate. Filter through a dry paper and determine reducing sugars as invert sugar in 50 cc. of the filtrate, as directed in **VIII, 25.** Invert a 75 cc. aliquot of the filtrate as directed under **VIII, 14.** Cool, nearly neutralize with sodium hydroxid solution, make up to 100 cc. and determine reducing sugars as invert sugar in the resulting solution, as directed under **VIII, 25.** Measure a 100 cc. aliquot of the filtrate into a 200 cc. flask, add 10 cc. of 25% hydrochloric acid and hydrolyze as directed under **VIII, 60.** Cool, neutralize with sodium hydroxid solution, make up to volume, filter through a dry paper and determine reducing sugars as invert sugar in 50 cc. of the filtrate as directed under **VIII, 25.** Calculate the

reducing sugars in each instance to per cent by weight of the original coffee. Calculate sucrose from the reducing sugars before and after inversion as directed in VIII 18, and calculate dextrin as follows: Subtract the reducing sugars after inversion from the reducing sugars after hydrolysis, multiply the difference by the factor 0.9561 to convert the result to dextrose and then by 0.9 to convert to dextrin.

In some instances the presence of sucrose in the water extract may be verified by polarization. The presence of dextrin in the water extract may be verified by polarization as directed under IX 25, and by the erythro-dextrin test [IX, 47] performed on the water extract previous to clarification with lead acetate.

23

EGG ALBUMEN AND GELATIN.—TENTATIVE.

Treat 100 grams of the whole coffee with 500 cc. of water and allow to stand with frequent stirring for 5 minutes. Filter and treat separate portions of the filtrate with (1) a strong solution of tannic acid; (2) Millon's reagent [XV, 9]; (3) by boiling. In the presence of egg albumen a more or less heavy precipitate will be formed in each case. As a confirmatory test, treat an aliquot of the filtrate with an excess of tannic acid solution, add a little salt if necessary to secure flocculation of the precipitate, filter and, without washing, introduce the paper and its contents into a Kjeldahl flask and determine nitrogen. By this method coffee not coated with albumen or gelatin will yield less than 10 mg. of nitrogen per 100 grams of sample.

24

CHICORY INFUSION.—TENTATIVE.

Cover 100-150 grams of the whole coffee with water, allow to soak 2-3 minutes, stirring frequently, and drain the aqueous washings through a coarse sieve. Wash the coffee upon the sieve with about 100 cc. of water and centrifugalize the combined washings. Decant the clear liquid from the sediment, drain almost dry upon filter paper, then mount the sediment in chloral hydrate [XXIV, 20 f.] and examine under the microscope for elements of chicory.

FATS AND WAXES.

25

Späth Method.—Tentative.

Treat 100-200 grams of the beans with low boiling petroleum ether for 10 minutes, pour off the petroleum ether and repeat the process. Filter the combined petroleum ether extracts, evaporate and determine the index of refraction and the saponification number of the residue, as directed under XXIII, 6 and 20.

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¹ Ann., 1908, 358: 327.

² Allen. Commercial Organic Analysis. 4th ed., 1909-14, 6: 607.

³ Forschb. über Lebensm., 1895, 2: 223.

XXVII. TEA.

1 DUST, STEMS AND FOREIGN LEAVES.—TENTATIVE.

Place 1 gram of the tea in a 300 cc. casserole, add 200 cc. of boiling water and allow to stand 15 minutes. This treatment will cause the leaves to unroll, and a macroscopic examination will reveal the presence or absence of dust or stems, while the leaves will be in condition for examination as to their form and structure¹.

2 PREPARATION OF SAMPLE.—TENTATIVE.

Grind the sample and pass it through a sieve having circular openings 0.5 mm. in diameter.

3 MOISTURE.—TENTATIVE.

Proceed as directed under IX, 2.

4 WATER EXTRACT².—TENTATIVE.

To 2 grams of the original sample in a 500 cc. Erlenmeyer flask add 200 cc. of hot water and boil over a low flame for an hour. The flask should be closed with a rubber stopper through which passes a glass tube 18 inches long for a condenser. The loss from evaporation should be replaced from time to time by the addition of hot water. Filter through a tared filter and wash the residue until the filtrate measures 300 cc., stirring the contents of the filter throughout the process to facilitate the filtering. Dry the filter paper and residue in the funnel in the steam oven until the excess of water is removed, transfer paper and contents to a tared weighing bottle and dry to constant weight at 100°C.

5 ASH.—OFFICIAL.

Proceed as directed under VIII, 4.

6 SOLUBLE AND INSOLUBLE ASH.—TENTATIVE.

Proceed as directed under IX, 17.

7 ASH INSOLUBLE IN ACID.—TENTATIVE.

Proceed as directed under XXIV, 5.

8 ALKALINITY OF THE ASH.—TENTATIVE.

Determine the alkalinity of the soluble and insoluble ash as directed under IX, 18 and 19.

9 PHOSPHORIC ACID IN THE ASH.—TENTATIVE.

Determine phosphoric acid (P_2O_5) in the soluble and insoluble ash as directed under XXVI, 11 and 12.

10 PETROLEUM ETHER EXTRACT.—TENTATIVE.

Proceed as directed under XXVI, 19.

11

PROTEIN.—TENTATIVE.

Determine nitrogen as directed under **I, 18, 21** or **23**. Subtract the percentage of nitrogen present as caffeine from the percentage of total nitrogen to obtain the percentage of nitrogen present as protein. Multiply this result by 6.25 to obtain the percentage of protein.

12

CRUDE FIBER.—TENTATIVE.

Proceed as directed under **VIII, 68**.

13

VOLATILE OIL.—TENTATIVE.

Add 100 grams of tea to 800 cc. of water, distil, extract the distillate several times with petroleum ether, transfer the combined petroleum ether extracts to a tared dish, evaporate at room temperature, dry in a desiccator and weigh.

CAFFEIN.

14

Modified Stahl Schmidt Method.—Tentative.

Proceed as directed under **XXVI, 15**.

TANNIN.

Proctor Modification of the Löwenthal Method³.—Tentative.

15

REAGENTS.

(a) *Potassium permanganate solution*.—Make up a solution containing 1.33 grams per liter and obtain its equivalent in terms of N/10 oxalic acid.

(b) *N/10 oxalic acid*.

(c) *Indigo carmine solution*.—Make up a solution containing 6 grams of indigo carmine (free from indigo blue) and 50 cc. of concentrated sulphuric acid per liter.

(d) *Gelatin solution*.—Soak 25 grams of gelatin for an hour in saturated sodium chlorid solution, heat until the gelatin is dissolved and make up to 1 liter after cooling.

(e) *Acid sodium chlorid solution*.—Acidify 975 cc. of saturated sodium chlorid solution with 25 cc. of concentrated sulphuric acid.

(f) *Powdered kaolin*.

16

DETERMINATION.

Boil 5 grams of the tea for 30 minutes with 400 cc. of water; cool, transfer to a 500 cc. graduated flask and make up to the mark. To 10 cc. of the infusion, filtered if not clear, add 25 cc. of the indigo carmine solution and about 750 cc. of water. Add from a burette the potassium permanganate solution, a little at a time while stirring, until the color becomes light green, then drop by drop, until the color changes to bright yellow or to a faint pink at the rim. Designate the number of cc. of permanganate used as "a".

Mix 100 cc. of the clear infusion of tea with 50 cc. of the gelatin solution, 100 cc. of the acid sodium chlorid solution and 10 grams of the powdered kaolin, and shake several minutes in a stoppered flask. After settling decant through a filter. Mix 25 cc. of the filtrate with 25 cc. of the indigo carmine solution and about 750 cc. of water and titrate with permanganate as before. The number of cc. of permanganate used subtracted from that obtained above, "a", gives the amount of permanganate required to oxidize the tannin. One cc. of N/10 oxalic acid is equivalent approximately to 0.004157 gram of tannin (gallotannic acid).

FACING.

17

GENERAL.—TENTATIVE.

Mineral pigments may be detected in the ash, or the tea may be shaken up with a large volume of water, and the water separated from the leaves by a sieve, when the insoluble mineral substances used in facing will settle and can be removed by filtration for further examination, as directed under **XI, 1**, the catechu and other soluble substances being in the filtrate.

18

PARAFFIN AND WAXY SUBSTANCES.—TENTATIVE.

Spread a quantity of the tea between 2 sheets of unglazed, white paper and place thereon a hot iron. Any greasy substance will stain the paper⁴.

PIGMENTS USED FOR COLORING OR FACING.

19

Read Method⁵.—Tentative.

Place 60 grams of the tea in a 60 mesh, 5-6 inch sieve, provided with a top. Sift a small quantity (approximately 0.1 gram) of the dust upon a piece of semi-glazed, white paper about 8 by 10 inches. To obtain the requisite amount of dust, it is sometimes necessary to rub the leaf gently against the bottom of the sieve, but this must not be done until the sieve has been well shaken over the paper. Place the paper on a plain, firm surface, preferably glass or marble, and crush the dust by pressing firmly upon it a flat steel spatula about 5 inches long. Repeat the crushing process until the tea dust is ground almost to a powder when particles of coloring matter, if present, become visible as streaks on the paper. Brush off the loose dust and examine the paper by means of a simple lens magnifying $7\frac{1}{2}$ diameters. In distinguishing these particles and streaks bright light is essential. In many cases the character of the pigment is indicated by the behavior of these streaks when treated with reagents and examined under a microscope. The crushed particles of natural leaf in either black or green tea appear in such quantity that there is no chance of mistaking them for coloring or facing material. This test should be repeated using black, semi-glazed paper for facings such as talc, gypsum, barium sulphate or clay.

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³ *Ibid.*, 13 (VII), p. 890.

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⁵ *Ibid.*; Proc. Eighth Intern. Cong. Appl. Chem., 1912, 18: 301.

XXVIII. BAKING POWDERS AND THEIR INGREDIENTS.

1 PREPARATION OF SAMPLE.—TENTATIVE.

Remove the entire sample from the package, mix carefully and pass through a 20–40 mesh sieve.

TOTAL CARBON DIOXID.

2 *General Method.—Tentative.*

Make the determination by the absorption method, employing any apparatus which gives accurate results when checked with pure calcite. Whatever apparatus is chosen, the tubes and materials used for absorbing and drying the carbon dioxid may be varied according to the preference of the analyst. Use 0.25–1 gram of sodium or calcium carbonate, according to the amount of absorbent employed, and in the case of baking powder 0.50–2 grams.

Method Using Knorr's Apparatus.—Tentative.

3 REAGENTS.

(a) 50% potassium hydroxid solution.

(b) Soda lime.—Finely granulated and freed from dust by sifting.

4 APPARATUS.

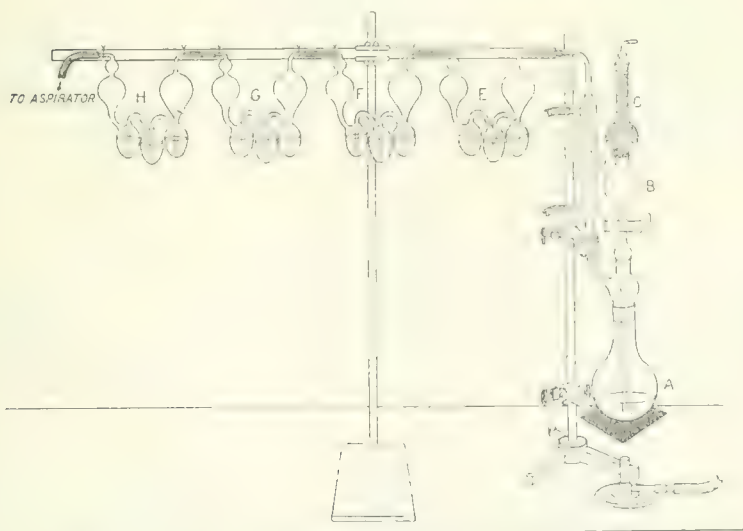


FIG. 14 KNORR'S APPARATUS FOR THE DETERMINATION OF CARBON DIOXID.

This consists of a flask (A), fitted by means of a ground-glass joint with a glass connection through the upper part of which passes a dropping funnel (B), and joined at the side with a Liebig condenser (D). The mouth of the dropping funnel

(B) is connected by means of a perforated stopper with a soda lime tube (C). The upper end of the Liebig condenser is connected by a rubber joint with a Geissler bulb (E), containing sulphuric acid for drying the gas passing into the next Geissler bulb (F), connected with (E), and containing strong potassium hydroxid solution (1 to 2). The bulb (F) is connected with a third Geissler bulb (G), containing sulphuric acid for the absorption of moisture escaping from F. A fourth Geissler bulb (H) is attached to G as a precaution to prevent moisture from the air being absorbed by G. H is connected with an aspirator. Many analysts prefer to replace the bulb (F) by 2 U-tubes filled with sifted soda lime.

5

DETERMINATION.

Place 0.5–2 grams of the baking powder, the amount depending upon the percentage of carbon dioxid present, in the flask (A), which must be perfectly dry. Close the flask with the stopper which carries the funnel tube, and the tube connecting with the absorption apparatus. Weigh separately the Geissler bulbs (F) and (G) and attach them to the apparatus. If 2 soda lime tubes are employed, weigh them separately and fill the first anew when the second increases materially in weight. Nearly fill the funnel tube (B) with hydrochloric acid (sp. gr. 1.1) and place the soda lime tube (C) in position. Then aspirate air through the Geissler bulbs at a rate of about 2 bubbles per second. Open the stopper of the funnel and allow the acid to run slowly into the flask, care being taken that the evolution of gas be so gradual as not to materially increase the current through the Geissler bulbs. After all the acid has been introduced, close the stop-cock in B, continue the aspiration and heat gradually the contents of the flask to boiling. While the flask is being heated the aspirator tube may be removed, although many analysts prefer, when using ground-glass joints, to aspirate during the entire operation. Continue the boiling for a few minutes after the water has begun to condense in D, then remove the flame, open the stop-cock in tube (B) and allow the apparatus to cool with continued aspiration. Remove the absorption bulbs (F) and (G) and weigh. The increase in weight is due to carbon dioxid.

Method Using Heidenhain's Apparatus.—Tentative.

6

REAGENTS.

(a) *Calcium chlorid*.—Use calcium chlorid dehydrated at 200°C., but not fused. Grind it coarsely in a coffee mill and sift through No. 18 wire gauze to remove the extremely coarse, and through No. 30 wire gauze to remove the very fine, particles.

(b) *Soda lime*.—Grind and sift the soda lime¹ for the weighed tubes as described above. It should not be too dry, as it must not absorb moisture to a greater degree than the calcium chlorid.

7

APPARATUS².

This consists of a cylinder (A), filled with soda lime to remove carbon dioxid from the air passing through the apparatus. A thick layer of cotton at the upper end prevents soda lime dust from being carried over. Connect the cylinder (A) by means of a perforated rubber stopper and a bent glass tube having a stop-cock (B) and a capillary constriction (C) with a short piece of rubber tubing to which is attached a short piece of glass tubing (E), fitted with a perforated rubber stopper. The latter fits tightly into the constriction of the funnel tube (D). The funnel of the latter is cylindrical in shape, $\frac{3}{4}$ inch in diameter at the upper end, $\frac{3}{8}$ inch at the lower end and 4 inches long, the rubber stopper of E fitting into the constriction.

The stem of the funnel tube (*D*) passes through a doubly perforated rubber stopper almost to the bottom of the evolution flask (*F*), which is ordinarily of 150 cc. capacity but, in the case of foaming liquids, may hold 300 cc. Through the second perforation in the stopper connect the evolution flask (*F*) with a reflux condenser (*G*), consisting of a $\frac{1}{4}$ inch glass tube around which is wound a small lead pipe carrying a current of cold water. To the upper end of the condenser attach a U-tube containing a little calcium chlorid (to be renewed when it has liquefied) to retain the bulk of the moisture. Connect this U-tube with a second U-tube (*H*), filled with coarse calcium chlorid, and this in turn with a third U-tube (*K*), filled at *I* with a 3 inch column of pumice stone impregnated with copper sulphate and completely dehydrated at 150°C., the remainder of the tube being filled with fine calcium chlorid. Connect the U-tube (*K*) with a bent glass tube having a stop-cock (*L*) which is closed when the apparatus is not in use. Next attach the absorption U-tubes (*M*) and (*N*) which are $\frac{1}{2}$ inch in diameter and 5 inches long, the first filled mainly with soda lime but containing a little calcium chlorid at the end where the air current

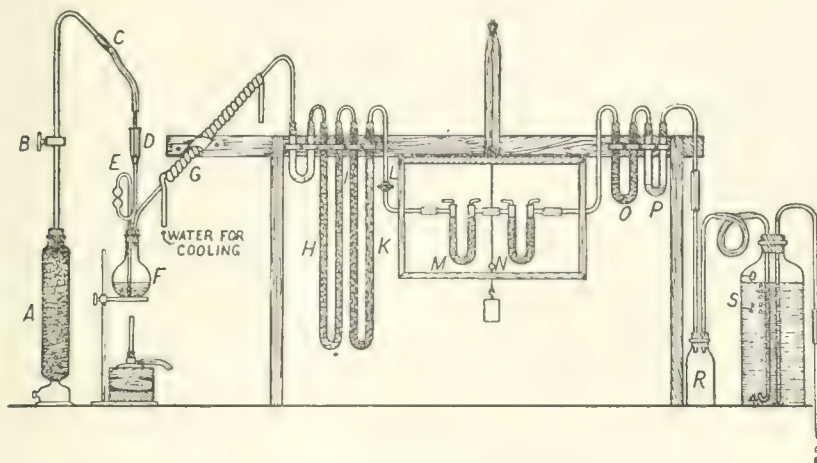


FIG. 15. HEIDENHAIN'S APPARATUS FOR THE DETERMINATION OF CARBON DIOXID.

enters, the second filled one half with soda lime and one half with calcium chlorid, the latter being placed at the side where the air current leaves. Connect *N* with a guard tube (*O*), filled with calcium chlorid on the side toward *N* and with soda lime on the side toward *P*, the latter being a small U-tube trapped with glycerol to indicate the passage of the air current. Connect *P* with a safety bottle (*R*), to receive any water which may be sucked back from the aspirator, and connect *R* with the aspirator (*S*), a 4 liter Mariette bottle.

The tubes (*M*) and (*N*) should hold about 20 grams, making the capacity of *M* for carbon dioxide almost 1 gram and that of *N* for moisture 0.2 gram. *M* should be refilled when its weight has increased 0.75 gram and *N* after an increase of 0.1 gram in weight.

Use the best grade of rubber for all connections, applying a trace of castor oil as a lubricant. For connections of the weighed tubes use rubber tubing boiled in weak lye, washed and dried. Apply also a little castor oil, which is thoroughly wiped off again before connecting the tubing.

Before using the apparatus fill *H* and *K* with carbon dioxide in order to saturate the alkalinity of the calcium chlorid and exhaust after several hours.

8

DETERMINATION.

In order to find the allowable rapidity of the air current employed during the determination, proceed as follows: Charge the apparatus exactly as for an analysis leaving out the carbonate. Begin to aspirate at the rate of about 50 cc. per minute. After 2 liters have been aspirated weigh the tubes, *M* and *N*. If they have lost in weight, repeat the experiment with 40 cc. per minute, and so on until the weight of the tubes remains constant. If the work has been done with due precaution, the first tube should have lost just as much as the second has gained. Do not exceed the safe speed thus found.

Weigh the tubes *M* and *N* at the air temperature of the balance room. Shortly before weighing open the tubes for a moment to allow equalization of air. Note the thermometer and barometer readings. Connect the tubes with the apparatus and test the tightness of the joints by closing *A* at the bottom, opening all the cocks, starting the aspirator, and observing *P*, in which the liquid should soon come to a standstill. Then disconnect the aspirator, close *B*, remove *F*, put in the substance, using about 1 gram of sodium carbonate or calcium carbonate or about 2 grams of baking powder, connect *F*, and start the condenser (*G*). Introduce 50 cc. of 10% hydrochloric acid through *D*, lifting *E* slightly and allowing only small quantities of the dilute acid to enter at a time. Light the burner under *F*, heat to boiling and reduce the flame to keep the liquid just at the boiling point. If no more air passes *P*, start the aspiration. When the water stops running from *S*, open *B* carefully and adjust the outflow of the aspirator by raising or lowering the syphon to one half the safe speed.

After *M* has become cool increase the current to the full safe speed and aspirate altogether 3 liters, continuing boiling to the end of the aspiration. After the tubes have assumed the temperature of the balance room, open for a moment and weigh. When extreme accuracy is desired, note again the thermometer and barometer readings and apply correction according to the following formula:

$$-(A^2 - A^1) \times T \text{ and } + (B^2 - B^1) \times B \text{ in which}$$

A^1 = the temperature at first weighing in degrees C.;

A^2 = the temperature at second weighing in degrees C.;

B^1 = the air pressure at first weighing in mm.;

B^2 = the air pressure at second weighing in mm.;

T and *B* are constants found from the following formulas:

$$T = V \times 0.0000039 \text{ gram;}$$

$$B = V \times 0.0000015 \text{ gram in which}$$

0.0000039 = change in weight of 1 cc. of air for 1°C.;

0.0000015 = change in weight of 1 cc. of air for 1 mm. pressure;

and the value of *V* is obtained from

$$V = \frac{G}{2.7} + \frac{F}{2.0} - \frac{G - F}{8.5},$$

representing the differential volume affected by temperature and pressure and being a constant for the tubes and in which

G = the weight of the empty tubes;

F = the weight of the fillings;

2.7 = the specific gravity of glass;

2.0 = the specific gravity of filling;

8.5 = the specific gravity of brass;

$$\frac{G}{2.7} + \frac{F}{2.0} = \text{volume of tubes and fillings;}$$

$$\frac{G + F}{8.5} = \text{volume of brass weights.}$$

9

RESIDUAL CARBON DIOXID³.—TENTATIVE.

Weigh 2 grams of the baking powder into a flask suitable for the subsequent determination of carbon dioxide, add 20 cc. of cold water and allow to stand 20 minutes. Place the flask in a metal drying cell surrounded by boiling water and heat, with occasional shaking, for 20 minutes.

To complete the reaction and drive off the last traces of gas from the semi-solid mass, heat quickly to boiling and boil for a minute. Aspirate until the air in the flask is thoroughly changed, and determine the residual carbon dioxide by absorption, as directed under 5 or 8.

The process described³, based on the methods of McGill⁴ and Catlin⁵, imitates as far as practicable the conditions encountered in baking but in such a manner that concordant results may be readily obtained on the same sample and comparable results on different samples.

10

AVAILABLE CARBON DIOXID.—TENTATIVE.

Subtract the residual carbon dioxide from the total.

11

ACIDITY.—TENTATIVE.

(For cream of tartar and its substitutes.)

Dissolve 1 gram of the sample in hot water and titrate with N/5 potassium hydroxid, using phenolphthalein as an indicator.

TARTARIC ACID, FREE OR COMBINED.

12

Wolff Method⁶.—Tentative.

(Applicable in the presence of phosphates.)

Shake repeatedly about 5 grams of the sample with about 250 cc. of cold water in a flask and allow the insoluble portion to subside. Decant the solution through a filter and evaporate the filtrate to dryness. Powder the residue, add a few drops of 1% resorcin solution and about 3 cc. of strong sulphuric acid and heat slowly. Tartaric acid is indicated by a rose-red color which is discharged on dilution with water.

TOTAL TARTARIC ACID.

13

Goldenberg-Geromont-Heidenhain Method.—Tentative.

(Applicable only in the absence of aluminium salts, calcium salts and phosphates.)

Into a shallow 6 inch porcelain dish weigh out 2 grams of the sample and sufficient potassium carbonate to combine with all the tartaric acid not in the form of potassium bitartrate. Mix thoroughly with 15 cc. of cold water and add 5 cc. of 99% acetic acid. Stir for 30 seconds with a glass rod bent near the end. Add 100 cc. of 95% alcohol, stir violently for 5 minutes, and allow to settle at least 30 min-

utes. Filter on a Gooch crucible with a thin layer of paper pulp and wash with 95% alcohol until 2 cc. of the filtrate do not change the color of litmus tincture diluted with water. Place the precipitate in a small casserole, dissolve in 50 cc. of hot water and add N/5 potassium hydroxid, leaving it still strongly acid. Boil for a minute. Finish the titration, using phenolphthalein as an indicator and correct the reading by adding 0.2 cc. One cc. of N/5 potassium hydroxid, under the above conditions, is equivalent to 0.02641 gram of tartaric anhydrid, 0.03001 gram of tartaric acid, or 0.03763 gram of potassium bitartrate. Standardize the N/5 potassium hydroxid by means of pure potassium bitartrate.

The accuracy of this method is indicated by the agreement of the percentages of potassium bitartrate in cream of tartar powders containing no free tartaric acid, obtained by calculation from the tartaric acid, with those obtained by calculation from the potassium oxid⁷.

FREE TARTARIC ACID.

14

Qualitative Test.—Tentative.

Extract 5 grams of the sample with absolute alcohol and evaporate the alcohol from the extract. Dissolve the residue in dilute ammonium hydroxid, transfer to a test tube, add a good sized crystal of silver nitrate and heat gently. Tartaric acid is indicated by the formation of a silver mirror. If desired, the absolute alcohol extract may be tested as directed under 12.

15

Quantitative Method.—Tentative.

Calculate the percentage of tartaric anhydrid combined with the potash as bitartrate, if any, and subtract this from the percentage of total tartaric anhydrid. The difference is the tartaric anhydrid originally added as the free acid, although, if the sample has been kept for a long time or has been improperly stored, a portion or all of this acid may exist at the time of analysis as the sodium salt resulting from the reaction in the can with the sodium bicarbonate. Multiply by 1.137 to obtain the percentage of tartaric acid.

16

POTASSIUM BITARTRATE.—TENTATIVE.

If, as is usually the case, potassium bitartrate is the only potassium salt present, multiply the percentage of total potash, determined as directed under 24, by 3.994.

STARCH.

17

Direct Inversion Method.—Tentative.

(For all baking powder ingredients free from lime.)

Weigh 5 grams of the powder into a 500 cc. graduated flask and proceed as directed under VIII, 60.

18

Indirect Method⁸.—Tentative.

(For phosphate, alum phosphate and all other baking powders containing lime.)

Mix 5 grams of the powder with 200 cc. of 3% hydrochloric acid in a 500 cc. graduated flask and allow the mixture to stand for an hour, with frequent shaking. Filter on an 11 cm. hardened filter, taking care that a clear filtrate is obtained. Rinse the flask once without attempting to remove all the starch, and wash the paper twice with cold water. Carefully wash the starch from the paper back into the flask with 200 cc. of water. Add 20 cc. of 25% hydrochloric acid and proceed as directed under VIII, 60.

The treatment with 3% hydrochloric acid, without dissolving the starch, removes effectively the lime, which otherwise would be precipitated as tartrate by the alkaline copper solution.

19

Modified McGill Method.—Tentative.

Digest 1 gram of the powder with 150 cc. of 3% hydrochloric acid for 24 hours at room temperature, with occasional shaking. Filter on a Gooch crucible, wash thoroughly with cold water and then once each, with alcohol and ether. Dry at 110°C. (4 hours is usually sufficient), cool and weigh. Burn off the starch, weigh again and determine the starch by difference.

The results by this method on cream of tartar powders and tartaric acid powders agree closely with those obtained by copper reduction. On phosphate, alum and alum-phosphate powders the results are usually satisfactory, but in some instances they may be over 2% too high.

ALUM IN THE PRESENCE OF PHOSPHATES⁹.

20

Qualitative Test.—Tentative.

(a) *In baking powder.*—Burn about 2 grams of the sample to an ash in a porcelain dish. Extract with boiling water and filter. Add to the filtrate a few drops of ammonium chlorid solution. A flocculent precipitate indicates alum.

(b) *In cream of tartar.*—Mix about 1 gram of the sample with an equal quantity of sodium carbonate, burn to an ash and proceed as in (a).

ASH¹⁰.

21

INSOLUBLE ASH AND PREPARATION OF SOLUTION.—TENTATIVE.

Char 5 grams of the sample in a platinum dish at a heat below redness. Boil the carbonaceous mass with dilute hydrochloric acid, filter into a 500 cc. graduated flask and wash with hot water. Return the residue, together with the paper, to the platinum dish and burn to a white ash. Boil again with hydrochloric acid, filter, wash, unite the 2 filtrates and dilute to 500 cc.

Incinerate the residue after the last filtration and determine the ash insoluble in acid.

22

IRON AND ALUMINIUM.—TENTATIVE.

Draw a 100 cc. aliquot of the solution, prepared as directed in 21, and separate silica, if necessary. Mix the solution with sodium phosphate solution in excess. Add ammonium hydroxid until a permanent precipitate is obtained, then hydrochloric acid, drop by drop, until the precipitate is dissolved. Heat the solution to about 50°C., mix with a considerable excess of 50% ammonium acetate solution and 4 cc. of 80% acetic acid.

As soon as the precipitate of aluminium phosphate, mixed with iron phosphate, has settled, collect on a filter, wash with hot water, ignite and weigh.

Fuse the mixed phosphates with 10 parts of sodium carbonate, dissolve in dilute sulphuric acid, reduce with zinc, and determine the iron by titration with a standard permanganate solution. In the same solution determine the phosphoric acid, as directed under I, 6 or 9. To obtain the weight of alumina (Al_2O_3) subtract the sum of the weights of ferric oxid (Fe_2O_3) and phosphorus pentoxid (P_2O_5) from the weight of the mixed phosphates.

23

CALCIUM.—TENTATIVE.

Heat the combined filtrate and washings, obtained in 22, to 50°C. and add an excess of ammonium oxalate solution. Allow to stand in a warm place until the precipitate has settled, filter, wash the precipitate with hot water, dry, ignite over a Bunsen burner and finally over a blast lamp. Cool in a desiccator and weigh as calcium oxid.

24

POTASSIUM AND SODIUM.—TENTATIVE.

Evaporate an aliquot of the solution, prepared as directed under 21, nearly to dryness to remove the excess of hydrochloric acid, dilute and heat to boiling. While still boiling add barium chlorid solution so long as a precipitate forms and then enough barium hydroxid solution to make the liquid strongly alkaline. As soon as the precipitate has settled, filter and wash with hot water, heat the filtrate to boiling, add sufficient ammonium carbonate solution (1 part of ammonium carbonate in 5 of 2% ammonium hydroxid solution) to precipitate all the barium, filter and wash with hot water. Evaporate the filtrate to dryness and ignite the residue below redness to remove ammonium salts. Add to the residue a little water and a few drops of ammonium carbonate solution. Filter into a tared platinum dish, evaporate, ignite below redness and weigh the mixed potassium and sodium chlorids.

Determine potassium in the mixed chlorids as directed in I, 45, beginning with "Digest the residue with hot water, filter through a small filter". Calculate the potassium so found to its equivalent of potassium chlorid and subtract this from the weight of the mixed chlorids to obtain the weight of sodium chlorid.

25

PHOSPHORIC ACID.—OFFICIAL.

Mix 5 grams of the sample with a little magnesium nitrate solution, dry, ignite, dissolve in dilute hydrochloric acid and dilute the solution to a definite volume. In an aliquot of the solution determine phosphoric acid as directed under I, 6 or 9.

26

SULPHURIC ACID¹¹.—TENTATIVE.

Boil 5 grams of the sample gently for 1½ hours with a mixture of 300 cc. of water and 15 cc. of concentrated hydrochloric acid. Dilute to 500 cc., draw off a 100 cc. aliquot, dilute considerably, precipitate with 10% barium chlorid solution, filter the precipitated barium sulphate on a Gooch, wash with hot water, dry, ignite and weigh.

27

AMMONIA.—TENTATIVE.

Introduce 2 grams of the sample into a distillation flask, add 300–400 cc. of water and an excess of sodium hydroxid solution, connect with a condenser and distil into a measured amount of standard acid. Titrate the excess of acid in the distillate with standard alkali, using methyl red or cochineal as an indicator.

Ammonia alum is often an ingredient of cream of tartar substitutes and baking powders, and ammonium carbonate is occasionally present in baking powders.

LEAD.

Method I. Colorimetric Method¹².—Tentative.

(Applicable in the absence of alum and phosphates. Approximate method for preliminary work.)

28

REAGENTS.

(a) *Sodium bisulphite solution.*—Dissolve 10 grams of anhydrous sodium carbonate in sufficient water to make 100 cc. and pass sulphur dioxid into the solution

until carbon dioxid is no longer evolved. Dilute a little of this solution with 10 volumes of water as needed in the determination.

(b) *10% potassium cyanid solution.*

(c) *Standard lead solution.*—Dissolve 1.6 grams of crystallized lead nitrate, previously dried over sulphuric acid, in a liter of water containing a few drops of dilute nitric acid. One cc. of this solution is equivalent to 1 mg. of metallic lead. Dilute 1 cc. of this solution to 100 cc. immediately before use in making up the color standards.

(d) *Lead-free tartrate solution.*—Dissolve 200 grams of tartaric acid in about 500 cc. of hot water, cool, add 40 cc. of the sodium bisulphite solution, heat to incipient boiling and test a few drops of the solution with potassium sulphocyanate solution to ascertain if all the iron is reduced to the ferrous state, repeating the treatment with about 10 cc. of the sodium bisulphite solution in case ferric iron is still present. Cool, add 20 cc. of the 10% potassium cyanid solution, and then strong ammonium hydroxid solution until the solution is distinctly alkaline to litmus paper. Boil until the solution is clear, cool, add 2 cc. of freshly prepared, colorless ammonium sulphid solution, dilute to 1 liter and allow to stand overnight. Filter to remove the precipitated sulphids, boil the filtrate until hydrogen sulphid is removed, cool and dilute to 1 liter with water.

29

PREPARATION OF SOLUTION.

(a) *Baking powder.*—Weigh 20 grams of the sample into a 250 cc. casserole, add water a little at a time with stirring until foaming ceases, then hydrochloric acid (1 to 1) a little at a time until all the carbonate is decomposed and finally 5 cc. excess of the hydrochloric acid. Cover with a watch glass and digest on a steam bath until all the starch is hydrolyzed as shown by testing 1 or 2 drops of the mixture with iodine. Filter through a folded filter and wash the filter several times with small portions of hot water. Treat the residue on the filter with several small portions of hot nitric acid (sp. gr. 1.2), collect the acid solution in a separate, small porcelain dish, evaporate this solution to dryness on a water bath and expel nitric acid by several treatments and evaporations with a few drops of concentrated hydrochloric acid. Rinse the contents of the dish through a small filter into the main solution and make up to 100 cc.

(b) *Tartaric acid and cream of tartar.*—Dissolve 100 grams of the sample in hot water, add 50 cc. of hydrochloric acid (1 to 1), filter into a liter graduated flask, wash the filter several times with water, and then treat the residue on the filter with several small portions of hot nitric acid (sp. gr. 1.2), collect the acid solution in a separate, small porcelain dish, evaporate this solution to dryness on a water bath and expel nitric acid by several treatments and evaporations with a few drops of concentrated hydrochloric acid. Rinse the contents of the dish through a small filter into the main solution, finally diluting the combined filtrates and washings to a liter.

30

DETERMINATION.

Introduce 50 cc. of the solution, prepared as directed in 29, into a beaker, add 2 cc. of the sodium bisulphite solution, heat to incipient boiling, and test a few drops of the solution with potassium sulphocyanate to determine if all the iron is reduced to the ferrous state, repeating the treatment with the sodium bisulphite solution if ferric iron is still present. Cool, add 1 cc. of the 10% potassium cyanid solution and neutralize to litmus with strong ammonium hydroxid solution; finally add an excess of 1 cc. of the last reagent. Boil gently until clear and colorless, cool

and make up to 100 cc. Treat with 2 drops of freshly prepared, colorless ammonium sulphid solution, mix and compare in a colorimeter with standard solutions, prepared by adding measured amounts of the standard lead solution to 50 cc. of the lead-free tartrate solution, diluting to 100 cc. and treating with 2 drops of freshly prepared colorless ammonium sulphid solution.

The final comparison should be made with a standard containing approximately the same amount of lead, and the addition of ammonium sulphid solution should be made to the standards and the solution of the sample at the same time, as the colors change on standing.

31*Method II.—Tentative.*

(Applicable to alum or phosphate baking powders or their ingredients.)

Weigh 100 grams of the sample into a 1.3 liter beaker and add an excess of hydrochloric acid (1 to 3) in small portions, keeping down excessive frothing with a little ether. Heat the mixture on a steam bath until the starch is hydrolyzed and the solution is quite limpid. Cool and add 200 cc. of 50% lead-free ammonium citrate solution. Place the beaker in a bath of cold water and add carefully ammonium hydroxid solution, in small portions with constant stirring, until the mixture is alkaline. If a precipitate forms, add sufficient ammonium citrate solution to dissolve it. Then add 15 cc. of saturated mercuric chlorid solution, dilute the mixture to about 1200 cc., saturate with hydrogen sulphid and allow to stand until the precipitate has settled (15–20 minutes). Filter and wash the precipitate with hydrogen sulphid water. Place the paper and precipitate in a small casserole, add 10 cc. of concentrated nitric acid and 2 cc. of concentrated sulphuric acid and heat on a hot plate until the mixed acids have been slowly driven off. Heat the residue in a muffle at low redness until the mercury salts have volatilized. Cool the casserole and leach the residue several times with 25% ammonium acetate solution, made slightly alkaline with ammonium hydroxid, pass the leachings through a small filter into a beaker and finally wash the residue and filter paper with a little hot water. Acidify the combined filtrate and washings with acetic acid, add an excess of potassium dichromate solution and allow to stand overnight. Filter on a tared Gooch, wash with water, dry for 30 minutes at 125°–150°C., cool and weigh as lead chromate. Calculate the weight of metallic lead. Conduct a blank determination upon all the reagents and correct the result accordingly.

32*Method III.—Tentative.*

(Applicable to alum or phosphate baking powders or their ingredients.)

Transfer 200 grams of the sample to a 3 liter Jena flask, add 300 cc. of concentrated nitric acid in small portions, shake thoroughly after each addition and heat the mixture slowly, shaking repeatedly. When brown fumes begin to appear at the mouth of the flask, discontinue heating and insert a stemless funnel in the neck of the flask. As soon as the action has moderated, place the flask on an asbestos gauze over a small Bunsen flame. When the action becomes weak, add slowly 90 cc. of concentrated sulphuric acid and continue heating until the fumes disappear. Then add 25 cc. of concentrated nitric acid from time to time with continued heating until all the starch is completely oxidized. Usually 3–4 additions of 25 cc. portions of nitric acid suffice. Finally expel the nitric acid as completely as possible. Cool, add 400 cc. of water, shake and allow to settle. The soluble sulphates of sodium, potassium, aluminium, iron, etc., go into solution, while calcium sulphate and most of the lead sulphate will be precipitated. Filter through an 18

cm. folded filter into a liter Erlenmeyer flask, rinse the 3 liter flask 2-3 times with small portions of water and pour the rinsings through the filter. Reserve the filtrate for the recovery of dissolved lead salts. Open the filter, containing the precipitate, over a 600 cc. beaker and wash the precipitate into it. Then transfer the contents of the beaker to a 2 liter Erlenmeyer flask together with whatever precipitate remains in the 3 liter flask. Dilute the contents of the flask so as to nearly fill the latter, stir thoroughly to dissolve the calcium sulphate, add 20 cc. of strong acetic acid and saturate the liquid thoroughly with hydrogen sulphid. Stopper the flask and set aside until the precipitate settles. Siphon off the supernatant liquid. When much calcium sulphate is present, one such treatment is not sufficient to dissolve all of it. In this case refill the flask with water, again acidify, saturate with hydrogen sulphid and allow to stand till the calcium sulphate is practically all dissolved and the residue of sulphids is dark colored. Solution may be hastened by the addition of lead-free sodium acetate to the water (50-75 grams to each 2 liters).

Treat the liquid containing the soluble sulphates separately to recover the trace of lead which it may contain. Partially neutralize with ammonium hydroxid solution just short of the point of producing a permanent precipitate of aluminium phosphate, saturate with hydrogen sulphid and allow the precipitate to settle. Some iron sulphid also will usually be precipitated. The sulphid precipitations should be made in very slightly acid solutions, otherwise lead sulphid will not be precipitated completely. Siphon off the supernatant liquid, transfer the precipitate to an 11 cm. filter and wash with hydrogen sulphid water. Transfer the first precipitate remaining in the 2 liter flask to a second 11 cm. filter and treat in the same way. Place the 2 filters with their contents in a 200 cc. Erlenmeyer flask, add 10 cc. of concentrated nitric acid and 5 cc. of concentrated sulphuric acid, insert a stemless funnel in the neck and heat to completely oxidize the material. When the nitric acid has all been expelled and the residue darkens, add more nitric acid until no such darkening occurs. Finally heat the residue till fumes of sulphur trioxid are given off, cool and add 15 cc. of water. Filter through a 7 cm. filter, rinse, then wash the filter twice with small portions of dilute sulphuric acid and finally with a little water. Place a clean 150 cc. beaker under the filter, dissolve the precipitate in 15-25 cc. of ammonium acetate solution [1 part of 99% acetic acid, 1 of water and 1 of ammonium hydroxid (sp. gr. 0.90); made neutral to litmus paper] and wash thoroughly with water.

Acidify the filtrate and washings with acetic acid, add an excess of potassium dichromate solution, heat on a steam bath and allow to cool and settle. Filter on a tared Gooch prepared with a thick layer of asbestos which has been previously dried at 125°C., wash with water, dry at about 125°C. and weigh as lead chromate.

33

Method IV.—Tentative.

(Applicable in the absence of alum and phosphates.)

Weigh 100 grams of the sample into a liter beaker and add an excess of hydrochloric acid (1 to 3) in small portions, keeping down excessive frothing with a little ether. Heat the mixture on a steam bath until the starch is hydrolyzed and the solution is quite limpid. Cool, add ammonium hydroxid solution until distinctly alkaline, dilute to about 800-900 cc. and saturate with hydrogen sulphid. Allow the mixture to stand for 3-4 hours or until the precipitate has settled, filter on a 12.5 cm. close-textured paper and wash the precipitate several times with hydrogen sulphid water. Place the filter paper and precipitate in a 100 cc. Erlenmeyer flask, add 5 cc. of concentrated sulphuric acid and 5 cc. of concentrated nitric acid and

METHODS OF ANALYSIS

heat on a hot plate, with occasional additions of small portions of concentrated nitric acid, until the mixture no longer blackens when evaporated to the point at which white fumes of sulphur trioxid appear. Cool, dilute with 20 cc. of water, warm until the ferric sulphate goes into solution, cool and then add 40 cc. of 95% alcohol by volume. Allow to stand overnight, filter on a Gooch and wash with 95% alcohol. Dissolve the lead sulphate remaining on the filter by washing with 20 cc. of 25% ammonium acetate solution, rendered slightly alkaline with ammonium hydroxid, collect the filtrate in a small beaker, passing it through the filter 3-4 times. Finally wash the filter with hot water, acidify the combined filtrate and washings with acetic acid, add an excess of potassium dichromate solution and allow to stand overnight. Filter on a small, tared Gooch, wash, dry for 30 minutes at 125°-150°C. and weigh as lead chromate. Calculate the metallic lead.

34

ARSENIC.—TENTATIVE.

Introduce 5 grams of the sample directly into the generator described under **XII, 2** (Fig. 7), add 10 cc. of water, a little at a time to prevent foaming over, and then 15 cc. of concentrated, arsenic-free hydrochloric acid, introducing it drop by drop until foaming ceases. Heat on a steam bath until a drop of the mixture, when diluted and treated with iodine solution, shows no blue color. Then dilute to about 30 cc. with water, add 4 cc. of potassium iodide solution and continue from this point as directed under **XII, 4**, beginning with "Heat to about 90°C.", except that the blank and the standards for comparison are made by the use of the arsenic-free hydrochloric acid of the same concentration as that used in the determination.

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XXIX. DRUGS.

CAFFEIN AND ACETANILID IN MIXTURES.

1

PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.

(a) If the sample is already in powder form, rub thoroughly in a mortar and keep in a tightly corked tube or flask. Powders in paper, cachet or capsule containers are frequently of such fineness as to require little further trituration except to produce a uniform product. On a tared 5.5 cm. filter weigh 0.3–0.5 gram of the sample or, if preferred, an amount equal to, or a multiple of, the average unit dose (previously ascertained by weighing collectively 20 or more such doses), wash with successive 5–10 cc. portions of the chloroform (30–50 cc. are usually sufficient) until the extraction is complete as indicated by the absence of any residue after evaporation of a small portion of the last washing. Collect the solution in a 200 cc. Erlenmeyer flask, connect the flask with a condenser by means of a cylindrical Kjeldahl connecting bulb¹ and distil until the volume is reduced to about 10 cc.

(b) If the caffein is present in the citrated form, or the composition of the mixture precludes complete extraction as directed in (a), weigh out the desired amount, transfer to a Squibb separatory funnel, add 50 cc. of the chloroform and 20 cc. of water, shake vigorously and, after clearing, draw off the lower layer through a small, dry filter into a 200 cc. Erlenmeyer flask. In the case of coated tablets and pills, ascertain their average weight, powder in a mortar and weigh out for each determination an amount equivalent to one or more tablets or pills. Repeat the extraction twice, using 50 cc. portions of the chloroform for each extraction. Distil the combined chloroform extracts to about 10 cc.

(c) In the case of dilute alcoholic solutions, evaporate a measured quantity on a steam bath until most of the alcohol has been expelled, or take an aliquot of the residue from an alcohol determination; transfer to a separatory funnel by pouring and rinsing with a minimum of water so that the final volume does not greatly exceed 20 cc., and then, in order to avoid any loss of acetanilid by hydrolysis during evaporation, add a little solid sodium bicarbonate and a drop of acetic anhydrid. (Should the preparation contain alkaloids, acidify with a few drops of dilute sulphuric acid immediately after acetylizaton to retain such basic material in solution.) Add 50 cc. of the chloroform, shake vigorously and, after clearing, draw off the chloroform layer through a filter into a 200 cc. Erlenmeyer flask. Repeat the extraction twice, using 50 cc. portions of the chloroform for each extraction, and distil the combined chloroform washings to a volume of about 10 cc.

CAFFEIN AND ACETANILID.—TENTATIVE.

2

REAGENTS.

(a) *Standard bromid-bromate solution.*—Dissolve 50 grams of potassium hydroxid in a small quantity of water, add a slight excess of bromin, dilute with water to dissolve any separated salts, boil to expel excess of bromin and dilute to 1 liter. Standardize the solution against recrystallized acetanilid and adjust the solution so that 1 cc. is equivalent to 5 or 10 mg. of acetanilid as desired.

(b) *Chloroform.*—Redistilled and residue-free. All corks used in the distillation should be treated previously with chloroform.

(C) *Wagner's reagent*.—Dissolve 2 grams of iodine and 6 of potassium iodide in a minimum amount of water and dilute to 100 cc.

3

CAFFEIN.—TENTATIVE.

Treat the chloroform extract, obtained in 1, with 10 cc. of sulphuric acid (1 to 10) and digest on a steam bath until the contents of the flask are reduced to 5 cc. Add 10 cc. of water and continue the digestion until the liquid is again reduced to 5 cc., then cool and transfer to a separatory funnel with a minimum of water, so that the final volume does not greatly exceed 20 cc. Add 50 cc. of the chloroform, extract in the usual way and, after clearing, withdraw the lower layer through a small, dry filter into a 200 cc. Erlenmeyer flask. Repeat the extraction with two 50 cc. portions of the chloroform. On the completion of the third extraction, distil the combined extracts down to about 10 cc., finally transferring the residual liquid, by washing with chloroform, to a tared beaker or crystallizing dish. Allow the solution to evaporate spontaneously, or by gentle heat and an air blast, to apparent dryness. Cool and allow to stand until the weight becomes constant.

Chloroform extracts in addition to caffeine and acetanilide contain oils, fats, waxes, resins, pigments and other substances from those preparations which contain powdered cinnamon, celery seed, ginger or other vegetable products. These appear either in suspension or solution after the caffeine-acetanilide mixture has been digested and contaminate the caffeine. Remove any suspended impurities by filtering through a small, moistened filter immediately after hydrolysis and prior to extraction with chloroform. Should the recovered caffeine be deeply colored or contaminated with foreign matter, purify it as follows: Dissolve in very dilute sulphuric acid (about 5 cc. of N/5 acid for every 100 mg. of caffeine), filter, if necessary, through a moistened filter, add 15–20 cc. of Wagner's reagent, sufficient at least to distinctly color the supernatant liquid, stir and allow to stand an hour, preferably in a refrigerator. Filter and wash the periodide with a few cc. of iodine solution, transfer both filter and precipitate to a separatory funnel, using not more than 20 cc. of water, decolorize with a crystal of sodium thiosulphate, then extract with three 50 cc. portions of chloroform and proceed as directed above.

4

ACETANILIDE.—TENTATIVE.

Transfer the solution of aniline sulphate, remaining in the separatory funnel in 3, to the Erlenmeyer flask used in effecting hydrolysis, then heat 10 minutes on a steam bath to expel all traces of chloroform. Wash the filter, used in the preceding operation to dry the chloroform solution of caffeine, with 5 cc. of water, adding the latter to the main solution of aniline sulphate. Add 10 cc. of concentrated hydrochloric acid, then run in the standard bromide-bromate solution until a faint yellow coloration remains, rotating the flask sufficiently to agglomerate the precipitated tribromaniline. Calculate the quantity of acetanilide from the number of cc. required to complete the precipitation.

Caffeine and acetanilide are the 2 principal ingredients of the preparation known as "acetanilide compound", a further constituent being sodium bicarbonate. The latter appears as the chloroform-insoluble residue and may be determined by titrating such residue, or one obtained by titrating a portion of the original mixture, with standard acid, using Congo red as an indicator. The bicarbonate may also be determined by igniting the original sample, or the chloroform-insoluble residue, with sulphuric acid and weighing the resulting sodium sulphate.

Should the "acetanilide compound" be combined with sodium bromide, the latter, in the absence of other halides, may be determined volumetrically by the Volhard method [III, 15].

CAFFEIN AND ACETPHENETIDIN (PHENACETIN) IN MIXTURES.

5

PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.

In the case of preparations containing acetphenetidin instead of acetanilid, but otherwise identical, make the gross separation of the caffein-acetphenetidin mixture as directed under 1.

6

CAFFEIN.—TENTATIVE.

Treat the chloroform extract, obtained as directed under 1, with 10 cc. of sulphuric acid (1 to 10) and digest on a steam bath until the liquid is reduced to about 5 cc. Dilute with 10 cc. of water and continue the digestion until the volume is again reduced to 5 cc., then add 10 cc. of water and continue heating until the residual liquid amounts to 8–10 cc. If, during the digestion, particles of acetphenetidin remain on the sides of the flask rinse them into the solution with a few drops of chloroform.

Great care must also be given to the degree of evaporation. Should the aqueous-acid solution and suspension of caffein-acetphenetidin be concentrated far beyond the limits indicated, more or less phenetidin sulphonate is likely to be formed, which later resists acetylation and conversion to acetphenetidin.

Cool and transfer with water to a separatory funnel, so that the final volume does not greatly exceed 20 cc. Then proceed as directed under 3.

7

ACETPHENETIDIN.—TENTATIVE.

Wash the filter, used to dry the chloroform in 6, with 5 cc. of water, receiving the water in the separatory funnel containing the solution of phenetidin sulphate. Treat with successive small portions of solid sodium bicarbonate until, after complete neutralization of free acid, an excess of the former remains at the bottom of the mixture. Add 50 cc. of chloroform and for every 0.10 gram of acetphenetidin, known or believed to have been present, 5 drops of acetic anhydride; shake vigorously, allow to clear, then withdraw the chloroform into a second separatory funnel containing 5 cc. of water. Shake this mixture and, after clearing, pass the solvent through a small, dry filter into a 200 cc. Erlenmeyer flask. Distil over about 40 cc. of the chloroform, make up the distillate to 50 cc. with chloroform, add this to the material in the first separatory funnel and extract again. Withdraw the chloroform layer to the second separatory funnel, wash and distil about 50 cc. (for use in the final extraction). Distil the chloroform down to about 10 cc., transfer with sufficient fresh solvent to a tared 50 cc. beaker or crystallizing dish, evaporate on the steam bath to apparent dryness, finally removing any considerable excess of acetic anhydride by repeated additions of 1 cc. of chloroform and a drop of alcohol. The reformed acetphenetidin should finally appear as a whitish, crystalline mass with a faint, acetous odor which disappears completely on standing some hours in the open, or in a vacuum desiccator over lime. Weigh from time to time until the final weight differs from the preceding by not more than 0.5 mg.

CAFFEIN AND ANTIPYRIN IN MIXTURES².

8

PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.

(a) Extract a weighed portion of the finely powdered sample on a filter with chloroform to separate the caffein and antipyrin from the usual excipients of tablet and pill combinations. Distil off the greater part of the chloroform and evaporate the remainder on the steam bath.

(b) In the case of alcoholic preparations, remove the alcohol from a measured amount of the sample by heating on a steam bath. Extract the residue with three 50 cc. portions of chloroform in a separatory funnel. Distil off the greater portion of the chloroform and evaporate the remainder on a steam bath.

9

ANTIPYRIN.—TENTATIVE.

Transfer the residue, obtained in 8, which should weigh about 0.25 gram, to a 150 cc. separatory funnel by means of two 5 cc. portions of alcohol-free chloroform, followed by 10 cc. of water. Add 1 gram of sodium bicarbonate and 10–15 cc. of N/5 iodine (or double the quantity of N/10 iodine), adding the latter in small portions and shaking the mixture vigorously after each addition. The iodine should then be in excess of that required to convert all the antipyrin into the mono-iod derivative. If not, add a little more and shake the mixture again. Remove the free iodine with a small crystal of sodium thiosulphate, add 15 cc. of chloroform, shaking vigorously for 1 minute. After clearing, draw off the chloroform solution into a second separatory funnel, wash with 5 cc. of water, filter through a small, dry filter into a tared 50 cc. beaker and evaporate to apparent dryness on the steam bath, using an air blast. Repeat the extraction with two (three, if N/10 iodine is used) 25 cc. portions of chloroform, wash, filter and evaporate each portion as above. Dry the nearly colorless, crystalline residue of caffeine and iodantipyrin 30 minutes at 105°C., cool and weigh. Designate this weight as "a".

Dissolve the composite residue in 5 cc. of glacial acetic acid, add 10 cc. of saturated sulphur dioxide solution, then transfer with hot water to a 400–500 cc. beaker until the final volume amounts to about 200 cc. Add sufficient silver nitrate solution to precipitate all the iodine (about 0.3 gram of silver nitrate); then a few drops of nitric acid, heat nearly to boiling and stir to agglomerate the silver iodide. Add 15 cc. of concentrated nitric acid, cover the beaker with a watch glass and boil gently for 5 minutes. Filter by decantation through a tared Gooch, wash the precipitate once with a little alcohol, then with two 100 cc. portions of boiling water and finally transfer the iodide to the crucible. Wash several times with hot water and again with alcohol to remove traces of organic matter, dry 30 minutes in an air bath at 110°C., cool and weigh. The weight of silver iodide multiplied by 0.8012 gives the weight of antipyrin.

10

CAFFEIN.—TENTATIVE.

Calculate the quantity of caffeine by multiplying the weight of silver iodide by 1.3374 and subtracting the product from the weight "a" above.

In the analysis of a mixture containing caffeine, antipyrin, acetanilide and sodium salicylate, the following steps are essential in effecting a separation: (1) Extraction of caffeine, acetanilide and antipyrin with chloroform from the aqueous soda solution; (2) Hydrolysis with sulphuric acid of the 3 substances thus separated preliminary to the determination of caffeine and antipyrin as directed in 9 and 10.

ACETANILID AND ACETPHENETIDIN (PHENACETIN) IN MIXTURES³.

ACETPHENETIDIN.—TENTATIVE.

11

REAGENTS.

(a) *Purified iodine*.—Dissolve 2 parts of resublimed iodine and 1 of potassium iodide in 1 of water, pour the clear solution into a large volume of water, filter and wash the finely precipitated iodine several times on a porous plate with water. Dry in the air and finally in a desiccator over sulphuric acid where it is kept in a glass-stoppered weighing bottle.

(b) *Standard sodium thiosulphate solution*.—Dissolve 30 grams of crystallized sodium thiosulphate in water and dilute to 1 liter. Standardize this solution against the purified iodine as follows: Weigh out about 0.3 gram of the purified iodine in a small, glass capsule (about $\frac{1}{2}$ inch high and $\frac{5}{8}$ inch diameter), provided with a closely fitting glass cap or stopper, and place the capsule in a 200 cc. Erlenmeyer flask containing 0.5 gram of potassium iodide dissolved in 10 cc. of water. After complete solution, titrate with the sodium thiosulphate solution, using 1 or 2 drops of starch solution as an indicator.

(c) *Standard iodine solution*.—Dissolve 40 grams of potassium iodide in the least possible quantity of water, add 30 grams of the purified iodine and, after solution, dilute to 1 liter. Standardize 25 cc. of this solution against the standard sodium thiosulphate solution.

12

DETERMINATION.

(1) Place 0.2 gram of the acetphenetidin-acetanilid mixture in a 50 cc. lipped Erlenmeyer flask, add 2 cc. of glacial acetic acid, heat gently over a wire gauze to complete solution and dilute with 40 cc. of water, previously warmed to 70°C. Transfer the clear liquid with two 10 cc. portions of warm (40°C.) water to a glass-stoppered, 100 cc. graduated flask containing 25 cc. of the standard iodine solution warmed to 40°C. Stopper, mix thoroughly, then add 3 cc. of concentrated hydrochloric acid, continue shaking until crystallization begins and then set aside to cool. If the ratio of acetphenetidin to acetanilid is equal to or greater than unity, crystalline scales will form almost immediately on the addition of acid. As the proportion of acetanilid increases, however, the periodid tends to remain in the liquid state. In such cases, gentle agitation or rotation of the flask in water, warmed not to exceed 40°C., hastens the formation of crystals. When the contents of the flask are at room temperature, fill with water to within 2-3 cc. of the mark, mix thoroughly and allow to stand overnight. Fill to the mark with water, mix thoroughly, allow to stand 30 minutes, filter through a 5.5 cm. dry, closely fitted filter into a 50 cc. graduated flask, rejecting, however, about 15 cc. of the first runnings but reserving them for the recovery of acetanilid. Transfer the 50 cc. aliquot to a 200 cc. Erlenmeyer flask and titrate with the standard sodium thiosulphate solution. Calculate the amount of acetphenetidin from the following formula:

Acetphenetidin = $I (0.0896 \times N)$ in which

0.0896 = the quantity of acetphenetidin contained in 1 cc. of a normal solution of this substance;

N = the normality of the standard sodium thiosulphate solution employed; and

I = the number of cc. of the standard sodium thiosulphate solution corresponding to the iodine combined with the acetphenetidin.

The formula of the precipitated periodid, which constitutes the basis for the above determination, is $(C_2H_5O.C_6H_4NH.COCH_3)_2HI.I_4$.

(2) The gravimetric determination of acetphenetidin may, if desired, be effected as follows: Filter off the periodid, preferably by suction, wash with 10-15 cc. of the standard iodine solution, then transfer together with the filter to a separatory funnel, using not over 50 cc. of water. Remove both free and added iodine with a few small crystals of sodium sulphite and extract the liquid with three 50 cc. portions of chloroform, washing each portion subsequently into a second separatory funnel with 5 cc. of water. After washing and clearing, filter the chloroform solution through

a dry 5.5 cm. filter into a 200 cc. Erlenmeyer flask, distil off most of the chloroform, transfer the residual solution (5–10 cc.), by means of a little chloroform, to a small, tared beaker or crystallizing dish, evaporate to dryness on a steam bath, cool and weigh.

For the identification of acetphenetidin, either alone or in admixture with acetanilid, the following test will be found of value³: To 1–2 mg. of the sample in a test tube add a drop of acetic acid, 0.5 cc. of water and 1 cc. of N/10 iodine, warm the mixture to about 40°C., then add a drop of concentrated hydrochloric acid. If acetphenetidin alone is present, its periodid separates almost immediately in the form of reddish brown leaflets or needle-like crystals. If the sample consists largely of acetanilid, the separation takes place on cooling and shaking the liquid. In the presence of considerable acetanilid, the periodid first separates as minute, oily globules, which, on vigorous shaking, gradually become crystalline. This test is so delicate that as little as 0.5 mg. of acetphenetidin may, if alone, be detected in the form of its characteristic periodid.

13

ACETANILID.—TENTATIVE.

(1) If the combined weight of the acetphenetidin-acetanilid mixture is known, determine that of the latter ingredient by difference; or, (2) Determine it directly from a second aliquot of the filtrate from the acetphenetidin periodid in **12** as follows:

Pipette 25–30 cc. of the clear liquid into a separatory funnel, decolorize with solid sodium sulphite and solid sodium bicarbonate in slight excess, add 1 or 2 drops of acetic anhydride, then extract with three 60 cc. portions of chloroform, passing the chloroform solution, when cleared, through a small, dry filter into a 200 cc. Erlenmeyer flask, and distil the chloroform, by the aid of gentle heat, to about 20 cc. Add 10 cc. of sulphuric acid (1 to 10) and digest on a steam bath until the residue has been reduced one half, add 20 cc. of water and continue the digestion for an hour; then add a second 20 cc. portion of water and 10 cc. of concentrated hydrochloric acid, titrate very slowly, drop by drop, with the standard bromide-bromate solution, **2 (a)**, until a faint yellow color remains. While adding this reagent, rotate the flask sufficiently to agglomerate the precipitated tribromanilin. Calculate the amount of acetanilid present.

If the preparation contains caffeine or antipyrin or both in addition to acetanilid and acetphenetidin, proceed as follows: (1) Digest the mixture by heating with dilute sulphuric acid to convert acetphenetidin and acetanilid to phenetidin and anilin sulphates, respectively; (2) Separate the caffeine and antipyrin by extraction with chloroform; (3) Re-form acetphenetidin and acetanilid by treating the solution of the corresponding sulphates with solid sodium bicarbonate in slight excess, then add a few drops of acetic anhydride and extract with chloroform⁴.

ACETPHENETIDIN (PHENACETIN) AND SALOL IN MIXTURES⁵.

ACETPHENETIDIN.

14

Acid Hydrolysis Method.—Tentative.

Weigh out on a tared 5.5 cm. filter an amount of the sample equal to, or a multiple of, the average weight of a unit dose and wash with sufficient successive, small portions of chloroform to extract completely all acetphenetidin and salol present in the mixture (about 40 cc.). Collect the solution in a tared, 100 cc. beaker and evaporate on a warm plate (50°–60°C.) to apparent dryness, using an air blast. Let stand 24 hours at room temperature to practically constant weight, then transfer the crystalline residue, by means of chloroform, to a 50 cc. lipped Erlenmeyer flask, evaporate the solvent by means of an air blast and gentle heat, add 10 cc.

of sulphuric acid (1 to 10) and evaporate on the steam bath until the volume is reduced one half. Add 10 cc. of water and continue the digestion as before, then add a second 10 cc. of water and evaporate to 5 cc. Transfer the residue with about 20 cc. of water to a small separatory funnel and extract with 15, 10 and 5 cc. of chloroform, washing each extract with 5 cc. of water in a second separatory funnel to recover traces of phenetidin sulphate possibly dissolved by the chloroform, finally rejecting the latter since it contains all the salol not previously eliminated during the digestion.

Add the wash water in the second separatory funnel to the solution of phenetidin sulphate in the first separatory funnel and proceed as directed under **7**, beginning with "Treat with successive small portions of solid sodium bicarbonate".

15*Alkaline Hydrolysis Method.—Tentative.*

On a small, tared filter weigh out an amount of the sample to contain not more than 0.10 gram of salol, exhaust with chloroform as directed in **14**, collect the solvent in a small, lipped Erlenmeyer flask and evaporate the chloroform by means of an air blast without heat. Add 10 cc. of 2.5% sodium hydroxid solution and heat 5 minutes on a steam bath. Cool quickly to room temperature in running water to prevent partial hydrolysis of the acetphenetidin. Transfer the liquid to a separatory funnel with a minimum of water, then rinse out the flask with the first 20 cc. portion of chloroform used in the extraction. Extract the alkaline solution with three 20 cc. portions of chloroform, wash each portion in a second separatory funnel with 5 cc. of water and pass the solution through a small, dry filter into a 200 cc. Erlenmeyer flask. Designate the combined alkaline solution and washings as *A*. Distil the combined chloroform extracts to about 5 cc. Transfer by means of a little chloroform to a small, tared beaker or crystallizing dish, evaporate on a steam bath with the aid of an air blast, cool and weigh the residual acetphenetidin at intervals until the weight becomes constant.

SALOL.**16***Acid Hydrolysis Method.—Tentative.*

Subtract the weight of acetphenetidin, as determined in **14**, from the combined weight of the 2 ingredients determined in **14**, to obtain the weight of salol.

17*Alkaline Hydrolysis Method.—Tentative.*

Place the combined alkaline solutions, *A*, under **15**, in a 500 cc. glass-stoppered bottle, dilute with water to about 200 cc., run in from a burette an excess (about 45 cc.) of N/7 potassium bromid-bromate, add 10 cc. of concentrated hydrochloric acid and shake 1 minute, then at intervals for 30 minutes. Add 10 cc. of 15% potassium iodid solution and shake at intervals for 15 minutes. Titrate the free iodine with standard sodium thiosulphate solution (preferably N/7), previously standardized against the N/7 bromid-bromate solution. One cc. of N/7 potassium bromid-bromate is equivalent to 2.55 mg. of salol. From the number of cc. of the N/7 bromid-bromate solution used, calculate the amount of salol on the basis of 12 atoms of bromine to 1 molecule of salol.

ACETANILID AND SODIUM SALICYLATE IN MIXTURES.**18****PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.**

Weigh an amount of the powdered sample equal to, or a multiple of, an average unit dose, transfer to a separatory funnel containing 10 cc. of water and, for every unit dose, add 0.10 gram of solid sodium bicarbonate. In the case of coated tablets

and pills, ascertain their average weight, powder in a mortar and weigh out an amount of the powder equivalent to one or more tablets or pills for each determination prior to treatment in the separatory funnel. In the examination of alcoholic preparations, distil off the alcohol from a measured volume on a steam bath, transfer to a separatory funnel with a minimum of water and add sufficient solid sodium bicarbonate (0.5 to 1.0 gram).

19**ACETANILID.—TENTATIVE.**

Extract the alkaline solution, obtained under **18**, with three 50 cc. portions of chloroform, wash each portion with 5 cc. of water in a second separatory funnel and collect the solvent, without previous drying, in a 200 cc. Erlenmeyer flask. Designate the aqueous solution as *A*. Distil off the chloroform very gently to about 5 cc., add 10 cc. of dilute sulphuric acid and completely hydrolyze on the steam bath. Proceed from this point as directed in **4**.

20**SODIUM SALICYLATE.—TENTATIVE.**

Acidify the aqueous solution of sodium salicylate, *A*, under **19**, with a few drops of concentrated hydrochloric acid and extract with sufficient (3–5) 25 cc. portions of chloroform to exhaust the salicylic acid present in the mixture. Treat each portion in a second separatory funnel with 20 cc. of water, containing 1 gram of anhydrous sodium carbonate for every 100 mg. of salicylic acid. Shake vigorously and, after clearing, wash each portion again in a second separatory funnel with 5 cc. of water, then add the washings to the main aqueous soda solution of sodium salicylate. Dilute to a known volume, transfer an aliquot, representing about 100 mg. of salicylic acid, to a 200 cc. Erlenmeyer flask, make up to 100 cc., heat nearly to boiling, then run in from a burette 25–40 cc. of strong (about *N*/5) iodine solution, sufficient to insure an excess during digestion and digest for an hour on a steam bath. Remove the free iodine with a few drops of sodium thiosulphate solution, decant the clear liquid through a tared Gooch, retaining most of the precipitate, tetraiodophenylenequinone ($C_6H_2I_2O$)₂, in the flask. To the latter add 50 cc. of boiling water, digest 10 minutes on the steam bath, then filter, wash gradually all the precipitate into a Gooch, using for this purpose and the final washing about 200 cc. of hot water. Dry to constant weight in an air bath at 100°C. Multiply the weight of the precipitate by 0.4654 to obtain the quantity of sodium salicylate present in the aliquot taken.

Should the mixture contain caffeine or antipyrin, or both, these substances will appear with the acetanilid in the first chloroform extract and may be determined as directed in the closely set type following **10**. Should the acetanilid be replaced by acetphenetidin in the mixture, the general procedure would not be materially altered, the acetphenetidin being weighed directly after recovery from its washed chloroform solution as separated from the sodium salicylate. If, instead of sodium salicylate, the mixture contains the free acid or its ammonium salt, add a larger quantity of sodium bicarbonate prior to extraction with chloroform to insure the fixation of salicylic acid.

In the analysis of a mixture of caffeine, acetanilid, sodium salicylate and codeine sulphate, the following procedure is recommended: (1) Extraction of caffeine, acetanilid and salicylic acid from the acidified solution; (2) Washing the chloroform solution with aqueous soda solution for the recovery of salicylic acid, preliminary to its treatment with iodine solution; (3) Separation of caffeine and acetanilid as directed under **3** and **4**; (4) Recovery of codeine from the solution of its sulphate after treatment with sodium bicarbonate and chloroform.

CAFFEIN, ACETANILID AND QUININ SULPHATE IN MIXTURES.

21

PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.

Transfer to a separatory funnel one or more average unit doses of the powdered sample, add 20 cc. of water and 50 cc. of chloroform, then 10 drops of dilute sulphuric acid and extract in the usual way. After clearing, wash the solvent in a second separatory funnel with 5 cc. of water prior to transferring to a 200 cc. Erlenmeyer flask. Repeat the foregoing operations with two 50 cc. portions of chloroform, finally distilling the combined chloroform solution to about 10 cc. by gentle heat.

22

CAFFEIN AND ACETANILID.—TENTATIVE.

Treat the chloroform residue obtained in 21 as directed under 3 and 4.

23

QUININ SULPHATE.—TENTATIVE.

Combine the wash water, used in the second separatory funnel in 21, with the solution of quinin bisulphate, add a slight excess of solid sodium bicarbonate, extract with three 50 cc. portions of chloroform, wash each portion with 5 cc. of water in a second separatory funnel and then pass through a dry filter into a 200 cc. Erlenmeyer flask. Distil by gentle heat to about 5 cc., evaporate on a steam bath to apparent dryness, dissolve the amorphous alkaloid in 5 cc. of neutral alcohol and titrate with N/50 hydrochloric acid to a faint red, using 2 drops of methyl red as an indicator. Heat on a steam bath until most of the alcohol has been expelled, adding, if necessary, sufficient acid to maintain the acid reaction. From the total number of cc. of acid employed in the titration calculate the quinin sulphate. One cc. of N/50 hydrochloric acid is equivalent to 8.73 mg. of quinin sulphate.

If the mixture contains acetphenetidin in place of acetanilid, proceed as outlined above, except that the separation of caffein and acetphenetidin is conducted as directed under 6 and 7.

CAFFEIN, ACETANILID AND CODEIN SULPHATE IN MIXTURES.

24

PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.

Proceed as directed under 21.

25

CAFFEIN AND ACETANILID.—TENTATIVE.

Proceed as directed under 22.

26

CODEIN SULPHATE.—TENTATIVE.

Proceed as directed under 23 to the point indicated by the sentence "Distil by gentle heat to about 5 cc.". Transfer the chloroform solution of codein with sufficient solvent to a small, tared beaker, evaporate to apparent dryness on a steam bath, add a few drops of alcohol to the amorphous residue, then a like amount of water and evaporate again. Finally cool and allow the usually crystalline product to stand until the weight becomes constant. The weight of this residue multiplied by 1.3144 gives the quantity of codein sulphate present.

This result should be checked volumetrically. Dissolve the residue in 3-5 cc. of neutral alcohol and titrate with N/50 sulphuric acid to a faint red, using methyl red as an indicator. From the number of cc. of standard acid employed calculate the amount of codein sulphate. One cc. of N/50 sulphuric acid is equivalent to 7.87 mg. of codein sulphate. The quantity of codein sulphate as found by weight will usually be slightly greater than that determined by titration.

CAFFEIN, ACETANILID, QUININ SULPHATE AND MORPHIN SULPHATE IN MIXTURES.

27

PREPARATION OF SAMPLE AND SOLUTION.—TENTATIVE.

Transfer to a separatory funnel an amount (containing not less than one fourth grain of morphin) of the powdered sample equal to, or a multiple of, a unit dose, add 20 cc. of water and 10 drops of dilute sulphuric acid, then extract with three 50 cc. portions of alcohol-free chloroform, wash each portion in a second separatory funnel with 5 cc. of water and add the combined washings to the alkaloidal solution in the first separatory funnel. Filter the chloroform extracts through a small, dry filter into a 200 cc. Erlenmeyer flask and distil by gentle heat to about 10 cc.

28

CAFFEIN AND ACETANILID.—TENTATIVE.

Treat the chloroform residue as directed under 3 and 4.

29

QUININ SULPHATE.—TENTATIVE.

Add to the solution of quinin and morphin sulphates, obtained in 27, 4–5 cc. of sodium hydroxid solution (1 to 10) and extract with four 40 cc. portions of chloroform, wash each portion with 5 cc. of water and pass the clear solvent through a small, dry filter into a 200 cc. Erlenmeyer flask. Remove the solvent by gentle distillation and titrate the residual quinin with N. 50 hydrochloric acid as directed under 23.

30

MORPHIN SULPHATE.—TENTATIVE.

Wash the filter, employed in 29, with 5 cc. of water and add to the aqueous alkaline solution of the alkaloid. Now add 0.5 gram of ammonium chlorid (or an amount slightly in excess of that required to free the morphin as well as convert all sodium hydroxid to sodium chlorid) and, to the resulting ammoniacal solution, add 45 cc. of chloroform and 5 cc. of alcohol, then extract in the usual way, washing the solvent in a second separatory funnel with 5 cc. of water. After clearing, pass the chloroform through a small, dry filter into a 200 cc. Erlenmeyer flask. Repeat the extraction with three 40 cc. portions of chloroform, washing and filtering as before, finally collecting all the solvent in an Erlenmeyer flask and distilling to about 10 cc. Transfer with chloroform to a small, tared beaker, evaporate to apparent dryness, add 0.5 cc. each of water and neutral alcohol, start crystallization by stirring with a glass rod and finally evaporate to dryness. Cool and allow to stand until the weight becomes constant.

Check the weight of morphin, thus determined, by titration with N/50 sulphuric acid, using a drop of methyl red as an indicator. Dissolve the alkaloid in 1–2 cc. of warm, neutral alcohol, then add the standard acid to a faint red. Evaporate most of the alcohol on a steam bath, adding, if necessary, sufficient acid to maintain the acid reaction. From the volume of acid used calculate the morphin sulphate. One cc. of N/50 sulphuric acid is equivalent to 7.58 mg. of morphin sulphate.

TRAGACANTH.

31

VOLATILE ACIDITY⁶.—TENTATIVE.

The quantity of volatile (acetic) acidity developed in the acid hydrolysis of gum tragacanth (*Astragalus gummifer*) affords a valuable index of the purity of this commodity when compared with results obtained by similar treatment of so-called "Indian gum" (*Cochlospermum gossypium* and *Sterculia urens*). The term "volatile acidity" expresses the number of cc. of N/10 potassium or sodium hydroxid required to neutralize the volatile (acetic) acid obtained by distilling with steam the products of the action of boiling aqueous phosphoric acid on 1 gram of the gum.

Treat 1 gram of the whole or powdered sample in a 700 cc. round-bottomed, long-necked flask for several hours in the cold with 100 cc. of water and 5 cc. of sirupy phosphoric acid until the gum is completely swollen. Boil gently for 2 hours under a reflux condenser. A very small amount of cellulose substance will remain undissolved. Now distil the hydrolyzed product with steam, using a spray trap⁷ to connect the distillation flask with the condenser and continue until the distillate amounts to 600 cc. and the acid residue to about 20 cc. Do not concentrate too far, as this would scorch the non-volatile, organic decomposition products and possibly contaminate the distillate. Titrate the distillate with N/10 potassium hydroxid, using 10 drops of phenolphthalein as an indicator, finally boiling the liquid under examination until a faint pink color remains. Correct the result by a blank determination and express the final result in terms of the number of cc. of N/10 alkali required, as in the above definition.

While tragacanth yields a practically colorless solution when boiled with aqueous phosphoric acid, Indian gum, on the other hand, gives a pink or rose solution. This reaction may be used as a preliminary test for the detection of Indian gum.

LEVANT WORMSEED.

32

SANTONIN.—TENTATIVE.

Extract 10 grams of the sample, ground to pass a 30 mesh sieve, in a Soxhlet extraction apparatus for 3 hours with chloroform. Distil off the chloroform until 7–8 cc. remain; add 100 cc. of 5% barium hydroxid solution and heat on a steam bath until the odor of chloroform has disappeared. Boil 5 minutes, cool and pass carbon dioxid (washed through sodium bicarbonate solution to remove traces of acid) until saturated. Filter on a small Büchner funnel, using suction, and wash twice with 10 cc. of water. Heat the filtrate on a steam bath, add 5 cc. of 25% hydrochloric acid and warm 5 minutes. Cool until lukewarm and extract with 20, 15 and 15 cc. of chloroform, passing the solvent through a small filter into a flask. Evaporate to dryness, removing the last traces of chloroform. Dissolve in 7.5 grams (9.5 cc.) of absolute alcohol, warming gently if necessary. Then add 42.5 cc. of water heated to 60°–70°C., stopper the flask and allow to cool. Start crystallization at this point by scratching the side of the flask with a rod or by seeding with a minute crystal of santonin. (Solutions containing a liberal amount of santonin, kept in a cool place for 24 hours, have been found in a supersaturated condition where this precaution was not observed.) Maintain the flask and contents at a temperature of 15°–17°C. for 24 hours. Filter and wash at 15°–17°C. with two 10 cc. portions of 15% alcohol by weight. Dry the flask and filter at 100°C., dissolve the santonin left in the flask and on the filter in chloroform and filter into a tared beaker. Wash the flask and paper thoroughly with chloroform, evaporate the combined filtrate and washings, dry at 100°C. to remove all traces of chloroform and weigh. To the weight found add 0.04 gram for the santonin dissolved in the dilute alcohol and multiply the total by 10 to obtain the per cent of santonin.

NITROGLYCERIN IN TABLETS.

33

PREPARATION OF SAMPLE.—TENTATIVE.

(a) Crush 25 tablets under 10 cc. of anhydrous ether in a 25 cc. cylinder by means of a stout glass rod. Rinse the rod with a little anhydrous ether, allow the insoluble material to settle and decant the solution into a 50 cc. graduated flask. Wash the residue repeatedly with 5 cc. portions of anhydrous ether, decant the washings into the flask until it is filled to the mark, stopper and mix well. Designate this solution as A.

Add 10 cc. of water to the residue, mix well and transfer the mixture to a small separatory funnel by means of a little water. Extract with 3 successive portions of 10, 5 and 5 cc. of ether. Collect the ether extracts in a 50 cc. beaker and designate this solution as *B*.

(b) Disintegrate 25 tablets in a small beaker with 10 cc. of water, breaking up any lumps with a glass rod, and transfer by means of a little water to a separatory funnel. Rinse the beaker with 10 cc. of ether and transfer this also to the funnel. Shake thoroughly, draw off the aqueous layer and transfer the ether through a funnel, containing a little cotton, to a 50 cc. graduated flask. Repeat the extraction with successive portions of ether until the flask is filled to the mark, stopper and mix well. Designate this solution as *C*.

In hand-made and soft compressed tablets, the method described under (a) is preferred, since the direct extraction of the dry crushed material with ether removes most of the nitroglycerin. In hard compressed tablets, the direct extraction is often not nearly so complete and, in such cases, the method described under (b) is to be preferred.

Nitrate Method⁸.—Tentative.

34

REAGENTS.

(a) *Phenoldisulphonic acid solution*.—Prepare as directed under IV, 14 (a).

(b) *Standard nitrate solution*.—Dissolve 0.7217 gram of potassium nitrate in 1 liter of water. Evaporate 10 cc. of this solution just to dryness in a porcelain dish on a steam bath. Cool and treat the residue with 2 cc. of the phenoldisulphonic acid solution, rubbing with a glass rod to insure intimate contact. After 5–10 minutes dilute to 250 cc. Each cc. of this solution contains 0.004 mg. of nitrogen. Add an excess of potassium hydroxid solution to an aliquot of this solution and dilute to 100 cc. (Do not use sodium or ammonium hydroxid.) It is advisable to prepare a standard of approximately the same color as the unknown.

35

DETERMINATION.

Place 20 cc. of the ether solution, *A* or *C* under 33, in a dried, tared 50 cc. beaker. Evaporate the solvent in a vacuum desiccator containing sulphuric acid. Apply the vacuum gradually, to prevent boiling. Allow the beaker to remain in the vacuum 30 minutes after the ether has evaporated. Weigh and calculate the ether extract per tablet. Treat the residue with 2 cc. of the phenoldisulphonic acid solution, rotating the beaker so that the reagent comes in contact with the entire inner surface. After 10 minutes add water and wash into a 100 cc. flask. Dilute to the mark and place 10 cc., representing 1 tablet, in a 100 cc. flask, add about 50 cc. of water and a few drops more of 20% potassium hydroxid solution than is required to neutralize the acid. Dilute to the mark and compare the color with that produced when a standard nitrate solution is similarly treated. Any convenient colorimeter or Nessler tubes may be used. Multiply the nitrate nitrogen found by 5.4 to obtain the equivalent of nitroglycerin.

When 33 (a) is used for the preparation of the sample, a correction, determined as directed in 37, should be made for the amount of nitroglycerin in *B* under 33, using all of *B* instead of an aliquot.

Nitrite or Modified Hay Method⁹.—Tentative.

36

REAGENTS.

(a) *Sulphanilic acid solution*.—Prepare as directed under IV, 12 (b).

(b) *Alpha-naphthylamin hydrochlorid solution*.—Prepare as directed under IV, 12 (c).

(c) *Standard nitrite solution*.—Weigh out 0.220 gram of dry silver nitrite [XV, 18 (c)], dissolve in a small quantity of hot water and decompose with a slight excess of sodium chlorid solution. When the solution becomes clear, dilute to 1 liter with nitrite-free water. Dilute 5 cc. of this solution to 1 liter with nitrite-free water. The second dilution, containing 0.0001 mg. of nitrous nitrogen per cc., is the standard to be used. [Cf. IV, 12 (d)]

37

DETERMINATION.

Place 5 cc. of the ether solution, *A* or *C* under 33, in a 50 cc. beaker dilute with 5–10 cc. of alcohol and add about 5 cc. of 0.5% alcoholic potassium hydroxid. Cover with a watch glass and allow to stand 10 minutes. Place on a steam bath, boil, remove the watch glass and, when most of the liquid has evaporated, add about 25 cc. of water and return to the steam bath until about half the liquid has evaporated or until the odor of alcohol can no longer be detected. Cool and dilute with nitrite-free water to 250 cc. Each cc. of this solution represents 0.01 of a tablet. Introduce an aliquot, representing 0.02–0.04 mg. of nitroglycerin, into a 100 cc. graduated flask, dilute with sufficient nitrite-free water to make the volume 90–95 cc., add a drop of concentrated hydrochloric acid, then 2 cc. of the sulphanilic acid solution and 2 cc. of the alpha-naphthylamin hydrochlorid solution. Complete the volume with nitrite-free water. Prepare at the same time and in the same manner standards containing known amounts of sodium nitrite. Stopper the flasks, mix well and compare the colors after 30 minutes, using any convenient colorimeter or Nessler tubes. Multiply the nitrite nitrogen found by the factor 8, which has been determined experimentally, to obtain the equivalent of nitroglycerin.

When 33 (a) is used for the preparation of the sample, a correction, determined as directed above, should be made for the amount of nitroglycerin in *B* under 33, using all of *B* instead of an aliquot.

PEPSIN IN LIQUIDS.—TENTATIVE.

38

REAGENTS.

(a) *Standard pepsin*.—Powder a good grade of U. S. P. pepsin and pass it through a No. 60 sieve; dry in vacuo over calcium chlorid, again pass through a sieve and preserve in a stoppered bottle. Ascertain the exact pepsin equivalent of the dry powder by the U. S. P. method¹⁰ and express in percentage based on the assumption that the U. S. P. product is 100% pure.

(b) *Standard pepsin solutions*.—Weigh off definite amounts of the standard pepsin into the requisite quantity of N/10 hydrochloric acid to make solutions containing 5 and 0.5 mg. of pepsin per cc. These should be freshly prepared.

(c) *Ricin solution*.—Grind commercial ricin, similar to the “Ricin Präparat nach Jacoby”, to a No. 60 powder, mix thoroughly, dry and keep in a desiccator. Digest 1 gram of this powder for an hour at 37.5°C. in 100 cc. of 5% sodium chlorid solution, cool, filter and use at once for the assay.

39

PREPARATION OF SOLUTIONS.

(a) *Dilute solution of the sample*.—Dilute the sample with a measured amount of N/10 hydrochloric acid until, upon digestion at 37.5°C., 1 cc. requires approximately 15 minutes to digest the precipitate obtained by mixing 2 cc. of the ricin solution and 0.5 cc. of N/10 hydrochloric acid. To 50 cc. of this diluted preparation add the requisite quantity of water or of N/5 hydrochloric acid to make the preparation of N/10 acid strength when diluted with N/10 acid to 90 cc. Preserve the sample in

a refrigerator. (Solid pepsin preparations may often be extracted with hydrochloric acid of appropriate strength and prepared for assay in the same manner.)

(b) *Dilute comparison solution of the sample.*—Add 1 cc. of N/10 hydrochloric acid to 9 cc. of the dilute solution of the sample.

(c) *Dilute inactive solution of the sample.*—Immerse a stoppered glass vessel, containing 45 cc. of the dilute solution of the sample and 5 cc. of N/10 hydrochloric acid, in boiling water for 15 minutes and filter.

(d) *Standard solution containing 0.5 mg. of active U. S. P. pepsin per cc.*—Immerse a stoppered test tube containing 18 cc. of the dilute solution of the sample in boiling water for 10 minutes and, after cooling, add 2 cc. of the standard pepsin solution, containing 5 mg. of pepsin per cc., and filter if necessary.

If the solutions to be tested are not clear, filter through hardened filters. If, however, they cannot thus be clarified, make check comparison tubes containing the same amounts of the preparation made up in the same way with 2 cc. of water in place of the ricin solution used in the determination.

40

DETERMINATION.

To each of 15 tubes, add from a burette 2 cc. of the ricin solution and 0.5 cc. of N/10 hydrochloric acid, heat to 37.5°C. and add the following quantities of the solutions:

To the first 5 tubes, add 0.00–1.00 cc. of the dilute comparison solution of the sample in 0.25 cc. increments, and 1.00–0.00 cc. of the dilute inactive solution of the sample in 0.25 cc. decrements. To the next 5 tubes, add 1.00–0.00 cc. of the dilute inactive solution of the sample in 0.25 cc. decrements and 0.00–1.00 cc. of the standard solution containing 0.5 mg. of active U. S. P. pepsin per cc. in 0.25 cc. increments. To the last 5 tubes, add 1.00–0.00 cc. of N/10 hydrochloric acid in 0.25 cc. decrements and 0.00–1.00 cc. of the standard pepsin solution containing 0.5 mg. of pepsin per cc. in 0.25 cc. increments.

By comparing any tube of the first group of 5 with the tubes in the remaining groups the degree of proteolytic activity of the dilute comparison solution of the sample may be matched against known amounts of standard pepsin both in ordinary acid medium, last group of 5, and in the same medium as the sample itself, second group.

Introduce the acid and the dilute inactive solution of the sample into the tubes first and then pour in the solutions to be tested as rapidly as possible from graduated pipettes, noting the total time consumed in the process after adding the pepsin.

After the addition of the solution to be tested, again immerse the test tubes in the 37.5°C. bath, preferably arranged in corresponding order in a partitioned square or oblong wire rack, such as is used in bacteriological work. Shake and examine the tubes from time to time for 1–2 hours, noting the time when the digestion begins and ends. In case of very weak solutions they may be allowed to digest overnight.

If the rate of digestion is the same in each group, the dilute comparison solution of the sample contains exactly 0.5 mg. of pepsin per cc. If the rate is more rapid in the first group than in the others, it is stronger, the comparative strength being closely indicated by the time of action in the tube containing less of the solution. If the rate of clearing is more rapid in the last group than in the second, some interfering substance is present and must be removed by dialysis, or by evaporation in vacuo at a low temperature until, upon re-examination and further dilution or concentration, the rate of digestion is identical or nearly so in each series.

Smaller quantities of pepsin may be determined in the same way by comparing

them with more dilute solutions of standard pepsin. Thus 0.05 mg. of U. S. P. pepsin can be readily detected by the nearly complete solvent action on the ricin precipitate in less than 2 hours. A marked action on the ricin within the same time is shown by 0.005 mg. For all practical purposes the absence of an appreciable solvent action after 4 hours digestion indicates the absence of pepsin. Express the result in per cent, assuming U. S. P. pepsin to be 100% pure and calculating the result according to the dilution found necessary in preparing the dilute solution of the sample.

TURPENTINE.

41

COLOR.—TENTATIVE.

Fill a 200 cc. flat-bottomed colorimeter tube, graduated in mm., to a depth of 40–50 mm. with the turpentine. Place the tube in a colorimeter and place on or under it a No. 2 yellow Lovibond glass. Over or under a second graduated tube in the colorimeter, place a No. 1 yellow Lovibond glass and run in the same turpentine until the color matches as nearly as possible the color in the first tube. Read the difference in depth of the turpentine in the 2 tubes. If this difference is 50 mm. or more, the turpentine is "standard".

42

SPECIFIC GRAVITY.—TENTATIVE.

Determine the specific gravity at $\frac{20^{\circ}\text{C.}}{4^{\circ}}$ by means of a pycnometer. The specific gravity may also be determined somewhat less accurately at any convenient temperature with a plummet, correcting the result by using the factor 0.00082 for each degree that the temperature of the determination differs from the standard temperature.

43

REFRACTIVE INDEX.—TENTATIVE.

Determine the refractive index at any convenient temperature with an accurate instrument and calculate the result to 20°C., using the correction factor 0.00045 for each degree that the temperature of the determination differs from 20°C.

44

DISTILLATION.—TENTATIVE.

Use an ordinary Engler flask (the internal diameter of the side tube must be 6–7 mm.) and condenser¹¹ and heat the flask in a glycerin or oil bath¹². Fit the flask with a thermometer reading 145°–200°C. Place 100 cc. of the turpentine in the flask, connect with the condenser and distil. Conduct the distillation so that the distillate passes over at the rate of 2 drops per second. Note the initial distilling temperature and the percentage distilling below 170°C.

POLYMERIZATION.—TENTATIVE.

45

REAGENT.

38N sulphuric acid.—Mix 140 grams of concentrated sulphuric acid with sufficient liquid, fuming sulphuric acid (about 10 grams) to obtain an acid slightly stronger than 38N. Determine the exact strength¹³ of this mixture and also of the concentrated acid as follows: Weigh out 6–8 grams in a bulb, having a capillary tube in the lower end and a tube with a stop-cock in the upper end, fitted with a platinum wire for suspending on a balance. (The bulb is filled by the aid of a slight vacuum, and the lower end of the capillary is emptied by closing the stop-cock simultaneously with the withdrawal of the capillary from the acid; after which it is wiped off first with a wet and then with a dry piece of cloth.) Run the acid into cold water, make up to volume and titrate an aliquot of the solution against standard alkali or add an

excess of ammonium hydroxid to an aliquot, evaporate to dryness, dry to constant weight at 120°–130°C. and weigh as ammonium sulphate. Calculate the sulphur trioxid content of the acid and add sufficient concentrated sulphuric acid to make it exactly 82.38% of SO_3 . The acid must be carefully protected against absorption of water from the air.

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DETERMINATION.

Place 20 cc. of the 38N sulphuric acid (100.92%) in a graduated, narrow-necked Babcock flask, stopper, place in ice water and cool. Add slowly 5 cc. of the turpentine. Mix the contents gradually, cool from time to time and do not allow the temperature to rise above 60°C. When the mixture no longer warms up on shaking, agitate thoroughly, place in a water bath and heat to 60°–65°C. for about 10 minutes, keeping the contents of the flask thoroughly mixed by vigorous shaking 5–6 times. Cool to room temperature and fill the flask with concentrated sulphuric acid until the unpolymerized oil rises into the graduated neck. Centrifugalize 4–5 minutes at about 1200 revolutions per minute, or allow to stand for 12 hours. Read the unpolymerized residue, notice its consistency and color and determine its refractive index.

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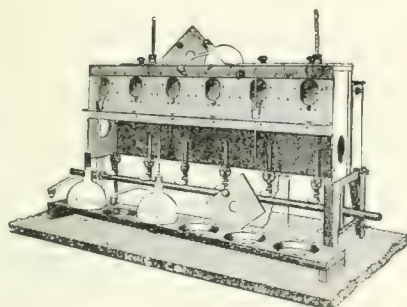
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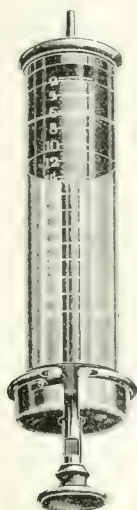
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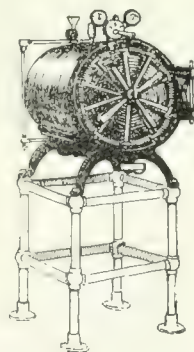
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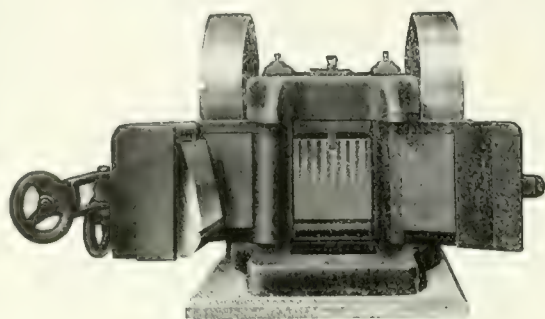
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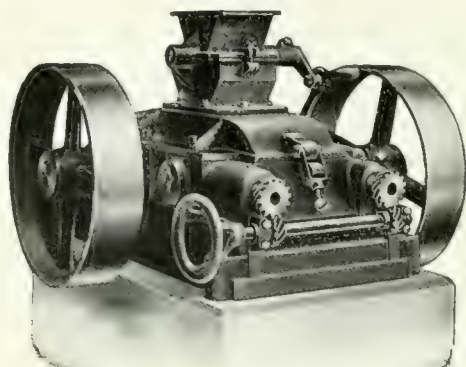
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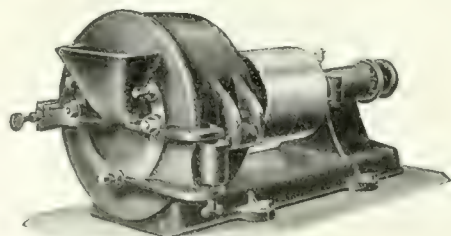
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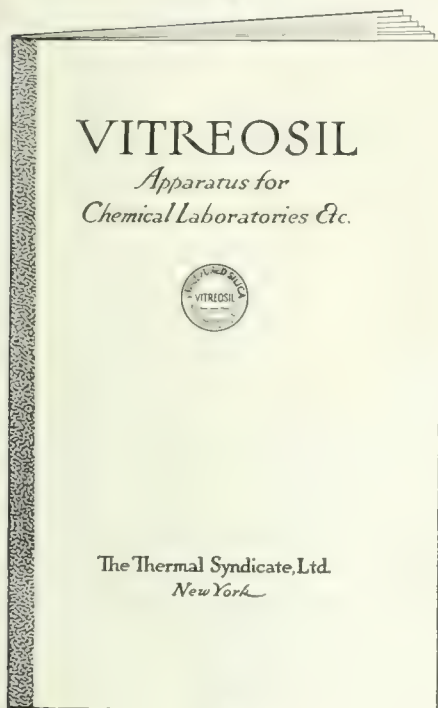
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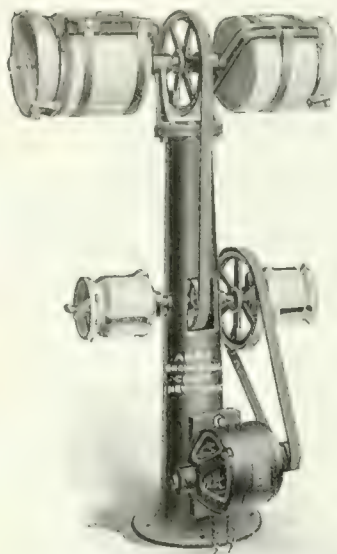
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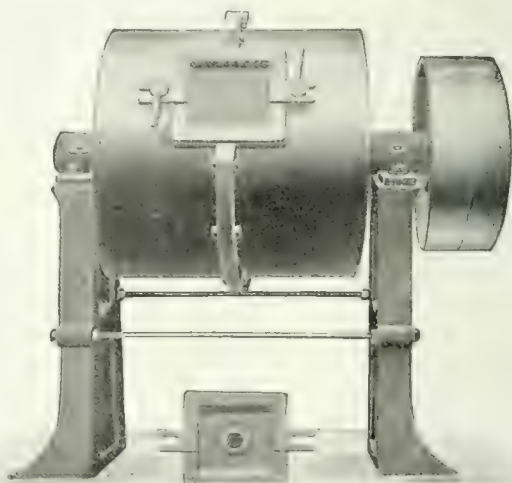
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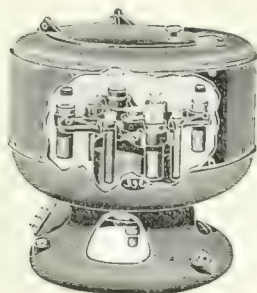
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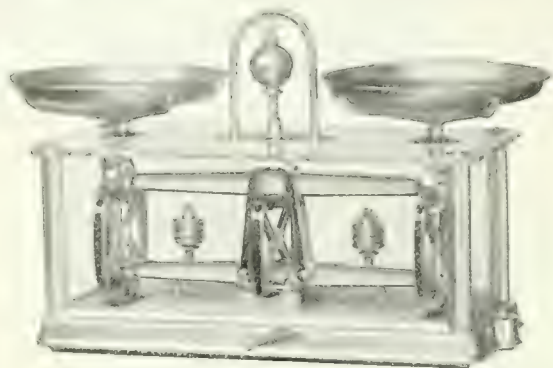
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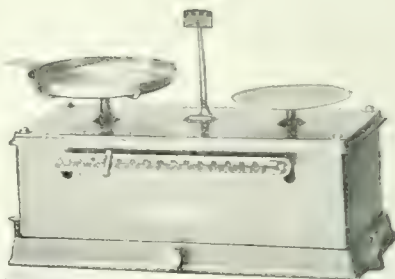
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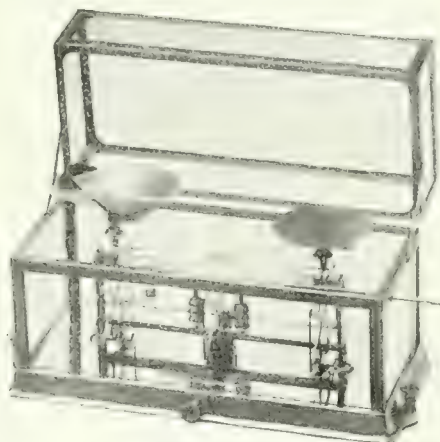
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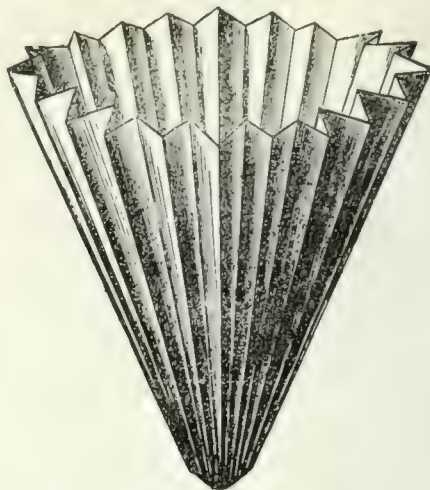
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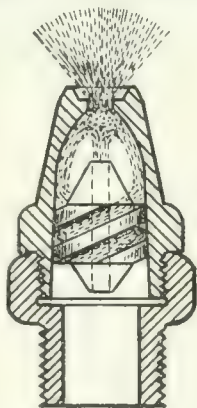


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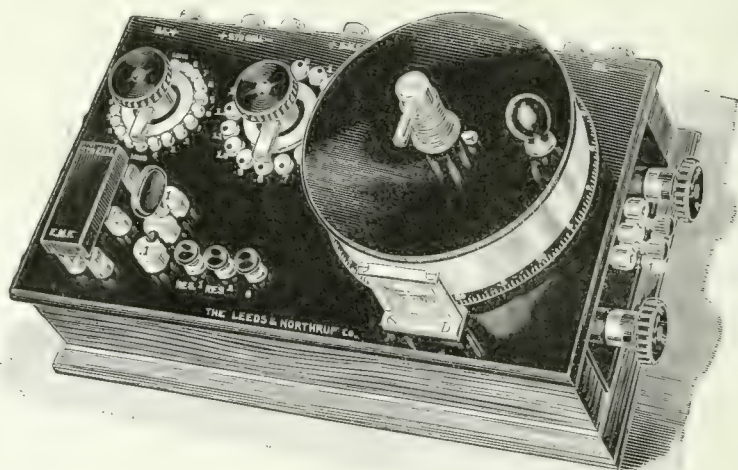
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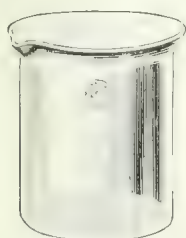
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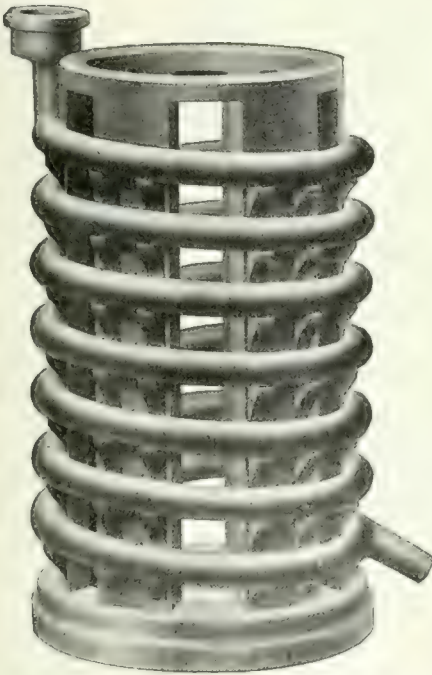
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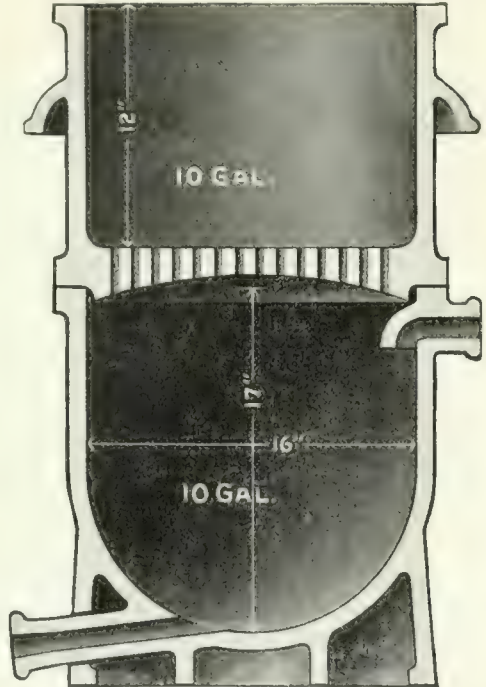
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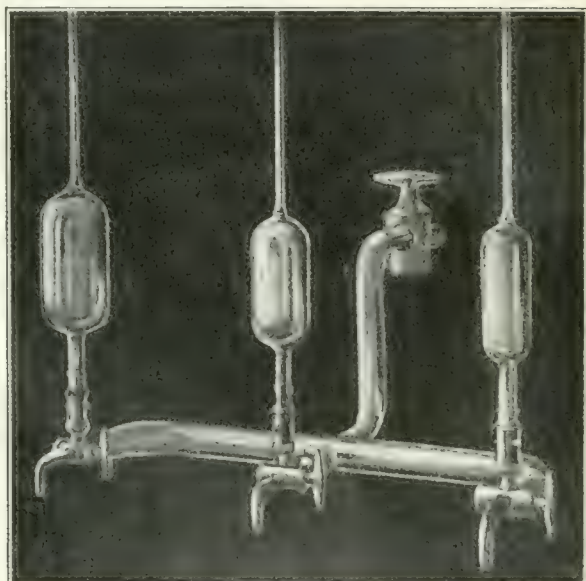
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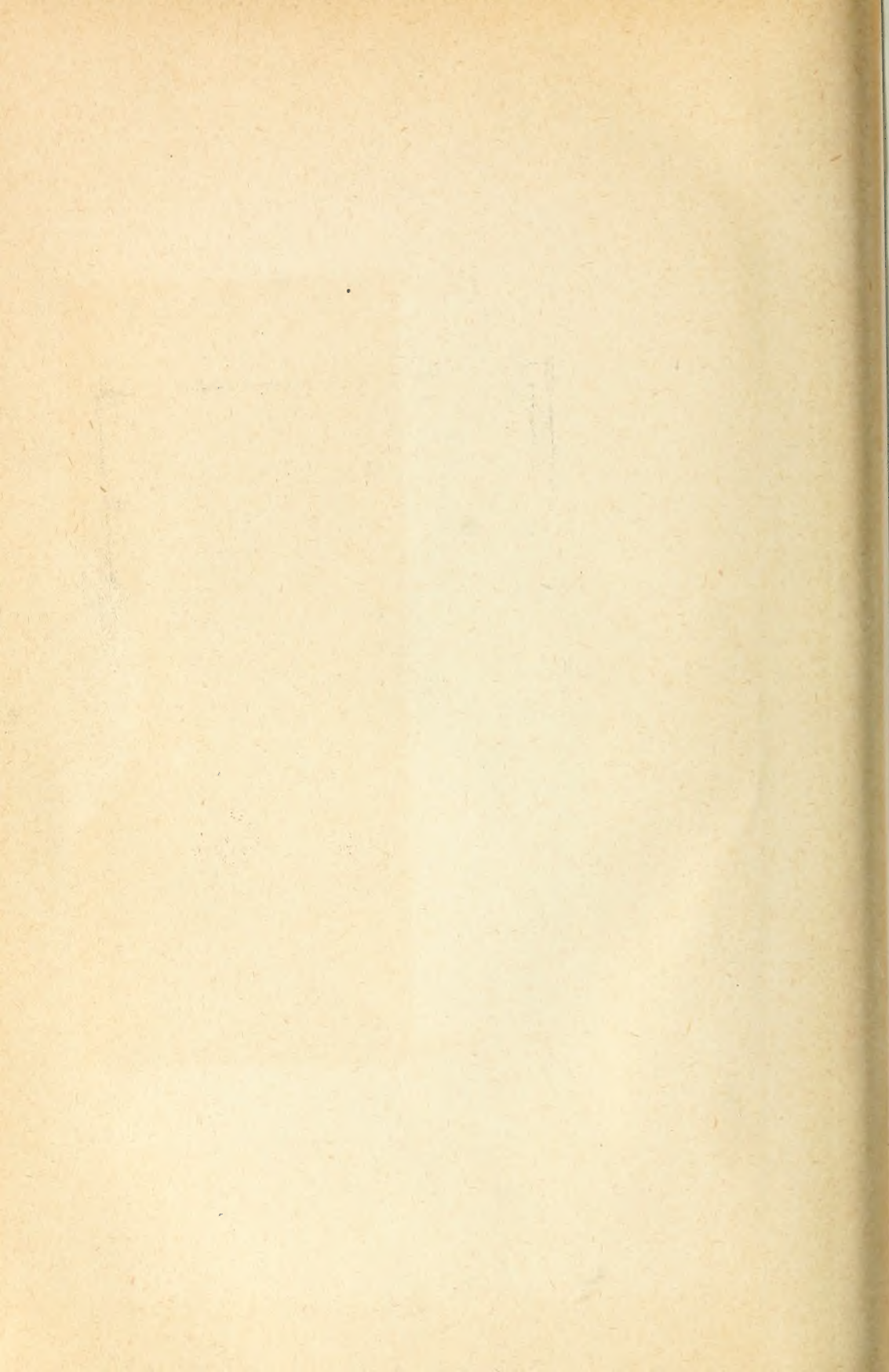
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